On the variability of the ecosystem response to elevated atmospheric CO₂ across spatial and temporal scales at the Duke Forest FACE experiment

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Abstract

While the significance of elevated atmospheric CO₂ concentration on instantaneous leaf-13 level processes such as photosynthesis and transpiration is rarely disputed, its integrated 14 effect at ecosystem level and at long-time scales remains a subject of debate. In part, the 15 uncertainty stems from the inherent leaf-to-leaf variability in gas exchange rates. By 16 combining 10 years of leaf gas exchange measurements collected during the Duke Forest 17 Free Air CO₂ Enrichment (FACE) experiment and three different leaf-scale stomatal 18 conductance models, the leaf-to-leaf variability in photosynthetic and stomatal 19 conductance properties is examined. How this variability is then reflected in ecosystem 20 water vapor and carbon dioxide fluxes is explored by scaling up the leaf-level process to 21 the canopy using model calculations. The main results are: (a) the space-time variability of 22 the photosynthesis and stomatal conductance response is considerable as expected. (b) 23 Variability of the calculated leaf level fluxes is dependent on both the meteorological 24 drivers and differences in leaf age, position within the canopy, nitrogen and CO_2 25 fertilization, which can be accommodated in model parameters. (c) Meteorological 26 variability is playing the dominant role at short temporal scales while parameter variability 27 is significant at longer temporal scales. (d) Leaf level results do not necessarily translate to 28 similar ecosystem level responses due to indirect effects and other compensatory 29 mechanisms related to long-term vegetation dynamics and ecosystem water balance. 30

Keywords: ecohydrological modeling, elevated CO₂, FACE, stomatal conductance model, spatio-temporal variability

33 *Running Title*: [Variability of ecosystem responses under elevated CO₂]

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34 **1 Introduction**

Elevated atmospheric CO_2 (eCO₂) has short and long term consequences as well as direct and indirect 35 effects on carbon and water fluxes and potentially on terrestrial biomass stocks. Specifically, the direct 36 short term effect of eCO_2 is an increase in carbon assimilation at the leaf scale, and a reduction of stomatal 37 conductance, as observed in the majority of species [Field et al., 1995; Medlyn et al., 2001; Long et al., 38 2004; Ainsworth and Rogers, 2007; Lawson et al., 2011]. Photosynthetic stimulation under eCO_2 is due to 39 the enhanced carboxylation efficiency of Rubisco (Ribulose - 1,5 - bisphosphate carboxylase oxygenase), 40 and is more pronounced in C3 than C4 plants, which are carbon limited under current ambient 41 42 concentrations of CO₂ (aCO₂) [Sage, 2004; Ainsworth and Rogers, 2007]. This leaf-level eCO₂ photosynthesis enhancement does not necessarily translate into similar increases in ecosystem carbon 43 assimilation [Ainsworth and Long, 2005; Leakey et al., 2009] or vegetation productivity and forest growth 44 [Leuzinger et al., 2011; Körner, 2013, 2015; Fatichi et al., 2014]. In the long run, the increase in carbon 45 assimilation is typically not followed by a respective increase in mineral nutrient availability, which 46 ultimately can limit ecosystem net primary production (NPP) [Oren et al., 2001; Luo et al., 2004; Finzi et 47 al., 2006; Norby et al., 2010]. Plants can also respond to this excess of carbon by down-regulating their 48 photosynthetic machinery [Paul and Foyer, 2001; Crous et al., 2008; Leakey et al., 2009; Ellsworth et al., 49 2012]. 50

Elevated atmospheric CO_2 may cause a reduction in stomatal aperture that directly leads to decreased 51 conductance to water vapor at the leaf scale [Field et al., 1995; Medlyn et al., 2001; Hetherington and 52 *Woodward*, 2003]. At the canopy scale, an increase in leaf area associated with eCO_2 increases transpiring 53 surface area but also within-canopy shading [Tor-ngern et al., 2015]. The mechanistic description of the 54 stomatal response to CO_2 and its signaling mechanism is an open question and may vary between species 55 [Mott, 1990; Assmann, 1999; Brodribb et al., 2009; Leakey et al., 2009]. For a given leaf area and forcing, 56 reduced stomatal conductance leads to (i) increased leaf temperatures due to a decrease in evaporative 57 cooling, and (ii) more favorable soil water conditions [Rawson, 1992; Hyvönen et al., 2007; De Kauwe et 58 al., 2013; Fatichi and Leuzinger, 2013; Keenan et al., 2013]. Consequently, the interplay between plant 59 responses to eCO_2 and hydrologic processes affects vegetation responses at the ecosystem scale that can 60 trigger changes in long-term global vegetation dynamics and feedbacks on climate [Betts et al., 2007; 61 62 Zaehle et al., 2007; Bonan, 2008; Sitch et al., 2008; Friedlingstein et al., 2014].

To unravel the mechanisms that affect the responses of ecosystems to eCO_2 , several free air CO_2 enrichment experiments (FACE) have been established since the early 1990s [*Owensby et al.*, 1993; *Lewin et al.*, 1994;

⁶⁵ Zanetti et al., 1996; Miglietta et al., 1998, 2001; Hendrey et al., 1999; Jordan et al., 1999; Reich et al.,

66 2001; Edwards et al., 2001; Norby et al., 2001; Okada et al., 2001; Bader et al., 2013; Drake, 2014]. The

major achievement of those experiments in reference to laboratory or chamber-based CO₂ enrichment experiments is the quantification of the effect of carbon fertilization (such as transpiration changes, enhanced water use efficiency, growth stimulation, transience of plant responses), in natural settings where most of the interactions between plant physiological, micrometeorological, hydrological and edaphic processes occur (but see [*Leuzinger et al.*, 2015]).

To overcome the limited scope of inference of FACE experiments, and their short time span (typically less 72 than a decade), terrestrial biosphere models have been used to assess the effects of eCO_2 on longer 73 timescales [Cramer et al., 1999; Krinner et al., 2005; Sitch et al., 2008; Scheiter et al., 2013]. Those models 74 integrate processes related to hydrology, plant physiology and forest demography, e.g., above- and 75 belowground carbon dynamics, species competition for light, water and mineral resources. However, the 76 theoretical potential of these models in quantifying responses of vegetation under eCO_2 and changing 77 climate can be hampered by the limited knowledge of physiological processes as well as "boundary 78 conditions" such as soil properties, vegetation composition and plant traits [Zaehle et al., 2014; Körner, 79 2015; Medlyn et al., 2015; Pappas et al., 2015a, 2015b]. 80

As these ecosystem models began confronting data from FACE experiments their predicted responses to 81 eCO_2 appeared to deviate from measurements at several time scales. Such differences have been attributed 82 to multi-species composition, local hydrological feedbacks, and feedbacks with leaf area, meteorological 83 and land surface processes such as rainfall interception and vegetation aerodynamic coupling with the 84 atmosphere [Curtis and Wang, 1998; Schäfer et al., 2002; Nowak et al., 2004; Friend and Kiang, 2005; 85 Ainsworth and Rogers, 2007; Dermody et al., 2007; Leakey et al., 2009; Williams et al., 2012; De Kauwe 86 et al., 2013; Zaehle et al., 2014; Medlyn et al., 2015]. What has not been explored is the variability in 87 responses to eCO₂ even within uniform ecosystems, which is hereafter termed as 'internal' variability. This 88 variability has both a spatial and a temporal component. The spatial variability of the responses within an 89 ecosystem can reflect differences in physiological properties among individual leaves, canopy position or 90 soil conditions, and the temporal variability of the responses relates primarily to exogenous meteorological 91 drivers (on short time scales) and endogenous shifts in slowly evolving states (e.g., leaf acclimation to 92 eCO₂, soil nitrogen depletion). The internal ecosystem variability can affect the assessment of whole 93 ecosystem level responses to eCO₂ (or at minimum its statistical significance) because such internal 94 variability must be integrated in space and time. 95

