

JNCI 15-1761R2

Article

**Evaluation of polygenic risk scores for breast and ovarian cancer risk prediction in
BRCA1 and *BRCA2* mutation carriers**

Karoline B. Kuchenbaecker, PhD^{0,1}

Lesley McGuffog,¹

Daniel Barrowdale, BSc¹

Andrew Lee,¹

Penny Soucy, PhD²

Sue Healey, BSc³

Joe Dennis, MSc¹

Michael Lush, PhD¹

Mark Robson, MD⁴

Amanda B. Spurdle, PhD⁵

Susan J. Ramus, PhD⁶

Nasim Mavaddat, PhD¹

Mary Beth Terry, PhD⁷

Susan L. Neuhausen, PhD⁸

Ute Hamann, PhD⁹

Melissa Southey, PhD¹⁰

Esther M. John, PhD¹¹

Wendy K. Chung, MD PhD ¹²

Mary B. Daly, MD ¹³

Saundra S. Buys, MD ¹⁴

David E. Goldgar, PhD ¹⁵

Cecilia M. Dorfling, MSc ¹⁶

Elizabeth J. van Rensburg, PhD ¹⁷

Yuan Chun Ding, PhD ⁸

Bent Ejlersen, MD ¹⁸

Anne-Marie Gerdes, MD ¹⁹

Thomas V. O. Hansen, PhD ²⁰

Susan Slager, PhD ²¹

Emily Hallberg, MPH ²¹

Javier Benitez, PhD ²²

Ana Osorio, PhD ²³

Nancy Cohen , MS ²⁴

William Lawler , ²⁴

Jeffrey N. Weitzel, MD ²⁵

Paolo Peterlongo, PhD ²⁶

Valeria Pensotti, PhD ²⁷

Riccardo Dolcetti, MD ²⁸

Monica Barile, MD ²⁹

Bernardo Bonanni, MD ²⁹

Jacopo Azzollini, MD ³⁰

Siranoush Manoukian, MD ³⁰

Bernard Peissel, MD ³⁰

Paolo Radice, PhD ³¹

Antonella Savarese, MD ³²

Laura Papi, MD ³³

Giuseppe Giannini, MD ³⁴

Florentia Fostira, ³⁵

Irene Konstantopoulou, PhD ³⁶

Julian Adlard, MBBS ³⁷

Carole Brewer, BSc ³⁸

Jackie Cook, FRCP ³⁹

Rosemarie Davidson, ⁴⁰

Diana Eccles, MD ⁴¹

Ros Eeles, PhD ⁴²

Steve Ellis, MSc ⁴³

EMBRACE, ⁴³

Debra Frost, ⁴³

Shirley Hodgson, FRCP ⁴⁴

Louise Izatt, PhD ⁴⁵

Fiona Lalloo, MBBS ⁴⁶

Kai-ren Ong, MD ⁴⁷

Andrew K. Godwin, PhD ⁴⁸

Norbert Arnold, PhD ⁴⁹

Bernd Dworniczak, ⁵⁰

Christoph Engel, MD ⁵¹

Andrea Gehrig, ⁵²

Eric Hahnen, PhD ⁵³

Jan Hauke, PhD ⁵³

Karin Kast, MD ⁵⁴

Alfons Meindl , PhD ⁵⁵

Dieter Niederacher, PhD ⁵⁶

Rita Katharina Schmutzler, MD ⁵³

Raymonda Varon-Mateeva, PhD ⁵⁷

Shan Wang-Gohrke, PhD ⁵⁸

Barbara Wappenschmidt, PhD ⁵³

Laure Barjhoux, M.Sc. ⁵⁹

Marie-Agnès Collonge-Rame, MD ⁶⁰

Camille Elan, MD ⁶¹

GEMO Study Collaborators, ⁶²

Lisa Golmard , PhD ⁶¹

Emmanuelle Barouk-Simonet, MD ⁶³

Fabienne Lesueur, PhD ⁶⁴

Sylvie Mazoyer, PhD ^{59*}

Joanna Sokolowska, MD ⁶⁵

Dominique Stoppa-Lyonnet, MD ⁶²

Claudine Isaacs, MD ⁶⁶

Kathleen B.M. Claes, PhD ⁶⁷

Bruce Poppe, MD ⁶⁷

Miguel de la Hoya, PhD ⁶⁸

Vanesa Garcia-Barberan, PhD ⁶⁸

Kristiina Aittomäki, MD ⁶⁹

Heli Nevanlinna, PhD ⁷⁰

Margreet G.E.M. Ausems, MD ⁷¹

J.L. de Lange, ⁷²

Encarna B. Gómez Garcia, MD ⁷³

HEBON, ⁷⁴

Frans B.L. Hogervorst, PhD ⁷⁵

Carolien M. Kets, MD ⁷⁶

Hanne E.J. Meijers-Heijboer, ⁷⁷

Jan C. Oosterwijk, PhD ⁷⁸

Matti A. Rookus, PhD ⁷⁹

Christi J. van Asperen, ⁸⁰

Ans M.W. van den Ouweland, PhD ⁸¹

Helena C. van Doorn, MD ⁸²

Theo A.M. van Os, MD ⁸³

Ava Kwong, MBBS ⁸⁴

Edith Olah, PhD ⁸⁵

Orland Diez , PhD ⁸⁶

Joan Brunet, MD ⁸⁷

Conxi Lazaro, PhD ⁸⁸

Alex Teulé, MD ⁸⁹

Jacek Gronwald, MD ⁹⁰

Anna Jakubowska, PhD ⁹⁰

Katarzyna Kaczmarek, MSc⁹⁰
Jan Lubinski, MD⁹⁰
Grzegorz Sukiennicki, MSc⁹⁰
Rosa B. Barkardottir, Cand.Sci⁹¹
Jocelyne Chiquette, MD⁹²
Simona Agata, PhD⁹³
Marco Montagna, PhD⁹³
Manuel R. Teixeira, MD⁹⁴
KConFab Investigators,⁹⁵
Sue Kyung Park,⁹⁶
Curtis Olswold, BSc²¹
Marc Tischkowitz, MD⁹⁷
Lenka Foretova, MD⁹⁸
Pragna Gaddam, BSc⁹⁹
Joseph Vijai, PhD¹⁰⁰
Georg Pfeiler, MD¹⁰¹
Christine Rappaport-Fuerhauser,¹⁰²
Christian F. Singer, MD¹⁰²
Muy-Kheng M. Tea, MD¹⁰²
Mark H. Greene, MD¹⁰³
Jennifer T. Loud, DNP¹⁰⁴
Gad Rennert, MD¹⁰⁵
Evgeny N. Imyanitov, PhD¹⁰⁶
Peter J. Hulick¹⁰⁷

John L. Hays, MD ¹⁰⁸

Marion Piedmonte, MA ¹⁰⁹

Gustavo C. Rodriguez, MD ¹¹⁰

Julie Martyn, PhD ¹¹¹

Gord Glendon, MSc ¹¹²

Anna Marie Mulligan, MD ¹¹³

Irene L. Andrulis, PhD ¹¹⁴

Amanda Ewart Toland, PhD ¹¹⁵

Uffe Birk Jensen, PhD ¹¹⁶

Torben A. Kruse, PhD ¹¹⁷

Inge Sokilde Pedersen, PhD ¹¹⁸

Mads Thomassen, PhD ¹¹⁷

Maria A. Caligo, PhD ¹¹⁹

Soo-Hwang Teo, PhD ¹²⁰

Raanan Berger, MD ¹²¹

Eitan Friedman, MD ¹²²

Yael Laitman , MSc ¹²³

Brita Arver, MD ¹²⁴

Ake Borg, PhD ¹²⁵

Hans Ehrencrona , MD ¹²⁶

Johanna Rantala, PhD ¹²⁷

Olufunmilayo I. Olopade, MD ¹²⁸

Patricia A. Ganz ¹²⁹

Robert L. Nussbaum, MD ¹³⁰

Angela R. Bradbury, MD ¹³¹

Susan M. Domchek, MD ¹³¹

Katherine L. Nathanson, MD ¹³¹

Banu K. Arun, MD ¹³²

Paul James, MBBS ¹³³

Beth Y. Karlan, MD ¹³⁴

Jenny Lester, MPH ¹³⁴

Jacques Simard, PhD ²

Paul D.P. Pharoah, BM ¹³⁵

Kenneth Offit, MD ¹³⁶

Fergus J. Couch, PhD ¹³⁷

Georgia Chenevix-Trench, PhD ⁵

Douglas F. Easton, PhD ¹

Antonis C. Antoniou, PhD ¹

⁰ The Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton,
Cambridge, UK

¹ Department of Public Health and Primary Care, University of Cambridge, UK

² Genomics Center, Centre Hospitalier Universitaire de Québec Research Center and
Laval University, 2705 Laurier Boulevard, Quebec City (Quebec), Canada

³ Department of Genetics, QIMR Berghofer Medical Research Institute, Herston

Road, Brisbane, Australia 4029

⁴ Clinical Genetics, Service, Department of Medicine, Memorial Sloan Kettering Cancer Center, 1275 York Avenue, New York, NY 10044, USA

⁵ Department of Genetics and Computational Biology, QIMR Berghofer Medical Research Institute, Herston Road, Brisbane, Australia 4029

