

## File S1

### Supporting Methods

**F2 population generation and sequencing:** An F2 population from an initial cross between plants *S. aethnensis* Pro2-21 (collected from Piano Provenzana, Mt. Etna, altitude 2036 m.; male parent) and *S. chrysanthemifolius* Ran1-5 (collected near Randazzo, Mt. Etna, altitude 763 m. see table 1 in (Chapman *et al.* 2013); female parent) was generated and grown in a growth room with the same temperature and light/dark cycle as the plants analysed in the initial transcriptomic investigation (Chapman *et al.* 2013). The parent plants were morphologically typical of the species and showed no evidence for introgression in a previous analysis (Chapman *et al.* 2013). Some admixture, however, cannot be ruled out, especially for the *S. aethnensis* parent, which was collected from a population close to the high altitude edge of the hybrid zone.

In total 147 F2 plants were grown, 64 of which were selected for RNA-seq analysis. F2 hybrid breakdown was observed in this cross, with 69 plants developing signs of necrosis or stunted growth, and not reaching maturity (collectively termed ‘necrotic’; Figure 1 A-D) and 78 healthy plants that grew to maturity (Figure 1 E-F) in the population. The 64 sequenced plants comprised 48 healthy and 16 necrotic F2s and apart from this designation were selected randomly.

RNA was extracted from leaf tissue of 3 week old seedlings using an RNeasy kit (Qiagen, Manchester, UK) with additional DNA removal using DNase (Qiagen), and from each an individual RNA-seq library was constructed. Sampling from seedlings reduces the variability associated with developmental and morphological differences that are more pronounced as the plants mature. These 64 libraries were then run (as a multiplex) across two lanes of a HiSeq2000 at the Wellcome Trust Centre for Human Genetics.

**Expression Analysis:** Comparisons were then made between the 48 healthy and 16 necrotic plants using t-tests, with significant differences being detected after applying an FDR of 5%. To determine if

certain categories of Gene Ontology (GO) were over-represented in the differentially expressed loci we carried out Fisher's Exact Tests in Blast2GO (Conesa *et al.* 2005; Gotz *et al.* 2008), applying a 5% FDR.

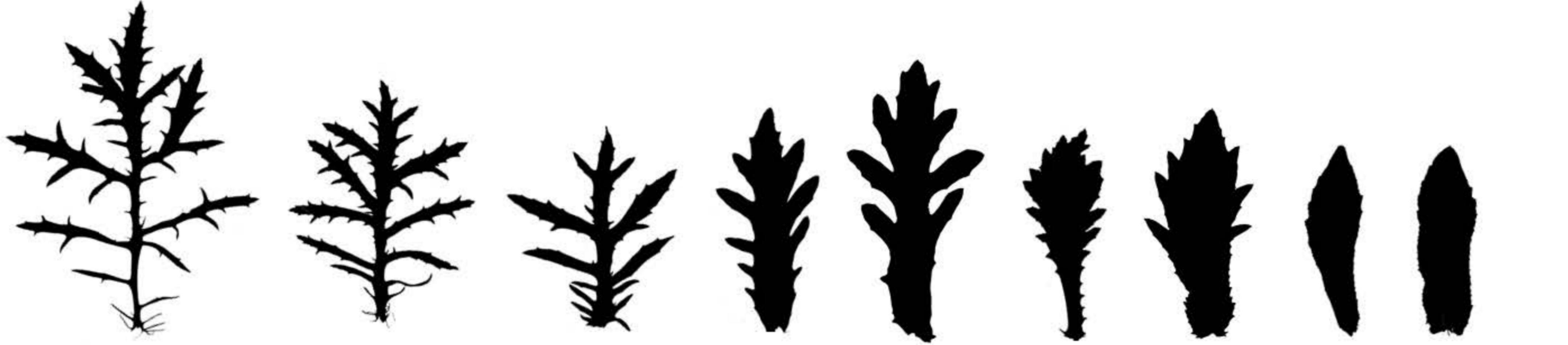
**Linkage Mapping:** SNPs from the loci that fulfilled the above criteria were identified using Proseq and imported into Microsoft Excel where consensus genotypes (per locus per individual) were produced. Up to 20% inconsistent genotyping within an individual was considered tolerable as this was likely a result of mis-mapping of a read by CLC-GW, or mis-scoring of a SNP by SAMtools. If more than 20% of SNPs showed an erroneous pattern then this locus was scored as missing data for this individual. Because a major aim of this portion of the analysis was to map highly differentiated outlier loci, we preferentially added to this list several outlier loci that did not fulfil criterion (2) above, i.e. less than five (but more than two) SNPs differentiated the parental alleles. For these loci, inconsistent SNP genotypes within a locus were not tolerated.

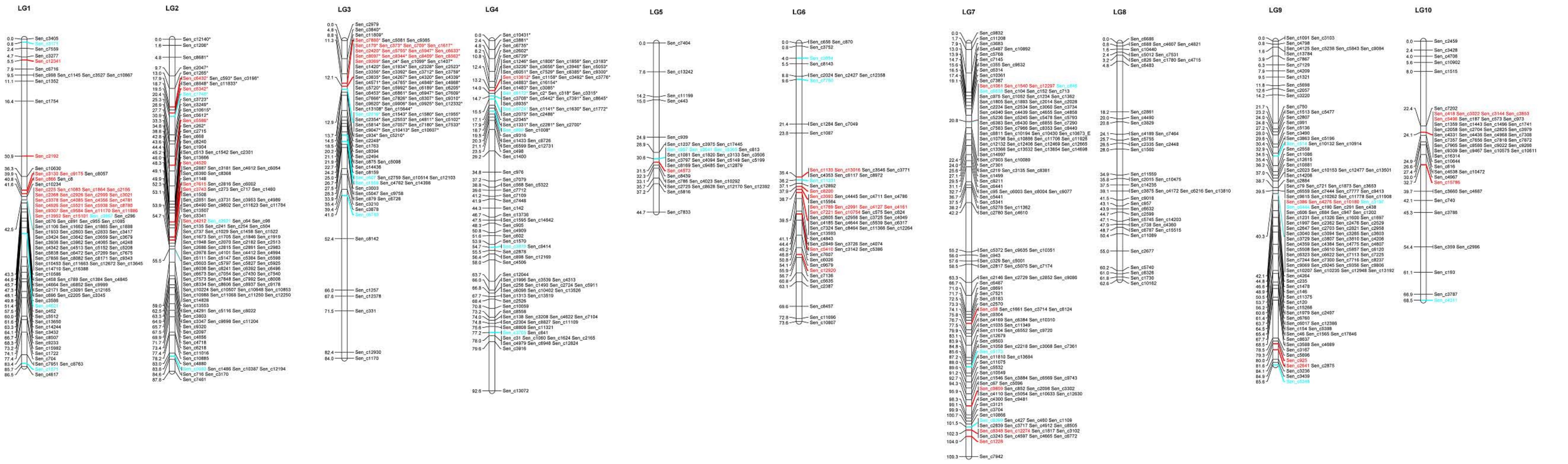
**Manual genotype verification:** Several markers on linkage group (LG) 4 were never homozygous in the F2 for alleles inherited from the *S. chrysanthemifolius* parent and so we sequenced twelve loci in or close to this region from the two parents and the two F1 plants to ensure that they were homozygous (for alternate alleles), and heterozygous, respectively. PCR amplification and sequencing followed the protocol outlined in (Chapman *et al.* 2013).