The scope of the current work is to investigate the role of this under-studied internal variability in the response of a pine plantation to eCO₂ at the leaf-level - and its up-scaled effect at the ecosystem scale. Using a combination of a decade of leaf gas-exchange and meteorological data re-analyzed for uniformity

at the Duke FACE site and stomatal and eco-hydrological modeling tools, we evaluate the variability of (a) 99 the reduction of stomatal conductance and transpiration, (b) the increase in water use efficiency, and (c) the 100 stimulation of carbon assimilation to eCO_2 at a range of temporal scales. The focus is on partitioning this 101 leaf-level scale variability in simulated fluxes into its two main sources: (1) temporal, where the effects of 102 meteorological drivers are explicitly considered; and (2) spatial, where the forest stand heterogeneity in the 103 response of photosynthetic capacity and stomatal response to eCO_2 is examined. The proposed 104 methodologies can be extended to variability of other factors such as those associated with edaphic 105 processes. However, for carbon-water relations, the spatial and temporal variability in factors describing 106 leaf-level gas exchange forms a logical starting point for all subsequent work. It is to be noted that while 107 short-term temporal variability in leaf-level photosynthetic and stomatal conductance is considered in 108 current Earth Systems Models, the importance of spatial variability and long-term trends in physiological 109 properties have been mostly neglected. 110

Data and Methods

112 **2.1 Data**

The data used were collected at the Duke FACE experiment [Schlesinger et al., 2006]. The site is located 113 in Orange County, near Durham, North Carolina, USA within the Duke Forest (79.09W, 35.98N, 168 m 114 a.s.l). The forest was established as a loblolly pine (Pinus taeda) plantation from 3-year old seedlings 115 [Pritchard et al., 2008] in 1983 after a clear cut in 1982. During the experiment, several deciduous 116 understory species were naturally established (e.g. Liquidambar styraciflua, Acer rubrum, Ulmus alata, 117 Cercis Canadensis, Cornus florida, Liriodendron tulipifera) [Schlesinger et al., 2006; Springer and 118 Thomas, 2007]. The soil in the area is a silt loam with an impermeable clay pan at about 30 cm [Oishi et 119 al., 2010]. Annual precipitation is about 1100 mm uniformly distributed across seasons and the mean annual 120 temperature is 15°C. A summary of the diurnal and seasonal fluctuations of meteorological data is given in 121 Figure 1. CO₂ fumigation was initiated in 1994 in a prototype plot and the full FACE site became 122 operational in 1996. In total, eight 30-m diameter circular plots were constructed (four aCO_2 ; four eCO_2) 123 [Feng et al., 2010]. Since 2005, the rings were split into quadrants and N fertilization (11.2 g of N m⁻²yr⁻¹ 124 as ammonium nitrate) was applied to half of them. The Duke Forest FACE experiment was terminated in 125 2010 and the site was de-commissioned. 126

During the experiment, a large number of leaf gas exchange measurements were conducted [Katul et al.,

128 2000; Crous and Ellsworth, 2004; Crous et al., 2008; Maier et al., 2008; Palmroth et al., 2013] and

129 collected in a common database [*Ellsworth et al.*, 2012]. Here, only data on loblolly pine are used. While

details on the sampling and measurement protocols can be found elsewhere (see citations above) the 130 framework common to all studies is briefly described as follows. All measurements were carried out on 131 intact leaves or leaves on cut branches (re-cut under water) using a portable photosynthesis system (Li-132 6400, Li-Cor, Lincoln, NB, USA) equipped with a standard leaf chamber. Leaves were sampled from 133 different levels inside the canopy (upper canopy sun-exposed and lower canopy shaded leaves), for all the 134 treatment combinations (CO₂, N fertilizations) and for different leaf age classes (current-year [new 135 leaves]/1-year-old [old leaves]). The dataset used here includes 485 curves, i.e. the responses of measured 136 CO_2 exchange (A_n) and transpiration rates (T) to variation in (inferred) internal CO_2 concentration or c_i 137 (hereafter referred to as $A_n - c_i$ curves). The measurements were unevenly distributed between different 138 treatment combinations and leaf age or level classes (Table 1). Meteorological and eddy-covariance CO₂ 139 and water vapor flux data were available for ambient conditions and were collected on a 22-m tall flux 140 tower (Fluxnet site US-Dk3) located within the same pine plantation from 1998 to 2008 [Oishi et al., 2010; 141 Paschalis et al., 2015]. Due to the close proximity of all replicate plots of the Duke-FACE site, the same 142 meteorological forcing is used for all of them. No major data gaps exist in the record and variable dependent 143 thresholds were used for quality control and outlier elimination in the records. 144

145 2.2 Models

The models used to describe leaf level gas-exchange and ecosystem level processes are briefly described. At the leaf level, the standard biochemical demand model for C3 photosynthesis and three different models describing the responses of stomatal conductance to environmental drivers and atmospheric CO₂ concentration are used. The outcome of these models is then introduced into a mechanistic process-based eco-hydrological model that resolves the essential hydrological and plant physiological processes at the ecosystem (or stand-level) scale.

152 2.2.1 Leaf Level Models

Net photosynthesis (A_n) is modeled according to the conventional biochemical demand formulation 153 [Farquhar et al., 1980] taking into account subsequent modifications introduced in a number of studies 154 [Collatz et al., 1991; Dai et al., 2004; Kattge and Knorr, 2007; Bonan et al., 2011]. Here, $A_n = A_g - R_d$, 155 where A_g is gross photosynthesis rate and R_d is leaf maintenance respiration (i.e. mitochondrial 156 respiration). The A_g is modeled as a function of the three limiting factors of photosynthesis: the Rubisco 157 limitation (J_c) , the light limitation (J_e) and the capacity of the leaf to export and utilize the products of 158 photosynthesis (J_s) . The term A_g depends on the biochemical parameters of photosynthesis, such as the 159 intrinsic quantum efficiency (φ), the maximum rate of Rubisco carboxylation (V_{cmax}) and the maximum 160 rate of electron transport (J_{max}) , and environmental conditions such as the absorbed photosynthetically 161

active radiation (*APAR*), the leaf temperature (T_s) and the concentration of CO₂ in the leaf intercellular space (c_i), or more precisely inside the chloroplasts (c_c) if a mesophyll conductance term is added. The term R_d is assumed to depend solely on V_{cmax} and leaf temperature. All the relevant equations used in this study can be found in full detail in [*Fatichi*, 2010; *Fatichi et al.*, 2012].

Stomata respond to environmental factors such as relative humidity (RH) or vapor pressure deficit (D), 166 atmospheric concentration of $CO_2(c_a)$ and soil moisture. The precise mechanisms that drive the regulation 167 of guard cell turgor pressure, and thus the stomatal aperture, are still a matter of open research [e.g., Buckley, 168 2005; Buckley and Mott, 2013; McAdam and Brodribb, 2014]. The mechanisms of the chemical/hormone 169 signaling for stomatal regulation are rather complex [Tardieu and Davies, 1993; Comstock, 2002; Kim et 170 al., 2010; Fatichi et al., 2016] and a commonly accepted mechanistic model for guard cell function does 171 not exist. Instead, several semi-empirical formulations that quantify the response of stomatal conductance 172 to changes in environmental conditions exist and are used in climate models. The three most common are 173 the Ball-Woodrow-Berry (or Ball-Berry) model [Ball et al., 1987], the Leuning model [Leuning, 1995], and 174 models based on optimality principles that compute stomatal conductance by maximizing carbon gain for 175 a unit of water loss [Cowan and Farquhar, 1977; Beringer et al., 1996; Mäkelä et al., 1996; Katul et al., 176 2010; Medlyn et al., 2011]. 177

178 The Ball-Berry model is defined as:

$$g_s = m_b \frac{A_n R H}{c_s} + g_0$$
 Eq 1

where m_b is a model parameter, c_s is the molar fraction of CO₂ at the leaf surface, and g_0 is a residual conductance, commonly related to imperfect stomatal closure and the cuticular conductance of leaves.