⁶ Department of Preventive Medicine, Keck School of Medicine, University of Southern California, California, USA

⁷ Department of Epidemiology, Columbia University, New York, NY, USA

⁸ Department of Population Sciences, Beckman Research Institute of City of Hope, Duarte, CA USA

⁹ Molecular Genetics of Breast Cancer, German Cancer Research Center (DKFZ), Im Neuenheimer Feld 580, 69120 Heidelberg, Germany

¹⁰ Genetic Epidemiology Laboratory, Department of Pathology, University of Melbourne, Parkville, Victoria, Australia

¹¹ Department of Epidemiology, Cancer Prevention Institute of California, 2201 Walnut Avenue, Suite 300, Fremont, CA 94538, USA

¹² Departments of Pediatrics and Medicine, 1150 St. Nicholas Avenue, Columbia University, New York, NY, 10032 USA

¹³ Department of Clinical Genetics, Fox Chase Cancer Center, 333 Cottman Avenue, Philadelphia, PA 19111, USA

¹⁴ Department of Medicine, Huntsman Cancer Institute, 2000 Circle of Hope, Salt Lake City, UT 84112, USA

¹⁵ Department of Dermatology, University of Utah School of Medicine, 30 North 1900 East, SOM 4B454, Salt Lake City, UT 84132, USA

- ¹⁶ Cancer Genetics Laboratory, Department of Genetics, University of Pretoria,
Private Bag X323, Arcadia 0007, South Africa
- ¹⁷ Cancer Genetics Laboratory, Department of Genetics, University of
Pretoria, Private Bag X323, Arcadia 0007, South Africa
- ¹⁸ Department of Oncology, Rigshospitalet, Copenhagen University Hospital,
Blegdamsvej 9, DK-2100 Copenhagen, Denmark
- ¹⁹ Department of Clinical Genetics, Rigshospitalet 4062, Blegdamsvej 9, København
Ø, Denmark
- ²⁰ Center for Genomic Medicine, Rigshospitalet, Copenhagen University Hospital,
Blegdamsvej 9, DK-2100 Copenhagen, Denmark
- ²¹ Department of Health Sciences Research, Mayo Clinic, 200 First Street SW,
Rochester, Minnesota, USA
- ²² (1) Human Genetics Group, Spanish National Cancer Centre (CNIO), Madrid, Spain;
(2) Biomedical Network on Rare Diseases (CIBERER), Madrid, Spain; (3) Human
Genotyping (CEGEN) Unit, Human Cancer Genetics Program, Spanish National Cancer
Research Centre (CNIO), Madrid, Spain
- ²³ (1) Human Genetics Group, Spanish National Cancer Centre (CNIO), Madrid, Spain;
(2) Biomedical Network on Rare Diseases (CIBERER), Madrid, Spain.
- ²⁴ City of Hope Clinical Cancer Genomics Community Research Network, 1500 East
Duarte Road, Duarte, CA 91010
- ²⁵ Clinical Cancer Genetics, City of Hope, 1500 East Duarte Road, Duarte, California
91010 USA
- ²⁶ IFOM, The FIRC (Italian Foundation for Cancer Research) Institute of Molecular
Oncology, c/o IFOM-IEO campus, via Adamello 16, 20139 Milan, Italy.

²⁷ (1) IFOM, the FIRC (Italian Foundation for Cancer Research) Institute of Molecular Oncology, Milan, Italy; (2) Cogentech Cancer Genetic Test Laboratory, Milan, Italy

²⁸ Centro di Riferimento Oncologico, IRCCS, Aviano, Italy; University of Queensland Diamantina Institute, Translational Research Institute, Brisbane, Australia

²⁹ Division of Cancer Prevention and Genetics, Istituto Europeo di Oncologia, Milan, Italy

³⁰ Unit of Medical Genetics, Department of Preventive and Predictive Medicine, Fondazione IRCCS (Istituto di Ricovero e Cura a Carattere Scientifico) Istituto Nazionale Tumori (INT), Milan, Italy

³¹ Unit of Molecular Bases of Genetic Risk and Genetic Testing, Department of Preventive and Predictive Medicine, Fondazione IRCCS (Istituto di Ricovero e Cura a Carattere Scientifico) Istituto Nazionale Tumori (INT), Milan, Italy

³² Unit of Genetic Counselling, Medical Oncology Department, Istituto Nazionale Tumori Regina Elena, Rome, Italy .

³³ Unit of Medical Genetics, Department of Biomedical, Experimental and Clinical Sciences, University of Florence, Florence, Italy

³⁴ Department of Molecular Medicine, University La Sapienza, Rome, Italy

³⁵ Molecular Diagnostics Laboratory, (INRASTES) Institute of Nuclear and Radiological Sciences and Technology, National Centre for Scientific Research Demokritos, Patriarchou Gregoriou & Neapoleos str., Aghia Paraskevi Attikis, Athens, GREECE

³⁶ Molecular Diagnostics Laboratory, INRASTES (Institute of Nuclear and Radiological Sciences and Technology), National Centre for Scientific Research Demokritos, Patriarchou Gregoriou & Neapoleos str., Aghia Paraskevi Attikis, Athens, GREECE

³⁷ Yorkshire Regional Genetics Service, Chapel Allerton Hospital, Leeds, UK

- ³⁸ Department of Clinical Genetics, Royal Devon & Exeter Hospital, Exeter, UK
- ³⁹ Sheffield Clinical Genetics Service, Sheffield Children's Hospital, Sheffield, UK
- ⁴⁰ Department of Clinical Genetics, South Glasgow University Hospitals, Glasgow, UK
- ⁴¹ University of Southampton Faculty of Medicine, Southampton University Hospitals NHS Trust, Southampton, UK
- ⁴² Oncogenetics Team, The Institute of Cancer Research and Royal Marsden NHS Foundation Trust, London UK
- ⁴³ Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Strangeways Research Laboratory, Worts Causeway, Cambridge, UK
- ⁴⁴ Medical Genetics Unit, St George's, University of London, UK
- ⁴⁵ Clinical Genetics, Guy's and St. Thomas' NHS Foundation Trust, London, UK
- ⁴⁶ Genetic Medicine, Manchester Academic Health Sciences Centre, Central Manchester University Hospitals NHS Foundation Trust, Manchester, UK
- ⁴⁷ West Midlands Regional Genetics Service, Birmingham Women's Hospital Healthcare NHS Trust, Edgbaston, Birmingham, UK
- ⁴⁸ Department of Pathology and Laboratory Medicine, 3901 Rainbow Boulevard, 4019 Wahl Hall East, MS 3040, University of Kansas Medical Center, Kansas City, Kansas, USA
- ⁴⁹ Department of Gynaecology and Obstetrics, University Hospital of Schleswig-Holstein, Campus Kiel, Christian-Albrechts University Kiel, Germany
- ⁵⁰ Institute of Human Genetics, University of Münster, Münster, Germany
- ⁵¹ Institute for Medical Informatics, Statistics and Epidemiology, University of Leipzig, Germany

- ⁵² Centre of Familial Breast and Ovarian Cancer, Department of Medical Genetics, Institute of Human Genetics, University Würzburg, Germany
- ⁵³ Center of Familial Breast and Ovarian Cancer, Centre for Integrated Oncology (CIO), Center for Molecular Medicine Cologne (CMMC), University Hospital Cologne, Medical Faculty, Cologne, Germany
- ⁵⁴ Department of Gynaecology and Obstetrics, University Hospital Carl Gustav Carus, Technical University Dresden, Germany
- ⁵⁵ Department of Gynaecology and Obstetrics, Division of Tumor Genetics, Klinikum rechts der Isar, Technical University Munich, Germany
- ⁵⁶ Department of Gynaecology and Obstetrics, University Hospital Düsseldorf, Heinrich-Heine University Düsseldorf, Germany
- ⁵⁷ Institute of Human Genetics, Campus Virchow Klinikum, Charite Berlin, Germany
- ⁵⁸ Department of Gynaecology and Obstetrics, University Hospital Ulm, Germany
- ⁵⁹ Bâtiment Cheney D, Centre Léon Bérard, 28 rue Laënnec, Lyon, France
- ^{59*} Lyon Neuroscience Research Center- CRNL, Inserm U1028, CNRS UMR5292, University of Lyon, Lyon, France
- ⁶⁰ Service de Génétique Biologique, CHU de Besançon, 25030 Besançon, France
- ⁶¹ Service de Génétique, Institut Curie, 26, rue d'Ulm, Paris Cedex 05, France
- ⁶² (1) Institut Curie, Department of Tumour Biology, Paris, France; Institut Curie, INSERM U830, Paris, France; (2) Université Paris Descartes, Sorbonne Paris Cité, France
- ⁶³ Oncogénétique, Institut Bergonié, 229 cours de l'Argonne, 33076 Bordeaux, France
- ⁶⁴ (1) Genetic Epidemiology of Cancer team, Inserm U900, Paris, France; (2) Institut Curie, 26 rue d'Ulm, Paris, France; (3) Mines ParisTech, Fontainebleau, France

⁶⁵ Laboratoire de génétique médicale, Nancy Université, Centre Hospitalier Régional et Universitaire, Rue du Morvan, 54511 cedex 1, Vandoeuvre-les-Nancy, France

