

Mechanical modelling of wood microstructure, an engineering approach

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The macroscopic elastic behaviour of wood derives from the mechanical performance of the cells which form its microstructure. Numerical finite element models are presented which relate the elastic properties of the wood continuum to local cell characteristics such as cell size, wall thickness, moisture content and microfibril angle. Preliminary results show good agreement with observed values.

Keywords: wood microstructure - finite element analysis - elastic properties

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1. Introduction

The macroscopic elastic behaviour of softwood timber is governed by nine orthotropic elastic constants in a coordinate system aligned locally with the grain direction. These are determined by the local microstructure of the wood cells. In this article, analytic and computational tools routinely applied to the analysis of large-scale engineering structures (laminated theory and finite element methodology) are used to predict these macroscopic elastic constants. This is done by treating the timber as a micro-cellular structure formed from composite laminated plate elements. The work described forms part of a broader interdisciplinary programme which seeks to predict the effect of specific cell-anatomical characteristics on the mechanical properties of radiata pine in order to improve its commercial value by the selection of genetic stock to give improved stiffness. Notwithstanding this specific application, the computer modelling techniques which have been developed are applicable to a broader range of problems involving the mechanics of wood deformation. These include studies of internal checking and cell collapse during drying and analysis of the non-linear mechanisms which occur at a cellular level during more extreme forms of wood processing such as slicing, splitting, tearing and sawing.

At high magnification, softwood timber is composed for the most part of axially aligned *tracheid* cells which occupy 95% of the timber volume. They resemble an irregular honeycomb of prismatic tubes of diameter 20-40 μm and of length 2-4 mm. In the rapidly growing earlywood portion of the growth cycle these tracheids have relatively large cross-sectional areas and thin walls. In the more slowly growing latewood stage the reverse is true, a difference in morphology which gives rise to the visible growth rings present in timber. The models which will be presented in this article relate primarily to studies of earlywood cells. An electron micrograph of a bundle of earlywood cells from *Pinus radiata* is shown in Figure 1(a). Each closed cell of the cellular array is itself formed from concentric multiple orthotropic plies or *lamellae* which are reinforced by helical windings of crystalline cellulose embedded in a matrix of polyose and lignin. The consequent laminated internal structure of the cell wall is apparent in Figure 1(b) which shows adjacent cell walls at somewhat greater magnification. In constructing a mechanical model for such a system, we seek to define the relationship between the macroscopic elastic properties of the timber continuum – i.e. the bundle of cells shown in Figure 1(a) – and local cell and cell-wall parameters such as cell size and geometry, wall thickness, moisture content and microfibril orientation. The models presented in this article attempt to account for all of these factors. In particular, they accommodate the effects both of the irregular nature of the cellular geometry *and* the complex laminated structure of the cell wall. Previous attempts at modelling wood microstructure have tended to concentrate *either* on an accurate representation of the cell wall at the expense of cellular geometry *or* on an accurate representation of the cellular geometry without modelling in any detail the structural complexity of the cell wall. Models of the former type, such as those of Cave [1-3] give good results for the axial modulus but cannot resolve cross-grain or shear behaviour. Cellular geometric models such as those of Gibson & Ashby [4] and Kahle & Woodhouse [5] which *do* predict cross-grain behaviour assume a

homogeneous structure for the cell wall and cannot readily be used to assess the effects of the cell-wall internal structure. The models presented in this article permit both aspects of the problem to be represented, albeit in an idealised fashion.

