

Immune biomarkers in the spectrum of childhood non-communicable diseases

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Abstract

A biomarker is an accurately and reproducibly quantifiable biological characteristic that provides an objective measure of health status or disease. Benefits of biomarkers include identification of therapeutic targets, monitoring of clinical interventions, and development of personalized (or precision) medicine. Challenges to the use of biomarkers include optimizing sample collection, processing and storage, validation, and often the need for sophisticated laboratory and bioinformatics approaches. Biomarkers offer better understanding of disease processes and should benefit the early detection, treatment and management of multiple non-communicable diseases (NCDs). This review will consider the utility of biomarkers in allergic and other immune mediated diseases in childhood. Typically, biomarkers are used currently to provide mechanistic insight or an objective measure of disease severity with their future role in risk stratification/disease prediction speculative at best. There are many lessons to be learned from the biomarker strategies used for cancer where biomarkers are in routine clinical use and industry wide standardized approaches have been developed. Biomarker discovery and validation in childhood disease lags behind that for adults; given the early onset and therefore potential lifelong impact of many NCDs there should be more studies incorporating cohorts of children. Many pediatric biomarkers are at the discovery stage with a long path to evaluation and clinical implementation. The ultimate challenge will be optimisation of prevention strategies that can be implemented in children identified as being at risk of an NCD through the use of biomarkers.

Keywords: biomarkers; non-communicable diseases; children; inflammation; microRNA; blood; urine; breath; mass spectrometry

Abbreviations:

ANCA	anti-neutrophil cytoplasmic antibodies
Anti-OmpC	antibody to outer membrane porin of <i>E.coli</i>
ASCA	Anti- <i>Saccharomyces cerevisiae</i> antibodies
ASD	autism spectrum disorder
BMI	body mass index
CD	Crohn's disease
CRP	C reactive protein
DHEAS	dehydroepiandrosterone sulfate
EBC	exhaled breath condensate
EMA	European Medicines Agency
EMP	endothelial microparticle
EPC	endothelial progenitor cell
FDA	Food and Drug Administration
FeNO	fractional exhaled nitric oxide
IBD	inflammatory bowel disease
ICS	inhaled corticosteroid
miRNA	microRNA
MS	mass spectrometry
mTOR	mechanistic target of rapamycin
NCD	non-communicable disease
PBMC	peripheral blood mononuclear cells
PMDA	Japanese Pharmaceutical and Medical Devices Agency
qRT-PCR	quantitative real time polymerase chain reaction
UC	ulcerative colitis

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Introduction

Non-communicable diseases (NCDs) are one of the major global challenges of the 21st century¹. NCDs have been termed a “slow-motion disaster”² and a global crisis³ as their prevalence increases in all countries, in all income groups, and at all ages. NCDs, often called chronic diseases, are generally considered as being one of four main types: cardiovascular diseases, cancers, chronic respiratory diseases, and diabetes. Allergic disease has been suggested as a fifth group given its high prevalence and early onset⁴, but neurocognitive, inflammatory bowel and other diseases are also important NCDs. Worldwide, two out of every three deaths each year are attributable to NCDs with one third of those who die being under the age of 60 years⁵. In all regions of the world the prevalence of NCDs is rising due to an aging population and the globalization of common risk factors⁵. The main risk factors for NCDs, accounting for two-thirds of all cases, are: tobacco use; food high in saturated and trans fats, salt and sugar; physical inactivity; and alcohol consumption⁶. NCDs not only directly threaten health, they influence economic development to compound the impact on health. The World Economic Forum ranks NCDs as one of the top global threats to economic development: a 10% increase of mortality by NCDs reduces annual economic growth by 0.5%⁷.

A shared feature of all NCDs is chronic low-grade inflammation promoted by modern diets, environmental pollutants, microbial exposure, and psychological and biological (e.g. oxidative or endoplasmic reticulum) stress^{1, 8}. This low-grade inflammation differs from classical inflammation, which occurs in response to a threat or injury and leads to tissue repair and restoration of the basal homeostatic state. The low-grade inflammation in NCDs is chronic; without effective treatment basal homeostasis cannot be restored and the damage may already be done⁹. Inflammatory pathways in NCDs are multi-factorial and part of a metabolic cascade, including cellular oxidative stress and insulin resistance, which induces allostatic overload, dysmetabolism, and ultimately chronic disease¹⁰. Changes in the gut microbiome have emerged as one of the pathways leading to chronic low-grade inflammation¹¹. Altered gut colonization and reduced microbiome diversity occur in response to both changed nutritional patterns and the built environment; a diverse microbiome is essential for normal immune development and regulation¹². There is global interest in gut dysbiosis in multiple NCD settings including for example beta-cell autoimmune disease¹³, atopic dermatitis¹⁴, and inflammatory bowel disease¹⁵. Despite the clear link between the gut microbiome and development of these diseases and obesity,

cardiovascular disease and metabolic disorders more generally, the causal relationship between alterations in the gut microbiome and ill health is likely to be complex. There may be multiple different pathways for different organisms; these pathways may or may not overlap in some of their stages. However, specific microbe-derived metabolites such as short chain fatty acids have emerged as examples whereby cross talk between the microbiome and host is achieved.

While NCDs are most prominent in adulthood, development in early life influences predisposition to NCDs. This starts as early as pregnancy, when maternal body composition and diet influences the infant's risk of NCDs later in life⁴. NCDs should therefore be studied using a life-course approach with overall risk depending on: the sequential effects of the developmental time-line with different metabolic trajectories, age-dependent decline in plasticity, and the subsequent differential responses to subsequent risk factors³⁰. Monitoring of these time-lines and trajectories would not only benefit the understanding of disease processes but enable risk stratification for disease intervention and even prevention. Biomarkers likely provide the necessary tool for this approach to be successful.

What are biomarkers?

A biomarker (biological marker) is a quantifiable biological characteristic that provides an objective measure of health status or disease. For disease, biomarkers have the potential for use in risk stratification, early detection, identifying treatment of choice and monitoring response to treatment, in surveillance, and in drug monitoring and development. Biomarkers are also used in other clinical scenarios such as microbial identification and diagnostics. A biomarker can be a gene, molecule, or other biological characteristic as long as it can be measured accurately and reproducibly and is a valid indicator of the process or outcome being "marked". More detailed discussion of the various but overlapping definitions of biomarkers that come from the National Institutes of Health Biomarkers Definitions Working Group and the World Health Organization, among others, can be found elsewhere³¹.

Traditionally, biomarkers have been used as indicators of a specific disease state and range from cardiac troponins for myocardial infarction, to genetic markers of cancer risk, to stool calprotectin for inflammatory bowel disease. Many of these biomarkers were first identified during targeted studies, typically of isolated molecules within *in vitro* cellular systems and/or

case control comparison of clinical specimens. The 'omics' era has led to unbiased discovery-based approaches to identify biomarkers, including panels (also called clusters or signatures) of biomarkers. Omics approaches, particularly proteomics, metabolomics and cellomics, but less so transcriptomics and epigenomics, are used in two broad ways: to better understand biological processes and for clinical biomarker discovery. While the latter is not intended to provide insight into disease processes, biological plausibility can help progress validation and implementation of any discovered biomarkers.

Biomarkers can also be used as surrogate or intermediate end points in translational and clinical research where the challenge is to know what the relationship is between any measure and the relevant clinical endpoint³¹. The use of biomarkers in this way might be most useful during clinical trials and for lifestyle, such as nutritional, interventions³². This is critical for those who work in the disease prevention arena so that the success, or otherwise, of an intervention can be considered before a clinical endpoint is achieved as this may take many years.

Among the omics approaches for biomarker discovery, proteomics is currently the most widely used. The proteome reflects splice variants and post-translational modifications of proteins that dictate structure, function, localization, maturation and turnover of proteins, all of which change rapidly in response to environmental signals³³. Sample types studied include serum, plasma, saliva, urine, exhaled breath, tissue biopsies, mucosal secretions, cerebrospinal fluid, and other biological specimens. The plasma proteome is of particular interest; not only is blood a routinely collected clinical sample but plasma is postulated to reflect the sum of multiple site-specific proteomes containing representatives of the entire set of more than 300,000 estimated human polypeptide species resulting from splice variants and post-translational modifications³⁴.

An emergent biomarker family measurable in the circulation is microRNA (miRNA). MicroRNAs are small endogenous non-coding RNAs with a critical post-transcriptional gene-regulatory role³⁵. Aberrant miRNA expression is implicated in disease, and disease and/or tissue specific miRNAs have been identified and these are surprisingly stable and can be relatively easily measured. Most progress in the use of miRNAs as biomarkers is in cancer where their utility has been considered for monitoring disease progression, outcomes, recurrence and metastasis³⁶.

Within paediatrics, biomarker discovery remains dominated by targeted approaches. Here we will review what is known about paediatric biomarkers for NCDs and identify emergent areas of study highlighting traditionally identified biomarkers as well as those revealed with untargeted approaches. Given the common genetic factors that contribute to inflammatory diseases as recently shown, for example, with paediatric autoimmune diseases³⁷, the challenge is to identify downstream proteins, metabolites, miRNAs, and other molecules that offer insight in a disease-specific manner. The microbiome might also serve as a biomarker as deep sequencing and transcriptomics approaches become more readily usable.

Biomarkers in non-communicable diseases in childhood

Allergy

Allergic disorders, including atopic dermatitis, food allergy, allergic rhinitis and asthma affect over one billion people across the globe and their prevalence is expected to quadruple by the 2050s³⁸. Atopic dermatitis, typically accompanied by IgE sensitization to food, and food allergy are the earliest onset NCDs and the most common chronic NCDs of childhood worldwide^{39, 40}. They are considered the first steps on the allergic march whereby atopic dermatitis and food allergy in infancy progress to asthma and allergic rhinitis in later childhood and into adult life^{41, 42}. As for many other NCDs, the development and deployment of biomarkers for monitoring disease severity, classifying and clarifying disease subsets, and guiding treatment decisions is an active area of translational research. Given the early age of onset and the prospect of life long disease there is also incredible value in predictive biomarkers usable around birth to identify children at risk of developing these diseases. Failing that, identifying those children with atopic dermatitis and/or food allergy who will progress to asthma and/or rhinitis would be of immense use.

Atopic dermatitis and food allergy. Much of the biomarker work within atopic dermatitis relates to monitoring disease severity and clinical improvement. Many investigators report an association between disease activity and circulating factors and then speculate that these could act as disease markers or new therapeutic targets. These circulating factors include: soluble adhesion molecules such as ICAM-1⁴³, the anti-bacterial peptide LL-37⁴⁴, interleukins such as interleukin (IL)-31⁴⁵ and IL-18⁴⁶, and numerous chemokines such as mucosa-associated epithelial chemokine (MEC/CCL28)⁴⁷, thymus and activation-regulated chemokine (TARC/CCL17)⁴⁸, and cutaneous T cell attracting chemokine (CTACK/CCL27)⁴⁹. However,

there are few evaluation studies to progress these molecules to clinical use. Candidates are usually analysed because of their role in disease pathogenesis so are common to multiple allergic and/or inflammatory disorders. While potentially suitable for disease monitoring⁵⁰, they are less likely to offer the specificity required for predictive biomarkers. Detailed analysis of CCL17 also underlines one of the real challenges of biomarker discovery in children. CCL17 circulating levels correlate inversely with age so are much higher at 0 – 1 year than in older age groups⁴⁸ highlighting the natural age-dependent variation in the abundance of proteins and other classes of molecules. This is a particular challenge for diagnostic and prognostic biomarkers but for predictive biomarkers implemented as early in life as possible this is unlikely to be a problem.

