Autologous Transfusion of Stored Red Blood Cells Increases Pulmonary Artery Pressure

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Abstract

Rationale: Transfusion of erythrocytes stored for prolonged periods is associated with increased mortality. Erythrocytes undergo hemolysis during storage and after transfusion. Plasma hemoglobin scavenges endogenous nitric oxide leading to systemic and pulmonary vasoconstriction.

Objectives: We hypothesized that transfusion of autologous blood stored for 40 days would increase the pulmonary artery pressure in volunteers with endothelial dysfunction (impaired endothelial production of nitric oxide). We also tested whether breathing nitric oxide before and during transfusion could prevent the increase of pulmonary artery pressure.

Methods: Fourteen obese adults with endothelial dysfunction were enrolled in a randomized crossover study of transfusing autologous, leukoreduced blood stored for either 3 or 40 days. Volunteers were transfused with 3-day blood, 40-day blood, and 40-day blood while breathing 80 ppm nitric oxide.

Measurements and Main Results: The age of volunteers was 41 ± 4 years (mean ± SEM), and their body mass index was 33.4 ± 1.3 kg/m². Plasma hemoglobin concentrations increased after transfusion of 40-day and 40-day plus nitric oxide blood but not after transfusing 3-day blood. Mean pulmonary artery pressure, estimated by transthoracic echocardiography, increased after transfusing 40-day blood (18 ± 2 to 23 ± 2 mm Hg; P < 0.05) but did not change after transfusing 3-day blood (17 ± 2 to 18 ± 2 mm Hg; P = 0.5). Breathing nitric oxide decreased pulmonary artery pressure in volunteers transfused with 40-day blood (17 ± 2 to 12 ± 1 mm Hg; P < 0.05).

Conclusions: Transfusion of autologous leukoreduced blood stored for 40 days was associated with increased plasma hemoglobin levels and increased pulmonary artery pressure. Breathing nitric oxide prevents the increase of pulmonary artery pressure produced by transfusing stored blood.

Clinical trial registered with www.clinicaltrials.gov (NCT 01529502).

Keywords: hemoglobins; pulmonary hypertension; nitric oxide; endothelium; echocardiography
Blood transfusion is a common medical intervention, with more than 14 million units of blood transfused annually in the United States (1). In developed countries, noninfectious hazards of transfusion, such as transfusion-related acute lung injury, have emerged as the leading complications of blood transfusion (2). In particular, the combination of transfusing red blood cells (RBC) stored for longer than 14 days and preexisting diseases, such as diabetes and atherosclerosis, in the recipient are associated with increased mortality (3–5). Conversely, other clinical studies have not reported adverse effects associated with transfusion of longer stored erythrocytes (6, 7).

The US Food and Drug Administration allows RBCs to be stored for up to 42 days (1). During storage, RBCs undergo progressive structural and functional changes that reduce oxygen-transport capacity and viability, leading to hemolysis and the release of microparticles, hemoglobin, heme, and iron into the circulation (8–10).

Nitric oxide (NO) is a potent, endogenous dilator of vascular smooth muscle. Circulating hemoglobin, both free in plasma and contained in shed microparticles, avidly scavenges NO via the dioxygenation reaction, depleting vascular NO levels and resulting in vasoconstriction (11). Because pulmonary vasoconstriction was produced in animal models by stored RBC transfusion and prevented by breathing NO (12), we previously tested whether breathing low levels of NO could selectively dilate the pulmonary circulation without producing systemic vasodilation or hypotension (13).

In awake mice with endothelial dysfunction, characterized by reduced vascular NO availability, transfusing cell-free hemoglobin produces marked systemic vasoconstriction and hypertension (14). In a subsequent study, Yu and coworkers (15) demonstrated that transfusing murine RBCs stored for 14 days induces systemic vasoconstriction and inflammation in diabetic mice with endothelial dysfunction and that breathing NO prevents the hypertension induced by transfusing stored RBCs. Breathing NO oxidizes plasma hemoglobin to methemoglobin, a form of hemoglobin that does not scavenge NO, blocking the vasoconstriction response (16). Hypertension and vasoconstriction were shown to be caused by transfusing the supernatant of stored murine RBCs, but transfusing washed 14-day stored RBCs did not cause vasoconstriction (15). In awake lambs, Baron and coworkers (12, 17) reported that autologous leukoreduced RBCs stored for 40 days induced pulmonary hypertension and inflammation, which was prevented by breathing NO.