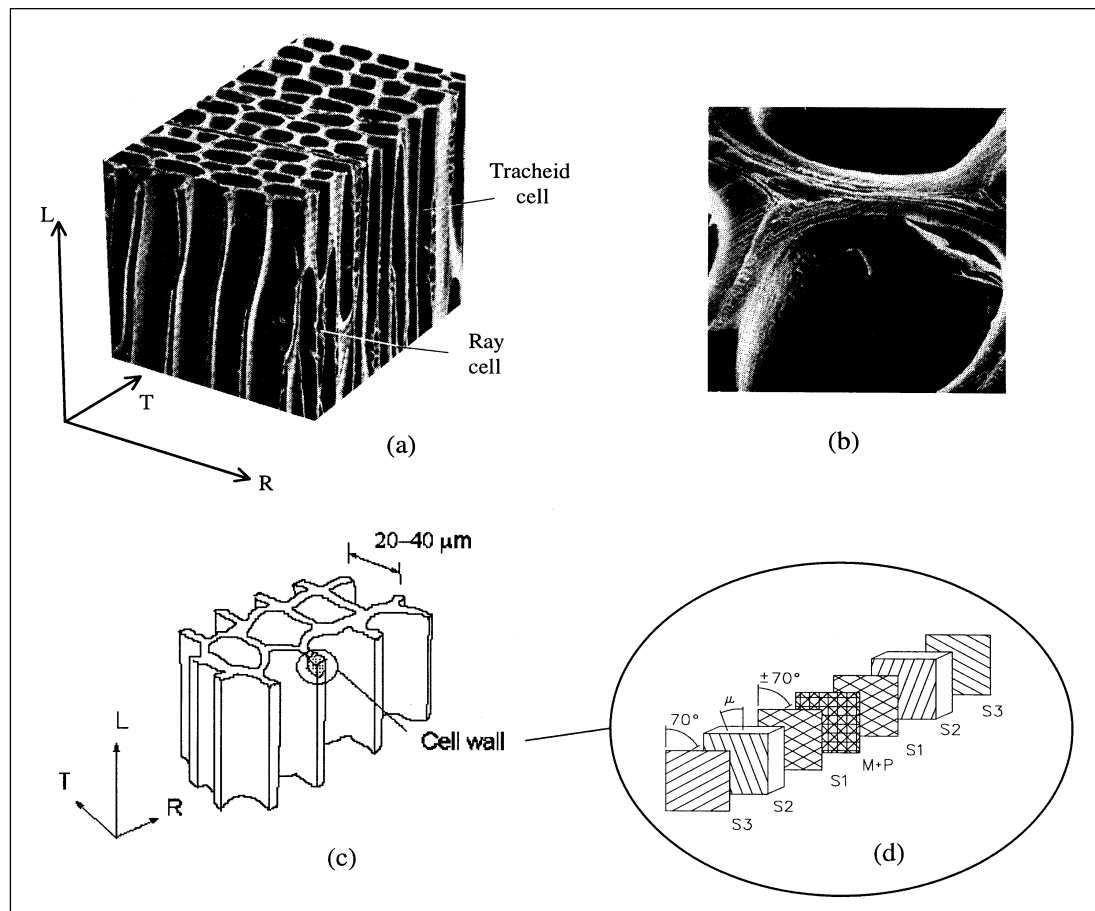


FIGURE 1: (a) Image of tracheid cells, (b) image of the cell wall, (c) model of cellular geometry, (d) model of cell-wall laminate

2. The theoretical model and simplifying assumptions

To form a quantitative mechanical model for the structure represented in Figure 1(a), simplifying assumptions are made at three distinct physical scales. The idealisation which is used at the largest of these scales is that of a prismatic bundle of cells of prescribed cross-sectional geometry and constant wall thickness, as sketched in Figure 1(c). This will subsequently be translated into a finite element (FE) model in which each cell wall division is modelled as a shell element (see Section 6, to follow). The internal structure of the cell-wall pair separating two adjacent cells is the next scale at which simplifying assumptions must be made. The idealisation used in the current analysis is illustrated in Figure 1(d). The cell wall pair, of thickness 4-8 μm, is represented as an elastic laminate with seven orthotropic plies. This is a substantial simplification but follows closely the convention that the several hundred individual lamellae which form the Secondary wall of a tracheid cell can be grouped into three larger entities; the S1, S2 and S3 layers. The Middle lamella which lies between the cells is then combined with the Primary cell wall of *each* cell to form a seventh central layer (M+P). Each of these layers has its own constitutive fractions - of cellulose, lignin and polyose - and characteristic microfibril direction(s). In the current exercise, the microfibril angle within the S1 and S3 layers is taken to be $\pm 70^\circ$ with a standard deviation of 12.5° . The mean microfibril angle μ in the S2 layer (see Figure 1(d)) which can vary within and between trees is specified as input to the current model, being incorporated into the cell wall with a standard deviation equal to one third of its mean. The middle lamella and primary cell wall are assumed to be transversely isotropic in the plane of the cell wall. Note that the sense of the helical microfibril windings is the same in adjacent cells and hence the microfibril angle reverses direction as the central plane of the cell-wall pair is traversed, giving an asymmetric laminate. There is no difficulty in incorporating such a laminate structure into a shell finite element model provided that elastic

constants are available for each ply of the laminate. This leads to consideration of the smallest scale of representation in the current model, that of a fundamental volume element within each cell wall layer.