181 The Leuning model is defined as:

$$g_s = m_l \frac{A_n P_{atm}}{(pc_s - \Gamma)(1 - D/D_0)} + g_0$$
 Eq 2

where m_l is a model parameter, P_{atm} is the atmospheric pressure, pc_s is the partial pressure of CO₂ at the 182 leaf surface, Γ is the photosynthetic CO₂ compensation point expressed as a partial pressure, and D_0 is a 183 second model parameter rescaling the sensitivity of stomata to vapor pressure deficit. It has been argued 184 that the partial pressure of CO_2 in the intercellular air space (pc_i) is a more reasonable assumption than at 185 the leaf surface [Mott and Morr, 1988; Assmann, 1999]. Here, the formulation using pc_s is retained for 186 comparison with previous results obtained for the same site [Katul et al., 2010]. However, we repeated the 187 entire analysis using pc_i (not reported here), and the results were virtually the same due to the strong 188 correlation between pc_i and pc_s . 189

190 The last model considered is based on the maximization problem

$$\operatorname*{argmax}_{g_{S}}\left(\int A_{n}dt - \lambda \int Tdt\right)$$
 Eq 3

where T is the transpiration rate and λ is a model parameter. When water availability does not vary 191 appreciably over the integration period, the integral operator is no longer necessary and the maximization 192 over the entire integration period can be achieved by maximizing the instantaneous Hamiltonian $A_n - \lambda T$ 193 as shown in Manzoni et al. [2013]. The optimization problem states that stomatal conductance adjusts such 194 that there is maximum carbon gain for a given water loss. The parameter λ is the Lagrange multiplier of the 195 optimization problem and can be shown to be equivalent to the marginal water use efficiency $\lambda =$ 196 $(\partial A_n/\partial g_s)/(\partial T/\partial g_s)$ [Manzoni et al., 2011b]. For the constant (or slowly evolving with respect to g_s) λ , 197 analytical relations between g_s and leaf metabolism, environmental conditions and c_s can be derived [Katul 198 et al., 2010; Manzoni et al., 2011b; Medlyn et al., 2011; Vico et al., 2013]. All of them predict a linear or 199 quasi-linearized scaling of $g_s \propto A_n c_s^{-1} D^{-1/2}$. Other optimality models, such as those that assume leaves 200 minimize the summed unit costs of transpiration and carboxylation predict similar responses [Prentice et 201 al., 2014] and are not treated here. However, to include all the limitations such as J_s and ensure that the 202 instantaneous Hamiltonian remains positive for all environmental conditions, especially when 203 photosynthetic limitations shift, the maximization problem is solved numerically using a steepest descent 204 algorithm (see also [Bonan et al., 2014]). 205

206 2.2.2 Ecosystem Model

To scale up the effects of eCO₂ from leaf to ecosystem, the ecohydrological/ecosystem model "Tethys-Chloris" (T&C hereinafter) [*Fatichi et al.*, 2012; *Fatichi and Leuzinger*, 2013; *Fatichi and Ivanov*, 2014; *Paschalis et al.*, 2015, 2016; *Pappas et al.*, 2016] is employed. T&C resolves the energy and water balance in the soil and at the land surface and models ecosystem vegetation dynamics. Carbon pools, energy exchanges, and water stores and fluxes are fully prognostic in the model.

Specifically, the model is forced with hourly meteorological variables (precipitation, temperature, incoming 212 shortwave/longwave radiation, wind speed, atmospheric pressure and relative humidity) and resolves the 213 radiation transfer through the canopy, the water fluxes in the soil and the canopy (interception, throughfall, 214 soil water flow, evaporation/sublimation and transpiration) and the carbon fluxes (photosynthesis and 215 respiration) at the canopy level. The model is fully mechanistic without adding a large degree of abstraction, 216 and most processes are modeled according to physically based formulations. The carbon and water cycles 217 are linked through stomatal regulations and the limitation that available soil water imposes on carbon 218 assimilation. The temporal dynamics of vegetation (e.g. leaf area index (LAI) evolution, biomass evolution) 219

are modeled by balancing the carbon gain (photosynthesis) and losses (respiration, tissue turnover).
 Vegetation dynamics are modeled using the concept of plant functional types (PFTs) or species-specific
 parameters. Vegetation is conceptualized as a series of carbon pools (leaves, fine roots, living sapwood,
 non-structural carbohydrates, fruits, heartwood, dead standing leaves) that evolve in time by partitioning
 the net primary production based on specific (but dynamic) allometric rules and phenology.

T&C uses the same leaf photosynthesis model earlier described. Stomatal conductance is modeled based on the Leuning formulation. A detailed model description and its entire mathematical formulation can be found elsewhere [*Fatichi*, 2010; *Fatichi et al.*, 2012] and is not repeated here.

228 2.3 Data Analysis

Parameters regulating the biochemical demand of photosynthesis were estimated using data from the leaf 229 gas exchange measurements. Only the parameters V_{cmax} and J_{max} were estimated, while the rest (e.g., 230 intrinsic quantum efficiency, temperature response in terms of activation energies and entropy factors, 231 Michaelis-Menten coefficients) were considered constant among all measured $A_n - c_i$ curves and identical 232 to the study by Paschalis et al., [2015] (intrinsic quantum efficiency $\varepsilon = 0.081 \mu mol CO_2/\mu mol photons$, 233 activation energy $H_a = 0.649$ kJ/mol, entropy factor $\Delta S = 72$ kJ/molK, Michaelis-Menten coefficients are 234 related to leaf temperature as in Fatichi, [2010]). This is a reasonable assumption since the degree of 235 variability of V_{cmax} and J_{max} is expected to be much larger in comparison to the remaining parameters 236 [*Miao et al.*, 2009]. Several methods for the estimation of V_{cmax} and J_{max} from $A_n - c_i$ curves exist [*Miao* 237 et al., 2009]. We chose to estimate the parameters by minimizing the sum of squared deviations 238 $\sum (A_n^m(V_{cmax}, J_{max}, c_i) - A_n^o(c_i))^2$, where A_n^m is the modeled net photosynthesis for an intercellular CO₂ 239 concentration c_i , and A_n^o is the observed net photosynthesis from the gas exchange measurements. With 240 this methodology, all measurements are used in the parameter estimation, instead of the common technique 241 of estimating V_{cmax} from the strictly Rubisco limited region of the curve, and J_{max} from the strictly light 242 limited region. Note that the region where co-limitations occur is ignored with this common technique. The 243 curves for which the goodness of fit $R^2 < 0.9$, or the ratio $J_{max}/V_{cmax} > 4$ were excluded from the 244 analysis. The results were grouped based on the treatments (CO₂/N fertilization), leaf age class and leaf 245 position inside the canopy (mainly height above the forest floor). Every $A_n - c_i$ curve was considered 246 independent. This assumption is reasonable given the large variability of photosynthetic capacity (i.e. V_{cmax} 247 and J_{max}) among leaves within a tree and at a given height above the forest floor [Luoma, 1997; Niinemets, 248 2007]. Inter-annual variability, which might be related to transient plant responses to the progressive 249 treatment effect, or seasonal differences were not computed [Crous and Ellsworth, 2004] in this evaluation. 250

The leaf exchange measurements were also used for the estimation of the parameters of the stomatal 251 responses. For every group, the parameters of the Ball-Berry and the Leuning model are estimated using a 252 least squares approach fitting the observations. Only values of $200 < c_s < 700$ [ppm] were used since they 253 provided a much better fit to the data (Figure 2). Very high and low values of c_s resulted in outliers. To 254 compute the variability within each group, a bootstrap sampling (i.e. random sampling with replacement) 255 of 100 realizations was used. The parameter value λ of the optimality model was computed numerically 256 approximating the marginal water use efficiency $\lambda = (\partial A_n / \partial g_s) / (\partial T / \partial g_s)$. For every gas exchange 257 curve, the λ was numerical determined using a second-order accurate central differencing given as $\lambda =$ 258 $(A_n(g_s + \delta, \theta) - A_n(g_s - \delta, \theta))/(2\delta aD)$, where $A_n(g_s, \theta)$ is the estimated net assimilation rate for the 259 photosynthesis parameters function of g_s and of a given set of environmental variables θ = 260 $[T_a, APAR, c_s, D]$ and a = 1.64 is the ratio between the molecular diffusivities of water vapor and CO₂. 261 The D is expressed in [Pa] and not as a molecular ratio for unit compatibility between this study and others 262 [e.g., Katul et al., 2010] i.e λ in [mol mol⁻¹ Pa⁻¹]. The δ is a fraction set to 1% of the measured stomatal 263 conductance g_s . Since an estimate of λ for every measurement can be calculated, there is no need for 264 bootstrap sampling in this case. In all of the cases reported, the residual conductance was neglected ($g_0 =$ 265 0). In the Leuning model, a constant value of the parameter $D_0 = 1000$ Pa was used for all cases. In the 266 supplementary material, additional results that investigated the importance of the marginal conductance g_0 267 and also the mesophyll conductance g_{mes} (i.e. conductance to CO₂ between the intercellular space and the 268 center of photosynthetic site in the chloroplasts) are provided. 269

To assess the differences between the mean values of each of the photosynthesis and stomata-related parameters (V_{cmax} , J_{max} , m_l , m_b , λ) according to the CO₂, N fertilization treatments, the leaf age class and the level of the samples within the canopy, an ANOVA was performed taking into account all the interactions. Covariation of the dependent variables (V_{cmax} , J_{max} , m_l , m_b , λ) of the ANOVA was neglected.