The utility of miRNAs for atopic dermatitis prognosis in children also has been explored⁵¹. Notably, both co-morbidities and sample type analysed affects the findings. Serum levels of miR-483-5p were increased in children with atopic dermatitis and were associated with other atopic conditions such as asthma or hayfever. For miR-203, urine levels were decreased but serum levels were increased in the children with atopic dermatitis⁵¹. There is also interest in plasma miRNA biomarkers in eosinophilic esophagitis (EoE) with several differentially expressed in paediatric EoE patients⁵². Some of these miRNAs mapped with glucocorticoid treatment and normalisation of eosinophil histology and cell counts so could offer a less invasive approach to monitor treatment efficacy⁵². There is much interest in miRNAs for better understanding disease pathogenesis, for informing treatment decisions, and as biomarkers. Most studies to date have focussed on profiling specific miRNAs in various allergic disease settings where they have been found to have a role in T cell, eosinophil and epithelial function⁵³. Hopefully we will see progress to using these as biomarkers in multiple disease settings.

Studies using omics approaches for atopic dermatitis are only just being reported. Examples include: a mass spectrometry study of serum to reveal changes in energy metabolism measurable as increased carnitine, free fatty acids and lactic acid, amongst others, in atopic dermatitis⁵⁴; and changes in the glycoproteome (glycosylation patterns are influenced by physiological status⁵⁵) by two-dimensional electrophoresis showing increased CD5L and decreased Apolipoprotein E with atopic dermatitis⁵⁶. Many of these are pilot discovery studies providing new mechanistic insight, such as a role for CD5L in protecting eosinophils from death by apoptosis⁵⁶, but in the longer term offer a potential source of biomarkers.

For food allergy in particular, biomarkers for predicting good outcomes to oral immunotherapy (OIT) would be highly beneficial and to date the focus has been on immunoglobulin and cytokine responses. It is well known that allergen specific IgE and IgG4 are modified by OIT but IgG1, IgG3 and IgA also change, with poor responders showing the smallest changes in these immunoglobulins⁵⁷. High allergen-specific IgA at outset of OIT or high allergen-specific IgG1 after rush phase might be useful biomarkers to predict positive immunological and clinical response to OIT⁵⁷. Individualising egg OIT based on an IgE-dependent dosing strategy and increasing maintenance dose individually based on egg white IgE level has been reported to improve clinical egg tolerance⁵⁸.

Predictive biomarkers for atopic dermatitis and food allergy are a highly desirable adjunct to allergic disease prevention strategies. There is a burgeoning literature focussed on analysis of samples collected at birth or in early infancy but much of this work is candidate driven with omics discovery approaches only just emerging. Raised fecal calprotectin at 2 months of age has been associated with increased risk of atopic dermatitis and asthma/asthmatic bronchitis by 6 years of age⁵⁹. This highlights the link between the microbiome, intestinal inflammation and allergic disease and the potential utility of monitoring gut health for predictive biomarkers in multiple disease settings. Similarly, nutritional status could be monitored through biomarkers⁶⁰ as could environmental exposures that effect disease manifestation⁶¹. Candidate approaches have also been undertaken using umbilical cord blood. Circulating factors studied in this context include soluble Fas ligand (sFasL) which was elevated in umbilical cord blood of newborns who developed atopic dermatitis⁶². The biological relevance of sFasL might relate to keratinocyte apoptosis, and subjective severity of atopic dermatitis in infancy correlated positively with sFasL levels at birth⁶². While neonatal levels of sFasL were higher than maternal levels, a positive relationship between maternal and umbilical cord blood levels indicates the value of analysing samples from mother and child for discovery of circulating biomarkers around the time of birth. Levels of the circulating allergy-related chemokines TARC/CCL17 and MDC/CCL22 at birth have also been associated with development of atopic dermatitis up to 3 years of age⁶³ and elevated total IgE over the first 6 years of life⁶⁴, respectively. Given the latter did not relate to specific IgE or clinical allergic disease it is unlikely a suitable standalone marker but might serve well as part of panel. Some other common observations that warrant further evaluation have emerged. The most notable of these relates to regulatory T cells where reduced numbers,

function, and/or FoxP3 expression at birth relate to atopic dermatitis and/or food allergy in early childhood⁶⁵⁻⁶⁷. However, a consensus methodological approach is required if the evaluation of this or any other cell type is to have any value as a biomarker. Genetic biomarkers might also have predictive utility. As an example, a filaggrin loss-of-function mutation associated with skin barrier dysfunction was linked to food allergy in older children especially following early evidence of food allergen sensitization and eczema⁶⁸.

The prospect of omics approaches applied at or near birth is particularly promising. Vernix is usually abundant on the skin of the newborn and can be collected non-invasively. The protein composition of vernix reflects skin functional responses with various families of proteins found in vernix including cytoskeletal proteins, cell adhesion molecules, cell junction proteins and transcription factors⁶⁹. A liquid chromatography tandem mass spectrometry analysis of vernix revealed a strong negative correlation between levels of polyubiquitin-C and calmodulin-like protein 5 and the development of atopic dermatitis at 2 years of age. These were suggested as candidate biomarkers for identifying newborns predisposed to development of atopic dermatitis⁶⁹.

Methodological and data analysis approaches pose the greatest immediate challenges to the clinical use of predictive biomarkers for atopic dermatitis and food allergy. However, the ultimate challenge is optimisation of prevention strategies that can be implemented in children identified as being at risk through use of biomarkers.

Allergic rhinitis. There is little biomarker work within pediatric allergic rhinitis. Recently, miR-146a expression in nasal mucosa and peripheral blood mononuclear cells (PBMCs) has been identified as a promising biomarker for pathogenesis and management of allergic rhinitis in children, including during sublingual immunotherapy (SLIT)⁷⁰. miR-146a was decreased in PBMCs and nasal mucosa of children with allergic rhinitis and inversely correlated with disease severity. This miRNA is highly expressed in regulatory T cells⁷¹ and FoxP3 mRNA in PBMCs from children with atopic dermatitis was also significantly decreased. Both miR-146a and FoxP3, as well as serum IL-10, levels increased with SLIT.

Asthma. Asthma, a chronic heterogeneous airway disease characterized by local inflammation and varying degrees of tissue remodeling, affects around 300 million people of all ages with increasing incidence⁷². It is the most common chronic disease of childhood.

Along with other allergic disorders, asthma begins in conjunction with IgE sensitization to allergens which most often occurs early in life. Recent years have seen much attention given to diagnosis and early intervention with alteration of the natural history of the disease and elimination of acute asthma exacerbations key goals.

Sub-phenotypes and associated pathogenetic mechanisms or endotypes of asthma are increasingly utilized to classify disease and better direct therapy. Single and combination biomarkers serve to identify these clusters with the presence of eosinophilic Th2 type inflammation perhaps the most important feature. Distinctions based on this parameter guide inhaled corticosteroid treatment (ICS) and biologic therapies⁷³⁻⁷⁵. Endobronchial tissue gene expression profiling revealed that asthmatic patients can be grouped as Th2-high, Th17-high and Th2/Th17-low and the clinical value of this should become apparent from follow on studies⁷⁶.

IgE and eosinophils have long standing roles as biomarkers of allergic disease and other single biomarkers that have emerged of value include nitric oxide and periostin. Serum levels of allergen-specific IgE guide identification of disease triggers and inform allergen avoidance strategies. Sputum eosinophil counts and ratios between peripheral blood eosinophils and neutrophils might cluster patients based on steroid responsiveness and are useful for identifying patients suitable for anti-IL-4, anti-IL-5 and anti-IL-13 therapy⁷⁷⁻⁸⁰. Nitric oxide is produced by the bronchial epithelium through the action of inducible nitric oxide synthase in response to IL-4 and IL-13⁸¹. It can be measured as fractional exhaled nitric oxide (FeNO) which is a biomarker for eosinophilic airway inflammation, is highly corticosteroid sensitive, and is relatively easily measured using point-of-care diagnostics⁸¹⁻⁸⁶. Periostin is an extracellular matrix protein secreted by airway epithelial cells and lung fibroblasts that is also induced by IL-4 and IL-13⁸⁷. It is elevated in air-liquid interface cultured nasal and bronchial epithelial cells isolated from asthmatic children⁸⁸. As a ligand for $\alpha_v\beta_3$ and $\alpha_v\beta_5$ integrins it supports adhesion and migration of epithelial cells and contributes to tissue remodeling⁸⁹. As a biomarker of type 2 inflammation and remodeling in asthma, serum levels of periostin, especially in combination with FeNO and blood eosinophil numbers, might predict the efficacy of anti-IgE antibody treatment⁹⁰. Serum periostin levels are also higher in asthmatic children⁹¹ and correlate with the response to inhaled corticosteroids, at least in adults⁹².

Serum miRNA 21 has been suggested as a novel biomarker for allergic inflammatory diseases as it was elevated among children with asthma or eosinophilic esophagitis⁹³. A profile of 10 miRNAs, including miR-638, has been associated with asthma in children aged 6 – 18 years and could distinguish those with severe asthma and with more severe acute exacerbations⁹⁴. More work is required to evaluate the disease-specific utility of these miRNAs⁶⁸.

There is an ever-growing demand for non-invasive techniques and biomarkers associated with disease control as well as prediction, identification and management of acute asthma exacerbations. Nasal lavage, serum, saliva, exhaled breath condensate, induced sputum and urine are some of the biological samples evaluated. Quite promising biomarker discovery approaches of exhaled breath include a range of mass spectrometry (MS) approaches including gas chromatography-MS (GC-MS), selected ion flow tube-MS (SIFT-MS), and ion mobility-mass spectrometry (IMS) which allow detection of volatile organic compounds and the study of the metabolome^{95,96}. Such techniques will also help reveal the underlying pathophysiology of acute asthma exacerbations.

There is little, if any, progress in biomarker discovery for predicting the development of asthma and other allergies. Early-life production of LPS-stimulated tumor necrosis factor- α (TNF α) but not IL-6, IL-10 or IL-12 by PBMCs from infants was associated with a higher risk for developing asthma⁹⁷. Combined exhaled volatile organic compounds, PBMC inflammatory gene expression (TLR4, catalase and TNF α), and clinical information (asthma predictive index) in preschool age children could predict asthma at 6 years of age⁹⁸. Similarly, inflammatory markers (IL-2, IL-4, IL-8 and IL-10) in exhaled breath condensate at 2 – 3 years of age were elevated among children who were persistent wheezers up to 5 years of age compared to children who had never wheezed^{99,100}. Further study is required to evaluate the efficacy of these as clinically useful biomarkers.

Autoimmunity

Autoantibodies are longstanding diagnostic biomarkers of organ-specific and non-specific autoimmune disorders and are part of diagnostic and classification criteria. With the ongoing discovery of novel autoantibodies, it has become evident that profiles of these biomarkers rather than single entities better distinguish disease phenotypes. The utility of autoantibody detection is not restricted to diagnosis but extends to prediction, prognosis and prevention of autoimmunity. This plethora of autoantibodies and the numerous

detection and analysis technologies now available dictate a need for standardization and harmonization of autoantibody nomenclature and testing. Requesting the appropriate test based on clinical features remains essential for optimal use of available biomarkers and needs close communication between the clinician and the laboratory specialist. Autoantibodies relevant to common autoimmune diseases in childhood are listed in Table 1.

Metabolic profiling of children with type 1 and type 2 diabetes has attracted interest as a means of investigating disease pathogenesis and identifying predictive biomarkers¹⁰¹. These blood based approaches, including from the umbilical cord, are all at the discovery stage. Progressing these to clinical implementation is an enormous challenge. For type 1 diabetes a large prospective study of children recruited before 4.5 months of age is underway (The Environmental Determinants of Diabetes in the Young; TEDDY¹⁰²). This involves nearly 9000 high risk children and will monitor development of islet autoimmunity and progression to type 1 diabetes to identify biomarkers of these. The metabolomic analysis of the samples collected is very carefully optimized¹⁰³. This study highlights that very large prospective studies using multicenter and systems biology approaches are likely required to identify disease pathways that can be targeted for disease prevention.