3. Modelling the cell wall lamellae

At a nano-structural level characteristic of the thickness of a single lamella, the wood cell wall is an interpenetrating network of carbohydrate and lignin macromolecules and water. The principal component of the wall is cellulose, a linear homopolymer consisting of β -1-4 linked glucose residues. Cellulose is present as highly crystalline microfibrils whose diameter is approximately 5 nm. These microfibrils are embedded in a matrix of polyoses (hemicelluloses) and lignin. There is good evidence to suggest that the polyoses, also polymers of sugar residues, are oriented in a similar fashion to the cellulose chains, and are intimately associated with the cellulose at the surface of the microfibrils, and form bridging links between microfibrils. Lignin on the other hand is a highly branched polymer of coniferyl alcohol, and has no preferred orientation. It is, however, not evenly distributed, but is thought to form tangential lamella, probably because of the lower degrees of molecular order in zones where successive lamellae have differing microfibrillar orientations. The polyose and lignin are covalently bonded at certain sites, and are very finely mixed. The microfibrils occur in concentric lamellae, each lamella approximately a single microfibril thick. Within a lamella the microfibrils are unevenly spaced, but are approximately parallel. In the secondary wall the microfibrils are also reasonably straight. In its natural state the cell wall is water-saturated. X-ray diffraction observations indicate that while amorphous cellulose is highly hygroscopic, the crystalline form is inaccessible to water. Isolated polyose fractions are also highly hydrophilic [6] though this affinity for water may well be modified *in situ* by the blocking of sorption sites by the lignin. The removal of moisture from the wall results in shrinkage, as well as increased stiffness.

4. A representative volume element

Although at this scale, the cell wall is truly a molecular complex, the wall lamellae can be considered as a first approximation to be continuous-fibre reinforced composites, and furthermore the microfibrils can reasonably be treated as straight and parallel. This allows the application of micromechanical tools developed to determine the overall properties of a heterogeneous material from the constituent properties, composition and structure of that material. Fundamental to all micromechanical homogenisation techniques is the concept of a representative volume element (RVE), or a portion of the heterogeneous material which can be considered to be in some way statistically indicative of the whole. In the case of the cell wall a suitable, albeit simple, RVE is shown in Figure 2(a). The central core of the RVE is a tangential agglomeration of microfibrils, which accounts for the uneven distribution of the microfibrils within a lamella, and is treated as a single entity because of the high degree of bonding provided by the associated polyoses. While crystalline cellulose has only monoclinic symmetry, the core is transversely isotropic, since there is no evidence to suggest that the crystallites are oriented in any transverse plane. Surrounding the crystalline core is a cellulose/polyose cortex, or sheath. This sheath is also transversely isotropic, and serves to account for the partial alignment of the polyoses in the microfibrillar direction, and to couple the core to the remainder of the matrix. This sheath is of uniform thickness since there is no reason to believe otherwise. The remainder of the RVE is occupied by an isotropic lignin/polyose matrix. This is unevenly distributed to account for the observed tangential lamellation. Since the matrix is a mixture of lignin and polyose, a second RVE is required to provide its properties.