In our previous study of healthy young volunteers (with intact endothelial function), transfusion of one unit of autologous, leukoreduced blood stored for 40 days neither changed systemic blood pressure nor impaired endothelial function, despite increasing plasma levels of cell-free hemoglobin. However, we did not assess pulmonary artery pressure (PAP) (10). In the present study, we selected overweight and obese volunteers with preexisting systemic endothelial dysfunction to assess the systemic and pulmonary vascular response to transfusion with stored autologous blood.

We hypothesized that transfusion of stored blood would increase pulmonary and/or systemic vasoconstriction by presenting a load of NO-scavenging hemoglobin in the presence of preexisting impaired endothelial function (18). We also tested the hypothesis that breathing NO, before and during transfusion, would prevent the vasoconstrictor effects of transfusing stored autologous blood.

**Methods**

**Population and Study Protocol**

The inclusion criteria to participate in this trial were age older than 18 and younger than 60 years; body mass index between 27 and 45 kg/m²; and evidence of impaired systemic endothelial function, as assessed by reduced postischemic vasodilation (natural log [Ln] of reactive hyperemia index [RHI] ≤0.75) (19).

The study protocol was a three-arm crossover study of autologous blood transfusion in volunteers with systemic endothelial dysfunction. Volunteers received in random order autologous leukoreduced blood stored between 2 and 3 days (3-d challenge), autologous leukoreduced blood stored for 38–42 days (40-d challenge), or autologous leukoreduced blood stored for 38–42 days while breathing NO (80 ppm; 40-d+NO challenge).

Venous blood was sampled before and 10 minutes, 1 hour, 4 hours, and 24 hours after completion of each blood transfusion to study plasma and serum biomarkers of hemolysis, inflammation, and organ injury.

**Echocardiography and Hemodynamic Measurements**

Transthoracic echocardiography was performed before blood transfusion and again 15 minutes before the end of the blood transfusion (see Figure E1 in the online supplement). In most of the volunteers, because a tricuspid regurgitation jet was not present for analysis, the pulmonary artery acceleration time (PAAT) was measured to estimate the mean PAP (20, 21). PAAT was defined as the interval between the onset of systolic pulmonary arterial flow and the peak flow velocity. PAAT was also normalized by the right ventricular ejection time (RVET; interval between the onset of right ventricular ejection and the time of systolic pulmonary arterial flow cessation). To estimate mean PAP, we used a validated formula (21). All measurements were
performed by the study cardiologist and reviewed by a second cardiologist, both of whom were masked to the treatment.

**Statistical Analysis**

We anticipated enrolling a total of 14 subjects in this crossover–randomized study (42 blood challenges). Sample size calculation and statistical analyses were performed using STATA-12 software (StataCorp LP, College Station, TX) value as detailed in the online supplement. Continuous variables are expressed as mean ± SEM or median and interquartile range, as appropriate. Statistical significance was reached with P less than 0.05.

Additional details on the study protocol, NO delivery, estimation of systolic and mean PAP, and statistical methods can be found in the online supplement.

**Results**

Fifty-seven volunteers were screened for a reduced RHI, and 14 were enrolled in the study (nine males and five females). Twelve volunteers completed the study, and two left the study before finishing all three phases (Figure 1). Demographic characteristics are shown in Table 1. Endothelial dysfunction was present in all participants, as reflected by a low RHI at the time of screening (LnRHI = 0.46 ± 0.03) (18), and baseline (before transfusion). Estimated mean PAP at baseline was 18 ± 1 mm Hg, and systolic PAP was 39 ± 2 mm Hg (Figure 2, Table 2).