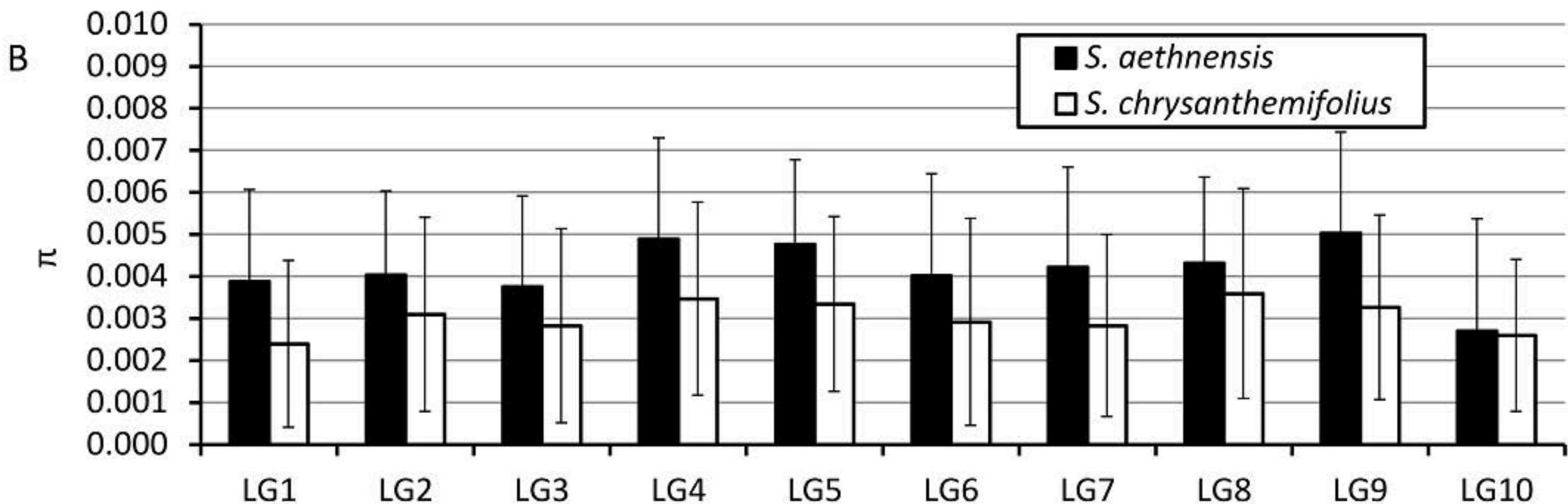
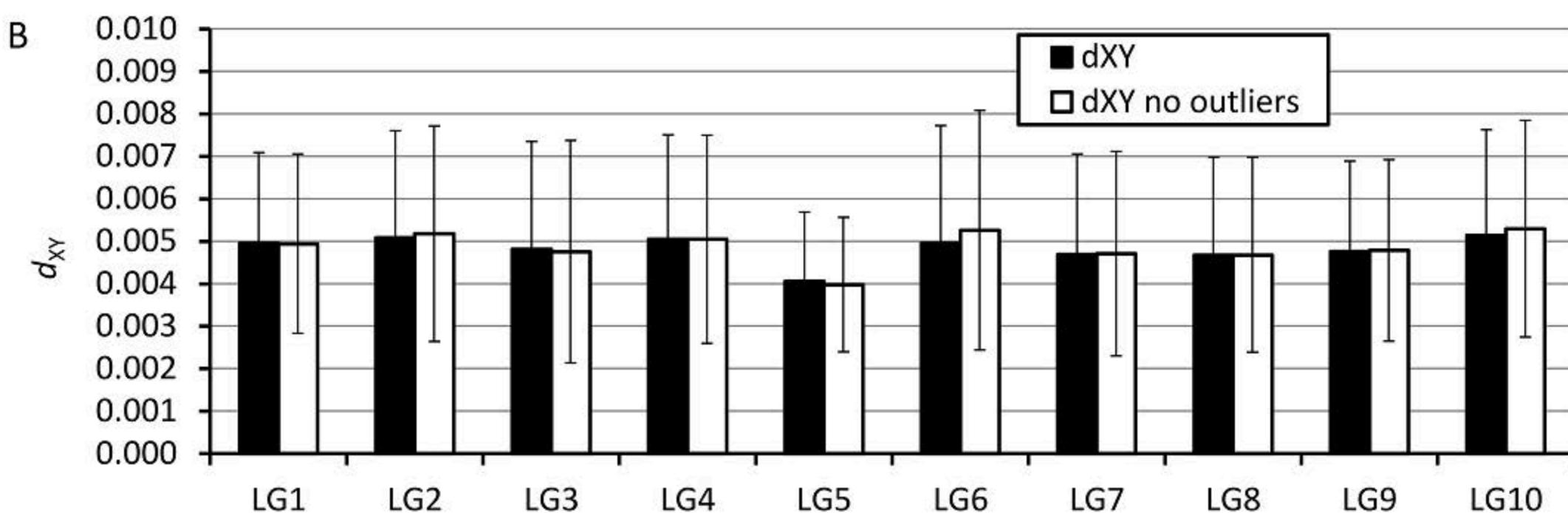
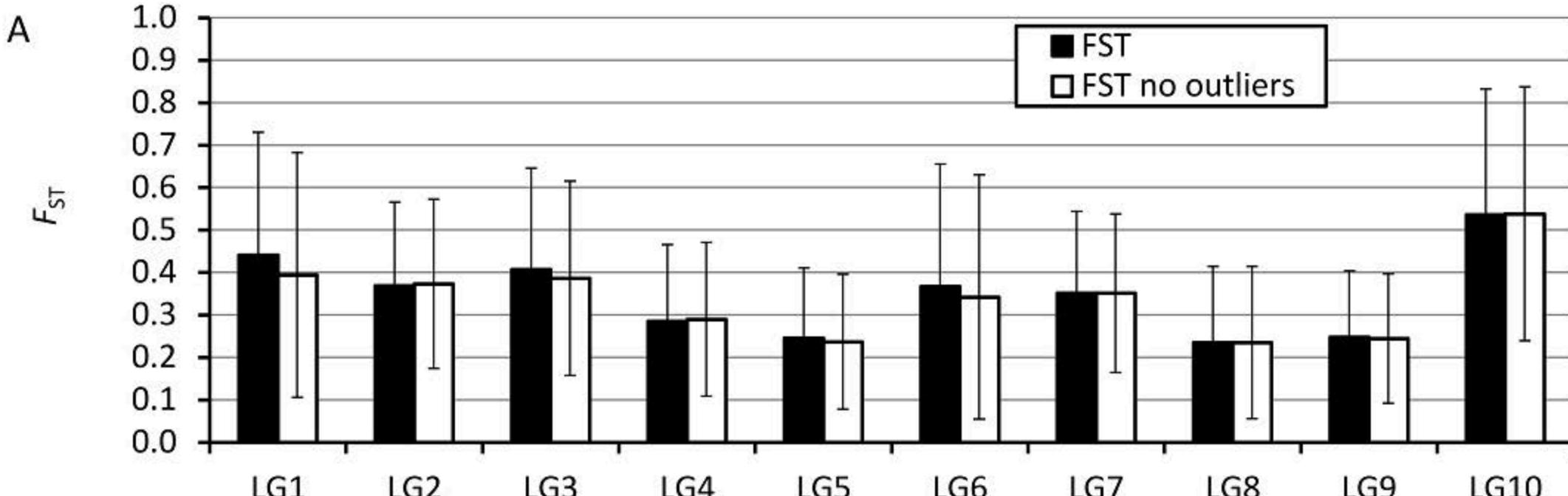
For six of the above loci and ten other loci exhibiting segregation distortion length differences caused by indels were identified between the parental alleles (either directly from the F2 transcriptome alignments, or from sequencing intron-containing regions of the loci from the parental individuals). For genotyping, loci were PCR amplified using a fluorescent '3rd primer' using protocols established for microsatellites (Chapman *et al.* 2008) and resolved on an ABI 3730 in the Department of Zoology, University of Oxford. Primer sequences are given in Table S2. Alleles were scored with GeneMarker (ver. 2.4.0; [www.softgenetics.com](http://www.softgenetics.com)).

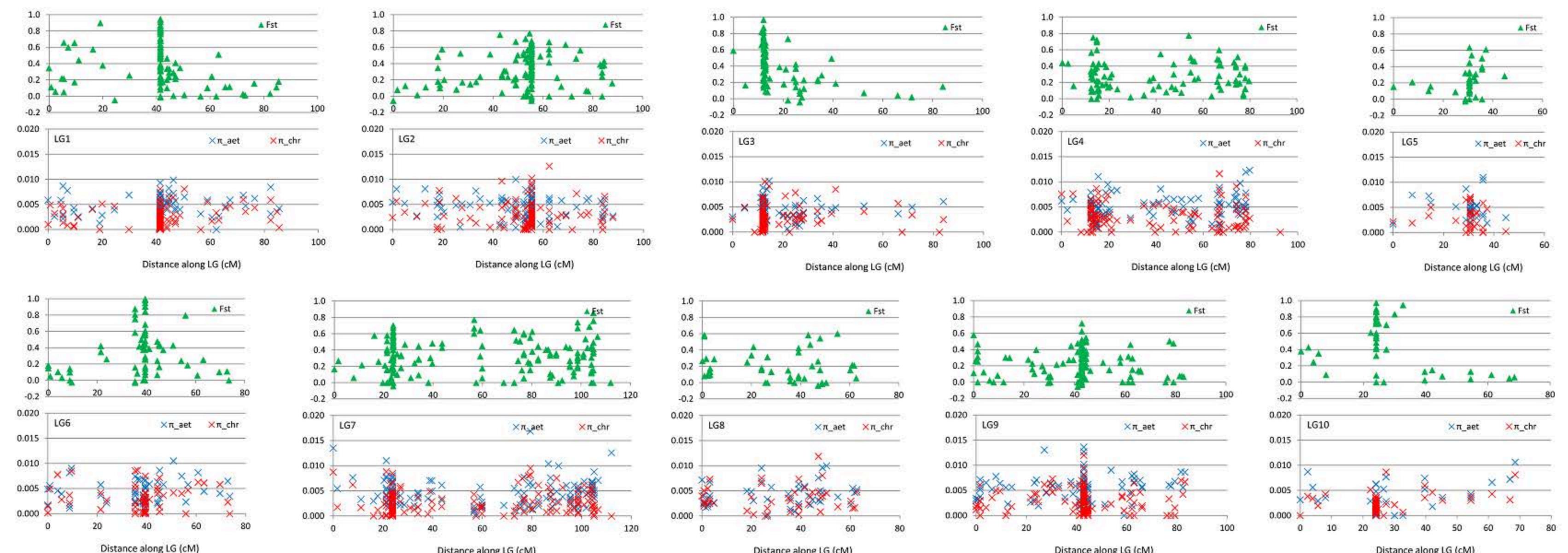
**Analysis of Polymorphism and Divergence:** Values were compared between LGs and between species, where appropriate, using Mann-Whitney U-tests, and the distribution of outlier markers per bin (5 and 10 cM bins) was compared to that expected based on marker density alone using a Fisher's Exact Test, in PAST (<http://folk.uio.no/ohammer/past/>). Analysing the data based on bins of a certain number of loci was precluded due to inability to resolve map positions uniquely (i.e. clustering of many loci).