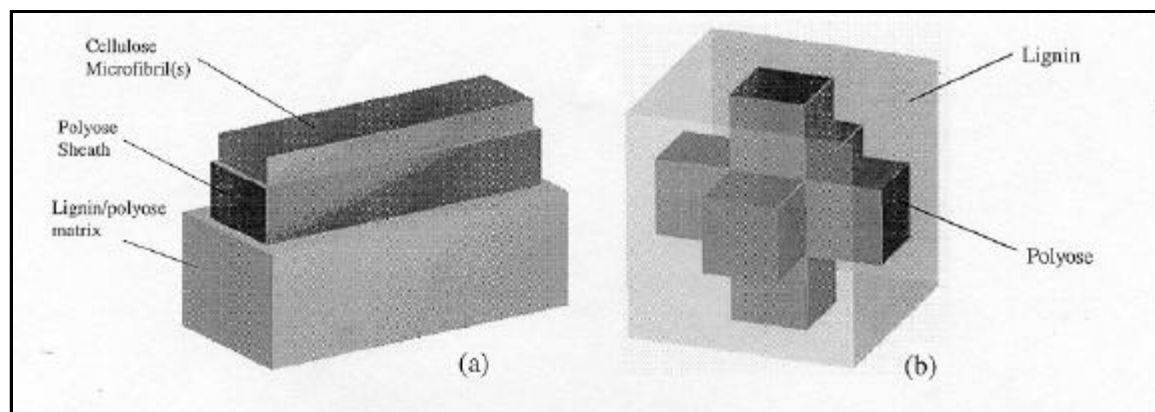


FIGURE 2: (a) Nanostructural RVE (extruded transverse section), (b) RVE for lignin/polyose matrix.

A simple model where the lignin is considered to be an inclusion in a polyose filler, or *vice versa*, is unacceptable because of the large difference in the mechanical properties of these two constituents. For this reason the RVE shown in Figure 2(b) is employed. It should be borne in mind that this is just a convenient model and does not significantly represent the actual cell wall nanostructure.

5. Homogenisation

Having chosen to treat the wall-lamella as heterogeneous continua, it remains to select a suitable homogenisation procedure to determine the properties of the equivalent homogeneous material. While much of what follows is applicable to other properties, for example the expansional coefficients or linear visco-elastic parameters, only the elastic moduli have been considered here. Numerous homogenisation techniques, of varying degrees of sophistication, exist in the literature. In this case the method developed by Chou and Carleone [7] was adopted, partly because it had been used before [8] and also because it is quite straightforward to implement. This method is directly applicable to layered media, but it can be adapted for use on a general orthotropic fibre reinforced RVE by making multiple homogenisation passes in orthogonal directions as illustrated in Figure 3.

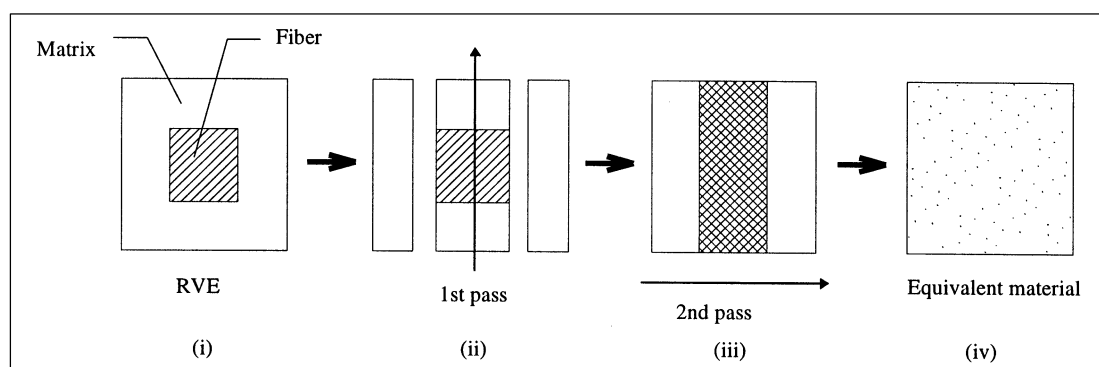


FIGURE 3: The homogenisation procedure for the cell wall RVE.

First the central portion is treated as a laminate, and is homogenised to yield an intermediate equivalent material. This material is then considered as part of a second laminate in an orthogonal direction. The homogenisation of this second laminate gives the equivalent material properties. While this multi-pass method rides roughshod over a number of assumptions made in the development of the theory, results using this method compare well with those obtained using a periodic FE homogenisation procedure (very much like the one used subsequently to find the equivalent elastic properties of the cellular structure), at least for the geometry and constituent properties chosen for the comparison (see Table 1).

<i>Equivalent Property</i>	<i>FE procedure</i>	<i>Multi-pass procedure</i>
$E_x=E_y$	1.91	2.03
E_z	50.4	50.4
ν_{xy}	0.256	0.212
$\nu_{zx}=\nu_{zy}$	0.184	0.184
G_{xy}	0.495	0.408
$G_{xz}=G_{yz}$	0.735	0.722

TABLE 1: Comparison of FE and multi-pass homogenisation results (values in GPa).