274 **2.4 Simulations**

The first set of simulations includes the quantification of the general patterns of responses for all three stomatal conductance formulations when they are coupled with the photosynthesis model. The second set of simulations is used for the quantification of the relative importance of the variability of model parameters and meteorological drivers on the leaf and subsequently on ecosystem responses to eCO₂.

279 2.4.1 Stomatal Conductance Model Responses to Meteorological Forcing

The effects of eCO₂ on A_n , T, and water use efficiency (defined as $WUE = A_n/T$) for various combinations of environmental forcing parameters summarized as vector $\boldsymbol{\theta}$ and for the three stomatal conductance models is analyzed here. The estimation of these responses is not trivial and depends on (a) the responses of the stomata to the metabolic and environmental conditions and (b) on the coupling between the leaf surface and the atmosphere. For this reason, the response patterns of each stomatal conductance model are first investigated separately. How these responses affect the leaf level energy balance and ultimately A_n , T, and WUE is then considered. In this first set of simulations, a full solution of the leaf energy balance coupled with the leaf biochemical module for photosynthesis and stomatal responses to environmental factors is carried out. The solution consists of an iterative scheme (a fixed point iteration for T_s), which can be computationally demanding.

- The value of A_n depends on absorbed PAR, the temperature of the leaf surface (T_s) and the CO₂ 290 concentration at the site of photosynthesis (c_c). For all the modeling experiments considered here the CO₂ 291 mesophyll conductance between the intercellular space (c_i) and the chloroplasts (c_c) is considered large 292 and thus $c_c \simeq c_i$, even though there is evidence against this argument [Niinemets et al., 2009; Flexas et al., 293 2012; 2015]. Including a constant mesophyll conductance would change the estimate of the biochemical 294 parameters (Section 2.3) but not the overall behavior of the models. The concentration c_i depends on the 295 strength of the coupling between the leaf interior and the atmosphere, which is determined by stomatal 296 conductance and T_s . The latter depends on the environmental forcing, the stomatal conductance (that affects 297 leaf evaporative cooling), and the geometric characteristics of the leaves [Vogel, 2009; Schymanski et al., 298 2013; Huang et al., 2015a] including leaf micro-roughness [Huang et al., 2015b]. Similar arguments can 299 be made for T and WUE. It clearly emerges that A_n , T, and WUE are affected by environmental drivers in 300 a nonlinear manner, and the choice of stomatal conductance model alone can be of major significance in 301 determining their values and variations. The responses of stomatal conductance for several combinations 302 of environmental factors are obtained by simultaneously solving the leaf surface energy balance and the 303 CO_2 flux from the atmosphere to the leaf interior. Moreover, λ , which depends on A_n , g_s and T, could also 304 be determined for all models as a complementary output. In the main text, the responses of λ and g_s to the 305 environmental forcing for all three models are presented in a condensed form, while in the supplementary 306 material, details concerning the responses of g_s , T, WUE, and also c_i/c_a to the environmental forcing are 307 featured. 308
- 309 2.4.2 Leaf-level Monte Carlo Simulations

To assess the effect of meteorological and parameter variability on the responses of leaves to eCO_2 , a Monte Carlo experiment was constructed. First, for each group representing a different combination of treatments, leaf age, and canopy level, a random parameter vector \boldsymbol{u} containing the photosynthetic and stomatal conductance parameters was drawn from the fitted distributions to the measured values. The distributions were assumed to be Gaussian with mean values and variances equal to the measured values, and the crosscorrelation between each parameter was neglected. Second, the leaf-level model for photosynthesis coupled

with the models for stomatal conductance was forced with the observed meteorological drivers for two 316 levels of atmospheric CO_2 concentrations (a CO_2 : 380 ppm; e CO_2 : 580 ppm) and for each of two light 317 environments: the upper canopy with sun-exposed leaves, and the lower canopy with shade-acclimated 318 leaves. For the upper canopy, PAR was set equal to the observed values above the canopy, and for the 319 bottom of the canopy, a reduction of PAR was generated assuming a Beer–Lambert light attenuation 320 $PAR_{shd} = PAR_{sun} \exp(-k \text{ LAI})$. In the leaf-level numerical experiments, values of LAI=4 and k=0.5 were 321 used as representative of the Duke pine forest [McCarthy et al., 2007]. In total, 30 different realizations 322 were sampled. In this simulation setup, only well coupled atmospheric conditions were taken into account 323 (i.e. $T_s = T_a$) for reasons that will be explained more thoroughly in the result section. 324

Three different statistics were used to quantify the effect of meteorological and parameter variability on the uncertainty of the responses to eCO₂:

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(b)
$$s_2 = \int_0^\tau T^e \mathrm{d}t / \int_0^\tau T^a \mathrm{d}t$$
, and

$$(c) s_3 = WUE^e(\tau) / WUE^a(\tau)$$

where $WUE^{a}(\tau) = \int_{0}^{\tau} A_{n}^{a} dt / \int_{0}^{\tau} T^{a} dt$, $WUE^{e}(\tau) = \int_{0}^{\tau} A_{n}^{e} dt / \int_{0}^{\tau} T^{e}$, $A_{n}^{a}(A_{n}^{e})$ is the net CO₂ assimilation under ambient(elevated) CO₂ and $T^{a}(T^{e})$ is the transpiration under ambient(elevated) CO₂. To analyze the effect of temporal scale on the leaf-level responses to eCO₂, three time scales (τ =1 hour, 1 day, and 1 year) were used. The terms s_{1} , s_{2} and s_{3} represent the eCO₂ effect on net CO₂ assimilation, transpiration and WUE, respectively.

For partitioning the variability of the responses to leaf-scale parameter and meteorological variability, a procedure similar in concept to the one-way ANOVA was used. The total variability of the responses of a statistic *s* is defined by the distribution of *s*, taking into account all the realizations (i.e. their union \cup) of the stomatal and photosynthesis parameters, i.e. the distribution of

339
$$S_{TOT} = \bigcup \{s^1(t), s^2(t), \dots, s^n(t)\}$$

for all n realizations and for every time step t, where the meteorological conditions are different. The number of time steps depends on the length of the meteorological records and on the time scale being examined. In every realization (*i* from 1 to *n*), the model parameters differ based on the Monte Carlo sampling procedure. Every time scale (τ =1 hour, 1 day, and 1 year) and every statistic (s_1 , s_2 , s_3) are examined separately. The variability due to the model parameters can be estimated by the distribution of the mean values of the responses of the statistic *s*, averaging all time steps *t*, for each realization *i*, i.e.

$$S_{PAR} = \left[\overline{s^1}, \overline{s^2}, \dots, \overline{s^n}\right],$$

where $\overline{s^i} = \frac{1}{m} \sum_{t=1}^{m} s^i(t)$. The variability introduced by the meteorological conditions can be estimated by the distribution of

$$S_{ENV} = \bigcup \{ s^1(t) - \overline{s^1} + \overline{s^{all}}, s^2(t) - \overline{s^2} + \overline{s^{all}}, \dots, s^n(t) - \overline{s^n} + \overline{s^{all}} \},$$

where $\overline{s^{all}} = \frac{1}{mn} \sum_{i=1}^{m} \sum_{i=1}^{n} S^{i}(t)$. In this study the standard deviation of the sets S_{TOT} , S_{PAR} , S_{ENV} was used as a metric of their variability.

352 2.4.3 Ecosystem Level Simulations

The purpose of this simulation is to investigate how the observed variability at the leaf scale (only) manifests itself at ecosystem scale, taking into account the major eco-hydrological feedbacks.