Interest in the use of biomarkers for early diagnosis and therapeutic intervention in rheumatoid arthritis has led to a number of studies investigating miRNA profiles in the joint and in peripheral blood. This has revealed shared profiles of overexpressed miRNAs but it is the signature measurable in blood including miR-24, miR-125a-5p, and miR-451, amongst others, that holds the most promise as biomarkers¹⁰⁴⁻¹⁰⁹. Recent work provided evidence for the use of 14-3-3n serum antibodies in RA^{110, 111}, and a diagnostic blood panel (including ANA, anti-Ro, anti-La, rheumatoid factor, and novel SjS autoantibodies such as anti-salivary gland protein 1 (SP1), anti-carbonic anhydrase 6 (CA6) and anti-parotid secretory protein (PSP)) for Sjögren syndrome¹¹²⁻¹¹⁴. Levels of DKK1, leptin, osteoprotegerin, osteopontin, and sclerostin are related to bone damage in psoriatic arthritis¹¹⁵. Serum calprotectin has been proposed as a biomarker for ANCA-associated vasculitis and more specifically for indicating relapsers among patients treated with rituximab^{116, 117}. Recently, activation of mechanistic target of rapamycin (mTOR), a ubiquitous serine/threonine kinase with a role in numerous cell processes including proliferation and survival, autophagy, and transcription, has been highlighted as a central pathway for the pathogenesis of systemic lupus erythematosus and

other autoimmune diseases. Blockade of mTOR and follow-up of this biomarker holds promise in personalized treatment of patients suffering from these diseases among others¹¹⁸. It is hoped that some of these biomarkers will improve diagnostic accuracy but most remain at the discovery phase. These studies are almost exclusively in adults and similar analyses should be pursued in pediatric populations.

Inflammatory bowel diseases

Inflammatory bowel diseases (IBDs) are chronic inflammatory disorders of the gastrointestinal tract; chief among these are ulcerative colitis (UC) and Crohn's disease (CD). IBDs arise from a complex interaction between genetics, environment, the gut microbiota and the mucosal immune response. Occurrence rates of CD and UC are generally reported to have increased for both adults and children¹¹⁹. The diagnosis of IBDs entails a multistep strategy using clinical, endoscopic and imaging modalities but there has been a longstanding interest in the development of biomarkers to better facilitate not only diagnosis but for monitoring treatment efficacy, disease severity, and the risk of complications. Reliable surrogate markers to avoid repeated endoscopy and to accurately report mucosal inflammation activity are particularly desirable. Circulating inflammatory markers such as CRP lack specificity for IBDs as they are also used in other clinical scenarios in which inflammation features such as autoimmunity and infection. The neutrophil-derived faecal inflammatory markers calprotectin and lactoferrin are measured as indicators of the local inflammatory burden within the gastrointestinal tract and are already in clinical use. These biomarkers are useful for delineating IBD from irritable bowel syndrome, for the management and monitoring of patients, and prediction of relapse but their utility in monitoring mucosal healing requires refinement and there is an agreed need for biomarker discovery strategies amongst gastroenterologists^{120, 121}.

Candidate biomarker approaches have been explored, for example, to show that measurement of a panel of antibodies to neutrophils, *Saccharomyces cerevisiae*, and *E. coli* outer membrane porin correlates with disease activity and the location and outcome of disease in children/young adults aged 1 – 21 years. Measures were highly specific for CD, especially the subset of patients at increased risk of surgery¹²². Serum anti-glycoprotein 2 (GP2) IgG and IgA might also be a novel marker of CD especially in children¹²³. More sophisticated 'omics' technologies for biomarker discovery using blood, intestinal mucosa,

and feces are also being pursued¹²⁴. Alongside classical proteomic approaches the use of imaging mass spectrometry on histological samples has been explored¹²⁵. Imaging mass spectrometry enables direct measurement of multiple individual species within a complex clinical sample through histology-directed mass spectral protein profiling¹²⁵. Such an approach requires tissue samples obtained during endoscopy. Blood or fecal biomarkers remain the ideal although the utility of exhaled breath condensate and exhaled nitric oxide for pediatric IBD has been considered¹²⁶. Analysis of exhaled breath is a well-established tool for non-invasive assessment of lung disease, yet there were some features that allowed differentiation of samples from children with asthma versus IBD, e.g. pH was lower with IBD than either asthma or controls¹²⁶. A larger study that would incorporate omics approaches is planned by this group.

Recently there has been a flurry of papers exploring miRNA signatures in IBD using blood, saliva and colonic tissue to reveal differential expression profiles associated with UC versus CD. Importantly these IBDs could be discriminated from other inflammatory disorders using machine learning techniques for expression profiling¹²⁷. There is now much interest in evaluating these in large, independent, clinically well characterized cohorts and it would be good to see such studies extended to include pediatric populations.

Other diseases

The role of the immune system and inflammation in metabolic and neurological disorders is being increasingly recognized. For example, Toll-like receptors link inflammation and oxidative stress to hypertension, insulin resistance and obesity, and are postulated to have a role in fetal programming of these diseases¹²⁸. Metabolic syndrome is a cluster of cardiometabolic features – insulin resistance, central obesity, dyslipidemia, and hypertension - that impose increased risk for cardiovascular disease (CVD) and type 2 diabetes mellitus. These features are increasing in prevalence secondary to the obesity epidemic. Life style interventions implemented in early childhood are of interest to prevent the rise in CVD and type 2 diabetes mellitus. The early detection of children at risk of the metabolic and cardiac derangements secondary to obesity is critical and readily and objectively quantifiable biomarkers would be ideal. Candidate biomarkers that have shown promise in the past include CRP but it lacks the necessary precision. Various adipokines, especially leptin and adiponectin, have also been investigated as biomarkers. Circulating adiponectin, but not leptin, tracked with health improvement, improved insulin sensitivity

and loss of fat mass, after lifestyle intervention in overweight and obese children, so adiponectin was postulated as a good biomarker for monitoring the efficacy of such lifestyle interventions¹²⁹.

Much of the biomarker work in children relates to obesity and cardiovascular risk, especially seeking markers of endothelial dysfunction, an early but reversible step in the development of atherosclerosis¹³⁰. Interest in endothelial progenitor cells (EPCs) and EMPs is driven by their correlation with cardiovascular risk functions and parameters of endothelial dysfunction¹³¹. They are postulated to reflect the balance between damage (EMPs) and regeneration (EPCs) and can be measured in blood using flow cytometry. As EPCs and EMPs independently predict microvascular endothelial dysfunction they could provide a useful measure of the effectiveness of intervention programmes in obese children. Fewer EPCs (reduced repair function) and more EMPs (greater damage) have been reported in obese children aged about 15 years old who had impaired endothelial microvascular function, raised systolic blood pressure, and increased arterial stiffness¹³². As with many other biomarker strategies, one of the main challenges to progressing EPCs and EMPs to clinical use relates to standardization of sample processing methods and choice of antigens for flow cytometric identification. Whether monitoring changes in EPCs and EMPs would be of use in younger children who physiologically adapt to increased blood flow demands remains to be seen. However, miR-125a-5p, 342-3p and 365-3p are potential plasma biomarkers for endothelial dysfunction in obese children aged 5 – 10 years¹³³.

Flow cytometry of circulating leukocytes, cytokine analysis in serum, and cytokines and miRNA from peripheral blood mononuclear cells in obese versus non-obese children revealed altered immune cell frequency (decreased invariant NKT cells), inflammatory environment (increased LPS-stimulated IL-1 β from PBMCs) and regulation of metabolic gene expression (increased TNF α and leptin, and decreased adiponectin) in the obese group¹³⁴. Much of this profile has been linked causally to the onset of metabolic disease in adults so offers some insight to disease trajectory in children. Dysregulated miR-33a and 33b were deemed of particular interest as future biomarkers¹³⁴. MicroRNAs are also of interest for their role in developmental, physiological and cognitive processes within the central nervous system¹³⁵.

While plasma is the preferred biological fluid for biomarker discovery in adults, there is probably greater value in exploring even less invasive samples for children. Saliva is of particular interest given the ease of collection. In a study of candidate biomarkers typical of obesity (CRP, insulin, leptin and adiponectin) in the saliva of 10 year old children, the biomarker profile was reminiscent of changes well documented in the circulation and was found to relate to insulin-resistance and systemic elevation of pro-inflammatory cytokines¹³⁶. Altered levels of salivary biomarkers, measured using a multiplex immunoassay approach, were identified in obese children to identify those at high risk of type 2 diabetes mellitus. Sample collection was highly standardized including fasting, time of collection, rinsing of the mouth, amount expectorated, temperature and processing controls, centrifugation to remove cells and cellular debris, careful curation of samples, and total protein assays. There is interest in developing this approach for longitudinal studies related to development of metabolism and other disorders in children and to monitor the success of interventions.

There are now studies emerging on metabolites as biomarkers in childhood. A biomarker discovery approach using untargeted global metabolite profiling with ultra-performance liquid chromatography/quadrupole orthogonal acceleration time of flight tandem micro mass spectrometry of fasted serum of 6 – 17 year old children of varying BMI identified 14 metabolites (e.g. bradykinin, linoleic acid, L-thyronine and naringenin) differentially expressed by BMI¹³⁷. These will be of use in future studies of the metabolic pathways altered with obesity that impact on risk of metabolic and other disorders to reveal novel therapeutic approaches and monitor interventions. Similarly, a metabolomic approach has been used in autism spectrum disorder (ASD) with the long-term goal being to validate any discovered biomarkers in larger clinical studies and implement them for precision medicine. Blood plasma from 4 – 6 year old children with ASD and age-matched controls was analysed with a variety of mass spectrometry methods and multiple orthogonal analytical methods were used¹³⁸. Some of the metabolites of interest had been associated previously with ASD (e.g. creatinine¹³⁹) and the study also revealed a variety of biomarkers associated with mitochondrial dysfunction (decreased citrate and increased succinate) which could relate to energy production and/or oxidative stress. Combined with the increasing interest in immunometabolism¹⁴⁰ there are likely to be data from global screens of energy pathways in relation to clinical phenotypes in the coming years.

Cancer

The focus of this review is immune-mediated NCDs of childhood. However, the use of biomarkers is most advanced for cancer so it is worthwhile considering the current status in this field. Heterogeneity within cancer types has driven much of the need for precision medicine within this clinical specialty. The approaches used for biomarker discovery and validation are similar to those already discussed ranging from traditionally identified single biomarkers to omics approaches. Breast cancer is an excellent example where well established single biomarkers – estrogen receptor (OR), progesterone receptor (PR) and human epidermal growth factor 2 (HER2) – are used routinely for prognosis and to target therapy¹⁴¹. Efforts are now focussed on further refining disease classification, especially molecular definitions to facilitate the use of rational therapies targeting specific molecular pathways¹⁴¹. Molecular profiling is already in use for breast cancer and for other cancer types such as metastatic colorectal cancer¹⁴².

Resurgent interest in cancer immunotherapies is accompanied by the need for complementary biomarkers. An example of this is the PD-1/PD-L1 axis¹⁴³. PD-L1 expression by cancer and immune cells is recognised to have a role in blocking anti-cancer immunity and PD-L1 targeted therapies such as an engineered humanised antibody have been developed for treatment of melanoma, lung, kidney, bladder and other cancers^{144,145}. Emerging data indicate that this therapy is most effective in patients with tumours that highly express PD-L1¹⁴⁴ which can be assessed immunohistochemically within the tumour biopsy¹⁴⁶. However, patients with PD-L1 low tumours also respond well to anti-PD-L1 therapy^{143, 145, 146}. Despite early promise, the implementation of PD-L1 as a predictive biomarker remains problematic but it does serve as a model for the general approach now being taken within the cancer field – parallel development of therapy and biomarker. Optimising precision medicine approaches remains under debate but is likely an iterative process: as more individuals have their disease managed this way the best timings and target populations will become apparent.