The geometry and constituent properties were chosen to be representative of those values for which the multi-pass procedure would be used. This method was used, in conjunction with data drawn from the literature, to determine the orthotropic moduli for the three secondary wall layers S1-S3, and for the compound middle lamella (M+P). The composition of the wall layers was calculated from Cave [2], assuming that the glucomannan and glucouronoarabinoxylan make up the polyose sheath. The oven-dry mass-fractions are given in Table 2. The densities of cellulose, polyose and lignin were taken to be 1600, 1500, and 1400 kg/m³, respectively.

TABLE 2: Cell wall layer compositions (oven-dry mass fractions).

[%]	M+P	S1	S2	S3
cellulose	16.0	44.6	50.4	43.8
polyose assoc. with cellulose	6.8	23.2	24.6	30.3
polyose assoc. with lignin	22.2	10.2	3.2	3.8
lignin	55.0	22.0	22.0	22.0

The nanostructural geometry was based on a tangential agglomeration of only 3 microfibrils, each having a 3.5 nm diameter. For lack of information, both the polyose sheath and the lignin/polyose matrix were uniformly distributed around the periphery of the microfibrillar core. Since the cellular FE model makes no attempt to incorporate the stresses developed by shrinkage, or the changes in layer thicknesses due to the removal of moisture, the effect of moisture content was limited to changes in constituent properties. However, the moduli computed are referred to the oven-dry dimensions. This was crudely accomplished by assuming that the RVE shrank (or swelled) only in the transverse direction, since the microfibrils are known to be unaffected by moisture, and that the change in total volume corresponded to the volume of moisture held by the wall constituents. The moisture content of the constituents (except for cellulose which was considered to remain dry) was determined by assuming that the relative humidity corresponding to the whole-wood moisture content (MC) of interest was constant throughout the wall. This relative humidity (RH) was found using data from the sorption curves of *Pinus radiata* sapwood [11]. The moisture contents of the polyose and lignin were then determined from the MC/RH curves given by Cousins [6,9,10]. The properties of the constituents at the determined moisture contents were taken from the work of Cave [3] and Cousins [6,9,10] with minor modifications. The nine engineering constants which are required to describe an orthotropic elastic material for each of the major cell wall layers were calculated using this procedure and are listed in Table 3. They are given at three representative whole-wood moisture contents. These moisture contents correspond to wood states commonly found in practice, namely green timber (130% MC), in-service structural timber in a normal atmosphere (12%), and oven-dry timber (0%). The subscripts refer to the a set of cartesian coordinates, the 3-axis being in the direction of the microfibrils, the 2-axis being normal to the lamella, and the 1-axis lying in the plane of the lamella and normal to the microfibrils.

TABLE 3: Calculated elastic constants for the cell wall layers of *Pinus radiata* (moduli in GPa).

MC (%)	Wall Layer	E_1	E_2	E_3	ν_{21}	ν_{31}	ν_{32}	G_{23}	G_{31}	G_{12}
130	S3	0.59	0.09	35.18	0.04	0.29	0.33	0.02	0.21	0.02
	S2	0.49	0.11	47.24	0.05	0.29	0.33	0.03	0.17	0.02
	S1	0.57	0.10	37.88	0.05	0.26	0.33	0.03	0.20	0.03
	M+P	1.38	0.15	12.78	0.03	0.24	0.28	0.04	0.52	0.09
12	S3	8.43	7.98	50.36	0.39	0.33	0.32	2.65	3.00	2.68
	S2	9.85	9.16	63.96	0.39	0.33	0.33	3.02	3.38	2.96
	S1	8.54	8.02	53.10	0.38	0.33	0.32	2.66	3.02	2.66
	M+P	5.07	5.12	18.43	0.38	0.31	0.31	1.78	2.11	1.88
0	S3	11.49	10.62	59.34	0.38	0.32	0.32	3.61	4.29	3.54
	S2	12.30	11.37	72.09	0.39	0.32	0.32	3.85	4.37	3.72
	S1	11.11	10.33	60.99	0.38	0.32	0.32	3.51	4.06	3.40
	M+P	8.02	7.78	26.10	0.37	0.31	0.31	2.72	3.93	2.84

6. The cellular FE model

The orthotropic elastic constants obtained from the analysis of the cell wall RVE are used as input for a finite element (FE) model of a prismatic cell bundle. Two different approaches have been adopted in representing the cell cross-sectional geometry. In the first, a model is formed for an unbounded array of identical cells, equivalent in some statistical sense to a collection of real cells. The simplest *FE-statistical* model of this type, and the only one used to date, is obtained by extracting statistical data - mean cell diameter, mean cell width, mean cell lumen area, etc. - from a micrograph of cell cross-sections and then using this data to construct a

regular cell array with the same statistics. The collection of such data by manual means is tedious but can be performed quite simply if an image processing program - such as METAMORPH - is used. The repeatability of a cyclic cellular unit in the resulting model means that an infinitely large collection of cells can be modelled, in this way, without approximation, by applying cyclic constraints (these correspond to a state of bulk uniform strain, direct or shear). A model of this type is shown in Figure 4.