Given the relatively high computational demand of T&C, only margin or end-member cases were sampled. 355 Specifically, two values of V_{cmax} were selected for the upper canopy representing approximately the 25% 356 and 75% percentiles of the measured V_{cmax} for all the cases of the upper canopy estimates (i.e. the 357 percentiles derived from the pool of estimated V_{cmax} values, lumping together all subcases of nitrogen 358 treatment, carbon fertilization and age class). A differentiation in canopy-level class was not prescribed in 359 those simulations, because T&C itself simulates a decline of V_{cmax} within the canopy proportional to a 360 nitrogen decay coefficient. The leaf age class was also excluded in this experiment. Similarly, two values 361 of the Leuning model parameter were sampled such that they represent approximately the 25% and 75% 362 percentiles of the observed values between all groups (i.e. the percentiles derived from the pool of parameter 363 estimates lumping together all subcases of nitrogen treatment, carbon fertilization, age class and level 364 within the canopy). Moreover, in the ecosystem level simulations, both overstory (pines) and understory 365 (deciduous hardwood species) vegetation are explicitly simulated to mimic the real ecosystem. For the 366 understory only, a single parametrization throughout the simulations was used, given its minor contribution 367 to the overall fluxes. The model parameter values for the deciduous understory species were taken from 368 Paschalis et al., [2015]. The set of statistical analyses described above are used to quantify the effects of 369 variability in the parameter values and the environmental drivers at the ecosystem scale. 370

371 **3 Results**

372 **3.1 Leaf Level**

3.1.1 Variability in V_{cmax}, J_{max} and Stomatal Conductance Within Canopy and Across Treatments 373 The variability in V_{cmax} and J_{max} (scaled to 25°C) within each group (canopy level/leaf age/treatment) is 374 large (Figure 3). This reflects the natural spatial variability, but also may integrate variability due to 375 measurement errors, and the transient behavior of long-term acclimations given that inter-annual variability 376 of the measurements was not explicitly taken into account. Despite the large within-group variability, 377 ANOVA suggests that both V_{cmax} and J_{max} of current-year needles decrease with decreasing light 378 availability within the canopy (low p-values of L and A treatments in Table 2). Both parameters depend on 379 nitrogen content per unit leaf area, which decreases with light availability within the canopy [Niinemets et 380 al., 1998, 2015; Grassi and Bagnaresi, 2001; Warren et al., 2003]. Compared to current-year foliage, 1-381 year-old needles, in most cases, have significantly lower V_{cmax} and J_{max} . Finally, V_{cmax} and J_{max} appear 382 non-responsive to eCO_2 (Table 2). This contradicts earlier arguments that a down-regulation of the 383 photosynthetic machinery of plants takes place under long-term exposure to eCO₂ [e.g. Medlyn et al., 1999]. 384 At the Duke FACE site, previous research suggested that plant structural adjustments occur after long 385 exposure to eCO₂, such as changes in the relation between leaf nitrogen concentration and photosynthetic 386 capacity or V_{cmax}, particularly in 1-year-old needles [Crous et al., 2008; Maier et al., 2008]. However, the 387 non-responsiveness of V_{cmax} and J_{max} has also been documented by Crous & Ellsworth [2004] who 388 identified a lack of a strong, statistically significant reduction of V_{cmax} and J_{max} at the Duke FACE. It 389 should be highlighted that this lack of response might be influenced by the transient long-term acclimation 390 to eCO_2 which was not taken into account (or at least can partially mask eCO_2 effect). Also, the potential 391 seasonal fluctuations of V_{cmax} at 25 C, were not taken into account [Zhang at al., 2014b], even though there 392 is contradictory evidence whether V_{cmax} varies strongly on a seasonal basis in the Duke Forest [Ellsworth, 393 2000; Juang et al., 2006; Ellsworth et al., 2012]... 394

The parameters of the stomatal responses to environmental factors were statistically different among the 395 various groups (Figure 4) as shown from the low p-values in ANOVA results in Table 2. However, these 396 differences are difficult to interpret as they are influenced by two major factors. First, there is considerable 397 uncertainty in the parameter estimation itself (Figure 2) [see also Yu et al., 2004; Medlyn et al., 2011]. 398 Second, all the results were obtained assuming that the marginal or residual conductance was equal to zero, 399 $g_0 = 0$, and different patterns are obtained when this assumption is removed as theoretically considered 400 elsewhere [Manzoni et al., 2011b]. For these reasons, we refrain from providing detailed explanations 401 402 regarding the among-groups differences in the values of these parameters, especially when patterns are not univocal. The results of the parameter estimation of all three g_s models (with $g_0 = 0$) are used in the following only to assess their variability. It is to be noted that the degree of variability between the g_s model parameter estimation when accounting for g_0 is comparable to those with $g_0 = 0$, despite the fact that parameter values are different (analysis not shown here).

407 3.1.2 Differences in Stomatal Conductance Model Behavior

The response of stomatal conductance as predicted by all three g_s models (Eqs 1-3) depends on the 408 environmental factors as shown in Figure 5 and in the Supplementary material. The key characteristic of 409 all the responses are: (a) a bell-shaped response in g_s to temperature; (b) a steep increase in g_s with 410 increased absorbed PAR, reaching a plateau for high irradiance; (c) a minimal influence of wind speed; (d) 411 a positive relation between g_s and relative humidity; and (e) a reduction in g_s under eCO₂. The first two 412 observations can be fully explained by the response of A_n to T_s and PAR, and the dependence of g_s on A_n . 413 The fact that wind speed (W_s) has minimal influence can be explained by the leaf shape (needles) that does 414 not allow the development of a thick laminar boundary layer [Huang et al., 2015a]. For this reason, only 415 well-coupled conditions were assumed in the subsequent Monte Carlo simulations. 416

The major functional difference between all the models is the response of g_s to RH. The Ball-Berry model 417 prescribes a linear relation between those variables (Eq. 1), in contrast to the Leuning model that expresses 418 g_s as a function of D. The g_s response to D is not a priori imposed and is an emergent outcome of the 419 stomatal optimization. These two models result in a nonlinear dependence of g_s on RH. This dependence 420 of g_s on RH results in similar responses of A_n , T, c_i/c_a and WUE between the optimality and Leuning 421 model formulations, but significant differences in comparison to the Ball-Berry model (Supplementary 422 423 material). For this reason, in the following, only two models (Ball-Berry and Leuning) are used for further analysis, given their current popularity in Earth System Models and the functional similarity between the 424 Leuning model and the optimality model (Figure 5). 425

Another difference between the three models is the estimated value of the marginal water use efficiency λ . 426 Roughly, λ is interpreted as the cost of water loss from stomata in units of carbon. It is of interest here 427 because it bridges the water and carbon economies of plants at the leaf scale. As λ increases, carbon 428 becomes easier to acquire, "cheaper", and at the same time water loss becomes more "expensive" [Katul et 429 al., 2009, 2010; Manzoni et al., 2011a, 2011b]. By definition, λ in the optimality model is constant on time 430 scales over which stomatal aperture fluctuates (usually sub-daily). In the Ball-Berry and Leuning models, 431 a value of λ can be computed and depends on both the environmental drivers and the level of CO₂ 432 enrichment. The λ patterns are substantially different between the models (Figure 6). The two g_s models 433 predict a relatively steady λ for a wide range of relative humidity conditions, suggesting close to optimal 434

behavior. This range of relative humidity (0.2 < RH < 0.9) corresponds to the most common values 435 expected for temperate and continental climates. However, discrepancies in predicted λ between the 436 Leuning and the Ball-Berry models appear at the RH extremes. In general, the Ball-Berry model predicts a 437 high value of λ for low RH, which suggest that this model assigns a "high" cost of water in a water-carbon 438 cost-benefit perspective, and for this reason, the predicted stomatal conductance for this model is much 439 lower (Supplementary material) than the other two g_s models at low RH. In contrast, the Leuning g_s model 440 predicts a high value of λ at high RH, suggesting lower than optimal stomatal conductance at low D (Figure 441 5). Interestingly, both g_s models predict a higher value of λ with increased temperature. Finally, the λ 442 computed from the Leuning modeled is more sensitive than its Ball-Berry counterpart in terms of $|\partial \lambda / \partial c_a|$. 443

444 3.1.3 Sensitivity to spatial and temporal variability

The key results from the leaf-level simulations are: (a) at short temporal scales (e.g. hourly) the main source 445 of the variability in the responses to eCO_2 is the environmental forcing; (b) as temporal scales increase, the 446 influence of variability in model parameters becomes predominant; and (c) the choice of the stomatal 447 conductance model can result in appreciable differences in the mean values of the responses of A_n , WUE, 448 T and their variability (Table 3, Figure 7). Here, the results of the leaf level variability of the responses to 449 eCO₂ for all combinations of treatments are featured (Table 3, Figure 7), whereas in the Supplementary 450 material, the case-by-case analysis is presented. As a metric of the variability of the responses to eCO₂, we 451 chose the standard deviation of the sets S_{TOT} , S_{PAR} and S_{ENV} (Table 3). In Figure 7, the variability of the 452 responses to eCO₂ can be interpreted by the range of the respective box plots. 453