⁶⁶ Lombardi Comprehensive Cancer Center, Georgetown University, 3800 Reservoir Road NW, Washington, DC, USA

⁶⁷ Center for Medical Genetics, Ghent University, De Pintelaan 185, 9000 Gent, Belgium

⁶⁸ Molecular Oncology Laboratory, Hospital Clinico San Carlos, IdISSC (El Instituto de Investigación Sanitaria del Hospital Clínico San Carlos), Martin Lagos s/n, Madrid, Spain

⁶⁹ Department of Clinical Genetics, Helsinki University Hospital, P.O. BOX 160 (Meilahdentie 2), 00029 HUS, Finland

⁷⁰ Department of Obstetrics and Gynecology, University of Helsinki and Helsinki University Hospital, Biomedicum Helsinki, P.O. BOX 700 (Haartmaninkatu 8), 00029 HUS, Finland

⁷¹ Department of Medical Genetics, University Medical Center Utrecht, P.O. Box 85090, 3508 AB Utrecht, The Netherlands

⁷² Department of Epidemiology. Netherlands Cancer Institute, P.O. Box 90203, 1006 BE, Amsterdam, The Netherlands

⁷³ Department of Clinical Genetics and GROW, School for Oncology and Developmental Biology, MUMC, P.O. Box 5800 6202 AZ Maastricht , The Netherlands

⁷⁴ The Hereditary Breast and Ovarian Cancer Research Group Netherlands (HEBON),Coordinating center: Netherlands Cancer Institute, Amsterdam, The Netherlands

⁷⁵ Family Cancer Clinic, Netherlands Cancer Institute, P.O. Box 90203, 1000 BE,

Amsterdam, The Netherlands

⁷⁶ Department of Human Genetics, Radboud University Nijmegen Medical Centre,
P.O. Box 9101, 6500HB Nijmegen, The Netherlands

⁷⁷ Department of Clinical Genetics, VU University Medical Centre, P.O. Box 7057,
1007 MB Amsterdam, the Netherlands

⁷⁸ Department of Genetics, University Medical Center, Groningen University,
Groningen, The Netherlands

⁷⁹ Department of Epidemiology, Netherlands Cancer Institute, P.O. Box 90203, 1000
BE, Amsterdam, The Netherlands

⁸⁰ Department of Clinical Genetics Leiden University Medical Center Leiden, P.O. Box
9600, 2300 RC Leiden, The Netherlands

⁸¹ Department of Clinical Genetics, Family Cancer Clinic, Erasmus University Medical
Center, P.O. Box 2040, 3000 CA Rotterdam, The Netherlands

⁸² Department of Gynaecology, Family Cancer Clinic, Erasmus MC Cancer Institute,
Room D4-20, PO Box 5201, 3008 AE Rotterdam, The Netherlands

⁸³ Department of Clinical Genetics, Academic Medical Center, P.O. Box 22700 1100
DE Amsterdam, The Netherlands

⁸⁴ The Hong Kong Hereditary Breast Cancer Family Registry; Cancer Genetics Center,
Hong Kong Sanatorium and Hospital, Hong Kong; Department of Surgery, The
University of Hong Kong, Hong Kong

⁸⁵ Department of Molecular Genetics, National Institute of Oncology, Budapest,
Hungary

⁸⁶ Oncogenetics Group, Vall d'Hebron Institute of Oncology (VHIO), Universitat
Autònoma de Barcelona, Vall d'Hebron University Hospital. Passeig Vall d'Hebron

119-129. Barcelona. Spain

⁸⁷ Genetic Counseling Unit, Hereditary Cancer Program, IDIBGI (Institut d'Investigació Biomèdica de Girona), Catalan Institute of Oncology. Av. França s/n. 1707 Girona, Spain

⁸⁸ Molecular Diagnostic Unit, Hereditary Cancer Program, IDIBELL (Bellvitge Biomedical Research Institute), Catalan Institute of Oncology. Gran Via de l'Hospitalet, 199-203. 08908 L'Hospitalet. Barcelona, Spain

⁸⁹ Genetic Counseling Unit, Hereditary Cancer Program, IDIBELL (Bellvitge Biomedical Research Institute), Catalan Institute of Oncology. Gran Via de l'Hospitalet, 199-203. 08908 L'Hospitalet. Barcelona, Spain

⁹⁰ Department of Genetics and Pathology, Pomeranian Medical University, Polabska 4, Szczecin, Poland.

⁹¹ 1) Laboratory of Cell Biology, Department of Pathology, hus 9, Landspítali-LSH v/Hringbraut, 101 Reykjavik, Iceland. 2) BMC (Biomedical Centre), Faculty of Medicine, University of Iceland, Vatnsmyrarvegi 16, 101 Reykjavik, Iceland

⁹² Unité de recherche en santé des populations, Centre des maladies du sein Deschênes-Fabia, Hôpital du Saint-Sacrement, 1050, chemin Sainte-Foy, Québec Québec, Canada

⁹³ Immunology and Molecular Oncology Unit, Veneto Institute of Oncology IOV - IRCCS, Via Gattamelata 64, Padua, Italy

⁹⁴ (1) Department of Genetics, Portuguese Oncology Institute of Porto, Rua Dr. António Bernardino de Almeida, 4200-072 Porto, Portugal; (2) Biomedical Sciences Institute (ICBAS), University of Porto, Porto, Portugal.

⁹⁵ Kathleen Cuninghame Consortium for Research into Familial Breast Cancer, Peter

MacCallum Cancer Center, Melbourne, Australia

⁹⁶ Department of Preventive Medicine, Seoul National University College of Medicine, Department of Biomedical Science, Seoul National University Graduate School, and Cancer Research Institute, Seoul National University, 103 Daehak-ro, Jongno-gu, Seoul 110-799, Korea

⁹⁷ Program in Cancer Genetics, Departments of Human Genetics and Oncology, McGill University, Montreal, Quebec, Canada

⁹⁸ Department of Cancer Epidemiology and Genetics, Masaryk Memorial Cancer Institute, Zlutý kopec 7, Brno, 65653 Czech Republic

⁹⁹ Clinical Cancer Genetics Laboratory, Memorial Sloan Kettering Cancer Center, New York, NY, USA

¹⁰⁰ Clinical Genetics Research Laboratory, Dept. of Medicine, Memorial Sloan Kettering Cancer Center, 1275 York Avenue, New York, NY 10044, USA

¹⁰¹ Department of Gynecology and Gynecological Oncology, Comprehensive Cancer Center, Medical University of Vienna, Währinger Gürtel 18-20, 1090 Vienna, Austria

¹⁰² Dept of OB/GYN, Medical University of Vienna, Vienna, Austria, Waehringer Guertel 18-20, A 1090 Vienna, Austria

¹⁰³ Clinical Genetics Branch, DCEG, NCI, NIH, 9609 Medical Center Drive, Room 6E-454, Rockville, MD, USA

¹⁰⁴ Clinical Genetics Branch, DCEG, NCI; 9609 Medical Center Drive, Room 6E-536, Rockville, MD, USA

¹⁰⁵ Clalit National Israeli Cancer Control Center and Department of Community Medicine and Epidemiology, Carmel Medical Center and B. Rappaport Faculty of Medicine, 7 Michal St., Haifa 34362, Israel

¹⁰⁶ N.N. Petrov Institute of Oncology, St.-Petersburg 197758, Russia

¹⁰⁷ Medical Director, Center for Medical Genetics, NorthShore University

HealthSystem, Clinical Assistant Professor of Medicine, University of Chicago Pritzker School of Medicine, 1000 Central Street, Suite 620, Evanston, IL 60201, US

¹⁰⁸ The Ohio State University Comprehensive Cancer Center Arthur C. James Cancer Hospital and Richard J. Solove Research Institute Biomedical Research Tower, Room 588, 460 West 12th Avenue, Columbus

¹⁰⁹ NRG Oncology, Statistics and Data Management Center, Roswell Park Cancer Institute, Elm St & Carlton St, Buffalo, NY 14263, USA

¹¹⁰ Division of Gynecologic Oncology, NorthShore University HealthSystem, Clinical Professor, Univ of Chicago, 2650 Ridge Avenue Suite 1507 Walgreens, Evanston, IL 60201, US

¹¹¹ ANZGOG, NHMRC Clinical Trials Centre, Locked Bag 77, Camperdown, NSW 1450, Australia

¹¹² Ontario Cancer Genetics Network: Lunenfeld-Tanenbaum Research Institute, Mount Sinai Hospital, Toronto, Ontario M5G 1X5

¹¹³ Laboratory Medicine Program, University Health Network, Toronto, Ontario, M5B 1W8, Department of Laboratory Medicine and Pathobiology, University of Toronto, Toronto, ON, Canada

¹¹⁴ Lunenfeld-Tanenbaum Research Institute, Mount Sinai Hospital, Toronto, Ontario M5G 1X5, Departments of Molecular Genetics and Laboratory Medicine and Pathobiology, University of Toronto, Ontario, Canada

¹¹⁵ Division of Human Cancer Genetics, Departments of Internal Medicine and Molecular Virology, Immunology and Medical Genetics, Comprehensive Cancer

Center, The Ohio State University, 998 Biomedical Research Tower, Columbus, OH,
USA