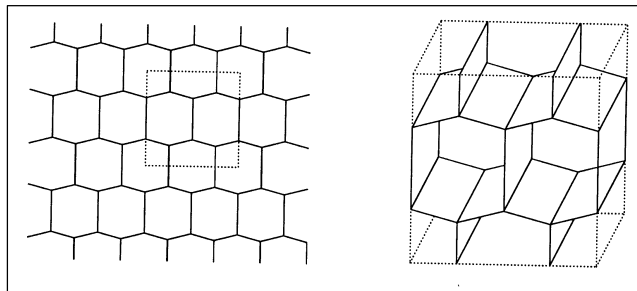


FIGURE 4: A cyclically repeated sub-unit of a regular array.

The second type of model is obtained by skeletising the original micrograph image and extruding the resulting planar cell arrangement to form a prismatic FE model based on the real cell geometry. The resulting model is no longer cyclical and a rigid “constant strain” constraint must be applied at its boundaries. Provided that a reasonable number of cells is used - of the order 100-200 - the edge effects involved in this approximation have been found to be small. An *FE-irregular(geometry)* model of this type and the micrograph from which it is derived are shown in Figure 5. The FE code ANSYS was used for the analysis of both types of model (“*statistical*” and “*irregular*”). In each case the double cell wall is treated as a “thick” composite shell element with the laminate structure of figure 1(d) and layer moduli of table III. By imposing a state of uniform strain on the boundary of the model - either in a cyclical sense or directly - the effective direct and shear moduli can be obtained from the computed overall ratios of load to deflection. If the cellular arrangement is typical of the continuum, these translate directly into orthotropic elastic moduli for a piece of timber with this particular cell geometry.

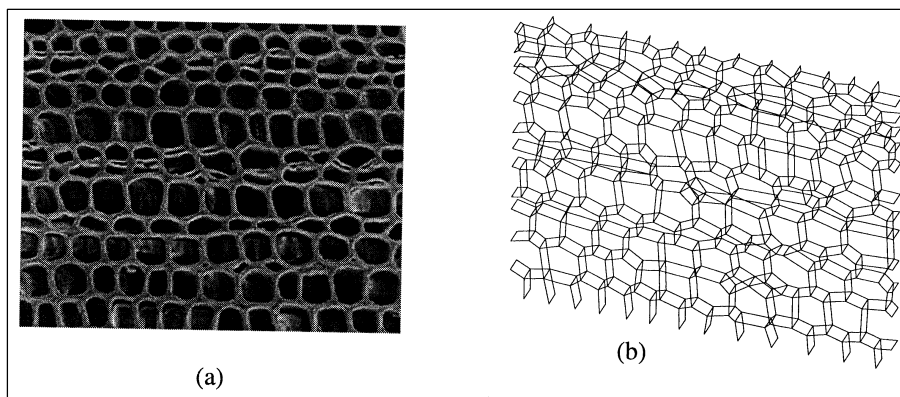


FIGURE 5: (a) Cellular geometry, (b) ANSYS FE model.

7. Comparison with analytic solutions for regular arrays

Validation of the above procedure by comparison with known solutions is difficult. In the particular case of a regular hexagonal array in which the elastic moduli in each cell-wall layer are set to the same (isotropic) values, the continuum moduli predicted by the above model should correspond to the analytic solutions of Gibson and Ashby [4]. This has been tested and found to be the case for a range of hexagonal geometries. Computed and predicted values of the three orthotropic Young’s Moduli for a cellular geometry very similar to that illustrated in Figure 4 are presented in Figure 6. They are plotted against an ordinate “ t/H ” which is a measure of the slenderness of the cell wall. In softwood a typical value for this parameter would lie in the range 0.1-0.2. The two analytic results, Gibson & Ashby (a) and Gibson & Ashby (b), are obtained by neglecting (theory (a)) and including (theory (b)) the shear deformation of the cell wall. The FE model includes this effect and corresponds very closely to theory (b), the more advanced of the two theoretical solutions.