As for the other diseases discussed here, progress in implementation of diagnostic and prognostic biomarkers in paediatric cancer populations lags behind that in adults¹⁴⁷. There are numerous studies identifying candidate biomarkers in small sample sets but these require validation. Examples include: surface-enhanced laser desorption /ionization-time-of-flight mass spectroscopy (SELDI-ToF-MS) to reveal platelet factor 4, connective tissue

activating peptide III and two fragments of complement C3a as potential circulating diagnostic biomarkers for acute lymphoblastic leukemia¹⁴⁸; and a variety of candidate biomarkers such as osteopontin and metallothionein reported by different groups for diagnosing various pediatric central nervous system tumours using either blood or cerebrospinal fluid¹⁴⁹. One particular lesson to be learned from oncology is the effort made to offset the disappointing follow on studies of promising new biomarkers typically related to the challenges of biomarker validation and implementation discussed herein¹⁵⁰.

Benefits and challenges of biomarkers

Biomarkers must consistently and accurately predict a biological process or clinical outcome of interest. Most biomarkers in clinical use today have come about through the traditional experimental route outlined above. Today, these and untargeted (typically omics) approaches are used for biomarker discovery. It is very easy to speculate about the longer-term clinical utility of a novel potential biomarker but the reality is a lengthy, painful, and expensive path from exploratory to a qualified biomarker. Translation of a biomarker is a very slow process – discovery, validation/evaluation, clinical trial and approval (Figure 1) can take over a decade at vast cost. Biomarker discovery typically relies on targeted or untargeted quantification of multiple analytes in complex samples in a case-control fashion to provide a list of candidate biomarkers. The next stage requires validation of these candidate biomarkers in large and independent cohorts of patients to reveal clinical utility. This requires cost effective and specific assays which might have to be developed from scratch; immunoassays are the standard as they readily lend themselves to translation but immunoblotting and mass spectrometry also provide valuable validation approaches. Mass spectrometry, in particular, enables high throughput approaches that can overcome the bottleneck between discovery and verification¹⁵¹. Finally, clinical implementation requires development of an assay for use in clinical laboratories, health economics assessment including identifying a willing manufacturer (i.e. costs versus benefits), and approval by appropriate regulatory agents (e.g. FDA)¹⁵². Bringing together relevant partners can overcome the cost and intellectual property hurdles to biomarker qualification. An example is the Predictive Safety Testing Consortium which relates to drug safety biomarkers and is a consortium between different pharmaceutical companies, academic institutions, and other partners under advisement of the FDA (U.S. Food and Drug Administration), its European

counterpart the EMA (European Medicines Agency), and PMDA (Japanese Pharmaceutical and Medical Devices Agency)¹⁵³.

The challenges facing the discovery, evaluation/validation, and clinical implementation of biomarkers are many and typically relate to sample choice and preparation, the instrumentation used to discover and validate biomarkers, and the downstream data and statistical analysis tools.

Sample. A simple to collect sample such as plasma, routinely used for a range of clinical diagnostics in hospitals and laboratories around the world, is an attractive option. However, global interrogation of the human plasma proteome is hindered by the most abundant plasma proteins that constitute some 99% of the plasma proteome³⁴. Successful biomarker discovery in plasma typically requires extensive sample fractionation and sophisticated analysis instrumentation such as mass spectrometry. One of the greatest challenges for clinical utility of biomarker analysis is sample heterogeneity due to natural physiological variation in health and disease (e.g. sex, age, time of day, lifestyle factors (Figure 2) and non-physiological factors (e.g. specimen handling). All of these elements must be standardized. Biological variation also limits the usefulness of pooling individual biological specimens for biomarker discovery¹⁵⁴.

Equipment and data analysis. Mass spectrometry is the cornerstone of proteomics and metabolomics providing high sensitivity, specificity, mass accuracy and good dynamic range, all while being relatively fast. High throughput technologies enable rapid identification of candidate biomarkers but many will require assay development *de novo* for validation and implementation phases although mass spectrometry is already in clinical use. Advances in proteomics and metabolomics have allowed many investigators to rapidly and precisely measure the size and relative abundance of vast numbers of proteins or metabolites in complex mixtures such as plasma, serum, saliva, and urine with the goal of biomarker discovery. Translating these to clinical utility can prove to be difficult. Sophisticated computational approaches are required to distil all the information available into development of suitable assays and then an accurate classifier score validated in complex clinical cohorts¹⁵². Poor choice of approach at this stage can lead to failure through either selection of the wrong biomarkers at the discovery stage or disappointment at the validation stage. This has led to the development of computational pipelines that support discovery,

validation, and implementation including machine learning frameworks being developed to support clinical deployment¹⁵⁵. Such pipelines have been developed in a number of areas including for cardiac transplantation where 5 biomarker candidates with biological relevant functions were identified and progressed to external validation¹⁵².

Multi-parameter flow cytometry, either fluorescence based or more recently through rare element labeled antibodies and mass cytometry, lends itself to biomarker discovery. However, this technology requires the use of antibodies or other labeling modalities to targets so retains an element of targeted discovery. However, machine learning strategies can remove the investigator biased constraints for identifying new cell subsets and other differentiating features. High dimensional mass cytometry enables measurement of some 40 features of an individual cell and has driven the need for single cell and population based computational strategies incorporating machine learning approaches to reveal multi-parametric relationships¹⁵⁶. There are an increasing number of computation tools to enable flow cytometry bioinformatics and support a range of activities related to data storage, retrieval, organisation and analysis for diagnosis and discovery¹⁵⁷. At the validation stage one of the greatest challenges is standardizing sample handling, reagents, instrument set up, and data analysis but there are efforts towards standardization on many fronts including the Human Immunology Project¹⁵⁸.

MicroRNAs can be measured easily using real time quantitative PCR which provides high precision signal amplification, and by microarrays and next generation sequencing platforms for high throughput to enhance the flow from discovery to validation to implementation. As with other technologies discussed herein one of the major challenges is standardization of procedures at sample collection, storage, RNA isolation, miRNA quality and quantity evaluation and pre-amplification if required. There are then challenges at the analysis stage as all methods introduce some level of bias so normalization strategies are applied to raw expression data and these are critical to meaningful data processing¹⁵⁹. As for mass spectrometry and flow cytometry the development of sophisticated usable data analysis pipelines is an ongoing challenge.

Summary

The utility of biomarkers is two-fold: to identify mechanistic pathways enabling better understanding of disease processes and revealing novel therapeutic targets; and to generate

diagnostic or prognostic biomarkers that have clinical impact. The goal for the latter is an objective, accurate and reproducible measurement that relates to the clinical scenario of interest. Such a biomarker must be accurate, sensitive and specific; it helps if it is physiologically relevant to the disease or biological/pathological state of interest; detection methods should be rapid, reliable, reproducible and easily interpreted; and ideally it should be measurable in a readily accessible biological fluid such as plasma, urine or saliva. The challenge is to translate a biomarker from the research environment to routine clinical use through the phases of discovery, validation/evaluation and implementation which can be an iterative process. Biomarker discovery for paediatric diseases lags behind that for adult diseases but the epidemic of NCDs should catalyse such activity to provide the much needed evidence-based approaches for risk stratification through the life-course.

What do we know?

- The prevalence of NCDs is increasing in all countries, in all income groups, and at all ages due to the globalization of common risk factors (tobacco use; food high in saturated and trans fats, salt and sugar; physical inactivity; and alcohol consumption).
- A shared feature of all NCDs is chronic low-grade inflammation promoted by modern diets, environmental pollutants, microbial exposure, and stress; inflammatory pathways in NCDs are multi-factorial and part of a metabolic cascade.
- A biomarker can be a gene, molecule, or other biological characteristic as long as it can be measured accurately and reproducibly to provide an objective measure of health status or disease with potential for use in risk stratification, early detection, identifying treatment of choice and monitoring response to treatment, in surveillance, and in drug monitoring and development.
- Many biomarkers already in clinical use were first identified during targeted studies with the 'omics' era now leading to unbiased discovery-based approaches to identify biomarkers (including panels, clusters or signatures) to better understand biological processes and for clinical biomarker discovery.
- Translation of a biomarker is a very slow process – discovery, validation/evaluation, clinical trial and approval can take over a decade at vast cost.

What is still unknown?

- Sample types for biomarker discovery, validation and clinical implementation include serum, plasma, saliva, urine, exhaled breath, tissue biopsies, mucosal secretions, cerebrospinal fluid, and other biological specimens; natural physiological variation and non-physiological factors contribute to sample heterogeneity and sample collection, processing and storage must be optimised.
- One approach does not fit all: mass spectrometry, flow cytometry, microarrays and next generation sequencing offer high throughput approaches to biomarker discovery and validation - sample analysis tools need optimising for the clinical scenario of interest.
- Analysis of complex data sets is challenging and poor choice of analytical approach can lead to failure; computational pipelines that support discovery, validation, and implementation including machine learning frameworks are required to support clinical deployment.
- Biomarker discovery and validation in childhood disease lags behind that for adults; given the early onset and therefore potential lifelong impact of many NCDs there should be more studies incorporating cohorts of children.

Table 1. Common autoimmune diseases of childhood and relevant autoantibodies.

Autoimmune Disease	Autoantibodies (Abs)	Age	Comments	References
Type 1 Diabetes	Anti-Insulin (IAA) Anti-Glutamate decarboxylase (GAD) Anti-Islet Cell (ICA) Anti-Insulinoma associated (IA-2) Anti-Zink-Transporter (ZnT8)	5-50 years 10-12 years 10.9 ± 3.9 years Adult	<ul style="list-style-type: none"> IAA are often the first specific detectable Abs among children, identified in ~ 50% of T1 diabetic children GAD usually appear <5 years following appearance of anti-insulin Abs All Abs can appear before onset of symptoms, prevalence rises after disease manifestation 	160-164
Coeliac disease	Anti-Gliadin Anti-Endomysium (EmA)/Tissue Transglutamin	9 month – 12.3 yrs Adult 0.5-92 yrs		165-167
Autoimmune thyroiditis	Anti-Thyroperoxidase Anti-Thyroglobulin	1-19 years		168, 169
Pediatric Lupus	Antinuclear antibodies (ANA) Anti-dsDNA Anti-Sm Anti-PCNA Anti-Ribosomal Protein (RPP) Anti-Nucleosome Anti-Histone Anti-Phospholipid Anti-SS-A (Ro) Anti-SS-B (La) Anti-U1-nRNP Anti-C1q	11.0 ± 3.6 years 4-18 years	<ul style="list-style-type: none"> High levels of anti-RPP point to children with different autoimmune diseases, or mixed connective tissue disease. 	170-172
Juvenile Idiopathic Arthritis	Rheumatoid Factor (RF) ANA	0.5-11.4 years	<ul style="list-style-type: none"> RF is identified in ~ 15% of cases. Anti-Cardiolipin in ~ 	173-175