¹¹⁶ Department of Clinical Genetics, Aarhus University Hospital, Brendstrupgaardsvej
21C, Aarhus N, Denmark

¹¹⁷ Department of Clinical Genetics, Odense University Hospital, Sonder Boulevard
29, Odense C, Denmark

¹¹⁸ Section of Molecular Diagnostics, Clinical Biochemistry, Aalborg University
Hospital, Reberbansgade 15, Aalborg, Denmark

¹¹⁹ Section of Genetic Oncology, Dept. of Laboratory Medicine, University and
University Hospital of Pisa, Pisa Italy

¹²⁰ Cancer Research Initiatives Foundation, Sime Darby Medical Centre, 1 Jalan
SS12/1A, Subang Jaya, 47500 Malaysia and University Malaya Cancer Research
Institute, University Malaya, 50603 Kuala Lumpur, Malaysia

¹²¹ The Institute of Oncology, Chaim Sheba Medical Center, Ramat Gan 52621, Israel

¹²² The Susanne Levy Gertner Oncogenetics Unit, Institute of Human Genetics, Chaim
Sheba Medical Center, Ramat Gan 52621, and Sackler Faculty of Medicine, Tel Aviv
University, Ramat Aviv 69978, Israel

¹²³ The Susanne Levy Gertner Oncogenetics Unit, Institute of Human Genetics, Chaim
Sheba Medical Center, Ramat Gan 52621, Israel

¹²⁴ Department of Oncology, Karolinska University Hospital, Stockholm, Sweden

¹²⁵ Department of Oncology, Clinical Sciences, Lund University and Skåne University
Hospital, Lund, Sweden

¹²⁶ Department of Clinical Genetics, Lund University Hospital, Lund, Sweden

¹²⁷ Department of Clinical Genetics, Karolinska University Hospital L5:03, Stockholm

S-171 76, Sweden

¹²⁸ 5841 South Maryland Avenue, MC 2115 Chicago, IL

¹²⁹ UCLA Schools of Medicine and Public Health, Division of Cancer Prevention & Control Research, Jonsson Comprehensive Cancer Center, 650 Charles Young Drive South, Room A2-125 HS, Los Angeles, CA 90095-6900, USA

¹³⁰ 513 Parnassus Ave., HSE 901E, San Francisco, CA. 94143 - 0794

¹³¹ Department of Medicine, Abramson Cancer Center, Perelman School of Medicine at the University of Pennsylvania, 3400 Civic Center Boulevard, Philadelphia, PA 19104, USA

¹³² Department of Breast Medical Oncology and Clinical Cancer Genetics Program, University Of Texas MD Anderson Cancer Center, 1515 Pressler Street, CBP 5, Houston, TX, USA

¹³³ Familial Cancer Centre, Peter MacCallum Cancer Centre, Locked Bag 1, A'Beckett Street, Melbourne, VIC 8006 AUSTRALIA

¹³⁴ Women's Cancer Program at the Samuel Oschin Comprehensive Cancer Institute, Cedars-Sinai Medical Center, 8700 Beverly Boulevard, Suite 290W, Los Angeles, CA, USA

¹³⁵ Department of Oncology, University of Cambridge, Cambridge, UK.

¹³⁶ Clinical Genetics Research Laboratory, Dept. of Medicine, Cancer Biology and Genetics, Memorial Sloan-Kettering Cancer Center, 1275 York Avenue, New York, NY 10044, USA

¹³⁷ Department of Laboratory Medicine and Pathology, and Health Sciences Research, Mayo Clinic, 200 First Street SW, Rochester, Minnesota, USA

Corresponding author: Dr Antonis Antoniou, Strangeways Research Laboratory,
Worts Causeway, Cambridge CB1 8RN, UK; email: antonis@srl.cam.ac.uk, tel.: +44
(0)1223 748630

Abstract

Background

Genome-wide association studies (GWAS) have identified 94 common single nucleotide polymorphisms (SNPs) associated with breast (BC) and 18 with ovarian cancer (OC) risks. Several of these are also associated with risk of BC or OC for women who carry a pathogenic mutation in the high-risk BC and OC genes *BRCA1* or *BRCA2*. The combined effects of these variants on BC or OC risk for *BRCA1* and *BRCA2* mutation carriers have not yet been assessed while their clinical management could benefit from improved personalized risk estimates.

Methods

We constructed polygenic risk scores (PRS) using BC and OC susceptibility SNPs identified through population-based GWAS: for BC (overall, oestrogen receptor (ER) positive, and ER-negative) and for OC. Using data from 15,252 female *BRCA1* and 8,211 *BRCA2* carriers, the association of each PRS with BC or OC risk was evaluated using a weighted cohort approach with time to diagnosis as the outcome and estimation of the hazard ratios (HR) per standard deviation increase in the PRS. All statistical tests were two-sided.

Results

The PRS for ER-negative BC displayed the strongest association with BC risk in *BRCA1* carriers (HR=1.27, 95% confidence interval (CI):1.23-1.31, $p=8.2 \times 10^{-53}$). In *BRCA2* carriers, the strongest association with BC risk was seen for the overall BC PRS (HR=1.22, 95%CI: 1.17-1.28, $p=7.2 \times 10^{-20}$). The OC PRS was strongly associated with OC risk for both *BRCA1* and *BRCA2* carriers. These translate to differences in absolute

risks (more than 10% in each case) between the top and bottom deciles of the PRS distribution, e.g., the OC risk was 6% by age 80 for *BRCA2* carriers at the 10th percentile of the OC PRS compared with 19% risk for those at the 90th percentile of PRS.

Conclusions

BC and OC PRS are predictive of cancer risks in *BRCA1* and *BRCA2* carriers. Incorporation of the PRS into risk prediction models has promise to better inform decisions on cancer risk management.

Introduction

Women who carry a pathogenic mutation in the *BRCA1* or *BRCA2* gene are at high risk of developing breast and ovarian cancers. The clinical management of healthy women with a *BRCA1* or *BRCA2* mutation involves a combination of frequent screening, risk-reducing surgeries and chemoprevention (1). Important decisions include whether or not to undergo preventive mastectomy and the age at which to undergo risk-reducing salpingo-oophorectomy (RRSO). These choices are invasive, have substantial side-effects, and are associated with adverse psychological effects (2-6). Improved personalized cancer risk estimates may help to identify women at particularly high risk or with high risk of disease at early ages who may benefit from early intervention as well as women at lower risk who may opt to delay surgery or chemoprevention (7). This could be achieved by incorporating risk-modifying factors into risk prediction.

Population-based genome-wide association studies have identified 94 common breast and 18 ovarian cancer susceptibility loci (8-10). While a smaller number of these loci were associated with risk in *BRCA1* and *BRCA2* mutation carriers at stringent statistical significance thresholds, the effect sizes in carriers are generally similar to those in the general population, once differences in the distributions of breast tumor estrogen receptor status in mutation carriers and non-carriers are taken into account (9, 11). Individually the identified breast and ovarian cancer risk-modifying variants confer only small to modest increases in risk. However, their effects can be combined into polygenic risk scores (PRS), which may be associated with much larger relative risks (12, 13). Prior to the clinical

implementation of these findings, it is important to assess the predictive utility of PRS in terms of discrimination, calibration, and potential for risk stratification (14).

Because women with *BRCA1* and *BRCA2* mutations are already at high risk of developing breast and ovarian cancers, the combined effects of risk-modifying variants could lead to much larger differences in the absolute risk of developing the disease as compared with the general population (12, 13, 15, 16). Earlier studies investigating the effect of PRS on the absolute risks of breast and ovarian cancer risks of *BRCA1* and *BRCA2* mutation carriers demonstrated potential for risk stratification (13, 17-19). However, these have been based on small numbers of SNPs (<15) and most were restricted to theoretical projections of the PRS association rather than empirical evaluations.

In this study we developed different PRS for breast and ovarian cancer as well as oestrogen receptor (ER)-specific PRS based on reported susceptibility loci from population-based studies, and evaluated their associations with risks for *BRCA1* and *BRCA2* carriers. We estimated absolute risks of developing breast and ovarian cancer for individuals with different values of the PRS in order to assess whether these PRS provide clinically useful risk stratification of mutation carriers.

Methods

Study population

Eligible study subjects included in the Consortium of Investigators of Modifiers of *BRCA1/2* (CIMBA) are female carriers of a pathogenic mutation in either

BRCA1 or *BRCA2* who are ≥ 18 years of age. Mutation carriers were recruited by 56 study centers in 26 countries. The majority were recruited through cancer genetics clinics, and enrolled into national or regional studies. We used data from 15,252 *BRCA1* (breast cancer=7,797; ovarian cancer=2,462) and 8,211 *BRCA2* (breast cancer=4,330; ovarian cancer=631) mutation carriers who were genotyped with the iCOGS array. Quality control has been described in detail elsewhere (11, 13, 18). Each of the host institutions recruited mutation carriers under protocols approved by local ethics review boards. Written informed consent was obtained from all subjects. Only samples of European ancestry were included in the present analysis.