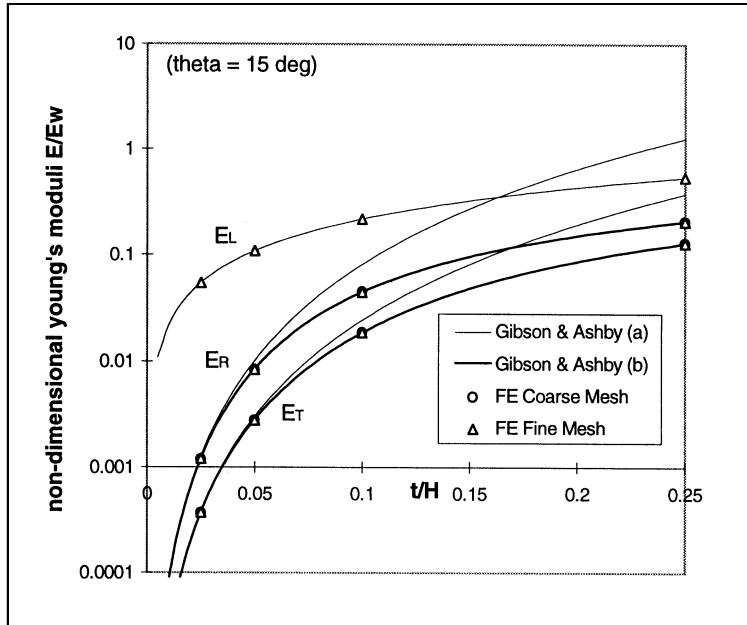


FIGURE 6: Computed and analytic moduli for a regular cell array with isotropic walls.

8. Results for “real” cell geometries

A number of parametric studies have been performed using the full model described above (i.e. including the internal structure of the cell wall as discussed in Sections 2-5) and varying cell parameters. Results from some of these are shown in Figure 7. The wood analysed is fictitious in the sense that the cell geometry has been taken from a particular micrograph of earlywood *Pinus radiata* and then used to test the effect of varying internal cell-wall parameters. Data is presented here for the particular case of a moisture content of 12 % and a basic density of 390 kg/m³.

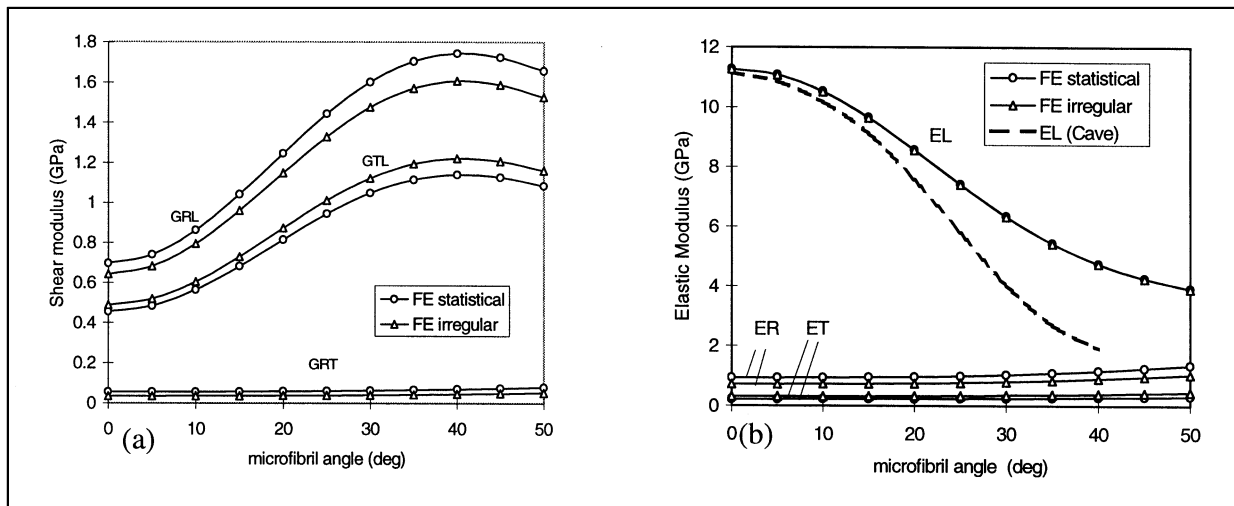


FIGURE 7: Calculated values of elastic constants; (a) radial, tangential and axial Young’s moduli (E_R , E_T and E_L), (b) shear moduli (G_{RL} , G_{TL} and G_{RT}).