	Anti-HMG anti-CCP/CCA	15.59 ± 4.13 years	30% of cases, in low titers and without association to clinical antiphospholipid syndrome <ul style="list-style-type: none"> • ANA in ~ 30% of cases • Anti-CCP/CCA are rarely positive 	
Juvenile Dermatomyositis (JDM)	Antibodies against: P155 Mi-2alpha Mi-2beta TIF1gamma MDA5 NXP2 SAE1 Ku PM100 PM75 Jo-1 SRP PL-7 PL-12 EJ OJ Ro-52	<18 years 1-80 years	<ul style="list-style-type: none"> • Approximately 10 % of children with JDM show myositis-specific autoantibodies • Children with anti-Jo-1 and/or anti-SRP tend to follow a chronic continuous disease course. • Anti-SRP are identified in polymyositis children with severe muscle weakness, and thinning or atrophy, and often are not responsive to single medication. • Children with anti-Mi-2 typically have mild to moderate disease and respond well to treatment. • Anti-p155 is the most common autoantibody found in children with JDM. 	176-178
Multiple Sclerosis	Anti-Myelin Oligodendrocyte Glycoprotein (MOG)	1.3 – 15.8 years	<ul style="list-style-type: none"> • Anti-Aquaporin for exclusion of optical neuromyelitis 	179

Disease	Biomarker	Sample type	Age	Analysis approach	Proposed use	Reference
Atopic Dermatitis Food Allergy	sICAM-1	serum	10 - 16 years	immunoassay	disease monitoring	43
	LL-37	serum	6 - 15 years	immunoassay	disease monitoring	44
	IL-31	serum	1 - 10 years	immunoassay	disease monitoring	45
	IL-18	serum	0 - 6 years	immunoassay	disease monitoring	46
	CCL28	serum	0 - 10 years	immunoassay	disease monitoring	47
	CCL17	serum	0 - >16 years	immunoassay	disease monitoring	48
		serum	birth	immunoassay	risk monitoring/disease prediction	63
	CCL27	serum	1 - 11 years	immunoassay	disease monitoring	49
	miR-203 miR-483-5p	serum/urine	0.5 - 6 years	global miRNA profiling & qRT-PCR	disease monitoring and progression	51
	carnitine free fatty acids lactic acid*	serum	0 - 3 years	Mass spectrometry	mechanistic insight/biomarker discovery	54
CD5L apolipoprotein E	plasma	2 - 7 years	2D elcetrophoresis	mechanistic insight/ biomarker discovery	56	
allergen-specific IgA and IgG	serum	5 - 12 years	DCP microarray	response to oral immunotherapy	57	
egg white IgE	serum	1 - 16 years	ImmunoCAP	response to oral immunotherapy	58	

	calprotectin	faeces	2 months	immunoassay	risk monitoring/disease prediction	59
	soluble FasL	plasma	birth	immunoassay	risk monitoring/disease prediction	62
	CCL22	plasma	birth	multiplex immunoarray	risk monitoring/disease prediction	64
	CD4+ regulatory T cells	whole blood/PBMCs	birth	flow cytometry, qRT-PCR, DNA methylation	risk monitoring/disease prediction	65 - 67
	polyubiquitin C calmodulin-like protein 5	vernix	birth	mass spectrometry	risk monitoring/disease prediction	69
	filaggrin	blood/saliva		DNA genotyping	risk monitoring/disease prediction	68
Allergic Rhinitis	miR-146a	nasal mucosa & PBMCs		qRT-PCR		70
	FoxP3	PBMCs	4 - 14 years		pathogenesis & management	
	IL-10	serum		immunoassay		
Asthma	FeNO/eosinophils	exhaled air/blood	6 - 19 years	NIOX® analyser/haematology analyser	disease phenotype/treatment response	90
	FeNO/eosinophils/periostin	exhaled air/blood	12-75years	NIOX® analyser/haematology analyser/immunoassay	treatment efficacy	83
	periostin	Airway biopsies	14 - 85	immunohistochemistry	mechanistic insight/biomarker discovery	87
		epithelial cells	6 - 16 years	cell culture/qRT-CR	mechanistic insight/biomarker discovery	88
		serum	6 - 15 years	immunoassay	mechanistic insight/biomarker discovery	91

	miR-21	serum		qRT-PCR	disease monitoring	93
	miR-638 & others	blood	6 - 18 years	global miRNA profiling	mechanistic insight/biomarker discovery	94
	TNF α	LPS-stimulated PBMCs	birth/3 months	cell culture/immunoassay	risk monitoring	97
	volatile organic compounds TLR4, catalase, TNF α clinical	EBC PBMCs	2 - 4 years	mass spectrometry qRT-PCR athma predictive index	risk monitoring	98
	IL-2 IL-4 IL-8 IL-10	EBC	2 - 3 years	multiplex immunoarray	risk monitoring	99
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IBD	ANCA ASCA anti-OmpC	serum	1 - 21 years	immunoassay	increased risk of surgery	122
	anti-glycoprotein 2	serum	2 -18 years	immunoassay	diagnosis	123
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Other diseases	Adiponectin	blood	5 - 15 years	radioimmunoassay	lifestyle intervention for obesity	129
	Endothelial progenitor cells Endothelial microparticles	blood	12 - 18 years	flow cytometry	monitoring endothelial dysfunction	132
	miR-125-5p miR-342-3p	plasma	5 - 10 years	global miRNA profiling/qRT-PCR	monitoring endothelial dysfunction	133

miR-365-3p						
CRP insulin adiponectin	saliva	11 years	multiplex immunoarray	intervention and prevention type 2 diabetes		136
bradykinin naringenin L-thyronine*	serum	6 - 17 years	mass spectrometry	mechanistic insight/biomarker discovery obesity		137
citrate succinate creatinine glutaric acid 3-aminoisobutyric acid p-hydroxyphenyllactate*	plasma	4 - 7 years	mass spectrometry	mechanistic insight/biomarker discovery autism spectrum disorder		138

Table 2. Summary of immune biomarkers in childhood non-communicable diseases.
*and others

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1. Prescott S, Nowak-Wegrzyn A. Strategies to prevent or reduce allergic disease. *Ann Nutr Metab* 2011; 59 Suppl 1:28-42.
2. Rosenbaum L, Lamas D. Facing a "slow-motion disaster"--the UN meeting on noncommunicable diseases. *N Engl J Med* 2011; 365:2345-8.
3. Beaglehole R, Bonita R, Horton R, Adams C, Alleyne G, Asaria P, et al. Priority actions for the non-communicable disease crisis. *Lancet* 2011; 377:1438-47.
4. Prescott SL. Early-life environmental determinants of allergic diseases and the wider pandemic of inflammatory noncommunicable diseases. *J Allergy Clin Immunol* 2013; 131:23-30.
5. Geneau R, Stuckler D, Stachenko S, McKee M, Ebrahim S, Basu S, et al. Raising the priority of preventing chronic diseases: a political process. *Lancet* 2010; 376:1689-98.
6. Yusuf S, Hawken S, Ounpuu S, Dans T, Avezum A, Lanas F, et al. Effect of potentially modifiable risk factors associated with myocardial infarction in 52 countries (the INTERHEART study): case-control study. *Lancet* 2004; 364:937-52.
7. Alwan A, Maclean DR, Riley LM, d'Espaignet ET, Mathers CD, Stevens GA, et al. Monitoring and surveillance of chronic non-communicable diseases: progress and capacity in high-burden countries. *Lancet* 2010; 376:1861-8.
8. Prescott SL, Clifton V. Asthma and pregnancy: emerging evidence of epigenetic interactions in utero. *Curr Opin Allergy Clin Immunol* 2009; 9:417-26.
9. Medzhitov R. Origin and physiological roles of inflammation. *Nature* 2008; 454:428-35.
10. Gregor MF, Hotamisligil GS. Inflammatory mechanisms in obesity. *Annu Rev Immunol* 2011; 29:415-45.
11. West CE, Renz H, Jenmalm MC, Kozyrskyj AL, Allen KJ, Vuillermin P, et al. The gut microbiota and inflammatory noncommunicable diseases: associations and potentials for gut microbiota therapies. *J Allergy Clin Immunol* 2015; 135:3-13; quiz 4.
12. Wold AE. The hygiene hypothesis revised: is the rising frequency of allergy due to changes in the intestinal flora? *Allergy* 1998; 53:20-5.
13. de Goffau MC, Luopajarvi K, Knip M, Ilonen J, Ruohtula T, Harkonen T, et al. Fecal microbiota composition differs between children with beta-cell autoimmunity and those without. *Diabetes* 2013; 62:1238-44.
14. Penders J, Gerhold K, Stobberingh EE, Thijs C, Zimmermann K, Lau S, et al. Establishment of the intestinal microbiota and its role for atopic dermatitis in early childhood. *J Allergy Clin Immunol* 2013; 132:601-7 e8.
15. Frank DN, St Amand AL, Feldman RA, Boedeker EC, Harpaz N, Pace NR. Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases. *Proc Natl Acad Sci U S A* 2007; 104:13780-5.
16. Cani PD, Geurts L, Matamoros S, Plovier H, Duparc T. Glucose metabolism: focus on gut microbiota, the endocannabinoid system and beyond. *Diabetes Metab* 2014; 40:246-57.

17. Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science* 2009; 324:1029-33.
18. Calder PC. Fuel utilization by cells of the immune system. *Proc Nutr Soc* 1995; 54:65-82.
19. Chacko BK, Kramer PA, Ravi S, Johnson MS, Hardy RW, Ballinger SW, et al. Methods for defining distinct bioenergetic profiles in platelets, lymphocytes, monocytes, and neutrophils, and the oxidative burst from human blood. *Lab Invest* 2013; 93:690-700.
20. Ravi S, Mitchell T, Kramer PA, Chacko B, Darley-Usmar VM. Mitochondria in monocytes and macrophages-implications for translational and basic research. *Int J Biochem Cell Biol* 2014; 53:202-7.
21. Ghesquiere B, Wong BW, Kuchnio A, Carmeliet P. Metabolism of stromal and immune cells in health and disease. *Nature* 2014; 511:167-76.
22. O'Sullivan D, van der Windt GJ, Huang SC, Curtis JD, Chang CH, Buck MD, et al. Memory CD8(+) T cells use cell-intrinsic lipolysis to support the metabolic programming necessary for development. *Immunity* 2014; 41:75-88.
23. Malinarich F, Duan K, Hamid RA, Bijin A, Lin WX, Poidinger M, et al. High mitochondrial respiration and glycolytic capacity represent a metabolic phenotype of human tolerogenic dendritic cells. *J Immunol* 2015; 194:5174-86.
24. Datta G, Kramer PA, Johnson MS, Sawada H, Smythies LE, Crossman DK, et al. Bioenergetic programming of macrophages by the apolipoprotein A-I mimetic peptide 4F. *Biochem J* 2015; 467:517-27.
25. Macintyre AN, Gerriets VA, Nichols AG, Michalek RD, Rudolph MC, Deoliveira D, et al. The glucose transporter Glut1 is selectively essential for CD4 T cell activation and effector function. *Cell Metab* 2014; 20:61-72.
26. Cao Y, Rathmell JC, Macintyre AN. Metabolic reprogramming towards aerobic glycolysis correlates with greater proliferative ability and resistance to metabolic inhibition in CD8 versus CD4 T cells. *PLoS One* 2014; 9:e104104.
27. Wahl DR, Byersdorfer CA, Ferrara JL, Opipari AW, Jr., Glick GD. Distinct metabolic programs in activated T cells: opportunities for selective immunomodulation. *Immunol Rev* 2012; 249:104-15.
28. Gerriets VA, Kishton RJ, Nichols AG, Macintyre AN, Inoue M, Ilkayeva O, et al. Metabolic programming and PDHK1 control CD4+ T cell subsets and inflammation. *J Clin Invest* 2015; 125:194-207.
29. Yin Y, Choi SC, Xu Z, Perry DJ, Seay H, Croker BP, et al. Normalization of CD4+ T cell metabolism reverses lupus. *Sci Transl Med* 2015; 7:274ra18.
30. Godfrey KM, Gluckman PD, Hanson MA. Developmental origins of metabolic disease: life course and intergenerational perspectives. *Trends Endocrinol Metab* 2010; 21:199-205.
31. Strimbu K, Tavel JA. What are biomarkers? *Curr Opin HIV AIDS* 2010; 5:463-6.