Polygenic risk scores

The effects of cancer susceptibility variants on cancer risks for mutation carriers were combined into PRS. The PRS for individual i was defined as the sum of the number of risk alleles across k variants weighted by the effect size of each variant:

$$PRS_i = \beta_1 g_{1i} + \dots + \beta_k g_{ki}$$

where g_{li} is the genotype of person i for variant l , expressed as the number of effect alleles (0,1, or 2) and β_l is the per-allele log risk ratio (Odds Ratio (OR) or Hazard Ratio (HR), **Supplementary Tables 1-6**) associated with the effect allele of SNP l .

The primary PRS were based on SNPs found to be associated with breast or ovarian cancer through GWAS in the general population. For breast cancer, we used the published PRS for overall breast cancer, ER-positive breast cancer and ER-

negative breast cancer (8, 20). In addition, we created updated PRS based on findings from population-based association and fine-mapping studies reported before April 2015 (**Supplementary Table 1**) (8, 10, 21-28). More details on the variant selection are provided in the **Supplementary Methods**.

We developed an ovarian cancer PRS by including the most strongly associated variant from each region associated at genome-wide statistical significance level with ovarian cancer risk in population-based studies or studies that combined population data and data from mutation carriers (**Supplementary Table 2**) (9, 23).

We also constructed secondary *BRCA1*- and *BRCA2*-specific PRS that were based on all variants showing evidence of association in *BRCA1* and *BRCA2* carriers, using the results and weights from the *BRCA1*- and *BRCA2*-specific GWAS (11-13). (**Supplementary Tables 3-6, Supplementary Methods**). However, the studies that led to the identification of these variants were based on the same dataset as the present analysis. Therefore, these *BRCA1*- and *BRCA2*-specific PRS cannot be independently validated in the present analysis. To reduce the bias from overfitting, we also constructed and evaluated unweighted versions of these PRS.

For the SNPs included in each PRS, we assessed whether there was evidence for pairwise interactions (**Supplementary Methods**).

Statistical analysis

To account for the non-random sampling of mutation carriers with respect to disease status, the association of each PRS with breast or ovarian cancer risk was

analysed using a weighted cohort Cox-regression with time to breast or ovarian cancer diagnosis, respectively, as the outcome (29) (**Supplementary Methods [Please be specific—Supplementary Methods, Results, or a particular table/figure?]**). We evaluated the associations of the breast cancer PRS (i.e. overall breast cancer PRS, ER-positive PRS and ER-negative PRS) with the risk for overall breast cancer for *BRCA1* and *BRCA2* mutation carriers. The ovarian cancer PRS was assessed for association with the risk of developing overall ovarian cancer for *BRCA1* and *BRCA2* mutation carriers. For these analyses, subjects were categorised into PRS percentile groups. To provide easily interpretable associations, the association analyses were repeated using continuous PRS predictors standardised to have mean 0 and variance 1. We assessed whether the HR per unit of the PRS varied with age by including a term for the interaction of the standardised PRS with age. We also fitted a Cox-regression that included separate PRS effects by age group.

To evaluate the ability of the PRS to discriminate between individuals developing breast or ovarian cancer at different ages, we computed the rank Harrell's c index (30) (**Supplementary Methods**).

Absolute age-specific cumulative risks of developing breast or ovarian cancer at different percentiles of the standardised PRS were calculated according to the approach described previously (15, 31) (**Supplementary Methods**).

Analyses were carried out in R using GenABEL (32) and in STATA v13.1 (33). Detailed methods are provided in **Supplementary Methods**.

Results

PRS associations with cancer risks

Using data from 15,252 *BRCA1* and 8,211 *BRCA2* carriers (Supplementary Table 7), there was no evidence for interaction between any two variants involved in any of the PRS after accounting for multiple testing (results not shown). All breast cancer PRS derived from population-based study results (**Supplementary Tables 1**) were statistically significantly associated with breast cancer risks for both *BRCA1* and *BRCA2* carriers (**Table 1**). Compared with the PRS developed by Mavaddat et al. (**Supplementary Table 9**), the updated breast cancer PRS displayed slightly stronger associations in *BRCA1* carriers but no improvements were seen in *BRCA2* carriers.

The PRS for ER-negative breast cancer displayed the strongest association with breast cancer risk in *BRCA1* carriers (per-standard-deviation (SD) HR=1.27, 95%CI: 1.23-1.31, $p=8.2 \times 10^{-53}$) (**Table 1**). Smaller HR estimates in *BRCA1*-breast cancer were seen for the PRS for overall breast cancer (HR=1.14, 95%CI: 1.11-1.17, $p=1.8 \times 10^{-18}$) and ER-positive breast cancer (HR=1.11, 95%CI: 1.08-1.15, $p=3.5 \times 10^{-13}$). In *BRCA2* carriers, the ER-negative breast cancer PRS displayed a smaller per-SD HR for breast cancer risk (HR=1.15, 95%CI: 1.10-1.20, $p=6.8 \times 10^{-10}$) compared to *BRCA1* carriers whereas the overall breast cancer PRS (HR=1.22, 95%CI: 1.17-1.28, $p=7.2 \times 10^{-20}$) and the ER-positive PRS (HR=1.22, 95%CI: 1.16-1.27, $p=4.0 \times 10^{-19}$) displayed stronger associations. The subsequent breast cancer analyses focus on the updated ER-negative breast cancer PRS for *BRCA1* carriers and the updated overall breast cancer PRS for *BRCA2* carriers.

Consistent with the above models, there were clear trends in risk by PRS for both *BRCA1* and *BRCA2* carriers when PRS was categorised by percentile (**Table 2**). The HR estimates were consistent with those predicted by the model in which PRS was fitted as a continuous covariate (**Figure 1**).

We also investigated whether the associations for the most strongly associated PRS differ by mutation type, as defined by the mutation functional effect (**Supplementary Methods**). There was marginal evidence of an interaction between the breast cancer risk PRS and class 2 mutations in *BRCA2* mutation carriers ($p=0.03$, with a slightly higher HR estimate for the PRS for class 2 mutation carriers).

The population-based ovarian cancer PRS was strongly associated with ovarian cancer risk in *BRCA1* carriers with a per-SD HR of 1.28 (95%CI: 1.22-1.34, $p=2.5 \times 10^{-26}$) (**Table 1**). The HR estimate was larger for ovarian cancer risk in *BRCA2* carriers: HR=1.49 (95%CI: 1.34-1.65, $p=8.5 \times 10^{-14}$). When we compared the HR estimates against the HRs predicted under a multiplicative polygenic model, only the HR estimate for *BRCA2* carriers for the 60-80% category was statistically significantly higher than the predicted value (**Figure 1**).

The unweighted *BRCA1*- and *BRCA2*-specific PRS for breast and ovarian cancer, constructed on the basis of association results in CIMBA, showed strong evidence of association with breast and ovarian cancer (**Supplementary Table 10**).

PRS x age interaction

There was evidence for a PRSxage interaction for the ER-negative breast cancer PRS for *BRCA1* carriers ($p=3 \times 10^{-6}$) and for the overall breast cancer PRS for *BRCA2* carriers ($p=0.01$) (**Table 3**). In the ovarian cancer analysis, a statistically

significant interaction with age was seen for the ovarian cancer PRS for *BRCA1* carriers ($p=0.003$). Each of these PRS showed stronger associations in younger age groups.

Discrimination

The ER-negative PRS had the highest value of Harrell's c , $c=0.58$ (95%CI: 0.57-0.59), for breast cancer in *BRCA1* carriers (**Table 4**). For breast cancer in *BRCA2* carriers, the highest values for Harrell's c were achieved by the population-based overall and ER-positive breast cancer PRS with values of $c=0.56$ (95%CI: 0.55-0.58) in each case. For ovarian cancer, the OC-PRS had $c=0.58$ (95%CI: 0.56-0.60) for *BRCA1* carriers and $c=0.63$ (95%CI: 0.60-0.67) for *BRCA2* carriers.

Predicted absolute risks by PRS percentile

We used the age-specific HR estimates to compute absolute cumulative breast and ovarian cancer risks for mutation carrier by PRS percentiles (**Figure 2**). We used the updated ER-negative PRS to predict breast cancer risk for *BRCA1* carriers and the updated overall breast cancer PRS to predict breast cancer risk for *BRCA2* carriers. *BRCA1* carriers at the 10th percentile of the PRS had a risk of 21% of developing breast cancer by age 50 and a 56% risk by age 80. In contrast, the *BRCA1* carriers at the 90th percentile of the PRS had a 39% breast cancer risk by age 50 and 75% by age 80. The ovarian cancer risk was 6% by age 80 for *BRCA2* carriers at the 10th percentile of the ovarian cancer PRS compared with 19% risk for those at the 90th percentile of PRS.

Discussion

This is the first evaluation of the combined effects of all known common breast and ovarian cancer susceptibility loci on cancer risks for women who carry a *BRCA1* or *BRCA2* mutation. We found strong evidence of association with cancer risks for PRS constructed using the results of population-based studies. These associations provide strong support for the hypothesis of a polygenic component for breast and ovarian cancer risks, respectively, that is largely shared between the general population and *BRCA1* and *BRCA2* mutation carriers. Moreover, the pattern of associations with the breast cancer subtype-specific PRS confirms the importance of tumour ER-status (11). The PRS based on SNPs associated with ER-negative disease in the general population displayed a much stronger association with overall breast cancer risk for *BRCA1* carriers than the ER-positive PRS, consistent with the observation that the predominant tumour subtype in *BRCA1* carriers is ER-negative (34, 35). In contrast, the majority of tumours in *BRCA2* carriers tend to be ER-positive. Consistent with this, the ER-positive PRS and the PRS for overall breast cancer constructed from general-population data exhibited stronger associations than the ER-negative PRS in *BRCA2* carriers.