**Pulmonary Artery Pressure Is Increased during Transfusion of 40-Day Stored Blood, Unchanged after 3 Days, and Decreased during 40-Day+NO**

Blood transfusion with 3-day challenge showed no systemic or pulmonary hemodynamic changes except for a transient decrease in heart rate following transfusion (Table 2; see Table E1).

Similarly, blood transfusion with 40-day challenge did not change systemic blood pressure, cardiac output, or RHI and only transiently decreased heart rate following transfusion. However, during transfusion, we measured a decreased PAAT and PAAT/RVET (P = 0.006 and P = 0.004, respectively) corresponding to an increase of mean and systolic PAP of 5 and 6 mm Hg, respectively (P = 0.009 and P = 0.006, respectively) (Figure 2, Table 2; see Table E1).

When volunteers breathed NO before and during transfusion of 40-day stored blood (40-d+NO challenge), the PAAT and PAAT/RVET increased (P = 0.004 and P = 0.009, respectively) and the mean PAP decreased by 5 mm Hg, and the systolic PAP decreased by 7 mm Hg (P = 0.005 and P = 0.004, respectively), whereas systemic vascular hemodynamic measurements were unchanged (Figure 2, Table 2; see Table E1).

When mean PAP values were compared, we found that during transfusion PAP tended to be greater in the 40-day challenge group as compared with PAP in the 3-day challenge group (40-d vs. 3-d; P = 0.054); also, PAP was lower in the 40-day+NO group compared with PAP in the 3-day group (40-d+NO vs. 3-d; P = 0.007); and, the PAP was markedly greater in the 40-day challenge group as compared with PAP in 40-day+NO (40-d vs. 40-d+NO; P = 0.001) (Figure 2, Table 2).

**Table 1. Baseline Characteristics**

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>n = 14</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male, n (%)</td>
<td>9 (64%)</td>
</tr>
<tr>
<td>Age, yr</td>
<td>41 ± 4</td>
</tr>
<tr>
<td>Race, n (%)</td>
<td></td>
</tr>
<tr>
<td>White</td>
<td>12 (86%)</td>
</tr>
<tr>
<td>African-American</td>
<td>1 (7%)</td>
</tr>
<tr>
<td>Asian</td>
<td>1 (7%)</td>
</tr>
<tr>
<td>Hispanic</td>
<td>0</td>
</tr>
<tr>
<td>ABO blood group, n (%)</td>
<td></td>
</tr>
<tr>
<td>A</td>
<td>3 (22%)</td>
</tr>
<tr>
<td>B</td>
<td>2 (14%)</td>
</tr>
<tr>
<td>AB</td>
<td>2 (14%)</td>
</tr>
<tr>
<td>O</td>
<td>7 (50%)</td>
</tr>
<tr>
<td>BMI, kg/m²</td>
<td>33.4 ± 1.3</td>
</tr>
<tr>
<td>LnRHI</td>
<td>0.46 ± 0.03</td>
</tr>
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</table>

Definition of abbreviations: BMI = body mass index; LnRHI = natural log of reactive hyperemia index.

Data are shown as mean ± SEM or count (%).
challenge groups was associated with the change (Δ) in mean PAP during blood transfusion from baseline (P < 0.001, correlation between log_{10} plasma NO consumption levels at 10 minutes vs. Δ mean PAP) (Figure 3D).

As expected, in the 40-day+NO challenge, there was no correlation between plasma NO consumption and plasma hemoglobin levels or between plasma NO consumption at 10 minutes and PAP (data not shown).