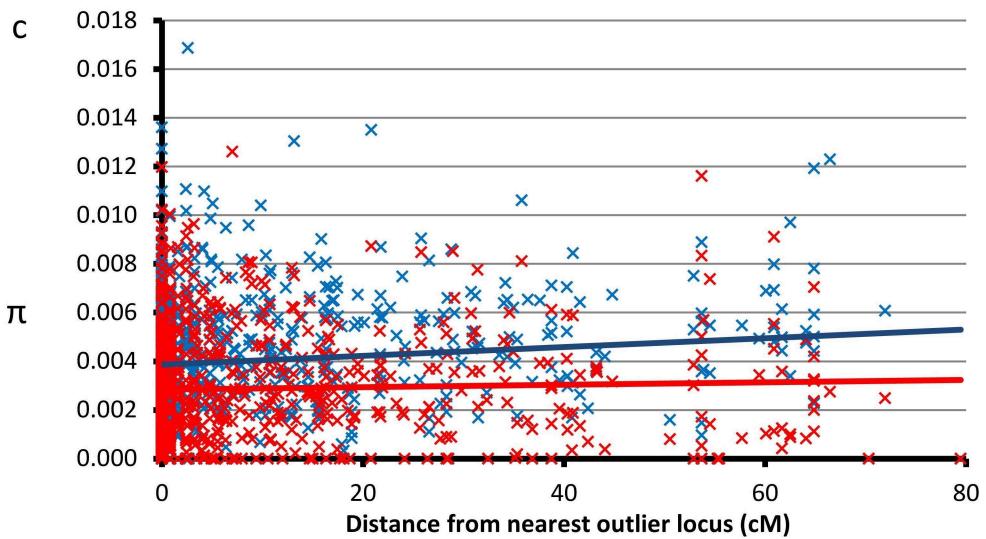
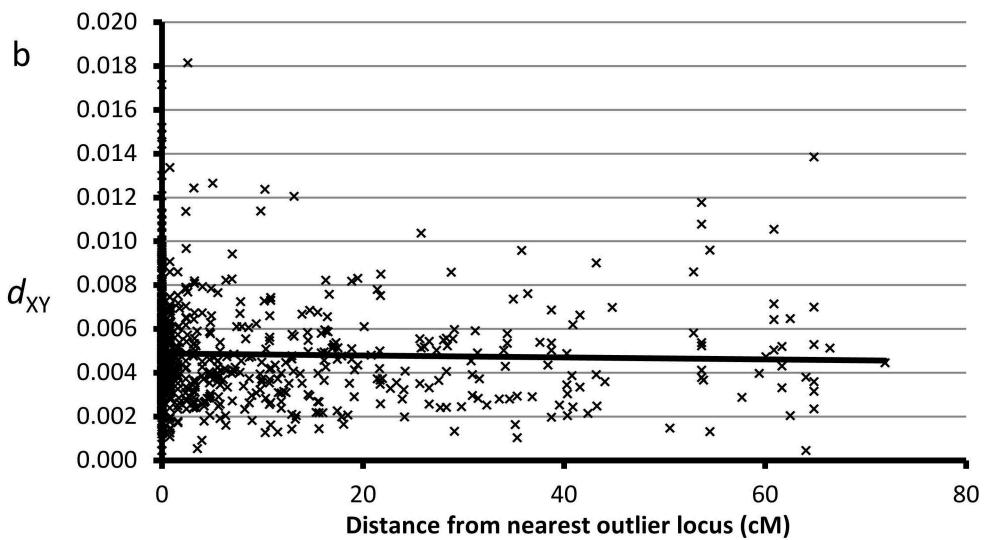
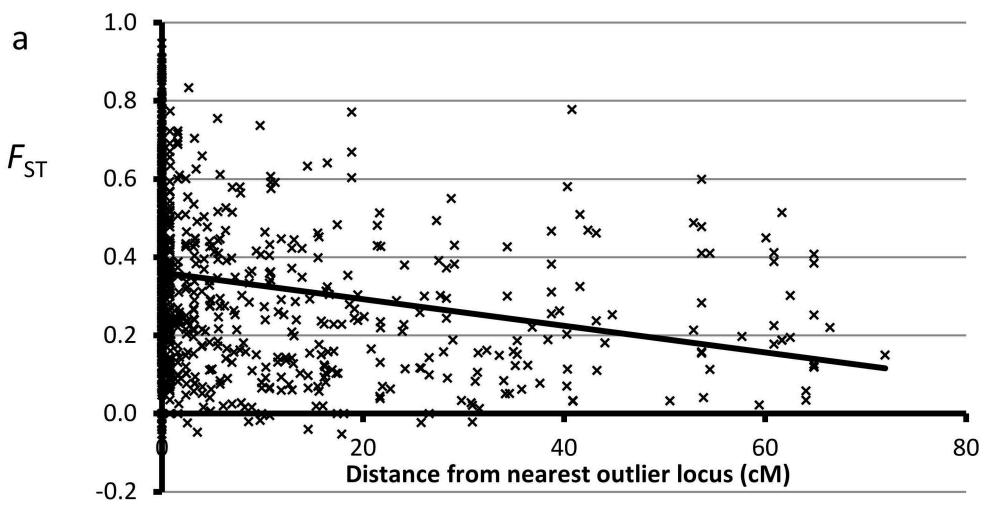
- Chapman, M. A., S. J. Hiscock and D. A. Filatov, 2013 Genomic divergence during speciation driven by adaptation to altitude. *Molecular Biology and Evolution* **30**: 2553-2567.
- Chapman, M. A., C. H. Pashley, J. Wenzler, J. Hvala, S. Tang *et al.*, 2008 A genomic scan for selection reveals candidates for genes involved in the evolution of cultivated sunflower (*Helianthus annuus*). *The Plant Cell* **20**: 2931-2945.
- Conesa, A., S. Gotz, J. M. Garcia-Gomez, J. Terol, M. Talon *et al.*, 2005 Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics* **21**: 3674-3676.
- Gotz, S., J. M. Garcia-Gomez, J. Terol, T. D. Williams, S. H. Nagaraj *et al.*, 2008 High-throughput functional annotation and data mining with the Blast2GO suite. *Nucleic Acids Research* **36**: 3420-3435.