Using the overall, ER-positive and ER-negative breast cancer PRS developed by Mavaddat, the per-SD HR estimates in mutation carriers were smaller than the corresponding per-SD OR estimates for breast cancer in the population-based study (20). These observations suggest that the relative extent, by which the SNPs modify

breast cancer risks in *BRCA1* and *BRCA2* mutation carriers is somewhat smaller than that in the general population, perhaps because a subset of SNPs do not combine multiplicatively with mutation status. Alternatively these observations may reflect a difference in the design: under a simple proportional hazards model the predicted odds ratio is larger than the corresponding rate ratio (HR), but this difference is usually small (36). Moreover, some overestimation cannot be ruled out entirely for the per-SD OR estimates from the population-based study due to a winner's curse effect. Interestingly, the HR estimate for the association of the ovarian cancer PRS with ovarian cancer risk was statistically significantly higher for *BRCA2* than for *BRCA1* mutation carriers. As a result, this PRS had also a higher discriminatory ability for ovarian cancer for *BRCA2* carriers compared to *BRCA1* mutation carriers.

Each of the most strongly associated PRS displayed statistically significant interactions with age, with the exception of the ovarian cancer PRS in *BRCA2* carriers, such that the HR per unit PRS decreased with increasing age. One possible explanation for the observed interaction between age and the ER-negative breast cancer PRS in *BRCA1* mutation carriers could be due to the use of the ER-negative breast cancer PRS from the general population to predict the risk of overall breast cancer risk for *BRCA1* mutation carriers. Although the majority of breast cancers in *BRCA1* mutation carriers are ER-negative, the proportion of ER-negative breast tumours decreases with increasing age at diagnosis (35). If the population-based ER-negative PRS were also associated primarily with ER-negative breast cancers in *BRCA1* mutation carriers, the ER-negative PRS would be more predictive of breast cancer in *BRCA1* carriers at younger ages. In contrast, in *BRCA2* carriers the proportion of ER-positive disease was found to decrease with increasing age at

diagnosis (35). Therefore, the overall PRS from the general population which is associated primarily with ER-positive breast cancers, may be more predictive of breast cancer in *BRCA2* mutation carriers at younger ages. Alternatively, it is possible that genetic risk modification has a stronger effect on developing early onset breast cancer.

A limitation of the present study is our inability to take family history into account because this information was not available for the majority of samples. Although the tests of association remain valid, it was therefore not possible to investigate how the associations vary by family cancer history.

Overall, the discrimination achieved by the PRS investigated in the current study was moderate. The highest discrimination was achieved by the ovarian cancer PRS in *BRCA2* carriers. We found the overall breast cancer PRS to have somewhat lower discriminatory ability in mutation carriers compared with the general population (20). However, given the different study designs, ER-tumour specificity in mutation carriers and different measures of relative risk, these model-performance estimates may not be directly comparable.

One possible explanation for the differences in the relative risk of the PRS between the mutation carriers and the population-based study is that not all variants identified in population-based studies are actually associated with risk in mutation carriers, perhaps as a result of functional redundancy (9). Conversely, variants that specifically modify risk in mutation carriers, examples of which have already been reported (13, 18), would not be included in PRS derived from population-based studies, and such variants might improve discrimination. On the other hand, because of the large sample sizes available in population-based studies,

the SNP selection and the logOR estimates used as weights for these PRS are likely to be more reliable than for PRS based on mutation carriers. We also derived *BRCA1*- and *BRCA2*-specific PRS that include variants discovered by population-based studies but only those showing evidence of association in mutation carriers. This approach makes use of the discovery power of population-based studies while accounting for possible mutation-carrier-specific differences in associations. However, the SNP selection and weights were based on results from the same dataset as that used in the present analysis. For this reason, we investigated the associations of mutation carrier-specific PRS without weights to reduce the possible overfitting. An analysis in an independent sample of mutation carriers will be required to assess whether these mutation-specific PRS outperform population-based PRS.

The present study demonstrates that there are large differences in the absolute cancer risks between *BRCA1* and *BRCA2* mutation carriers with higher versus lower values of the PRS. These differences are much greater than those found in population-based studies (20, 37) because the average risks conferred by *BRCA1* and *BRCA2* mutations are already high (17, 18). The clinical management of healthy women with a *BRCA1* or *BRCA2* mutation involves a combination of frequent screening, risk-reducing surgery and possibly chemoprevention (1) which can be associated with substantial side effects. In particular, RRSO leads to premature menopause, is associated with increased morbidity and has implications for family planning (38, 39). Therefore, the timing of RRSO has to be carefully considered. There are no widely accepted risk thresholds for RRSO in mutation carriers: RRSO is recommended to all carriers on the basis of their average risk. The current NCCN guidelines recommend RRSO for *BRCA1* carriers at ages 35-40 and *BRCA2* carriers at

ages 40-45 (40). The average cumulative risk of ovarian cancer by age 40 for *BRCA1* mutation carriers has been estimated as 2.8% (41). However, on the basis of our analyses, the cumulative risk of ovarian cancer for those at the lowest 1% of the PRS by age 40 is predicted to be 0.7%, and 20% of *BRCA1* mutation carriers are predicted to have a risk of ovarian cancer of <1.3% by age 40. Therefore, the current results may be used to develop risk-based thresholds for RRSO recommendations. One possibility would be to assume that women with *BRCA1* mutations would not be offered RRSO until their cumulative risk of ovarian cancer approaches or exceeds 2.8%. A similar rule has recently been recommended for the counselling of women with mutations in moderate-risk genes (42). The ages at which women with *BRCA1* mutations would reach a cumulative risk of ovarian cancer of 2.8% are 48 years for those at the 1st percentile of the PRS, and 46, 45, 44 and 43 years for those at the 5th, 10th, 20th and 30th percentile of the PRS, respectively. For these women, deferring oophorectomy to these ages as opposed to the recommended ages 35-40 may be preferable for childbearing, and to avoid very early menopause. Another option would be to use risk-based thresholds defined for the general population. For example, a 10% lifetime risk of ovarian cancer is often cited as a recommended threshold for RRSO (43). Based on our results, *BRCA2* carriers at the 10th percentile of the ovarian cancer PRS have an estimated 6% lifetime risk and approximately 38% of *BRCA2* mutation carriers have a lifetime risk of ovarian cancer which is <10%. Women at this lower end of the risk spectrum might opt to delay RRSO to near or after the natural menopause, in order to avoid the harmful longer term adverse effects of a surgically induced premature menopause, and this also provides a longer period for child bearing. Therefore, the PRS may be informative in guiding women

with *BRCA1* and *BRCA2* mutations on the optimal timing of RRSO, and can identify women at lower risk who may opt for less intensive interventions, such as salpingectomy with delayed oophorectomy.

Decisions in relation to breast cancer prevention could also be influenced by refined risk estimates. For example, the *BRCA1* carriers at the 90th percentile of the ER-negative breast cancer PRS had an estimated breast cancer risk of 19% by age 40 and 39% by age 50, compared with 5% by age 40 and 21% by age 50 for carriers at the 10th percentile of the PRS. As with RRSO, there are currently no widely accepted risk-thresholds for offering risk reducing bilateral mastectomy (RRBM) for women with *BRCA1* and *BRCA2* mutations. However, studies in non-mutation carriers have shown that the uptake and timing of RRBM is directly related to the magnitude of breast cancer risk (44) and similar arguments may be applicable to mutation carriers. To provide comprehensive risk prediction, the PRS should be combined with other risk factors, including family history. Such a model would form the foundation for the development of risk-based clinical management guidelines for mutation carriers. In parallel, it will be necessary to perform risk communication studies to assess the acceptability of risk stratification in women with *BRCA1* and *BRCA2* mutations.

In conclusion, the results demonstrate that these PRS could be useful in risk prediction for mutation carriers. Incorporating these PRS into risk prediction models for *BRCA1* and *BRCA2* mutation carriers, together with other risk modifiers, may allow for more personalised risks for *BRCA1* and *BRCA2* mutation carriers and ultimately facilitate better management of mutation carriers.

Funding

This work was supported by grant C12292/A11174 from Cancer Research – UK. Funding for the iCOGS infrastructure came from: the European Community's Seventh Framework Programme under grant agreement n° 223175 (HEALTH-F2-2009-223175) (COGS), Cancer Research UK (C1287/A10118, C1287/A 10710, C12292/A11174, C1281/A12014, C5047/A8384, C5047/A15007, C5047/A10692, C8197/A16565), the National Institutes of Health (CA128978) and Post-Cancer GWAS initiative (1U19 CA148537, 1U19 CA148065 and 1U19 CA148112 - the GAME-ON initiative), the Department of Defence (W81XWH-10-1-0341), the Canadian Institutes of Health Research (CIHR) for the CIHR Team in Familial Risks of Breast Cancer, Komen Foundation for the Cure, the Breast Cancer Research Foundation, and the Ovarian Cancer Research Fund . R01-CA122443 and P50-CA136393.