Specifically, the total variability of the net CO₂ assimilation effect at the hourly time scale varies from 1-454 1.7 for the Ball-Berry g_s model and from 1-1.5 for the Leuning g_s model. For this estimation, only the 455 hours when A_n^e and $A_n^a > 1 \mu \text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ were used. This variability can be explained almost entirely by 456 the environmental forcing. Furthermore, it is trivial to show that the ratio of the instantaneous water use 457 efficiency (WUE) in eCO₂ and aCO₂ for the Ball-Berry model equals the ratio of the concentrations 458 [eCO₂]/[aCO₂] and is independent of model parameters and environmental forcing (for the Ball-Berry 459 model and $g_0 = 0$, the instantaneous $WUE = A_n/g_s D = c_s/m_b RH$ and thus $WUE^e/WUE^a =$ 460 [eCO2]/[aCO2]). For the Leuning model, the only contribution to the variability of the ratio of WUE comes 461 from the dependence of the CO₂ compensation point Γ on leaf temperature, and thus for the instantaneous 462 ratio of WUEs, the only source of variability originates from the environmental forcing. Finally, concerning 463 the transpiration reduction under eCO_2 at the hourly time scale, the variability of the reduction ratio is larger 464 for the Ball-Berry model, while its mean value is smaller. This result seems rather insensitive to the light 465 environment and is the same for the upper and lower levels of the canopy. 466

As the temporal scale increases, the variability of the responses diminishes primarily because the environmental forcing variability is reduced by the longer averaging interval. Conversely, the variability in model parameters affects all temporal scales. In other words, at the hourly scale, almost the entire variability depends on the environmental forcing, but at longer scales (e.g. the inter-annual scale), the variability of the responses depends progressively more on model parameter variability.

The leaf-level responses to eCO_2 have a pronounced seasonal and diurnal cycle (Figure 8). Carbon assimilation enhancement due to eCO_2 is larger during the summer according to both g_s conductance models. Environmental variables such as temperature and radiation are most favorable during this period (Figure 1), strengthening the effect of higher atmospheric CO₂ on photosynthesis. The effect of eCO_2 on transpiration reduction is more pronounced during winter. The composite effect of seasonality in meteorology and the reduced net CO₂ assimilation during winter result in a distinct seasonal pattern of transpiration reductions due to eCO_2 .

The diurnal cycle of A_n enhancement and T reduction predicted by both g_s models is similar. The effect 479 of eCO₂ on the transpiration reduction is less strong during midday, when A_n is larger. The only difference 480 in terms of diurnal patterns is the late afternoon drop in A_n stimulation (and T reduction) predicted by the 481 Leuning model. The reason for this is the sensitivity to D and the temporal lag of vapor pressure deficit D 482 with PAR and T_a (mainly due to boundary-layer growth and heat/water vapor storage as discussed in 483 [Matheny et al., 2014; Zhang et al., 2014]). Given that the temporal shift between RH and D to T_a and PAR 484 is not the same, the difference between the two model predictions of A_n stimulation and T reduction can 485 be attributed to differences in the diurnal cycles of *RH* and *D*. 486

487 **3.2 Ecosystem Level**

488 3.2.1 T&C Simulation Results

Beyond the direct effects and the role played by of environmental drivers on CO₂ and water fluxes [Holtum 489 and Winter, 2010; De Kauwe et al., 2013], several indirect effects exist when upscaling from leaf to 490 ecosystem level. Those effects can create compensatory mechanisms between the enhancement of net 491 assimilation and reduction of water and carbon fluxes that complicate such upscaling [Fatichi et al., 2016b]. 492 For instance, it is noted that eCO₂ can result in leaf area index (LAI) increases due to NPP enhancement 493 [Kergoat, 2002; Norby et al., 2005; Dermody et al., 2007; McCarthy et al., 2007]. Increased foliage area 494 results in larger transpiration rates that can deplete soil water, and increases canopy interception, which can 495 further reduce water availability in the soil. However, the eCO_2 may also lead to an increased WUE, 496 primarily due to reduced transpiration per unit leaf area that can lead to favorable soil water conditions, and 497 thus result in an even stronger stimulation of productivity. The degree to which the effects of decreased 498

transpiration per unit leaf area and the increased foliage area compensated for each other is a difficultquestion that can be addressed by ecosystem models.

To assess how such indirect effects impact the variability of photosynthesis stimulation, transpiration 501 reduction, NPP and WUE increase at the ecosystem level due to elevated atmospheric CO₂, the T&C model 502 was used. The model performs reasonably well in reproducing the effects of eCO₂ on water and carbon 503 fluxes as well as vegetation dynamics at the Duke Forest (Supplementary Material). In Figure 9, the indirect 504 effects of eCO₂ at the ecosystem level are shown. In general, eCO₂, leads to a ~20% increase in leaf area 505 (comparable to observed changes [McCarthy et al., 2006, 2007], which in turn reduces the fraction of the 506 foliage exposed to direct light. It also leads to higher modeled soil moisture values, although differences 507 between the CO_2 treatments are unlikely to be large enough to modify plant water stress for most of the 508 time. The effect of increased WUE on soil moisture is more pronounced during periods of drought. Finally, 509 there is an increase in water losses due to enhanced evaporation from interception for an eCO_2 state, but 510 those losses are low in comparison to the total precipitation at the Duke Forest. The strength of the CO_2 511 effect is dependent on the model parameters, with V_{cmax} being more influential than the parameters related 512 513 to stomatal conductance.

The effects of the meteorological and parameter variability on the variability of carbon assimilation 514 enhancement, transpiration reduction, and WUE increase are similar in magnitude and patterns to the leaf 515 level results (Figure 10). In particular, most of the variability at the short temporal scales can be explained 516 by the environmental drivers, whereas model parameter variability can explain most of the variability of 517 the ecosystem responses in the longer-term. The range of temporal scales at which high-frequency 518 meteorological variability can influence long-term variability depends on the potential of the 519 meteorological forcing to be integrated into slow dynamics such as soil moisture dynamics [Paschalis et 520 al., 2015]. For the Duke Forest site, where water availability is rarely stressing vegetation, high-frequency 521 meteorological forcing cannot significantly impact soil water stress, and this is the reason why there is a 522 reduction in the contribution of meteorological variability with scale. For the pines, the magnitude of the 523 response at both the leaf and ecosystem levels is similar. This means that either the indirect effects of LAI, 524 soil moisture and interception are minimal, or they compensate each other. Fatichi et al., [2016b] showed 525 that the importance of these indirect effects is negatively dependent on the wetness index (annual 526 precipitation/annual potential evapotranspiration) of the site. Given the large wetness index of the Duke 527 FACE, the indirect effects are likely small in this location. It should be noted that in contrast to the leaf 528 level results, at the ecosystem level, the mean values of the response at the annual scale are different from 529 the mean value of the responses at the short (hourly and daily) scales. This effect can be explained by the 530

fact that the annual scale integrates the effect of the dynamically evolving leaf area, and thus the phenology
of the vegetation, which is absent from the leaf level results.

Finally, there is a difference between the responses of the pines and the hardwood species in the simulations. For the hardwood species, the eCO₂-induced reduction in transpiration is smaller and this leads to a smaller WUE enhancement in comparison to the pines. The reason for this behavior is two-fold. First, the reduction in transpiration is larger during winter in Duke FACE (Figure 8), when the hardwood species shed their leaves. Secondly, the hardwoods have a smaller degree of coupling between the leaf and the atmosphere due to their larger leaf sizes that increases the leaf boundary layer resistance, as shown by other studies [*De Kauwe et al.*, 2013; *Medlyn et al.*, 2015; *Huang et al.*, 2015a].

540 **4 Discussion**

The uncertainty of leaf and ecosystem level responses to eCO₂ was explored using long-term gas exchange measurements and models for leaf- and ecosystem scales. The goal was to shed light on the uncertainty of the responses across temporal scales due to variability in meteorological drivers, the uncertainty of the parameters related to leaf biochemistry and stomatal conductance, and the functional relation used to link stomatal conductance and net assimilation. Potentially, other factors can be included (e.g. soil hydraulic properties, root-density distribution, etc.) but those selected here are deemed to be common to most of the Earth System models.