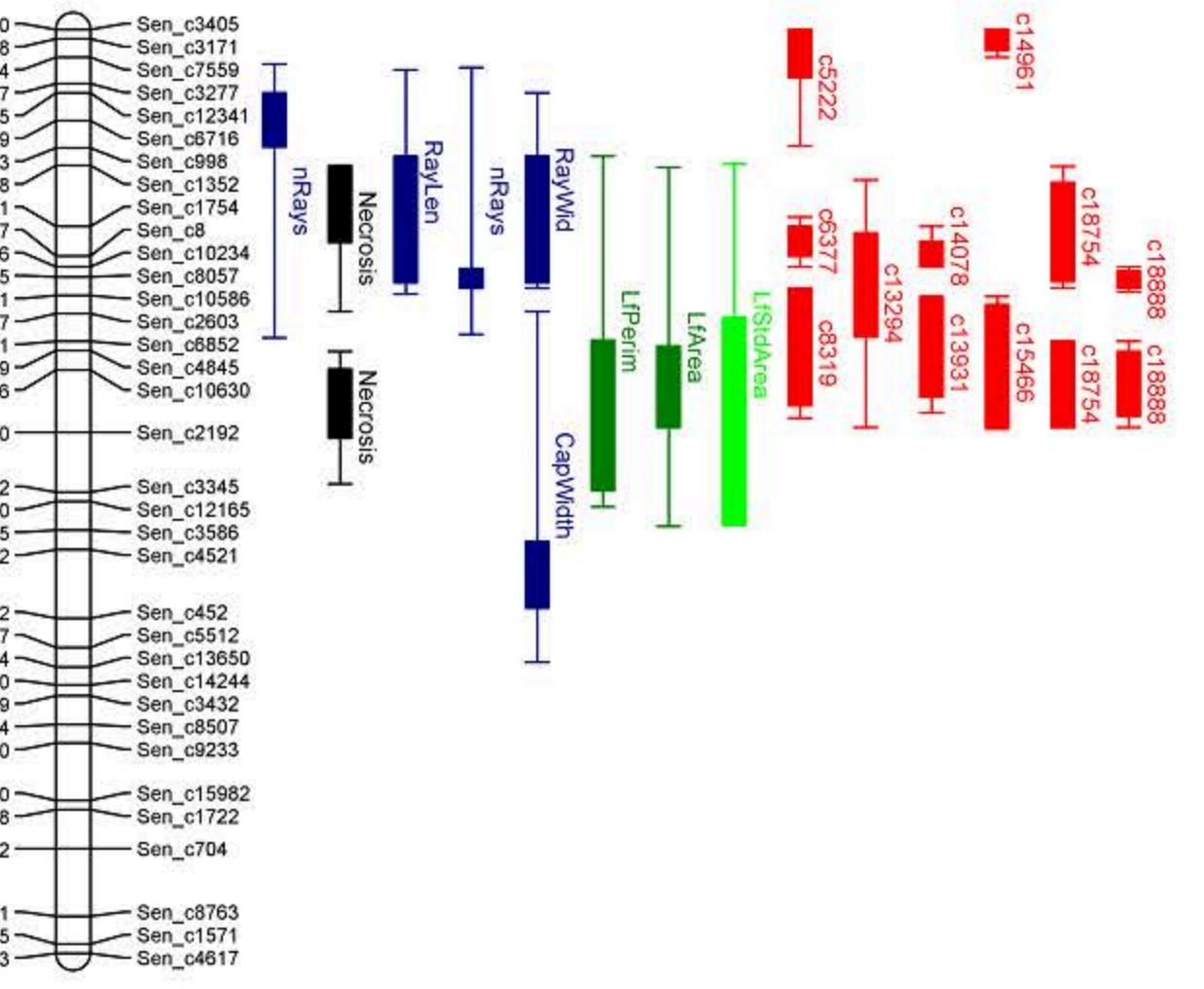




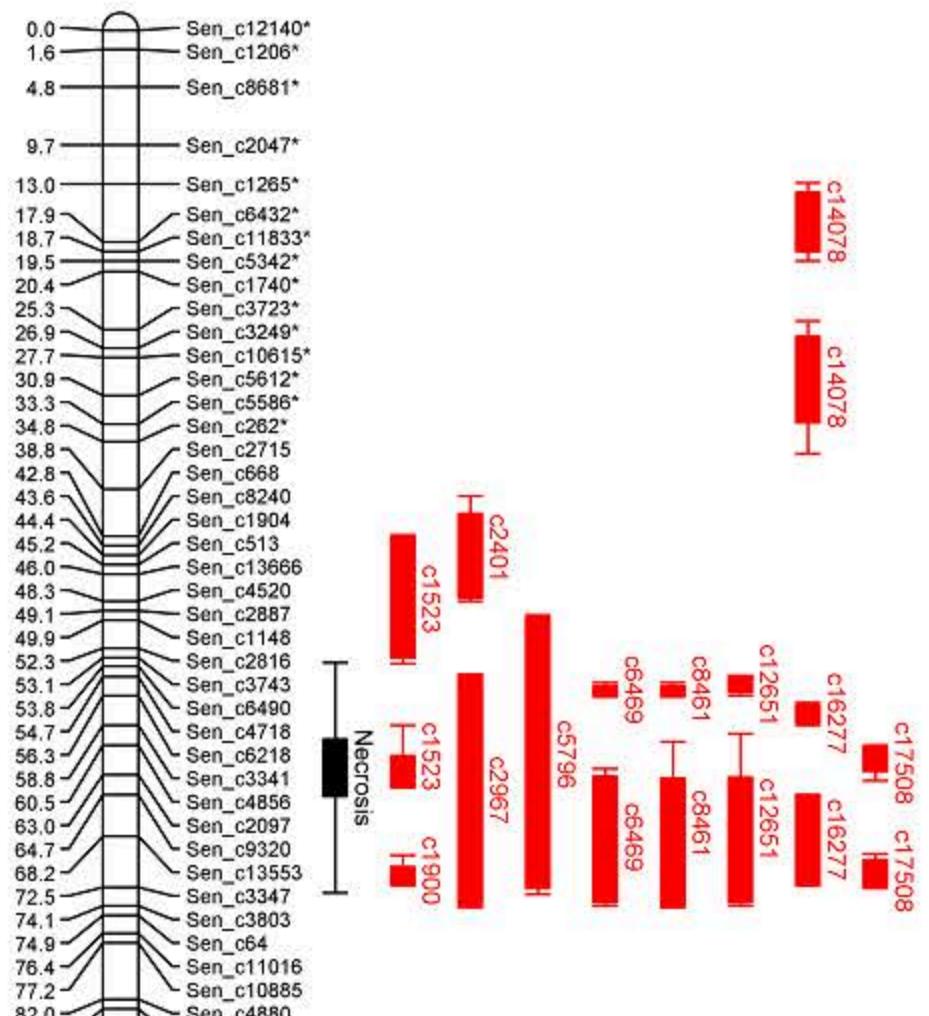




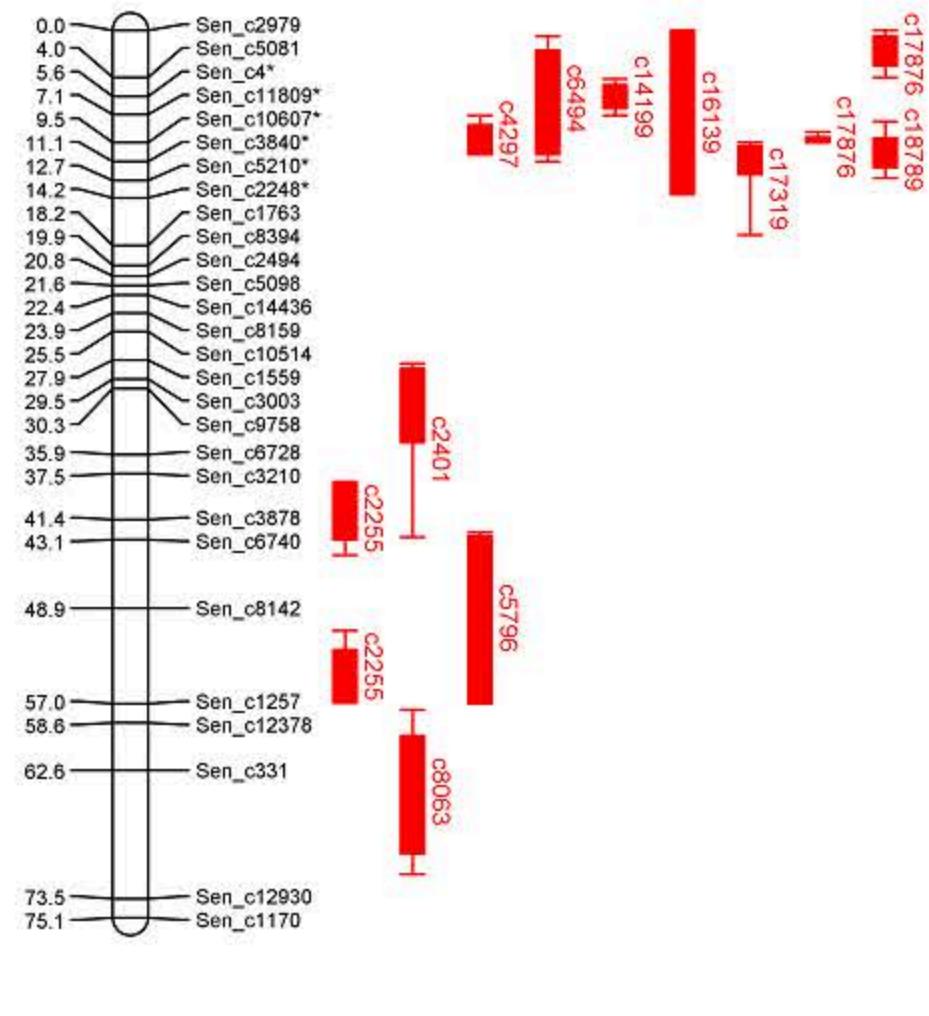