### Supernatant Levels of Hemoglobin in the PRBC Units Are Increased with Storage Time and Correlate Closely with Supernatant NO Consumption

Supernatant hemoglobin and NO consumption levels in the RBC units of the 40-day and 40-day+NO challenges were approximately four times higher than those in the 3-day RBC units (Figures 4A and 4B). NO consumption in the supernatant of the RBCs units of the 3-day, 40-day, and 40-day+NO challenges closely correlated with supernatant levels of hemoglobin (P < 0.001, correlation between log_{10} supernatant hemoglobin vs. log_{10} supernatant NO consumption) (Figure 4C). The Δ mean PAP in the 3-day and 40-day challenge groups correlated with the levels of NO consumption in the supernatant of the transfused RBC unit (P = 0.003, correlation of the log supernatant NO consumption vs. Δ mean PAP) (Figure 4D). In the 40-day+NO challenge group the Δ mean PAP did not correlate with the level of NO consumption in the supernatant of the transfused RBC units.

### Endothelial Dysfunction Is Associated with Low Plasma Levels of NO Metabolites; Breathing NO Immediately Increases Plasma Nitrate Levels but Not Plasma Nitrite Levels

At baseline (before transfusion), endothelial dysfunction in our overweight-obese volunteers was characterized by a reduced RHI (decreased availability of endothelial NO) (22) and confirmed by extremely low plasma levels of nitrite and nitroso compounds compared with prior reports in the literature (see Table E3) (10, 23, 24). Other NO metabolites were comparable with the levels that we previously reported in young healthy human volunteers (see Table E3) (10).

Levels of NO metabolites in plasma and erythrocytes showed similar trends after transfusion with 3-day and 40-day challenges (see Figure E4). There was no significant decrease in plasma and erythrocyte nitrate levels after transfusion with 3-day and 40-day challenges. Plasma nitrite levels increased slightly, whereas erythrocyte nitrite levels halved immediately following transfusion, and there was no change in the plasma or erythrocyte levels of nitroso compounds after transfusion with either 3-day or 40-day stored RBCs.

After transfusion with 40-day+NO stored blood, nitrate levels in plasma and erythrocytes increased immediately after inhalation of NO and returned to baseline levels at 24 hours (see Figures E4A–E4D). Plasma levels of nitrite increased only after 24 hours in the 40-day+NO challenge group (see Figure E4). Concentrations of other NO metabolites (RXNO, NOx) in plasma and erythrocytes after NO breathing did not significantly differ from

### Table 2. Hemodynamic and Transthoracic Echocardiography Variables before and during Transfusion

<table>
<thead>
<tr>
<th></th>
<th>3 Days (n = 9)</th>
<th></th>
<th>40 Days (n = 11)</th>
<th></th>
<th>40 Days+NO (n = 12)</th>
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<tbody>
<tr>
<td></td>
<td>Pre</td>
<td>During</td>
<td>Pre</td>
<td>During</td>
<td>Pre</td>
</tr>
<tr>
<td>PAAT, ms</td>
<td>132 ± 9</td>
<td>129 ± 8</td>
<td>127 ± 8</td>
<td>110 ± 6*</td>
<td>132 ± 7</td>
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<tr>
<td>PAAT/RVET, mm Hg</td>
<td>0.40 ± 0.03</td>
<td>0.39 ± 0.02</td>
<td>0.40 ± 0.03</td>
<td>0.32 ± 0.02*</td>
<td>0.39 ± 0.02</td>
</tr>
<tr>
<td>Mean PAP, mm Hg</td>
<td>17 ± 2</td>
<td>18 ± 2</td>
<td>18 ± 2</td>
<td>23 ± 2*</td>
<td>17 ± 2</td>
</tr>
<tr>
<td>Systolic PAP, mm Hg</td>
<td>38 ± 3</td>
<td>39 ± 3</td>
<td>40 ± 3</td>
<td>46 ± 3*</td>
<td>38 ± 3</td>
</tr>
<tr>
<td>CO, L/min</td>
<td>5.3 ± 0.4</td>
<td>5.0 ± 0.3</td>
<td>4.5 ± 0.4</td>
<td>4.6 ± 0.3</td>
<td>4.5 ± 0.3</td>
</tr>
<tr>
<td>LVEF, %</td>
<td>65 ± 1</td>
<td>68 ± 2</td>
<td>63 ± 1</td>
<td>63 ± 1</td>
<td>64 ± 2</td>
</tr>
<tr>
<td>SV, ml</td>
<td>87 ± 8</td>
<td>82 ± 7</td>
<td>77 ± 8</td>
<td>78 ± 8</td>
<td>75 ± 6</td>
</tr>
<tr>
<td>LnRHI</td>
<td>0.74 ± 0.09</td>
<td>0.87 ± 0.06</td>
<td>0.74 ± 0.07</td>
<td>0.90 ± 0.06</td>
<td>0.67 ± 0.07</td>
</tr>
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</table>