32. Calder PC. Biomarkers of immunity and inflammation for use in nutrition interventions: International Life Sciences Institute European Branch work on selection criteria and interpretation. *Endocr Metab Immune Disord Drug Targets* 2014; 14:236-44.
33. Rifai N, Gerszten RE. Biomarker discovery and validation. *Clin Chem* 2006; 52:1635-7.
34. Anderson NL, Anderson NG. The human plasma proteome: history, character, and diagnostic prospects. *Mol Cell Proteomics* 2002; 1:845-67.
35. Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell* 2009; 136:215-33.
36. He Y, Lin J, Kong D, Huang M, Xu C, Kim TK, et al. Current State of Circulating MicroRNAs as Cancer Biomarkers. *Clin Chem* 2015; 61:1138-55.
37. Li YR, Li J, Zhao SD, Bradfield JP, Mentch FD, Maggadottir SM, et al. Meta-analysis of shared genetic architecture across ten pediatric autoimmune diseases. *Nat Med* 2015; 21:1018-27.
38. Global Atlas of Allergy. In: Akdis CA, Agache I, eds: *European Academy of Allergy and Clinical Immunology*, 2014.
39. Odhiambo JA, Williams HC, Clayton TO, Robertson CF, Asher MI. Global variations in prevalence of eczema symptoms in children from ISAAC Phase Three. *Journal of Allergy and Clinical Immunology* 2009; 124:1251-8.e23.
40. Prescott SL, Pawankar R, Allen KJ, Campbell DE, Sinn JK, Fiocchi A, et al. A global survey of changing patterns of food allergy burden in children. *World Allergy Organization Journal* 2013; 6:1-12.
41. Alduraywish SA, Lodge CJ, Campbell B, Allen KJ, Erbas B, Lowe AJ, et al. The march from early life food sensitization to allergic disease: a systematic review and meta-analyses of birth cohort studies. *Allergy* 2016; 71:77-89.
42. Thomsen SF. Epidemiology and natural history of atopic diseases. *European Clinical Respiratory Journal* 2015; 2:10.3402/ecrj.v2.24642.
43. Halmerbauer G, Frischer T, Koller DY. Monitoring of disease activity by measurement of inflammatory markers in atopic dermatitis in childhood. *Allergy* 1997; 52:765-9.
44. Leung TF, Ching KW, Kong AP, Wong GW, Chan JC, Hon KL. Circulating LL-37 is a biomarker for eczema severity in children. *J Eur Acad Dermatol Venereol* 2012; 26:518-22.
45. Ezzat MH, Hasan ZE, Shaheen KY. Serum measurement of interleukin-31 (IL-31) in paediatric atopic dermatitis: elevated levels correlate with severity scoring. *J Eur Acad Dermatol Venereol* 2011; 25:334-9.
46. Hon KL, Leung TF, Ma KC, Wong CK, Wan H, Lam CW. Serum concentration of IL-18 correlates with disease extent in young children with atopic dermatitis. *Pediatr Dermatol* 2004; 21:619-22.
47. Ezzat MH, Sallam MA, Shaheen KY. Serum mucosa-associated epithelial chemokine (MEC/CCL28) in atopic dermatitis: a specific marker for severity. *Int J Dermatol* 2009; 48:822-9.
48. Fujisawa T, Nagao M, Hiraguchi Y, Katsumata H, Nishimori H, Iguchi K, et al. Serum measurement of thymus and activation-regulated

- chemokine/CCL17 in children with atopic dermatitis: elevated normal levels in infancy and age-specific analysis in atopic dermatitis. *Pediatr Allergy Immunol* 2009; 20:633-41.
49. Hon KL, Leung TF, Ma KC, Li AM, Wong Y, Fok TF. Serum levels of cutaneous T-cell attracting chemokine (CTACK) as a laboratory marker of the severity of atopic dermatitis in children. *Clin Exp Dermatol* 2004; 29:293-6.
 50. Kataoka Y. Thymus and activation-regulated chemokine as a clinical biomarker in atopic dermatitis. *J Dermatol* 2014; 41:221-9.
 51. Lv Y, Qi R, Xu J, Di Z, Zheng H, Huo W, et al. Profiling of serum and urinary microRNAs in children with atopic dermatitis. *PLoS One* 2014; 9:e115448.
 52. Lu TX, Sherrill JD, Wen T, Plassard AJ, Besse JA, Abonia JP, et al. MicroRNA signature in patients with eosinophilic esophagitis, reversibility with glucocorticoids, and assessment as disease biomarkers. *J Allergy Clin Immunol* 2012; 129:1064-75 e9.
 53. Rebane A. microRNA and Allergy. *Adv Exp Med Biol* 2015; 888:331-52.
 54. Huang Y, Chen G, Liu X, Shao Y, Gao P, Xin C, et al. Serum metabolomics study and eicosanoid analysis of childhood atopic dermatitis based on liquid chromatography-mass spectrometry. *J Proteome Res* 2014; 13:5715-23.
 55. Kim YS, Hwang SY, Oh S, Sohn H, Kang HY, Lee JH, et al. Identification of target proteins of N-acetylglucosaminyl-transferase V and fucosyltransferase 8 in human gastric tissues by glycomic approach. *Proteomics* 2004; 4:3353-8.
 56. Kim WK, Hwang HR, Kim do H, Lee PY, In YJ, Ryu HY, et al. Glycoproteomic analysis of plasma from patients with atopic dermatitis: CD5L and ApoE as potential biomarkers. *Exp Mol Med* 2008; 40:677-85.
 57. Mayumi S, Kamemura N, Nagao M, Irahara M, Kagami S, Fujisawa T, et al. Differential response in allergen-specific IgE, IgGs and IgA levels for predicting outcome of oral immunotherapy. *Pediatr Allergy Immunol* 2016.
 58. Vickery BP, Pons L, Kulis M, Steele P, Jones SM, Burks AW. Individualized IgE-based dosing of egg oral immunotherapy and the development of tolerance. *Ann Allergy Asthma Immunol* 2010; 105:444-50.
 59. Orivuori L, Mustonen K, de Goffau MC, Hakala S, Paasela M, Roduit C, et al. High level of fecal calprotectin at age 2 months as a marker of intestinal inflammation predicts atopic dermatitis and asthma by age 6. *Clin Exp Allergy* 2015; 45:928-39.
 60. Oh SY, Chung J, Kim MK, Kwon SO, Cho BH. Antioxidant nutrient intakes and corresponding biomarkers associated with the risk of atopic dermatitis in young children. *Eur J Clin Nutr* 2010; 64:245-52.
 61. Schoeters GE, Den Hond E, Koppen G, Smolders R, Bloemen K, De Boever P, et al. Biomonitoring and biomarkers to unravel the risks from prenatal environmental exposures for later health outcomes. *Am J Clin Nutr* 2011; 94:1964S-9S.

62. Su KW, Chen PC, Wang IJ. Cord blood soluble Fas ligand and pediatric atopic dermatitis. *Allergy Asthma Proc* 2011; 32:366-71.
63. Miyahara H, Okazaki N, Nagakura T, Korematsu S, Izumi T. Elevated umbilical cord serum TARC/CCL17 levels predict the development of atopic dermatitis in infancy. *Clin Exp Allergy* 2011; 41:186-91.
64. Folsgaard NV, Chawes BL, Bonnelykke K, Jenmalm MC, Bisgaard H. Cord blood Th2-related chemokine CCL22 levels associate with elevated total-IgE during preschool age. *Clin Exp Allergy* 2012; 42:1596-603.
65. Bullens DM, Seys S, Kasran A, Dilissen E, Dupont LJ, Ceuppens JL. Low cord blood Foxp3/CD3gamma mRNA ratios: a marker of increased risk for allergy development. *Clin Exp Allergy* 2015; 45:232-7.
66. Hinz D, Bauer M, Roder S, Olek S, Huehn J, Sack U, et al. Cord blood Tregs with stable FOXP3 expression are influenced by prenatal environment and associated with atopic dermatitis at the age of one year. *Allergy* 2012; 67:380-9.
67. Smith M, Tourigny MR, Noakes P, Thornton CA, Tulic MK, Prescott SL. Children with egg allergy have evidence of reduced neonatal CD4(+)CD25(+)CD127(lo/-) regulatory T cell function. *J Allergy Clin Immunol* 2008; 121:1460-6, 6 e1-7.
68. Venkataraman D, Soto-Ramirez N, Kurukulaaratchy RJ, Holloway JW, Karmaus W, Ewart SL, et al. Filaggrin loss-of-function mutations are associated with food allergy in childhood and adolescence. *J Allergy Clin Immunol* 2014; 134:876-82 e4.
69. Holm T, Rutishauser D, Kai-Larsen Y, Lyutvinskiy Y, Stenius F, Zubarev RA, et al. Protein biomarkers in vernix with potential to predict the development of atopic eczema in early childhood. *Allergy* 2014; 69:104-12.
70. Luo X, Hong H, Tang J, Wu X, Lin Z, Ma R, et al. Increased Expression of miR-146a in Children With Allergic Rhinitis After Allergen-Specific Immunotherapy. *Allergy Asthma Immunol Res* 2016; 8:132-40.
71. Park H, Huang X, Lu C, Cairo MS, Zhou X. MicroRNA-146a and microRNA-146b regulate human dendritic cell apoptosis and cytokine production by targeting TRAF6 and IRAK1 proteins. *J Biol Chem* 2015; 290:2831-41.
72. Asher MI, Montefort S, Bjorksten B, Lai CK, Strachan DP, Weiland SK, et al. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC Phases One and Three repeat multicountry cross-sectional surveys. *Lancet* 2006; 368:733-43.
73. Corren J, Lemanske RF, Hanania NA, Korenblat PE, Parsey MV, Arron JR, et al. Lebrikizumab treatment in adults with asthma. *N Engl J Med* 2011; 365:1088-98.
74. Fahy JV. Type 2 inflammation in asthma--present in most, absent in many. *Nat Rev Immunol* 2015; 15:57-65.
75. Woodruff PG, Modrek B, Choy DF, Jia G, Abbas AR, Ellwanger A, et al. T-helper type 2-driven inflammation defines major subphenotypes of asthma. *Am J Respir Crit Care Med* 2009; 180:388-95.