The work addressed the consequences of this variability as guided by a set of questions pertinent to the development of the next generation of ecosystem models and FACE experiments: (a) Are there essential differences between various modeling parametrizations for stomatal responses to environmental factors? (b) How reliable are the observed changes in carbon assimilation enhancement, transpiration reduction and acclimation of plant to eCO₂ given such 'internal' ecosystem variability? (c) How can this variability be incorporated in the next generation of dynamic vegetation models?

Concerning (a), from the results of the present research, it is apparent that the selection of the model that 554 describes the responses of stomata to environmental drivers and atmospheric CO₂ concentrations is crucial. 555 Despite the equivalently good fit of all g_s models to the observed data (Figure 2), significant differences 556 between the results exist depending on the model selection. It has been recently shown that the relation 557 between stomatal conductance and net CO₂ assimilation can have strong implications for ecosystem 558 vegetation dynamics and potentially feed-back on the global climate [Bonan et al., 2014; Sato et al., 2015]. 559 Note, for instance, that all the existing models decrease stomatal conductance in response to eCO_2 560 regardless of the parameter values or environmental forcing (Figure 5). Measurements show that for certain 561

species, the g_s response to eCO₂ may be small or entirely absent [*Brodribb and McAdam*, 2013], even 562 though contradictory results also exist [Franks and Britton-Harper, 2016]. Furthermore, even when 563 accounting for the observed variability of responses as derived from observations, the value of 564 WUE^{e}/WUE^{a} is almost prescribed in current models (Figure 7). These findings reinforce the quest for 565 mechanistic representation of stomatal conductance [Damour et al., 2010; Medlyn et al., 2011; Fatichi et 566 al., 2016c]. Several recent studies have attempted mechanistic description of stomatal aperture regulation 567 [Buckley et al., 2003; Peak and Mott, 2011; Mott and Peak, 2013], which can eventually eliminate 568 ambiguities associated with empirical (Ball-Berry and Leuning) or phenomenological (optimality) models. 569 However, a mechanistic model of stomatal response to environmental drivers and plant water status suitable 570 for imminent implementation in Earth System models is not available yet, despite its desirability. 571

Concerning (b), the results here show that a detailed characterization of leaf- and ecosystem-level responses 572 to eCO_2 cannot be separated by temporal scale over which such responses are being evaluated. Specifically, 573 the variability of the responses to eCO_2 at short temporal scales, commonly referred to as instantaneous, is 574 dependent on meteorological variability. Given that meteorological variability is 'irreducible' in a natural 575 system, long-term data are essential for robust quantification of 'instantaneous' ecosystem responses (in a 576 statistical sense). At longer temporal scales, the primary source of variability originates from the variation 577 of the parameters describing the photosynthetic capacity of leaves and the sensitivity of their stomatal 578 responses to environmental drivers. Given the high leaf-to-leaf variance in such parameters even at small 579 spatial scales, as shown in the work here, it becomes necessary to evaluate the robustness of the ecosystem 580 responses at long (e.g. annual) time scales. At such time scales, variability in environmental drivers is 581 reduced but variability in leaf parameters persists and introduces significant uncertainty in fluxes and stores. 582 It should be noted that in the present study, the parameter variability ought to be limited since the study 583 region was a uniform single-species plantation. In unmanaged ecosystems, species diversity and the uneven 584 age of the plants would enhance the variability contribution of the "spatial" uncertainty of the parameters 585 [e.g., Pappas et al., 2016]. 586

The issue of spatial parameter heterogeneity and/or of subtle underlying trends in the parameters along with 587 the large variability in boundary conditions (e.g., species diversity, soil hydraulic properties) can hamper 588 statistical inference of response signals to eCO₂, especially at the ecosystem scale where direct and indirect 589 effects on plant productivity and hydrology co-exist [Holtum and Winter, 2010; Piao et al., 2013; Zaehle 590 et al., 2014; Fatichi et al., 2016b]. For this reason, discrepancies between the results of various FACE 591 experiments should not be surprising [Nowak et al., 2004; Körner, 2006; Norby and Zak, 2011; De Kauwe 592 et al., 2013] and likely only a combination of data analysis from field studies and modeling may identify 593 the causes for different responses and the real uncertainty bounds of the results. 594

Concerning (c), modeling procedures that incorporate both meteorological variability and the spatial 595 variability in the photosynthetic capacity, stomatal responses to environmental variables and generally all 596 the parameters related to plant functioning is essential. While meteorological variability is already 597 incorporated in most modeling studies at the hourly or sub-hourly scale, the spatial variability of the 598 parameters related to plant functioning is more difficult to address. The emergence of trait-based approaches 599 in vegetation modeling can be regarded as a reasonable solution [Scheiter et al., 2013; Bodegom et al., 600 2014; Pappas et al., 2016]. Such an approach can inherently incorporate the variability and stochasticity of 601 the plant traits within and between ecosystems. Using such approaches to quantify variability of the 602 responses of ecosystems is also straightforward, since the deterministic framework of PFTs is loosened and 603 diversity/variability of plants traits can be explicitly taken into account with stochastic analyses. This 604 approach has only been recently incorporated into large-scale ecosystem models but it is promising to 605 address uncertainty of spatially and potentially temporally variable parameters [Pavlick et al., 2013; 606 607 Scheiter et al., 2013; Sakschewski et al., 2015].

608 **5** Conclusions

The leaf and ecosystem level variability of the responses of carbon gain, transpiration and water use 609 efficiency to elevated CO2 was considered for a wide range of temporal scales at the Duke FACE 610 experiment. Using an extensive data set consisting of more than 500 $A_n - c_i$ curves collected over a 10 611 year period, we estimated the variability of the parameters related to the photosynthetic machinery of the 612 leaves and the response of their stomata to environmental drivers, and the impacts of carbon and nitrogen 613 fertilization on those parameters. Combining the results of the data analysis with modeling approaches at 614 the leaf (three stomatal conductance models; Ball-Berry, Leuning and optimality model) and the ecosystem 615 scale model T&C, the variability of the responses due to meteorological drivers and model parameter 616 uncertainty was partitioned and analyzed. 617

618 The key findings are:

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to environmental drivers and CO_2 concentrations is large, complicating the identification of transient patterns, such as acclimation to eCO_2 or nutrient feedbacks, and suggesting that extensive data collection is essential for such identification, even in a uniform plantation.

(a) The variability of the parameters describing photosynthetic capacity and the responses of stomata

(b) While the three commonly used stomatal conductance models describe the gas-exchange
 measurements reasonably for ambient and enriched atmospheric CO₂, there are fundamental
 differences between their responses to environmental drivers and their interpretation of the

626 627 marginal water use efficiency. Those differences may lead to important long-term consequences in simulated plant water stress and thus vegetation growth and mortality.

- (c) The variability in the responses to elevated CO₂ of photosynthesis stimulation, transpiration
 reduction and water use efficiency enhancement depends on the scale at which the responses are
 being evaluated. Variability in leaf-level fluxes at short temporal scales can be mainly attributed to
 meteorological drivers. As temporal scales increase, the contribution of the uncertainty in the model
 parameters becomes predominant.
- (d) The ecosystem-level responses to elevated CO₂ were comparable to those at the leaf level at the
 Duke FACE. Indirect effects related to the impact of leaf-level photosynthesis stimulation and
 stomatal conductance reduction, such as LAI increase and more favorable soil water conditions
 exist, however, compensatory effects tended to weaken these indirect effects, producing an overall
 response similar to the leaf-level response.

Quantification of the effects of increased atmospheric CO₂ concentrations are essential for understanding the coupled bidirectional effects of anthropogenic carbon emissions on global vegetation and climate. As both stomatal regulation in response to environmental drivers and internal ecosystem variability play major roles, the evidence presented in this study may spur research for a mechanistic models of stomatal functioning, which will be essential for the next generation of terrestrial ecosystem models.

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	Ambient CO ₂				Elevated CO ₂			
	Upper		Lower		Upper		Lower	
	New	Old	New	Old	New	Old	New	Old
no N Fertilization	53	79	45	37	33	64	26	34
N Fertilization	31	9	29	7	10	9	9	9

Table 1: Number of $A - c_i$ curves included at the data analysis

Table 2: p-values of the 4-way ANOVA for V_{cmax} , J_{max} , m_b , m_l , λ , according to the CO₂ fertilization (C), Nitrogen fertilization (N), Canopy level (L) and Age class (A). Values below 5%, where the difference of the mean values of each of the dependent variables (V_{cmax} , J_{max} , m_b , m_l , λ) are statistically significant based on the discrete dependent variables (C, N, L, A) marked as bold. Covariation between the dependent variables is neglected.