Definition of abbreviations: CO = cardiac output; LnRHI = natural log of reactive hyperemia index; LVEF = left ventricular ejection fraction; NO = nitric oxide; PAAT = pulmonary artery acceleration time; PAP = pulmonary artery pressure; RVET = right ventricular ejection time; SV = stroke volume.

Data are shown as mean ± SEM. *P < 0.01 paired t test values pretransfusion versus during transfusion.

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the values after 3-day or 40-day transfusions (without NO breathing).

**Discussion**

We investigated the effects of transfusing stored autologous RBCs on systemic and pulmonary hemodynamics and systemic endothelial function in obese adult volunteers with endothelial dysfunction. The principal finding of our study was that autotransfusion of one unit of leukoreduced blood stored for 40 days increased the PAP, as estimated by transthoracic echocardiography. Inhalation of 80 ppm NO prevented the increase in PAP induced by transfusing 40-day stored blood. A second important finding was the absence of systemic hypertension, inflammation, or organ injury after transfusion of 40-day stored blood in our volunteers with preexisting systemic endothelial dysfunction. We previously reported that diabetic mice with endothelial dysfunction were sensitized to systemic vasoconstriction after the transfusion of stored syngeneic blood (15). We also noted that both diabetic and high-fat diet-fed mice developed an enhanced systemic inflammatory response after stored blood transfusion (15, 25). Thus, we were surprised that our obese volunteers with a markedly reduced RHI did not show any systemic vasoconstriction or inflammation after stored 40-day blood transfusion. Similar findings of an absence of inflammation have been reported after transfusion of stored blood into healthy (nonobese) volunteers (10, 26) and intensive care unit patients (27).

We measured the PAAT by transthoracic echocardiography and calculated an estimated mean and systolic PAP (20, 21). Our volunteers with endothelial dysfunction exhibited an increased PAP at baseline (before transfusion). These observations are in agreement with those of McQuillan and coworkers (27), who reported that patients with a higher body mass index had an increased systolic PAP. It is possible that the combination of obesity and systemic endothelial dysfunction placed our volunteers at higher risk for pulmonary vasoconstriction when challenged with a stored blood transfusion. Because transfusion with blood stored for 40 days did not change the cardiac output, the increase of PAP that we measured is most likely explained by an increase of pulmonary vascular resistance caused by

![Figure 3. Plasma levels of hemoglobin (Hb), plasma nitric oxide (NO) consumption, and changes of mean pulmonary artery pressure (PAP). (A) Plasma Hb increased after 40-day transfusion challenge (*P < 0.01 plasma Hb at 1-h and 4-h measurements compared with baseline [= before transfusion]), and after 40-day+NO challenge (**P < 0.01 plasma Hb at 10-min, 1-h, 4-h, 24-h measurements compared with baseline in 40-d challenge). No significant changes were found after challenge with 3-day transfusion. (B) Plasma NO consumption increased only at 4 hours after transfusion of 40-day stored blood (**P = 0.001). (C) Plasma Hb concentration at baseline, 10 minutes, and 4 hours after transfusion in volunteers receiving 3-day and 40-day challenges correlates with NO consumption (P < 0.001). (D) Plasma NO consumption at 10 minutes after 3-day and 40-day challenges correlates with Δ mean PAP (mm Hg) (P < 0.001). Data are presented as mean ± SEM. In A and B, a two-way analysis of variance for repeated measures with post hoc Bonferroni correction. In C and D, measurements of plasma Hb and NO consumption were transformed into a log10 scale.
pulmonary vasoconstriction. Without inserting a pulmonary artery catheter, we are unable to determine whether transfusion altered the pulmonary capillary wedge pressure. However, because no change in PAP was measured following transfusion with blood stored for 3 days, it is unlikely that the wedge pressure was changed by transfusing one unit of 40-day RBCs.