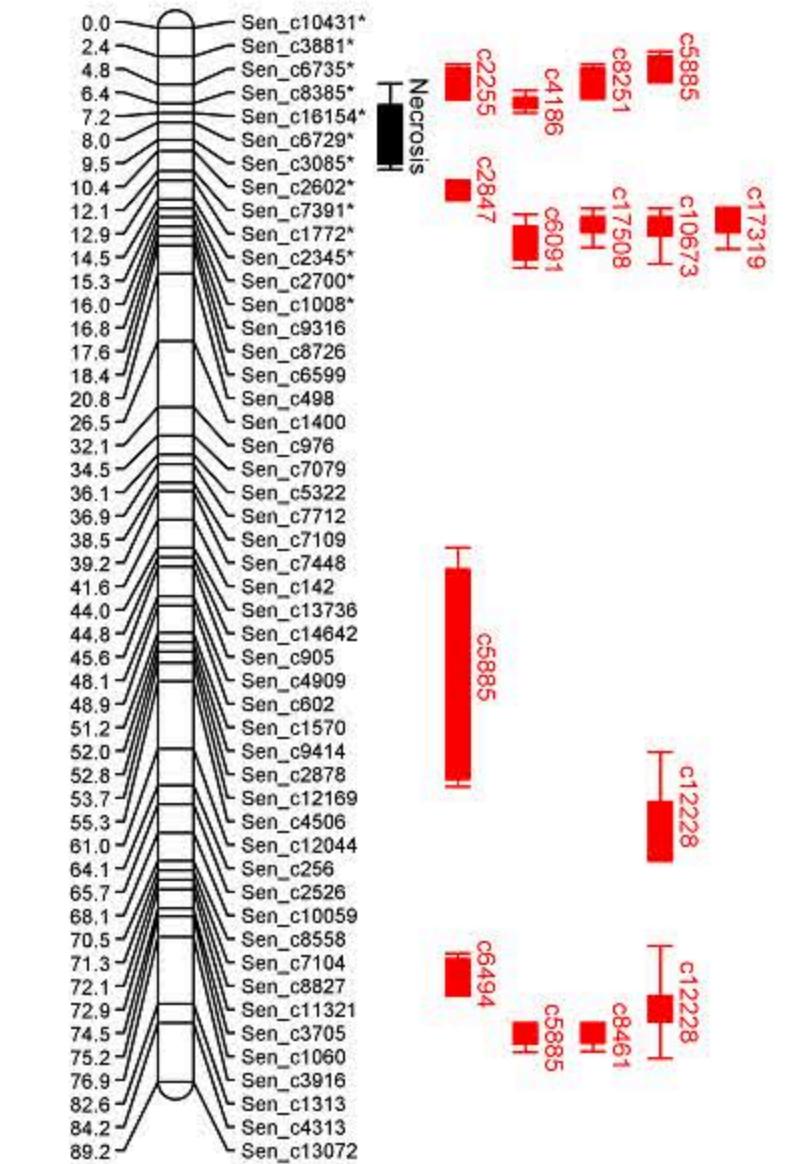
LG2



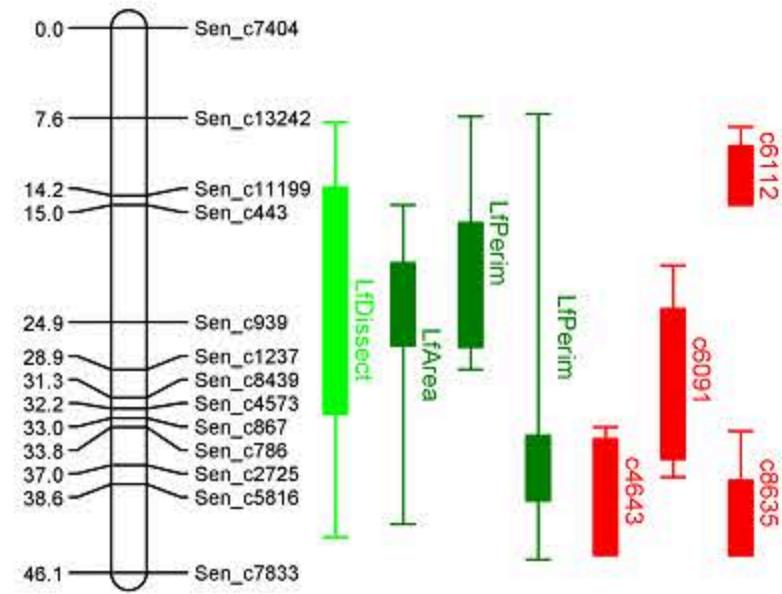
LG3



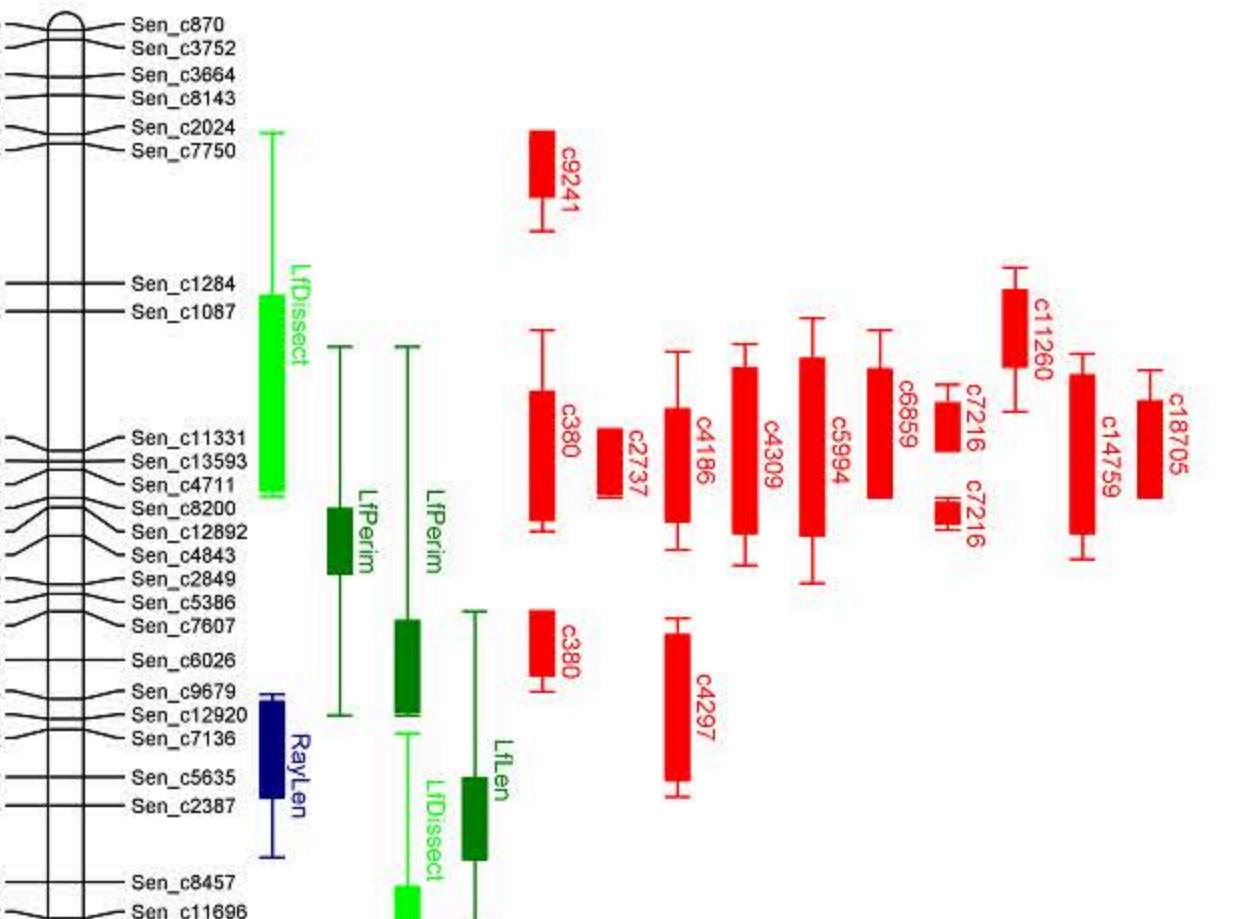