76. Choy DF, Hart KM, Borthwick LA, Shikotra A, Nagarkar DR, Siddiqui S, et al. TH2 and TH17 inflammatory pathways are reciprocally regulated in asthma. *Sci Transl Med* 2015; 7:301ra129.
77. Bel EH, Wenzel SE, Thompson PJ, Prazma CM, Keene ON, Yancey SW, et al. Oral glucocorticoid-sparing effect of mepolizumab in eosinophilic asthma. *N Engl J Med* 2014; 371:1189-97.
78. Green RH, Brightling CE, McKenna S, Hargadon B, Parker D, Bradding P, et al. Asthma exacerbations and sputum eosinophil counts: a randomised controlled trial. *Lancet* 2002; 360:1715-21.
79. Jayaram L, Pizzichini MM, Cook RJ, Boulet LP, Lemiere C, Pizzichini E, et al. Determining asthma treatment by monitoring sputum cell counts: effect on exacerbations. *Eur Respir J* 2006; 27:483-94.
80. Pavord ID, Korn S, Howarth P, Bleecker ER, Buhl R, Keene ON, et al. Mepolizumab for severe eosinophilic asthma (DREAM): a multicentre, double-blind, placebo-controlled trial. *Lancet* 2012; 380:651-9.
81. Guo FH, Uetani K, Haque SJ, Williams BR, Dweik RA, Thunnissen FB, et al. Interferon gamma and interleukin 4 stimulate prolonged expression of inducible nitric oxide synthase in human airway epithelium through synthesis of soluble mediators. *J Clin Invest* 1997; 100:829-38.
82. Hanania NA, Wenzel S, Rosen K, Hsieh HJ, Mosesova S, Choy DF, et al. Exploring the effects of omalizumab in allergic asthma: an analysis of biomarkers in the EXTRA study. *Am J Respir Crit Care Med* 2013; 187:804-11.
83. Modena BD, Tedrow JR, Milosevic J, Bleecker ER, Meyers DA, Wu W, et al. Gene expression in relation to exhaled nitric oxide identifies novel asthma phenotypes with unique biomolecular pathways. *Am J Respir Crit Care Med* 2014; 190:1363-72.
84. Smith AD, Cowan JO, Filsell S, McLachlan C, Monti-Sheehan G, Jackson P, et al. Diagnosing asthma: comparisons between exhaled nitric oxide measurements and conventional tests. *Am J Respir Crit Care Med* 2004; 169:473-8.
85. Wenzel S, Ford L, Pearlman D, Spector S, Sher L, Skobieranda F, et al. Dupilumab in persistent asthma with elevated eosinophil levels. *N Engl J Med* 2013; 368:2455-66.
86. Gogate S, Katial R. Pediatric biomarkers in asthma: exhaled nitric oxide, sputum eosinophils and leukotriene E4. *Curr Opin Allergy Clin Immunol* 2008; 8:154-7.
87. Takayama G, Arima K, Kanaji T, Toda S, Tanaka H, Shoji S, et al. Periostin: a novel component of subepithelial fibrosis of bronchial asthma downstream of IL-4 and IL-13 signals. *J Allergy Clin Immunol* 2006; 118:98-104.
88. Lopez-Guisa JM, Powers C, File D, Cochrane E, Jimenez N, Debley JS. Airway epithelial cells from asthmatic children differentially express proremodeling factors. *Journal of Allergy and Clinical Immunology* 2012; 129:990-7.e6.
89. Conway SJ, Izuhara K, Kudo Y, Litvin J, Markwald R, Ouyang G, et al. The role of periostin in tissue remodeling across health and disease. *Cell Mol Life Sci* 2014; 71:1279-88.

90. Konradsen JR, Skantz E, Nordlund B, Lidegran M, James A, Ono J, et al. Predicting asthma morbidity in children using proposed markers of Th2-type inflammation. *Pediatric Allergy and Immunology* 2015; 26:772-9.
91. Song JS, You JS, Jeong SI, Yang S, Hwang IT, Im YG, et al. Serum periostin levels correlate with airway hyper-responsiveness to methacholine and mannitol in children with asthma. *Allergy* 2015; 70:674-81.
92. Kanemitsu Y, Matsumoto H, Izuhara K, Tohda Y, Kita H, Horiguchi T, et al. Increased periostin associates with greater airflow limitation in patients receiving inhaled corticosteroids. *J Allergy Clin Immunol* 2013; 132:305-12 e3.
93. Sawant DV, Yao W, Wright Z, Sawyers C, Tepper RS, Gupta SK, et al. Serum MicroRNA-21 as a Biomarker for Allergic Inflammatory Disease in Children. *Microna* 2015; 4:36-40.
94. Midyat L, Gulen F, Karaca E, Ozkinay F, Tanac R, Demir E, et al. MicroRNA expression profiling in children with different asthma phenotypes. *Pediatr Pulmonol* 2015.
95. Shorter JH, Nelson DD, McManus JB, Zahniser MS, Sama SR, Milton DK. Clinical study of multiple breath biomarkers of asthma and COPD (NO, CO(2), CO and N(2)O) by infrared laser spectroscopy. *J Breath Res* 2011; 5:037108.
96. Smith D, Spanel P, Herbig J, Beauchamp J. Mass spectrometry for real-time quantitative breath analysis. *J Breath Res* 2014; 8:027101.
97. Halonen M, Lohman IC, Stern DA, Ellis WL, Rothers J, Wright AL. Perinatal tumor necrosis factor-alpha production, influenced by maternal pregnancy weight gain, predicts childhood asthma. *Am J Respir Crit Care Med* 2013; 188:35-41.
98. Klaassen EM, van de Kant KD, Jobsis Q, van Schayck OC, Smolinska A, Dallinga JW, et al. Exhaled biomarkers and gene expression at preschool age improve asthma prediction at 6 years of age. *Am J Respir Crit Care Med* 2015; 191:201-7.
99. van de Kant KD, Jansen MA, Klaassen EM, van der Grinten CP, Rijkers GT, Muris JW, et al. Elevated inflammatory markers at preschool age precede persistent wheezing at school age. *Pediatr Allergy Immunol* 2012; 23:259-64.
100. Zhang YL, Luan B, Wang XF, Qiao JY, Song L, Lei RR, et al. Peripheral blood MDSCs, IL-10 and IL-12 in children with asthma and their importance in asthma development. *PLoS One* 2013; 8:e63775.
101. Frohnert BI, Rewers MJ. Metabolomics in childhood diabetes. *Pediatr Diabetes* 2015.
102. Group TS. The Environmental Determinants of Diabetes in the Young (TEDDY) Study. *Ann N Y Acad Sci* 2008; 1150:1-13.
103. Lee HS, Burkhardt BR, McLeod W, Smith S, Eberhard C, Lynch K, et al. Biomarker discovery study design for type 1 diabetes in The Environmental Determinants of Diabetes in the Young (TEDDY) study. *Diabetes Metab Res Rev* 2014; 30:424-34.
104. Churov AV, Oleinik EK, Knip M. MicroRNAs in rheumatoid arthritis: Altered expression and diagnostic potential. *Autoimmunity Reviews* 2015; 14:1029-37.

105. Madsen R, Rantapaa-Dahlqvist S, Lundstedt T, Moritz T, Trygg J. Metabolic responses to change in disease activity during tumor necrosis factor inhibition in patients with rheumatoid arthritis. *J Proteome Res* 2012; 11:3796-804.
106. van Wietmarschen HA, Dai W, van der Kooij AJ, Reijmers TH, Schroen Y, Wang M, et al. Characterization of rheumatoid arthritis subtypes using symptom profiles, clinical chemistry and metabolomics measurements. *PLoS One* 2012; 7:e44331.
107. Churov AV, Oleinik EK, Knip M. MicroRNAs in rheumatoid arthritis: Altered expression and diagnostic potential. *Autoimmun Rev* 2015; 14:1029-37.
108. Kim SW, Ramasamy K, Bouamar H, Lin AP, Jiang D, Aguiar RC. MicroRNAs miR-125a and miR-125b constitutively activate the NF-kappaB pathway by targeting the tumor necrosis factor alpha-induced protein 3 (TNFAIP3, A20). *Proc Natl Acad Sci U S A* 2012; 109:7865-70.
109. Murata K, Furu M, Yoshitomi H, Ishikawa M, Shibuya H, Hashimoto M, et al. Comprehensive microRNA analysis identifies miR-24 and miR-125a-5p as plasma biomarkers for rheumatoid arthritis. *PLoS One* 2013; 8:e69118.
110. Van Schaardenburg D MM, Gui Y, Turk S, Maksymowych WP, Marotta A. Change in 14-3-3' expression in early RA patients treated with dmards corresponds with change in DAS28 and good EULAR responses. *Arthritis Rheumatol* 2014:S837.
111. Maksymowych WP, Marotta A. 14-3-3eta: a novel biomarker platform for rheumatoid arthritis. *Clin Exp Rheumatol* 2014; 32:S-35-9.
112. Jasek M MK, Suresh L, Ambrus JJ, Pardo D. Sjo (TM) an advanced diagnostic panel for detection of Sjogren's Syndrome autoantibodies, arthritis. *Rheumatol* 2014:S1110.
113. Shen L, Suresh L, Lindemann M, Xuan J, Kowal P, Malyavantham K, et al. Novel autoantibodies in Sjogren's syndrome. *Clin Immunol* 2012; 145:251-5.
114. Vermeersch P, Bossuyt X. Prevalence and clinical significance of rare antinuclear antibody patterns. *Autoimmun Rev* 2013; 12:998-1003.
115. Chandran V TA, Gladman DD. Neutrophil-lymphocyte ratio as a marker of disease activity in psoriatic arthritis. *Arthritis Rheumatol* 2014:S704.
116. Draibe JB PR, Merkel PA, Salama AD, Investigators R-I. Serum calprotectin and disease relapse in ANCA-associated vasculitis. *Arthritis Rheumatol* 2014:S818.
117. Stone JH, Merkel PA, Spiera R, Seo P, Langford CA, Hoffman GS, et al. Rituximab versus cyclophosphamide for ANCA-associated vasculitis. *N Engl J Med* 2010; 363:221-32.
118. Perl A. mTOR activation is a biomarker and a central pathway to autoimmune disorders, cancer, obesity, and aging. *Ann N Y Acad Sci* 2015; 1346:33-44.
119. Malmborg P, Grahnquist L, Lindholm J, Montgomery S, Hildebrand H. Increasing incidence of paediatric inflammatory bowel disease in northern Stockholm County, 2002-2007. *J Pediatr Gastroenterol Nutr* 2013; 57:29-34.

120. Kopylov U, Rosenfeld G, Bressler B, Seidman E. Clinical utility of fecal biomarkers for the diagnosis and management of inflammatory bowel disease. *Inflamm Bowel Dis* 2014; 20:742-56.
121. Lewis JD. The utility of biomarkers in the diagnosis and therapy of inflammatory bowel disease. *Gastroenterology* 2011; 140:1817-26 e2.
122. Zholudev A, Zurakowski D, Young W, Leichtner A, Bousvaros A. Serologic testing with ANCA, ASCA, and anti-OmpC in children and young adults with Crohn's disease and ulcerative colitis: diagnostic value and correlation with disease phenotype. *Am J Gastroenterol* 2004; 99:2235-41.
123. Grzybowska-Chlebowczyk U, Wos H, Sieron AL, Wiecek S, Augusciak-Duma A, Koryciak-Komarska H, et al. Serologic investigations in children with inflammatory bowel disease and food allergy. *Mediators Inflamm* 2009; 2009:512695.
124. Huang H, Vangay P, McKinlay CE, Knights D. Multi-omics analysis of inflammatory bowel disease. *Immunol Lett* 2014; 162:62-8.
125. M'Koma AE. Diagnosis of inflammatory bowel disease: Potential role of molecular biometrics. *World J Gastrointest Surg* 2014; 6:208-19.
126. Huang Y, Lemberg DA, Day AS, Dixon B, Leach S, Bujanover Y, et al. Markers of inflammation in the breath in paediatric inflammatory bowel disease. *J Pediatr Gastroenterol Nutr* 2014; 59:505-10.
127. Hubenthal M, Hemmrich-Stanisak G, Degenhardt F, Szymczak S, Du Z, Elsharawy A, et al. Sparse Modeling Reveals miRNA Signatures for Diagnostics of Inflammatory Bowel Disease. *PLoS One* 2015; 10:e0140155.
128. Thompson JA, Webb RC. Potential role of Toll-like receptors in programming of vascular dysfunction. *Clin Sci (Lond)* 2013; 125:19-25.
129. Cambuli VM, Musiu MC, Incani M, Paderi M, Serpe R, Marras V, et al. Assessment of adiponectin and leptin as biomarkers of positive metabolic outcomes after lifestyle intervention in overweight and obese children. *J Clin Endocrinol Metab* 2008; 93:3051-7.
130. Davignon J, Ganz P. Role of endothelial dysfunction in atherosclerosis. *Circulation* 2004; 109:III27-32.
131. Sabatier F, Camoin-Jau L, Anfosso F, Sampol J, Dignat-George F. Circulating endothelial cells, microparticles and progenitors: key players towards the definition of vascular competence. *Journal of Cellular and Molecular Medicine* 2009; 13:454-71.
132. Bruyndonckx L, Hoymans VY, Frederix G, De Guchtenaere A, Franckx H, Vissers DK, et al. Endothelial progenitor cells and endothelial microparticles are independent predictors of endothelial function. *J Pediatr* 2014; 165:300-5.
133. Khalyfa A, Kheirandish-Gozal L, Bhattacharjee R, Khalyfa AA, Gozal D. Circulating miRNAs as Potential Biomarkers of Endothelial Dysfunction In Obese Children. *Chest* 2015.
134. Carolan E, Hogan AE, Corrigan M, Gaotswe G, O'Connell J, Foley N, et al. The impact of childhood obesity on inflammation, innate immune cell frequency, and metabolic microRNA expression. *J Clin Endocrinol Metab* 2014; 99:E474-8.