This work was supported by grant UM1 CA164920 from the National Cancer Institute. The content of this manuscript does not necessarily reflect the views or policies of the National Cancer Institute or any of the collaborating centers in the Breast Cancer Family Registry (BCFR), nor does mention of trade names, commercial products, or organizations imply endorsement by the US Government or the BCFR. BFBOCC is partly supported by: Lithuania (BFBOCC-LT): Research Council of Lithuania grant LIG-07/2012. BIDMC is supported by the Breast Cancer Research Foundation. BRCA-gene mutations and breast cancer in South African women (BMBSA) was supported by grants from the Cancer Association of South Africa (CANSA) to Elizabeth J. van Rensburg. SLN was partially supported by the Morris and Horowitz Families Endowed Professorship. This work was partially supported by Spanish Association against Cancer (AECC08), RTICC 06/0020/1060, FISPI08/1120, Mutua

Madrileña Foundation (FMMA) and SAF2010-20493. City of Hope Clinical Cancer Genetics Community Network and the Hereditary Cancer Research Registry, supported in part by Award Number RC4CA153828 (PI: J. Weitzel) from the National Cancer Institute and the Office of the Director, National Institutes of Health. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health. Funds from Italian citizens who allocated the 5x1000 share of their tax payment in support of the Fondazione IRCCS Istituto Nazionale Tumori, according to Italian laws (INT-Institutional strategic projects '5x1000') to SM and from FiorGen Foundation for Pharmacogenomics to LP. The CIMBA data management and data analysis were supported by Cancer Research – UK grants C12292/A11174 and C1287/A10118. SH is supported by an NHMRC Program Grant to GCT. ACA is a Cancer Research -UK Senior Cancer Research Fellow. GCT is an NHMRC Senior Principal Research Fellow. This research has been co-financed by the European Union (European Social Fund – ESF) and Greek national funds through the Operational Program "Education and Lifelong Learning" of the National Strategic Reference Framework (NSRF) - Research Funding Program of the General Secretariat for Research & Technology: SYN11_10_19 NBCA. Investing in knowledge society through the European Social Fund. The DKFZ study was supported by the DKFZ. EMBRACE is supported by Cancer Research UK Grants C1287/A10118 and C1287/A11990. D. Gareth Evans and Fiona Lalloo are supported by an NIHR grant to the Biomedical Research Centre, Manchester. The Investigators at The Institute of Cancer Research and The Royal Marsden NHS Foundation Trust are supported by an NIHR grant to the Biomedical Research Centre at The Institute of Cancer Research and The Royal Marsden NHS Foundation Trust. Ros Eeles and

Elizabeth Bancroft are supported by Cancer Research UK Grant C5047/A8385. Ros Eeles is also supported by NIHR support to the Biomedical Research Centre at The Institute of Cancer Research and The Royal Marsden NHS Foundation Trust. The authors acknowledge support from The University of Kansas Cancer Center (P30 CA168524) and the Kansas Bioscience Authority Eminent Scholar Program. A.K.G. was funded by 5U01CA113916, R01CA140323, and by the Chancellors Distinguished Chair in Biomedical Sciences Professorship. The German Consortium of Hereditary Breast and Ovarian Cancer (GC-HBOC) is supported by the German Cancer Aid (grant no 109076, Rita K. Schmutzler) and by the Center for Molecular Medicine Cologne (CMMC). The study was supported by the Ligue Nationale Contre le Cancer; the Association "Le cancer du sein, parlons-en!" Award; the Canadian Institutes of Health Research for the "CIHR Team in Familial Risks of Breast Cancer" program and the French National Institute of Cancer (INCa). CI received support from the Non-Therapeutic Subject Registry Shared Resource at Georgetown University (NIH/NCI grant P30-CA051008), the Fisher Center for Familial Cancer Research, and Swing Fore the Cure. Bruce Poppe is a senior clinical investigator of FWO. Was supported by a grant RD12/0036/0006 and 12/00539 from ISCIII (Spain), partially supported by European Regional Development FEDER funds. The HEBCS was financially supported by the Helsinki University Hospital Research Fund, Academy of Finland (266528), the Finnish Cancer Society and the Sigrid Juselius Foundation. The HEBO study is supported by the Dutch Cancer Society grants NKI1998-1854, NKI2004-3088, NKI2007-3756, the Netherlands Organization of Scientific Research grant NWO 91109024, the Pink Ribbon grant 110005 and the BBMRI grant NWO 184.021.007/CP46. HEBO thanks the registration teams of the Comprehensive

Cancer Centre Netherlands and Comprehensive Centre South (together the Netherlands Cancer Registry) and PALGA (Dutch Pathology Registry) for part of the data collection. HRBCP is supported by The Hong Kong Hereditary Breast Cancer Family Registry and the Dr. Ellen Li Charitable Foundation, Hong Kong. Hungarian Breast and Ovarian Cancer Study was supported by Hungarian Research Grants KTIA-OTKA CK-80745, OTKA K-112228 and the Norwegian EEA Financial Mechanism Hu0115/NA/2008-3/OP-9. ICO: Contract grant sponsor: Asociación Española Contra el Cáncer, Spanish Health Research Fund; Carlos III Health Institute; Catalan Health Institute and Autonomous Government of Catalonia. Contract grant numbers: ISCIIIRETIC RD06/0020/1051, RD12/0036/008, PI10/01422, PI10/00748, PI13/00285, PIE13/00022, 2009SGR290 and 2014SGR364. The IHCC was supported by Grant PBZ_KBN_122/P05/2004. The ILUH group was supported by the Icelandic Association “Walking for Breast Cancer Research” and by the Landspítali University Hospital Research Fund. This work was supported by the Canadian Institutes of Health Research for the “CIHR Team in Familial Risks of Breast Cancer” program, the Canadian Breast Cancer Research Alliance-grant #019511 and the Ministry of Economic Development, Innovation and Export Trade – grant # PSR-SIIRI-701. IOVHBOCS is supported by Ministero della Salute and “5x1000” Istituto Oncologico Veneto grant. This study was in part supported by Liga Portuguesa Contra o Cancro. kConFab is supported by a grant from the National Breast Cancer Foundation, and previously by the National Health and Medical Research Council (NHMRC), the Queensland Cancer Fund, the Cancer Councils of New South Wales, Victoria, Tasmania and South Australia, and the Cancer Foundation of Western Australia. KOHBRA is supported by a grant from the National R&D Program for Cancer Control,

Ministry for Health, Welfare and Family Affairs, Republic of Korea (1020350). MAYO is supported by NIH grants CA116167, CA128978 and CA176785, an NCI Specialized Program of Research Excellence (SPORE) in Breast Cancer (CA116201), a grant from the Breast Cancer Research Foundation, and a generous gift from the David F. and Margaret T. Grohne Family Foundation. Jewish General Hospital Weekend to End Breast Cancer, Quebec Ministry of Economic Development, Innovation and Export Trade. MODSQUAD was supported by MH CZ - DRO (MMCI, 00209805) and by the European Regional Development Fund and the State Budget of the Czech Republic (RECAMO, CZ.1.05/2.1.00/03.0101) to LF, and by Charles University in Prague project UNCE204024 (MZ). MSKCC is supported by grants from the Breast Cancer Research Foundation, the Robert and Kate Niehaus Clinical Cancer Genetics Initiative, and the Andrew Sabin Research Fund. 1R01 CA149429-01. The research of Drs. MH Greene and PL Mai was supported by the Intramural Research Program of the US National Cancer Institute, NIH, and by support services contracts NO2-CP-11019-50 and NO2-CP-65504 with Westat, Inc, Rockville, MD. For CIMBA PRS paper: The research of Drs. MH Greene and JT Loud was supported by the Intramural Research Program of the US National Cancer Institute, NIH, and by support services contracts NO2-CP-11019-50 and NO2-CP-65504 with Westat, Inc, Rockville, MD. NICCC is supported by Clalit Health Services in Israel. Some of it's activities are supported by the Israel Cancer Association and the Breast Cancer Research Foundation (BCRF), NY. This work has been supported by the Russian Federation for Basic Research (grants 14-04-93959 and 15-04-01744). This study was supported by National Cancer Institute grants to the NRG Oncology Administrative Office and Tissue Bank (CA 27469), the NRG Oncology Statistical and Data Center (CA 37517),

and NRG Oncology's Cancer Prevention and Control Committee (CA 101165). Drs. Greene, Mai and Savage were supported by funding from the Intramural Research Program, NCI. OSUCCG is supported by the Ohio State University Comprehensive Cancer Center. This work was supported by the ITT (Istituto Toscano Tumori) grants 2011-2013. Ministry of Science, Technology and Innovation, Ministry of Higher Education (UM.C/HIR/MOHE/06) and Cancer Research Initiatives Foundation. This project was partially funded through a grant by the Israel cancer association and the funding for the Israeli Inherited breast cancer consortium. SWE-BRCA collaborators are supported by the Swedish Cancer Society. UCHICAGO is supported by NCI Specialized Program of Research Excellence (SPORE) in Breast Cancer (CA125183), R01 CA142996, 1U01CA161032 and by the Ralph and Marion Falk Medical Research Trust, the Entertainment Industry Fund National Women's Cancer Research Alliance and the Breast Cancer research Foundation. OIO is an ACS Clinical Research Professor. Jonsson Comprehensive Cancer Center Foundation; Breast Cancer Research Foundation. UCSF Cancer Risk Program and Helen Diller Family Comprehensive Cancer Center. UKFOCR was supported by a project grant from CRUK to Paul Pharoah. National Institutes of Health (NIH) (R01-CA102776 and R01-CA083855; Breast Cancer Research Foundation; Susan G. Komen Foundation for the cure, Basser Research Center for BRCA. Frieda G. and Saul F. Shapira BRCA-Associated Cancer Research Program; Hackers for Hope Pittsburgh. Kate Lawrenson is funded by Ovarian Cancer Research Fund (OCRF) grant number 258807 and an Ann Schreiber Program of Excellence award from the Ovarian Cancer Research Fund (POE/USC/01.12). Janet Lee and Howard Shen are funded by National Institute of Health grant number 5 U19 CA148112-02. Tassja Spindler is funded by National