Transfusion of blood stored for prolonged periods into either lambs or dogs produces acute pulmonary hypertension (12, 28–30). Baron and coworkers (12) developed a model of autologous blood transfusion in the awake lamb and reported that top-load transfusion of one unit of blood stored for 40 days induced transient pulmonary vasoconstriction and an increased PAP. Treating lambs with a low dose of L-NAME to mimic endothelial dysfunction markedly augmented the pulmonary vasoconstriction following transfusion with stored blood (12). In a subsequent study of lambs resuscitated with autologous stored blood after hemorrhagic shock, Baron and colleagues (17) reported enhanced pulmonary vasoconstriction and hypertension. Moreover, Solomon and coworkers (28) described persistent pulmonary hypertension in a septic canine model subjected to an exchange transfusion with stored canine donor blood. In elegant studies, Stevens and coworkers showed that the relevant hydrostatic pressure for alveolar edema formation is the transcapillary pressure, which depending on the model may or may not correlate with PAP (31, 32). In Solomon’s canine study, the increase of pulmonary hypertension after stored blood transfusion was associated with more severe lung damage than the fresh blood transfused control, evidenced by increased necrosis, hemorrhage, and thrombosis (28). Similarly, Baron and coworkers (17) reported that two of six animals transfused with stored blood developed severe pulmonary edema. Thus, any connection between the increased PAP and lung edema, necrosis, hemorrhage, and thrombosis especially in patients with acute lung injury receiving stored blood should be closely examined.

To determine the degree of depletion of NO in plasma after blood transfusion and the benefits of breathing NO, we measured NO consumption in both stored RBC supernatant and the recipient’s plasma after transfusion. During storage, we found that the NO consumption of the supernatant was markedly increased, as others have reported (9, 17). Supernatant NO consumption closely correlated with the level of plasma hemoglobin in the supernatant. The increase of PAP during transfusion correlated with both the

Figure 4. Plasma hemoglobin (Hb) and plasma nitric oxide (NO) consumption in the supernatant of red blood cell (RBC) unit, and changes of mean pulmonary artery pressure (PAP). (A) Supernatant Hb levels contained in the 40-day and 40-day+NO stored blood units are higher than those in the supernatant of the 3-day stored units (P < 0.01). (B) Supernatant NO consumption is higher in the 40-day and 40-day+NO stored units as compared with the NO consumption of the 3-day stored units (P < 0.01). (C) Supernatant NO consumption correlates with supernatant Hb levels from all RBC units (P < 0.001). (D) Supernatant NO consumption correlates with change of mean PAP after 3-day and 40-day challenges (P = 0.003). Despite high NO consumption levels of the 40-day+NO stored units, breathing NO decreases mean PAP. Data presented as median and interquartile range. In A and B, the P value was calculated with a Kruskal-Wallis rank test. In C and D, measurements of Hb in the supernatant and NO consumption were transformed to a log10 scale. Note that there are three fewer points in D compared with Figure 3D: two samples from the 3-day stored blood units and one sample from the 40-day stored unit were not obtained.
level of NO consumption of the supernatant in the transfused RBC unit and with the level of NO consumption in the transfusion recipient’s plasma measured 10 minutes after completing the transfusion. It is likely that the increase of PAP during transfusion of blood stored for 40 days is caused by the transfusion of hemoglobin contained in the blood storage unit and by hemolysis of circulating transfused erythrocytes. The peak level of plasma hemoglobin in our volunteers occurred between 1 and 4 hours after the transfusion, consistent with ongoing hemolysis after transfusion.