135. Stoicea N, Du A, Lakis DC, Tipton C, Arias-Morales CE, Bergese SD. The MiRNA Journey from Theory to Practice as a CNS Biomarker. *Front Genet* 2016; 7:11.
136. Goodson JM, Kantarci A, Hartman ML, Denis GV, Stephens D, Hasturk H, et al. Metabolic disease risk in children by salivary biomarker analysis. *PLoS One* 2014; 9:e98799.
137. Farook VS, Reddivari L, Chittoor G, Puppala S, Arya R, Fowler SP, et al. Metabolites as novel biomarkers for childhood obesity-related traits in Mexican-American children. *Pediatr Obes* 2015; 10:320-7.
138. West PR, Amaral DG, Bais P, Smith AM, Egnash LA, Ross ME, et al. Metabolomics as a tool for discovery of biomarkers of autism spectrum disorder in the blood plasma of children. *PLoS One* 2014; 9:e112445.
139. Whiteley P, Waring R, Williams L, Klovrsa L, Nolan F, Smith S, et al. Spot urinary creatinine excretion in pervasive developmental disorders. *Pediatr Int* 2006; 48:292-7.
140. Norata Giuseppe D, Caligiuri G, Chavakis T, Matarese G, Netea Mihai G, Nicoletti A, et al. The Cellular and Molecular Basis of Translational Immunometabolism. *Immunity* 2015; 43:421-34.
141. Kos Z, Dabbs DJ. Biomarker assessment and molecular testing for prognostication in breast cancer. *Histopathology* 2016; 68:70-85.
142. Tran NH, Cavalcante LL, Lubner SJ, Mulkerin DL, LoConte NK, Clipson L, et al. Precision medicine in colorectal cancer: the molecular profile alters treatment strategies. *Ther Adv Med Oncol* 2015; 7:252-62.
143. Fusi A, Festino L, Botti G, Masucci G, Melero I, Lorigan P, et al. PD-L1 expression as a potential predictive biomarker. *Lancet Oncol* 2015; 16:1285-7.
144. Herbst RS, Soria JC, Kowanetz M, Fine GD, Hamid O, Gordon MS, et al. Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients. *Nature* 2014; 515:563-7.
145. Patel SP, Kurzrock R. PD-L1 Expression as a Predictive Biomarker in Cancer Immunotherapy. *Mol Cancer Ther* 2015; 14:847-56.
146. Ilie M, Hofman V, Dietel M, Soria JC, Hofman P. Assessment of the PD-L1 status by immunohistochemistry: challenges and perspectives for therapeutic strategies in lung cancer patients. *Virchows Arch* 2016.
147. Hirsch S, Marshall LV, Carceller Lechon F, Pearson AD, Moreno L. Targeted approaches to childhood cancer: progress in drug discovery and development. *Expert Opin Drug Discov* 2015; 10:483-95.
148. Shi L, Zhang J, Wu P, Feng K, Li J, Xie Z, et al. Discovery and identification of potential biomarkers of pediatric acute lymphoblastic leukemia. *Proteome Sci* 2009; 7:7.
149. Russell MD, Young AM, Karri SK. Biomarkers of pediatric brain tumors. *Front Pediatr* 2013; 1:7.
150. McShane LM, Altman DG, Sauerbrei W, Taube SE, Gion M, Clark GM, et al. REporting recommendations for tumor MARKer prognostic studies (REMARK). *Nat Clin Pract Oncol* 2005; 2:416-22.
151. Whiteaker JR, Lin C, Kennedy J, Hou L, Trute M, Sokal I, et al. A targeted proteomics-based pipeline for verification of biomarkers in plasma. *Nat Biotechnol* 2011; 29:625-34.
152. Cohen Freue GV, Meredith A, Smith D, Bergman A, Sasaki M, Lam KK, et al. Computational biomarker pipeline from discovery to clinical

- implementation: plasma proteomic biomarkers for cardiac transplantation. *PLoS Comput Biol* 2013; 9:e1002963.
153. Goodsaid FM, Frueh FW, Mattes W. The Predictive Safety Testing Consortium: A synthesis of the goals, challenges and accomplishments of the Critical Path. *Drug Discov Today Technol* 2007; 4:47-50.
 154. Whiteaker JR, Zhang H, Zhao L, Wang P, Kelly-Spratt KS, Ivey RG, et al. Integrated pipeline for mass spectrometry-based discovery and confirmation of biomarkers demonstrated in a mouse model of breast cancer. *J Proteome Res* 2007; 6:3962-75.
 155. Shin H, Markey MK. A machine learning perspective on the development of clinical decision support systems utilizing mass spectra of blood samples. *J Biomed Inform* 2006; 39:227-48.
 156. Diggins KE, Ferrell PB, Jr., Irish JM. Methods for discovery and characterization of cell subsets in high dimensional mass cytometry data. *Methods* 2015; 82:55-63.
 157. O'Neill K, Aghaeepour N, Spidlen J, Brinkman R. Flow cytometry bioinformatics. *PLoS Comput Biol* 2013; 9:e1003365.
 158. Maecker HT, McCoy JP, Nussenblatt R. Standardizing immunophenotyping for the Human Immunology Project. *Nat Rev Immunol* 2012; 12:191-200.
 159. Meyer SU, Pfaffl MW, Ulbrich SE. Normalization strategies for microRNA profiling experiments: a 'normal' way to a hidden layer of complexity? *Biotechnol Lett* 2010; 32:1777-88.
 160. Yu L, Robles DT, Abiru N, Kaur P, Rewers M, Kelemen K, et al. Early expression of antiinsulin autoantibodies of humans and the NOD mouse: evidence for early determination of subsequent diabetes. *Proc Natl Acad Sci U S A* 2000; 97:1701-6.
 161. Batstra MR, van Driel A, Petersen JS, van Donselaar CA, van Tol MJ, Bruining GJ, et al. Glutamic acid decarboxylase antibodies in screening for autoimmune diabetes: influence of comorbidity, age, and sex on specificity and threshold values. *Clin Chem* 1999; 45:2269-72.
 162. Decochez K, Keymeulen B, Somers G, Dorchy H, De Leeuw IH, Mathieu C, et al. Use of an islet cell antibody assay to identify type 1 diabetic patients with rapid decrease in C-peptide levels after clinical onset. *Belgian Diabetes Registry. Diabetes Care* 2000; 23:1072-8.
 163. Pilia S, Casini MR, Cambuli VM, Ibba A, Civolani P, Zavattari P, et al. Prevalence of Type 1 diabetes autoantibodies (GAD and IA2) in Sardinian children and adolescents with autoimmune thyroiditis. *Diabet Med* 2011; 28:896-9.
 164. Lampasona V, Petrone A, Tiberti C, Capizzi M, Spoletini M, di Pietro S, et al. Zinc transporter 8 antibodies complement GAD and IA-2 antibodies in the identification and characterization of adult-onset autoimmune diabetes: Non Insulin Requiring Autoimmune Diabetes (NIRAD) 4. *Diabetes Care* 2010; 33:104-8.
 165. Grodzinsky E, Ivarsson A, Juto P, Olcen P, Falth-Magnusson K, Persson LA, et al. New automated immunoassay measuring immunoglobulin A antigliadin antibodies for prediction of celiac disease in childhood. *Clin Diagn Lab Immunol* 2001; 8:564-70.
 166. Salmi TT, Collin P, Korponay-Szabo IR, Laurila K, Partanen J, Huhtala H, et al. Endomysial antibody-negative coeliac disease: clinical

- characteristics and intestinal autoantibody deposits. *Gut* 2006; 55:1746-53.
167. Dahlbom I, Olsson M, Forooz NK, Sjöholm AG, Truedsson L, Hansson T. Immunoglobulin G (IgG) anti-tissue transglutaminase antibodies used as markers for IgA-deficient celiac disease patients. *Clin Diagn Lab Immunol* 2005; 12:254-8.
 168. Kabelitz M, Liesenkotter KP, Stach B, Willgerodt H, Stablein W, Singendonk W, et al. The prevalence of anti-thyroid peroxidase antibodies and autoimmune thyroiditis in children and adolescents in an iodine replete area. *Eur J Endocrinol* 2003; 148:301-7.
 169. Torok KS, Arkachaisri T. Autoimmune thyroiditis in antinuclear antibody positive children without rheumatologic disease. *Pediatr Rheumatol Online J* 2010; 8:15.
 170. McGhee JL, Kickingbird LM, Jarvis JN. Clinical utility of antinuclear antibody tests in children. *BMC Pediatr* 2004; 4:13.
 171. Chiang LL, Lin YT, Chan HY, Chiang BL. Differential manifestations of prepubescent, pubescent and postpubescent pediatric patients with systemic lupus erythematosus: A retrospective study of 96 Chinese children and adolescents. *Pediatr Rheumatol Online J* 2012; 10:12.
 172. Cozzani E, Drosera M, Gasparini G, Parodi A. Serology of Lupus Erythematosus: Correlation between Immunopathological Features and Clinical Aspects. *Autoimmune Dis* 2014; 2014:321359.
 173. Hugle B, Hinze C, Lainka E, Fischer N, Haas JP. Development of positive antinuclear antibodies and rheumatoid factor in systemic juvenile idiopathic arthritis points toward an autoimmune phenotype later in the disease course. *Pediatr Rheumatol Online J* 2014; 12:28.
 174. Gupta R, Thabab MM, Vaidya B, Gupta S, Lodha R, Kabra SK. Anti-cyclic citrullinated peptide antibodies in juvenile idiopathic arthritis. *Indian J Pediatr* 2010; 77:41-4.
 175. Wittemann B, Neuer G, Michels H, Truckenbrodt H, Bautz FA. Autoantibodies to nonhistone chromosomal proteins HMG-1 and HMG-2 in sera of patients with juvenile rheumatoid arthritis. *Arthritis Rheum* 1990; 33:1378-83.
 176. Yu HH, Chang HM, Chiu CJ, Yang YH, Lee JH, Wang LC, et al. Detection of anti-p155/140, anti-p140, and antiendothelial cells autoantibodies in patients with juvenile dermatomyositis. *J Microbiol Immunol Infect* 2014.
 177. Muro Y, Ishikawa A, Sugiura K, Akiyama M. Clinical features of anti-TIF1-alpha antibody-positive dermatomyositis patients are closely associated with coexistent dermatomyositis-specific autoantibodies and anti-TIF1-gamma or anti-Mi-2 autoantibodies. *Rheumatology (Oxford)* 2012; 51:1508-13.
 178. Tansley SL, McHugh NJ, Wedderburn LR. Adult and juvenile dermatomyositis: are the distinct clinical features explained by our current understanding of serological subgroups and pathogenic mechanisms? *Arthritis Res Ther* 2013; 15:211.
 179. Hacohen Y, Absoud M, Deiva K, Hemingway C, Nytrova P, Woodhall M, et al. Myelin oligodendrocyte glycoprotein antibodies are associated with a non-MS course in children. *Neurol Neuroimmunol Neuroinflamm* 2015; 2:e81.