LG



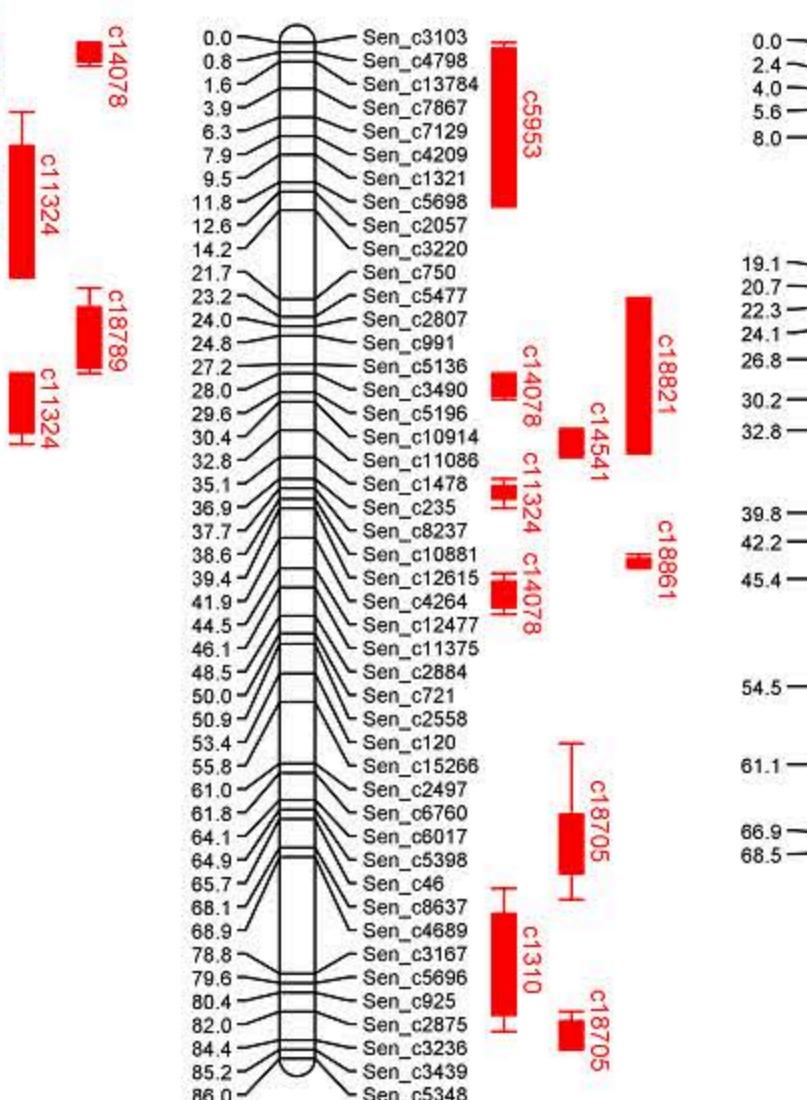
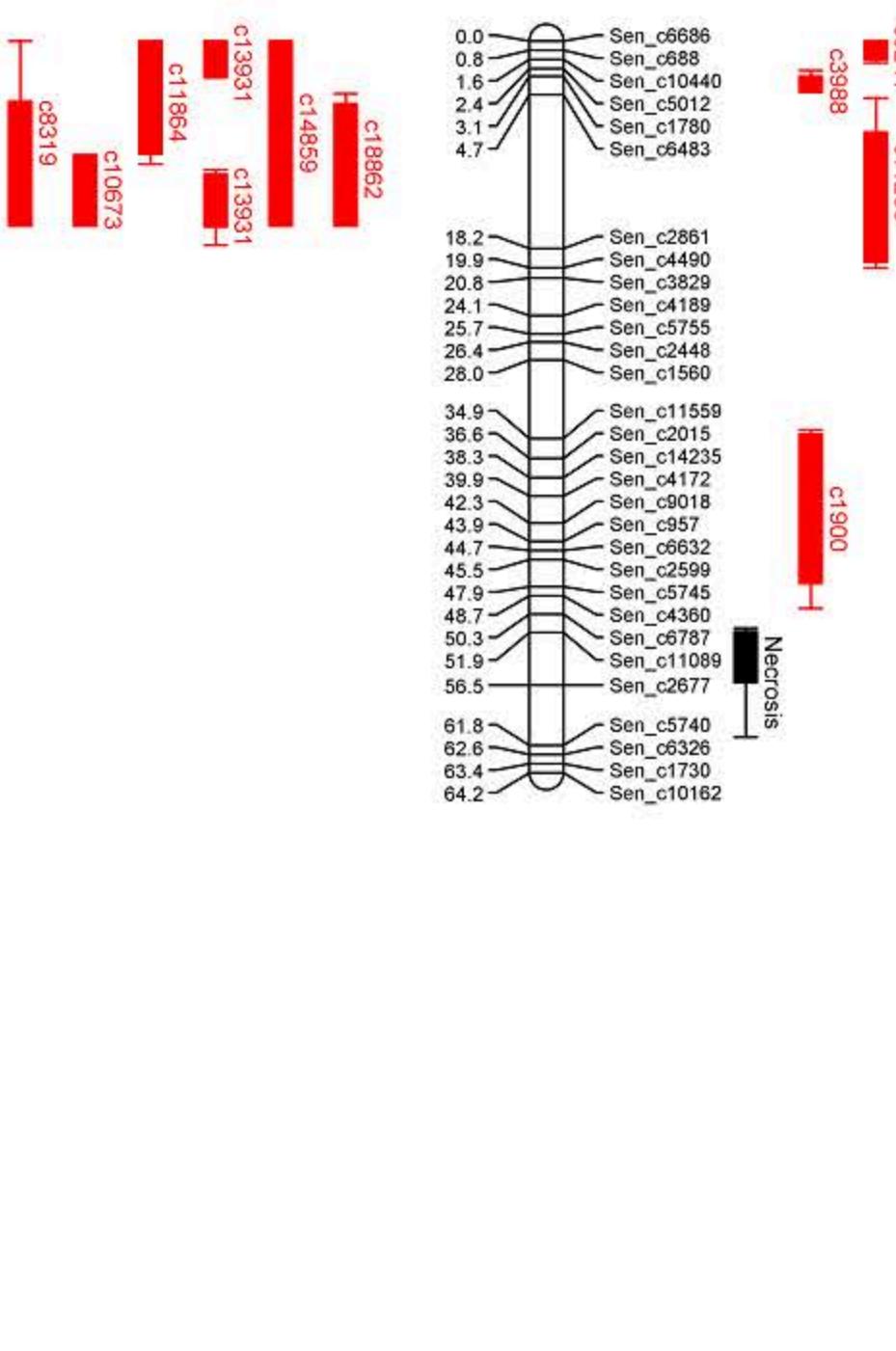
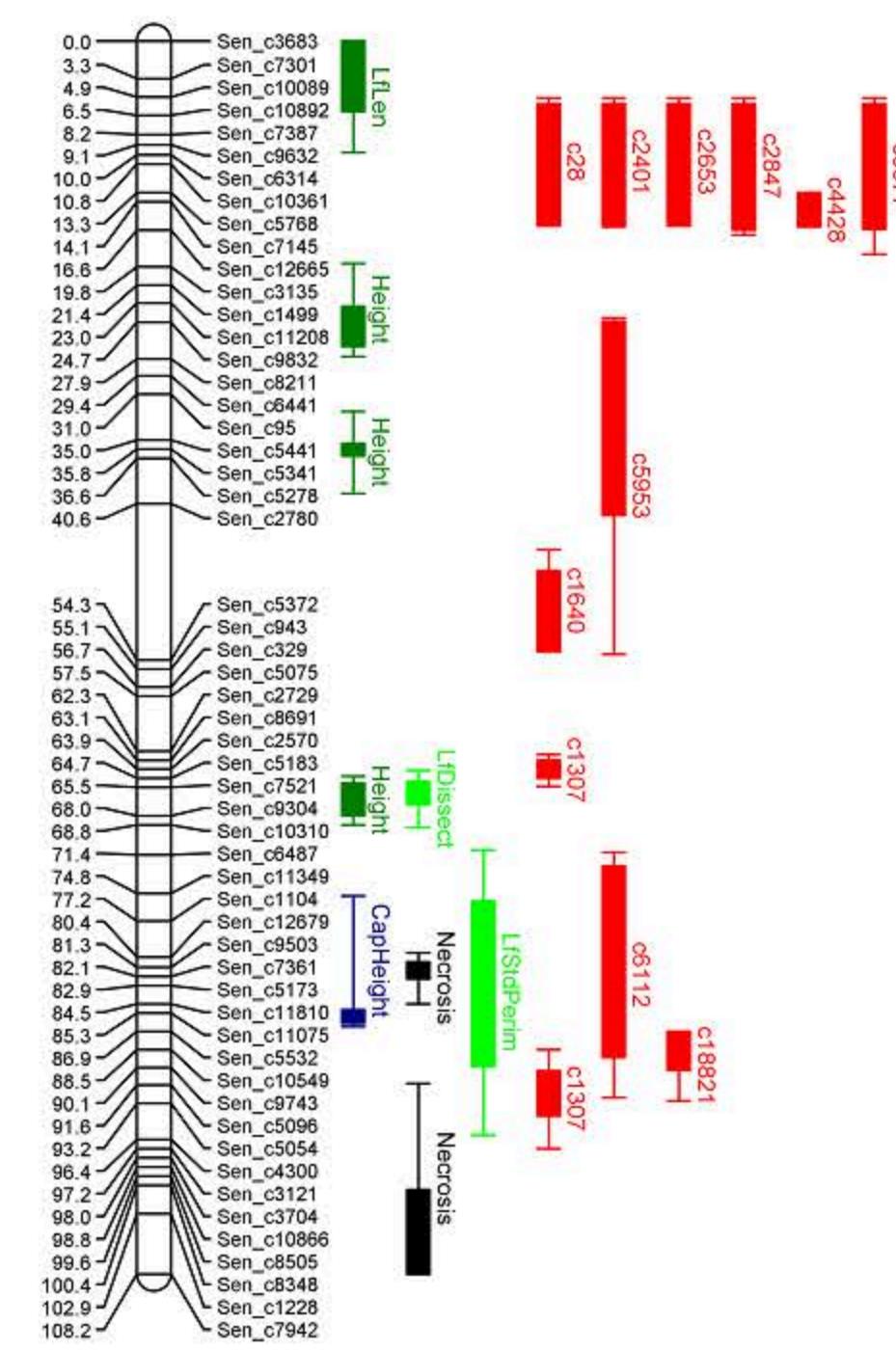
LG5