	V _{cmax}	J_{max}	m_b	m_l	λ
С	0.8198	0.8632	<0.0001	<0.0001	0.2187
Ν	0.9713	0.0348	<0.0001	<0.0001	<0.0001
L	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
А	0.0002	<0.0001	<0.0001	<0.0001	<0.0001
C*N	0.2814	0.2215	<0.0001	<0.0001	0.0283
C*L	0.6183	0.5745	<0.0001	0.0003	0.2553
C*A	0.8797	0.9018	<0.0001	<0.0001	<0.0001
N*L	0.2490	0.5534	<0.0001	<0.0001	0.0057
N*A	0.1084	<0.0001	<0.0001	<0.0001	0.4524
L*A	0.0011	0.1440	<0.0001	<0.0001	0.2224
C*N*L	0.4530	0.9175	<0.0001	<0.0001	0.8188
C*N*A	0.0972	0.9791	0.0022	<0.0001	0.3420
C*L*A	0.8968	0.5492	<0.0001	<0.0001	0.0565
N*L*A	0.1342	0.8288	<0.0001	<0.0001	0.2688
C*N*L*A	0.6623	0.3119	<0.0001	0.4495	0.8237

Table 3: Standard deviations of the simulated ratios of net photosynthesis (A_n) , water use efficiency (WUE) and transpiration (T) for two canopy levels (upper sun leaves, lower shaded leaves), three time scales (1 hour, 1 day, 1 year) according to the Ball-Berry and the Leuning stomatal conductance model. σ_{TOT} , σ_{PAR} , σ_{ENV} represent the standard deviations of the sets S_{TOT} , S_{PAR} , S_{ENV} respectively. The three sets lump the variability due to CO₂ and N treatments and leaf age. The same data used in this table are used for the illustration of the variability of the responses in Figure 7.

	A_n^e/A_n^a			W	WUE ^e /WUE ^a				T^e/T^a	
	σ_{TOT}	σ_{PAR}	σ_{ENV}	σ_{TOT}	σ_{PAR}	σ_{ENV}	σ_{TOT}	σ_{PAR}	σ_{ENV}	
	Sun Leaves									
Ball-Berry										
1 hour	0.240	0.042	0.236	0.000	0.000	0.000	0.162	0.036	0.158	
1 day	0.166	0.037	0.162	0.036	0.010	0.034	0.182	0.032	0.179	
1 year	0.039	0.035	0.018	0.022	0.018	0.013	0.040	0.036	0.018	
Leuning										
1 hour	0.149	0.020	0.148	0.025	0.000	0.025	0.086	0.016	0.084	
1 day	0.113	0.018	0.112	0.017	0.006	0.016	0.074	0.014	0.073	
1 year	0.022	0.019	0.011	0.014	0.012	0.006	0.021	0.018	0.009	
	Shaded Leaves									
Ball-Berry										
1 hour	0.171	0.038	0.168	0.000	0.000	0.000	0.117	0.034	0.113	
1 day	0.594	0.290	0.559	0.200	0.065	0.190	0.872	0.502	0.791	
1 year	0.052	0.049	0.018	0.055	0.023	0.050	0.053	0.050	0.020	
Leuning										
1 hour	0.112	0.018	0.110	0.025	0.000	0.025	0.063	0.015	0.062	
1 day	0.096	0.019	0.094	0.025	0.003	0.025	0.074	0.013	0.072	
1 year	0.018	0.015	0.009	0.010	0.008	0.006	0.016	0.014	0.009	

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Figure 1: Meteorological forcing observed at the Duke Forest. In the left panel the diurnal cycle air temperature (T_a), windspeed (W_s), photosynthetic active radiation (*PAR*) and vapor pressure deficit (*D*) for summer (June-July-August), and in the left panel the seasonal cycle of the same variables is shown. Axes colors (black, blue purple and red) correspond to the color lines of the respective variables (T_{α} , W_s , *PAR*, *D*).



Figure 2: Typical agreement between measured and modeled stomatal conductance (g_s) for the three stomatal conductance models. Grey points represent the entire data set with no filtering, whereas red dots represent only the data points for which the CO₂ concentration at the leaf surface (c_s) was in the range: 200 ppm< c_s < 700 ppm.



Figure 3: Boxplots showing the variability of V_{cmax} (upper panels) and J_{max} (lower panels) at 25°C for every class of CO₂, N fertilization, leaf age and canopy level. Boxes refer to the 25%-75% percentiles and whiskers to the 5%-95% percentiles. The left panels represent plants grown under ambient CO₂, and the right panels plants grown under elevated CO₂. Panels marked as (A) represent new needles and (B) old needles. Blue background color represents unfertilized and yellow background color represents N-fertilized trees. Blue boxplots represent measurement taken at the upper part of the canopy and grey boxplot measurements at the lower part.



Figure 4: Same as Figure 3, but for the parameters related to the stomatal conductance models. m_b is the parameter of the Ball-Berry stomatal conductance model, m_l is the parameter related to the Leuning model and λ is the marginal water use efficiency used in the optimality model.



Figure 5: Modeled stomatal conductance according to the Leuning (upper panels), Ball-Berry (middle panels) and the optimality model (lower panels). For each subplot all meteorological variables were held constant and equal to the reference state ($T_a=25^{\circ}$ C, $PAR = 200 \text{ Wm}^{-2}$ (~915 μ mol m⁻² s⁻¹), $W_s = 2 \text{ ms}^{-1}$, RH = 0.6 [-]), except the variable of interest featured on the corresponding x-axis. The parameters used are: $V_{cmax} = 80 \text{ }\mu\text{mol }\text{m}^{-2}\text{s}^{-1}$, $J_{max} = 160 \text{ }\mu\text{mol }\text{m}^{-2}\text{s}^{-1}$, $m_l = 6$, $m_b = 5$, $\lambda = 10^{-8} \text{ [mol mol}^{-1}\text{Pa}^{-1}$]



Figure 6: Marginal water use efficiency λ [µmolCO₂ molH₂O⁻¹Pa⁻¹] (for this calculation the unit convention follow [*Katul et al.*, 2010]) as predicted by the Leuning and the Ball-Berry models for several combinations of meteorological forcing (left panels) and CO₂ concentrations (right panels).



Figure 7: Boxplots representing the total variability (left green boxplots), variability due to model parameters only (middle purple boxplots) and variability due to meteorological forcing only (right blue boxplots) of the ratios $\frac{A_n^e}{A_n^a}, \frac{WUE^e}{WUE^a}, \frac{T^e}{T^a}$, estimated based on the Ball-Berry and the Leuning models for A_n , WUE, and T due to eCO_2 integrated at the time scales of 1 hour (upper panels), 1day (middle panels) and 1 year (lower panels) for leaves exposed to full light (left 2 panels) and shaded leaves (right 2 panels).



Figure 8: Median values of the diurnal (upper panels) and seasonal (lower panels) cycles of the net assimilation stimulation (left) and transpiration reduction (right) modeled using the Ball Berry (red lines) and the Leuning (black lines) model, for the hourly (continuous lines) and daily (dashed lines) time scale.



Figure 9: Modeled temporal evolution of the eCO2 effects: (a) LAI enhancement, (b) reduction of the fraction of canopy area exposed to sun, (c) soil moisture enhancement (d) increase in evaporation losses from interception for the overstory pines. The various combinations of V_{cmax} and m_l are shown in the legend. Arrows pointing up relate to high values and arrows pointing down to low values of the respective parameter. The blue errorbars in (a) represent the time evolution of the observed LAI enhancement. The range of the errorbars represent the standard deviation of the measured LAI enhancement within a year. The arrow in (a) represents the occurrence of an ice storm that caused damages to the vegetation.



Figure 10: Ecosystem level variability of the responses of A_n , WUE, aboveground NPP and T for the overstory pines (left panel) and the understory hardwood species (right panel) as simulated by the T&C model. The boxplot represent the total variability for 3 different time scales (1hour, 1day and 1 year) and the dots the mean value of the ecosystem response according to the combination of the V_{cmax} and m_l parameters. The dashed lines represent the average response of the pines.