Breathing NO is an efficacious and safe method to selectively reduce pulmonary hypertension. Minneci and coworkers (16) demonstrated that inhaled NO reversed the pulmonary hypertension produced by hemoglobin infusion in dogs. Many thousands of patients with acute pulmonary hypertension have been successfully treated with inhaled NO (13, 33). In children with pulmonary endothelial dysfunction proved by a lack of response to acetylcholine infusion, and pulmonary hypertension following extracorporeal bypass, Wessel and coworkers (34) showed that PAP was normalized by breathing NO. In our study, breathing NO before and during transfusion abolished the acute pulmonary pressure increase induced by transfusing blood stored for 40 days in volunteers with endothelial dysfunction. Thus, we believe that inhaled NO prevented or reversed any vasoconstriction caused by NO scavenging by hemoglobin. We reviewed other possible sources for the increase of PAP with stored blood transfusion, such as reduced red cell deformability (35), and presumably increased viscosity. There is limited evidence that NO exposure can reduce red cell deformity (36). If reactive oxygen species are elevated by iron released by stored blood and help produce vasoconstriction, it is possible that inhaled NO could scavenge oxygen free radicals (37). These results suggest that patients with endothelial dysfunction that receive stored blood transfusions could be treated with inhaled NO to prevent an increased pulmonary pressure.

Finally, to further investigate the response to inhaled NO after transfusion of stored blood in volunteers with endothelial dysfunction, we measured NO metabolites in plasma and erythrocytes. We report that baseline levels of nitrite were three times lower than the levels we measured in healthy young volunteers without endothelial dysfunction (10). Unlike our previous study in healthy volunteers, volunteers with endothelial dysfunction showed no change of NO metabolite levels after transfusion of either 3-day or 40-day stored blood challenges. In particular, plasma nitrite levels, despite a slight increase immediately after transfusion of 3-day or 40-day blood, remained at extremely low levels when compared with healthy individuals. It is likely that the diseased endothelium of our obese volunteers was incapable of increasing NO synthesis after transfusing free hemoglobin. This is in contrast to our previous study of healthy volunteers where circulating nitrite levels were increased after stored blood transfusion. Breathing NO produced an immediate increase of erythrocytic and plasma levels of nitrate but not nitrite. Thus, it is unlikely that breathing NO produced its effects via nitrite release.

Our study has several limitations. First, this is an exploratory study conducted with only 14 human volunteers. Larger clinical trials of stored blood transfusion may eventually clarify whether an increase in PAP manifests after multiple transfusions with stored blood and if this translates to increased adverse pulmonary and cardiac events.

Second, we did not explore the effects of breathing NO in the absence of a transfusion in overweight volunteers with systemic endothelial dysfunction, we also did not study the effects of a volume challenge (saline or albumin) on systemic and pulmonary hemodynamics, and we did not investigate the effects of breathing NO during transfusion of 3-day stored blood. These additional control groups need to be examined in future studies. Third, without continuous monitoring of PAP by a pulmonary catheter, we do not know if the peak of in vivo hemolysis at 1–4 hours correlates with an increased PAP. Future studies should investigate whether in vivo hemolysis causes persistent pulmonary vasoconstriction. Lastly, we did not evaluate other possible noxious effects of stored blood transfusion, such as iron and heme toxicity, microparticles, protein oxidation, or lipid peroxidation of the RBC membrane.

In conclusion, in our randomized crossover trial of autologous leukoreduced stored blood transfusion in overweight and obese adult volunteers with systemic endothelial dysfunction, we found that transfusion of leukoreduced erythrocytes stored for 40 days was associated with an increased PAP, most likely caused by NO depletion by plasma hemoglobin. Selective pulmonary vasodilation induced by breathing 80 ppm NO prevented the increase of PAP after the transfusion of 40-day stored red cells. This study demonstrates an increase in PAP following transfusion of 40-day stored blood and provides a safe and practical pharmacologic intervention to avoid the increase of PAP. Clinical trials of stored blood transfusion should prospectively search for pulmonary hypertension and any adverse effect of augmenting pulmonary edema, especially in critically ill patients with endothelial dysfunction and acute lung injury.

Author disclosures are available with the text of this article at www.atsjournals.org.

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