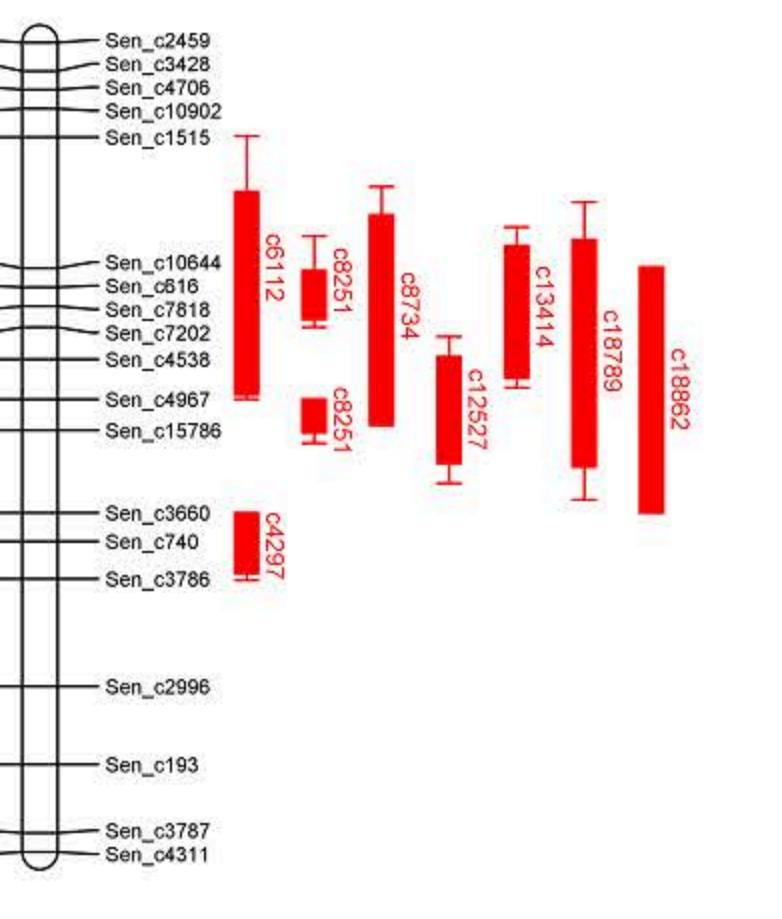
LG6



LGT



10



**Table S1 – Numbers of sequence reads produced per individual.**

Species	Individual	N reads	Species	Individual	N reads
<i>S. chrysanthemifolius</i>	R1-5	12591356	F2	F2_76	5944820
<i>S. aethnensis</i>	P2-21	25237815	F2	F2_77	5796127
F2	F2_2	5946413	F2	F2_78	6150781
F2	F2_6	5858567	F2	F2_80	6010262
F2	F2_7	5649616	F2	F2_81	5676708
F2	F2_8	5410637	F2	F2_82	6486196
F2	F2_9	5953172	F2	F2_83	6405013
F2	F2_12	6549678	F2	F2_85	5264016
F2	F2_14	5993052	F2	F2_86	5636553
F2	F2_15	6149542	F2	F2_89	6318809
F2	F2_16	5715691	F2	F2_90	5959283
F2	F2_17	5779442	F2	F2_91	5453964
F2	F2_22	5492286	F2	F2_93	6068437
F2	F2_24	6129351	F2	F2_100	6176081
F2	F2_25	6231166	F2	F2_104	6627566
F2	F2_27	6137433	F2	F2_108	5124406
F2	F2_29	5671652	F2	F2_111	5765542
F2	F2_30	5411550	F2	F2_115	6041251
F2	F2_32	5859679	F2	F2_116	5753443
F2	F2_35	5680176	F2	F2_119	5445717
F2	F2_42	5650940	F2	F2_120	5512703
F2	F2_44	5750924	F2	F2_122	5779572
F2	F2_45	5639667	F2	F2_124	6381769
F2	F2_47	6621453	F2	F2_126	6413537
F2	F2_49	6342642	F2	F2_131	5892944
F2	F2_52	5077913	F2	F2_134	6291519
F2	F2_53	6065291	F2	F2_136	6476101
F2	F2_54	5534249	F2	F2_137	5346393
F2	F2_64	5809735	F2	F2_138	6328657
F2	F2_65	5710319	F2	F2_142	6111139
F2	F2_66	5822554	F2	F2_143	7008111
F2	F2_69	6315459	F2	F2_145	5585352
F2	F2_71	5960569	F2	F2_147	6021070

**Table S2 – Primers used in the targeted genotyping and sequencing.**

A. PCR primers used to amplify and sequence loci in and near the region of extreme transmission ratio distortion on LG4.

Locus*	F primer	R primer
4_c10431	GGCTTCCTCGAGAACCTGCG	CCAATGTCTGACCCGAACTT
4_c6735	CCGGTAGTGGGAAGTTGAAA	TCCATTCAAGATGGGATGAT
4_c1856†	GGAACCTGTGGCTCCAGTAG	GGGTTCCCCATAAAGTTGATCG
4_c6172†	AAAGCAAGCAAGTGCATCT	GGTTCTGGCTTGAAGCTTG
4_c2	ACAACACCAGTCCATGCAC	TGTCACCATCAAGTTGTTCCA
4_c2075§	TGGTTGATTTACRATGCACA	CCAAACTGCAGCTGTTGTA
4_c2700§	CAATTGGGGCAGCTTGTAA	AACCTCCAWCGTGTGAAAC
4_c1331	TGGTCATATGGAAGCGGTTT	TGAAGCCAGGGAGAAGGAAT
4_c809	ATTTTACACCGCCGTTGAC	GGAAGCCATTCTGACCAAA
4_c8726	ACGCTTACCAAGCTCCATTG	ACCGGGGAAGAAATTGAAAC
4_c6599	TCAATTGTGTTAGTTGGAGTTGG	CGTGCCTCTAYATTAGCTG
4_c12731	ATAACCCGAGGGCACAAATGAG	AATGGCATTAAACGCTCGG

<sup>†</sup> locus was not homozygous in the parents but was heterozygous in the F1

§ locus was not heterozygous in the F1

#### B. PCR primers used to amplify and genotype loci showing segregation distortion

Locus*	F primer**	R primer
2_c5342	CACGACGTTGAAACGACATCGACTCGKAATCCCTT	CGCTCCATCGTACAACAAAAA
2_c6432	CACGACGTTGAAACGACGGAGATGGAGCTTGGAAATG	TCCACCTCTTGCTCCTTG
3_c4848	CACGACGTTGAAACGACCAAACACGTACACCTCACC	CCTAACCAAGCTCATGGTTT
3_c5992	CACGACGTTGAAACGACGCCAGATGCCAGCTCTTGAAGG	TGCAATACTAATACGCAGTCCA
3_c6205	CACGACGTTGAAACGACGGATCTCGAATTGATTGATT	ACCACAAACCCCTAAAACC
3_c6633	CACGACGTTGAAACGACGGATCTCGAATTGATTGATT	CGGATTATAGGCGGTAAWA
3_c7609	CACGACGTTGAAACGACTTGTCTCAATTGGCAGSA	CCCCTACTGCTACTGCTTTTC
3_c8097	CACGACGTTGAAACGACTGAAAGTTGGCAGTGGTTG	GGGGCACACTTCCACCTAAT
3_c8562	CACGACGTTGAAACGACATCCAGCCCCAAAAATAAC	TTGGTGGGTGTGAAGTGAAA
3_c13171	CACGACGTTGAAACGACGCTGTTGGGTATTTGGTGA	TGTGAAGAATTTCATGAGATAGGG
4_c809	CACGACGTTGAAACGACATTACCCGCCGTGAC	CCATACACTGAAACTGCGTTC
4_c1856	CACGACGTTGAAACGACGGAACCTGTGGCTCAGTAG	CGAACACATTAAACGAACATTAG
4_c6172	CACGACGTTGAAACGACAAAGCAAGCAAGTGCATCT	ATAGAACAGTCGAAACCGTTG
4_c6735	CACGACGTTGAAACGACCCGTAGTGGGAAGTTGAAA	TTTTGTTACGCACCAATATG
4_c8726	CACGACGTTGAAACGACACGCTTACCAAGCTCCATTG	GAAAATGAGTGTGCAGGTAAA
4_c12731	CACGACGTTGAAACGACATAACCCGAGGGCACAATGAG	CCCAAAATYAACCCATTCTATTA

length difference uncovered, so was genotyped on a subset of the F2 (see below)

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length difference uncovered, so was genotyped on a subset of the F2 (see below)

\*LG is given at the start of primer name

\*\*note the universal addition to the F primer (CACGACGTTGTAAAACGAC)

2301 loci differentially-expressed in necrotic F2 (reduced to most specific terms)