In terms of its influence on wood stiffness, the most important cell-wall parameter is the mean microfibril angle in the S2 layer (the angle μ of Figure 1(d)). This varies from tree to tree but also within each tree. In Canterbury-grown *Pinus radiata*, for example, μ can vary from 45-50° in the corewood to 10-20° in the outerwood (beyond say the tenth growth ring) causing large variations in wood stiffness in boards cut from the same tree [12]. This macroscopic effect is illustrated in Figure 7 which shows the variation with microfibril angle of the direct and shear moduli for the assumed cell geometry as μ is varied in the range 0-50°. Shown also in Figure 7(b) is Cave’s result for the axial modulus [2]. This is clearly close to the values predicted by the current model at low microfibril angles but diverges at larger values. The discrepancy at higher microfibril

angles can be attributed to the axial contributions of the S1 and S3 layers, which are included in the FE representation but not in Cave’s analysis, and which become more significant as the microfibril angle increases. A unique feature of the current approach is its ability to relate transverse values of the elastic moduli directly to microfibril data. The variation is not great, E_R and E_T increasing by approximately 30% as the microfibril angle ranges from 0 to 50°. The values obtained for the shear moduli (Figure 7(b)) and Poisson’s ratios (not presented here) show somewhat greater variation.

Comprehensive validation of the current model will be possible only through comparison of predicted and measured orthotropic moduli for clearwood samples of known microfibril orientation and cellular geometry. No such data exists at present but a measurement programme with this objective is currently in hand [13]. We can, however, check that the values predicted by the FE model are at least in broad agreement with those which *have* been measured for similar species. A comparison of this type is presented in Table 4. This shows computed values for the nine orthotropic elastic constants obtained using the statistical and irregular meshes already discussed. Comparison is made with measured values [14] for Norway and Sitka spruce. It is almost certain that these latter data are collected from outerwood with a low microfibril angle. For comparison, computed data is presented only for microfibril angles in the range 0-10°. The comparison is not entirely valid since the measured data derives from wood which includes both earlywood and latewood bands whereas the FE models currently simulate an “equivalent” earlywood material (i.e. having the same basic density). Nonetheless, the correspondence is generally quite close and appears to be somewhat improved when the irregular model is used.

TABLE 4: Computed and measured orthotropic elastic constants (basic density 390 kg/m³, moisture content 12 %, moduli in GPa).

	E_L	E_R	E_T	G_{LT}	G_{LR}	G_{TR}	ν_{TR}	ν_{LR}	ν_{LT}
FE-statistical ($\mu=0^\circ$)	11.3	.94	.22	.46	.70	.055	.36	.25	.25
($\mu=10^\circ$)	10.5	.94	.22	.56	.86	.055	.36	.31	.31
FE-irregular ($\mu=0^\circ$)	11.3	.72	.31	.49	.64	.034	.46	.25	.25
($\mu=10^\circ$)	10.5	.73	.31	.60	.79	.035	.46	.31	.31
Sitka spruce [14]	11.6	.90	.50	.72	.75	.039	.25	.37	.47
Norway spruce [14]	10.7	.71	.43	.62	.50	.023	.31	.38	.51

9. Conclusions

Finite element models have been constructed which permit a realistic micro-structural representation of softwood timber taking into account cell geometric effects and the complex laminated structure of the cell wall. Such models are able to predict macroscopic timber stiffness for a given cellular geometry and assumed distribution of microfibril angle within the cell wall. The orthotropic elastic moduli predicted in this way have been compared to theoretical predictions for regular arrays of uniform cells (with isotropic cell walls) and to existing published data for softwood of low microfibril angle. Close correspondence is achieved in both instances. Further validation of this approach to the prediction of macroscopic wood stiffness requires the collection of measured orthotropic data for the elastic moduli of fully characterised wood samples. Such a programme of measurement is currently in hand and will be reported at a later date.

10. Acknowledgements

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