Fisher's Exact Test										
Test-Set: 2301DE_ttest.txt										
Test all Gene Ontology terms if they are enriched in a test group when compared to a reference group using Fisher's Exact Test with Multiple Testing Correction of FDR (Benjamini and Hochberg).										
GO Term	Name	Type	FDR	single test p-Value	# in test group	# in reference group	# non annot test	# non annot reference group	Over/Under	
GO:0003735	structural constituent of ribosome	F	4.20E-27	1.70E-30	90	103	1811	12607	over	
GO:0022627	cytosolic small ribosomal subunit	C	4.30E-17	4.80E-20	32	11	1869	12699	over	
GO:0005730	nucleolus	C	3.20E-16	4.00E-19	107	235	1794	12475	over	
GO:0022625	cytosolic large ribosomal subunit	C	1.80E-14	2.50E-17	32	17	1869	12693	over	
GO:0001510	RNA methylation	P	2.30E-07	7.20E-10	40	72	1861	12638	over	
GO:0005886	plasma membrane	C	9.50E-06	3.60E-08	440	2268	1461	10442	over	
GO:0010103	stomatal complex morphogenesis	P	1.00E-05	4.10E-08	44	102	1857	12608	over	
GO:0010075	regulation of meristem growth	P	2.90E-05	1.40E-07	49	128	1852	12582	over	
GO:0009664	plant-type cell wall organization	P	2.30E-04	1.20E-06	48	136	1853	12574	over	
GO:0006412	translation	P	2.80E-04	1.60E-06	129	538	1772	12172	over	
GO:0009506	plasmodesma	C	4.20E-04	2.60E-06	131	555	1770	12155	over	
GO:0043481	anthocyanin accumulation in tissues in response to UV light	P	1.20E-03	9.00E-06	27	60	1874	12650	over	
GO:0009098	flower development	P	1.50E-03	1.20E-05	143	641	1758	12069	over	
GO:0007169	transmembrane receptor protein tyrosine kinase signaling pathway	P	3.80E-03	3.40E-05	28	70	1873	12640	over	
GO:0042254	ribosome biogenesis	P	5.70E-03	5.80E-05	55	196	1846	12514	over	
GO:0008356	asymmetric cell division	P	9.00E-03	9.50E-05	9	8	1892	12702	over	
GO:0005576	extracellular region	C	9.00E-03	9.50E-05	164	794	1737	11916	over	
GO:0009965	leaf morphogenesis	P	1.00E-02	1.10E-04	39	125	1862	12585	over	
GO:0019199	transmembrane receptor protein kinase activity	F	1.70E-02	2.00E-04	12	18	1889	12692	over	
GO:0009555	adaxial/abaxial pattern specification	P	1.80E-02	2.20E-04	19	43	1882	12667	over	
GO:0009505	plant-type cell wall	C	2.30E-02	2.90E-04	47	171	1854	12539	over	
GO:0048767	root hair elongation	P	2.80E-02	3.60E-04	35	115	1866	12595	over	
GO:0009825	multidimensional cell growth	P	2.80E-02	3.60E-04	25	70	1876	12640	over	
GO:0010817	regulation of hormone levels	P	2.80E-02	3.70E-04	68	281	1833	12429	over	
GO:0019843	rRNA binding	F	3.00E-02	4.00E-04	16	34	1885	12676	over	
GO:0016878	acid-thiol ligase activity	F	3.10E-02	4.10E-04	12	20	1889	12690	over	
GO:0032440	2-alkenal reductase [NAD(P)] activity	F	3.30E-02	4.50E-04	47	175	1854	12535	over	
GO:0008236	serine-type peptidase activity	F	3.90E-02	5.60E-04	30	95	1871	12615	over	
GO:0006805	xenobiotic metabolic process	P	4.20E-02	6.20E-04	5	2	1896	12708	over	
GO:0006655	phosphatidylglycerol biosynthetic process	P	4.40E-02	6.70E-04	18	44	1883	12666	over	

592 loci over-expressed in necrotic F2 (reduced to most specific terms)

Fisher's Exact Test										
Test-Set: 592OEinnecrotic_ttest.txt										
GO Term	Name	Type	FDR	single test p-Value	# in test group	# in reference group	# non annot test	# non annot reference group	Over/Under	
GO:0006623	protein targeting to vacuole	P	4.00E-05	1.70E-08	18	94	451	14048	over	
GO:0005777	peroxisome	C	1.50E-04	1.20E-07	23	177	446	13965	over	
GO:0016192	vesicle-mediated transport	P	8.00E-03	7.90E-06	36	473	433	13669	over	
GO:0016773	phosphotransferase activity, alcohol group as acceptor	F	1.10E-02	1.20E-05	62	1048	407	13094	over	
GO:0008756	o-succinylbenzoate-CoA ligase activity	F	4.40E-02	6.60E-05	4	4	465	14138	over	
GO:0016301	kinase activity	F	4.40E-02	6.70E-05	67	1233	402	12909	over	
GO:0005886	plasma membrane	C	4.50E-02	7.60E-05	120	2588	349	11554	over	

1709 loci under-expressed in necrotic F2 (reduced to most specific terms)

Fisher's Exact Test										
Test-Set: 1709UEinnecrotic_ttest.txt										
Test all Gene Ontology terms if they are enriched in a test group when compared to a reference group using Fisher's Exact Test with Multiple Testing Correction of FDR (Benjamini and Hochberg).										
GO Term	Name	Type	FDR	single test p-value	# in test group	# in reference group	# non annot test	# non annot reference group	Over/Under	
GO:0003735	structural constituent of ribosome	F	2.90E-34	1.60E-37	87	106	1345	13073	over	
GO:0005730	nucleolus	C	3.60E-24	4.00E-27	104	238	1328	12941	over	
GO:0022627	cytosolic small ribosomal subunit	C	4.90E-21	7.50E-24	32	11	1400	13168	over	
GO:0022625	cytosolic large ribosomal subunit	C	2.60E-18	4.70E-21	32	17	1400	13162	over	
GO:0001510	RNA methylation	P	3.40E-11	9.40E-14	40	72	1392	13107	over	
GO:0010103	stomatal complex morphogenesis	P	5.60E-09	2.00E-11	43	103	1389	13076	over	
GO:0010075	regulation of meristem growth	P	2.80E-07	1.40E-09	45	132	1387	13047	over	
GO:0009664	plant-type cell wall organization	P	3.10E-07	1.60E-09	46	138	1386	13041	over	
GO:0009506	plasmodesma	C	1.40E-05	1.10E-07	110	576	1322	12603	over	
GO:0043481	anthocyanin accumulation in tissues in response to UV light	P	1.50E-05	1.30E-07	26	61	1406	13118	over	
GO:0006412	translation	P	5.40E-05	5.40E-07	105	562	1327	12617	over	
GO:0007169	transmembrane receptor protein tyrosine kinase signaling pathway	P	5.10E-04	5.80E-06	25	73	1407	13106	over	
GO:0009825	multidimensional cell growth	P	9.00E-04	1.10E-05	24	71	1408	13108	over	
GO:0019843	rRNA binding	F	1.10E-03	1.30E-05	16	34	1416	13145	over	
GO:0009505	plant-type cell wall	C	1.20E-03	1.40E-05	42	176	1390	13003	over	
GO:0032440	2-alkenal reductase [NAD(P)] activity	F	1.70E-03	2.30E-05	42	180	1390	12999	over	
GO:0009965	leaf morphogenesis	P	3.40E-03	4.70E-05	33	131	1399	13048	over	
GO:0006655	phosphatidylglycerol biosynthetic process	P	4.60E-03	6.60E-05	17	45	1415	13134	over	
GO:008356	asymmetric cell division	P	5.80E-03	9.00E-05	8	9	1424	13170	over	
GO:0048767	root hair elongation	P	7.10E-03	1.10E-04	30	120	1402	13059	over	
GO:0048437	floral organ development	P	8.30E-03	1.40E-04	69	384	1363	12795	over	
GO:0005576	extracellular region	C	8.70E-03	1.40E-04	128	830	1304	12349	over	
GO:0007020	microtubule nucleation	P	1.00E-02	1.70E-04	19	60	1413	13119	over	
GO:0009220	pyrimidine ribonucleotide biosynthetic process	P	1.20E-02	2.20E-04	26	101	1406	13078	over	
GO:0006913	nucleocytoplasmic transport	P	1.30E-02	2.30E-04	32	138	1400	13041	over	
GO:0009955	adaxial/abaxial pattern specification	P	1.30E-02	2.30E-04	16	46	1416	13133	over	
GO:0010817	regulation of hormone levels	P	1.40E-02	2.70E-04	55	294	1377	12885	over	
GO:0042545	cell wall modification	P	1.60E-02	3.10E-04	29	122	1403	13057	over	
GO:0048439	flower morphogenesis	P	1.80E-02	3.40E-04	16	48	1416	13131	over	
GO:0009933	meristem structural organization	P	2.00E-02	3.90E-04	42	209	1390	12970	over	
GO:0060771	phyllotactic patterning	P	2.10E-02	4.20E-04	4	1	1428	13178	over	
GO:0000462	maturation of SSU-rRNA from tricistronic rRNA transcript (SSU-rRNA, 5.8S rRNA, LSU-rRNA)	P	2.10E-02	4.20E-04	4	1	1428	13178	over	
GO:0048438	floral whorl development	P	2.60E-02	5.20E-04	58	325	1374	12854	over	
GO:0045036	protein targeting to chloroplast	P	2.90E-02	6.00E-04	16	51	1416	13128	over	
GO:0016441	posttranscriptional gene silencing	P	3.40E-02	7.20E-04	38	189	1394	12990	over	
GO:0009855	determination of bilateral symmetry	P	3.60E-02	7.80E-04	22	87	1410	13092	over	
GO:0047668	amygdalin beta-glucosidase activity	F	4.10E-02	9.40E-04	3	0	1429	13179	over	
GO:0008460	dTDP-glucose 4,6-dehydratase activity	F	4.10E-02	9.40E-04	3	0	1429	13179	over	
GO:0031957	very long-chain fatty acid-CoA ligase activity	F	4.10E-02	9.40E-04	3	0	1429	13179	over	
GO:0080082	esculin beta-glucosidase activity	F	4.10E-02	9.40E-04	3	0	1429	13179	over	
GO:0080081	4-methylumbelliferyl-beta-D-glucopyranoside beta-glucosidase activity	F	4.10E-02	9.40E-04	3	0	1429	13179	over	
GO:0010159	specification of organ position	P	4.10E-02	9.40E-04	3	0	1429	13179	over	
GO:0009240	isopentenyl diphosphate biosynthetic process	P	4.40E-02	1.00E-03	35	173	1397	13006	over	
GO:0045338	farnesyl diphosphate metabolic process	P	4.80E-02	1.20E-03	4	2	1428	13177	over	
GO:0048830	adventitious root development	P	4.80E-02	1.20E-03	4	2	1428	13177	over	
GO:0046658	anchored to plasma membrane	C	4.90E-02	1.20E-03	13	39	1419	13140	over	