Institute of Health grant number CA173531-01. Work was performed within the USC Norris Comprehensive Cancer Center which is supported by a Cancer Center Support Grant (award number P30 CA014089) from the National Cancer Institute. Victorian Cancer Agency, Cancer Australia, National Breast Cancer Foundation. Dr Karlan is funded by the American Cancer Society Early Detection Professorship (SIOP-06-258-01-COUN) and the National Center for Advancing Translational Sciences (NCATS), Grant UL1TR000124.

Notes

The funders had no role in the design of the study; the collection, analysis, and interpretation of the data; the writing of the manuscript; and the decision to submit the manuscript for publication.

Contributors

KBK and ACA drafted the initial manuscript. KBK performed the statistical analyses. ACA, KBK, DFE, GCT, FC, and KO conceived and designed the study. LM and DB are the CIMBA database managers. GCT initiated and coordinates CIMBA. KBK, JD, and ML carried out the bioinformatics. All authors except KBK, DB, LM, ML, JD and AL acquired phenotypic data and DNA samples or performed SNP genotyping. All authors read and approved the final manuscript.

Acknowledgments

We wish to acknowledge Maggie Angelakos, Judi Maskiell, Gillian Dite, Helen Tsimiklis. We wish to thank members and participants in the New York site of the Breast Cancer Family Registry for their contributions to the study. We wish to thank

members and participants in the Ontario Familial Breast Cancer Registry for their contributions to the study. BFBOCC-LT acknowledge Vilius Rudaitis, Laimonas Griškevičius, Ramūnas Janavičius (if not in the authorship). BFBOCC-LV acknowledge Drs Janis Eglitis, Anna Krilova and Aivars Stengrevics. BMBSA wish to thank the families who contribute to the BMBSA study. We wish to thank Yuan Chun Ding and Linda Steele for their work in participant enrollment and biospecimen and data management. We thank Bent Ejlersen for the recruitment and genetic counseling of participants. We thank Alicia Barroso, Rosario Alonso and Guillermo Pita for their assistance. Alessandra Viel of the CRO Aviano National Cancer Institute, Aviano (PN), Italy; Laura Ottini of the "Sapienza" University, Rome, Italy; Liliana Varesco of the IRCCS AOU San Martino - IST Istituto Nazionale per la Ricerca sul Cancro, Genoa, Italy; Maria Grazia Tibiletti of the Ospedale di Circolo-Università dell'Insubria, Varese, Italy; Antonella Savarese of the Istituto Nazionale Tumori Regina Elena, Rome, Italy; Stefania Tommasi of the Istituto Nazionale Tumori "Giovanni Paolo II" - Bari, Italy; Irene Feroce of the Istituto Europeo di Oncologia, Milano, Italy; Alessandra Viel of the CRO Aviano National Cancer Institute, Aviano (PN), Italy; Liliana Varesco of the IRCCS AOU San Martino - IST Istituto Nazionale per la Ricerca sul Cancro, Genoa, Italy; Maria Grazia Tibiletti of the Ospedale di Circolo-Università dell'Insubria, Varese, Italy; Laura Ottini of the "Sapienza" University, Rome, Italy; Aline Martayan of the Istituto Nazionale Tumori Regina Elena, Rome, Italy; Stefania Tommasi of the Istituto Nazionale Tumori "Giovanni Paolo II" - Bari, Italy. The CIMBA data management and analysis is funded through Cancer Research- UK grant C12292/A11174. ACA is a Senior Cancer Research - UK Research Fellow. RE is supported by NIHR support to the Biomedical Research Centre at The Institute of Cancer Research and The Royal

Marsden NHS Foundation Trust. We thank Ms. JoEllen Weaver and Dr. Betsy Bove for their technical support. Genetic Modifiers of Cancer Risk in BRCA1/2 Mutation Carriers (GEMO) study National Cancer Genetics Network «UNICANCER Genetic Group», France. We wish to pay a tribute to Olga M. Sinilnikova, who with Dominique Stoppa-Lyonnet initiated and coordinated GEMO until she sadly passed away on the 30th June 2014, and to thank all the GEMO collaborating groups for their contribution to this study. GEMO Collaborating Centers are: Coordinating Centres, Unité Mixte de Génétique Constitutionnelle des Cancers Fréquents, Hospices Civils de Lyon - Centre Léon Bérard, & Equipe «Génétique du cancer du sein», Centre de Recherche en Cancérologie de Lyon: Olga Sinilnikova†, Sylvie Mazoyer, Francesca Damiola, Laure Barjhoux, Carole Verny-Pierre, Mélanie Léone, Nadia Boutry-Kryza, Alain Calender, Sophie Giraud; and Service de Génétique Oncologique, Institut Curie, Paris: Dominique Stoppa-Lyonnet, Marion Gauthier-Villars, Bruno Buecher, Claude Houdayer, Etienne Rouleau, Lisa Golmard, Agnès Collet, Virginie Moncoutier, Muriel Belotti, Antoine de Pauw, Camille Elan, Catherine Nogues, Emmanuelle Fourme, Anne-Marie Birot. Institut Gustave Roussy, Villejuif: Brigitte Bressac-de-Paillerets, Olivier Caron, Marine Guillaud-Bataille. Centre Jean Perrin, Clermont–Ferrand: Yves-Jean Bignon, Nancy Uhrhammer. Centre Léon Bérard, Lyon: Christine Lasset, Valérie Bonadona, Sandrine Handallou. Centre François Baclesse, Caen: Agnès Hardouin, Pascaline Berthet, Dominique Vaur, Laurent Castera. Institut Paoli Calmettes, Marseille: Hagay Sobol, Violaine Bourdon, Tetsuro Noguchi, Audrey Remenieras, François Eisinger. CHU Arnaud-de-Villeneuve, Montpellier: Isabelle Coupier, Pascal Pujol. Centre Oscar Lambret, Lille: Jean-Philippe Peyrat, Joëlle Fournier, Françoise Révillion, Philippe Vennin†, Claude Adenis. Centre

Paul Strauss, Strasbourg: Danièle Muller, Jean-Pierre Fricker. Institut Bergonié, Bordeaux: Emmanuelle Barouk-Simonet, Françoise Bonnet, Virginie Bubien, Nicolas Sevenet, Michel Longy. Institut Claudius Regaud, Toulouse: Christine Toulas, Rosine Guimbaud, Laurence Gladiéff, Viviane Feillel. CHU Grenoble: Dominique Leroux, Hélène Dreyfus, Christine Rebischung, Magalie Peysselon. CHU Dijon: Fanny Coron, Laurence Faivre. CHU St-Etienne: Fabienne Prieur, Marine Lebrun, Caroline Kientz. Hôtel Dieu Centre Hospitalier, Chambéry: Sandra Fert Ferrer. Centre Antoine Lacassagne, Nice: Marc Fréney. CHU Limoges: Laurence Vénat-Bouvet. CHU Nantes: Capucine Delnatte. CHU Bretonneau, Tours: Isabelle Mortemousque. Groupe Hospitalier Pitié-Salpêtrière, Paris: Florence Coulet, Chrystelle Colas, Florent Soubrier, Mathilde Warcoin. CHU Vandoeuvre-les-Nancy: Johanna Sokolowska, Myriam Bronner. CHU Besançon: Marie-Agnès Collonge-Rame, Alexandre Damette. Creighton University, Omaha, USA: Henry T. Lynch, Carrie L. Snyder. We wish to thank the technical support of Ilse Coene en Brecht Crombez. We acknowledge Alicia Tosar and Paula Diaque for their technical assistance. HEBCS would like to thank Dr. Kristiina Aittomäki, Taru A. Muranen, Drs. Carl Blomqvist and Kirsimari Aaltonen and RNs Irja Erkkilä and Virpi Palola for their help with the HEBCS data and samples. The Hereditary Breast and Ovarian Cancer Research Group Netherlands (HEBON) consists of the following Collaborating Centers: Coordinating center: Netherlands Cancer Institute, Amsterdam, NL: M.A. Rookus, F.B.L. Hogervorst, F.E. van Leeuwen, S. Verhoef, M.K. Schmidt, N.S. Russell, J.L. de Lange, R. Wijnands; Erasmus Medical Center, Rotterdam, NL: J.M. Collée, A.M.W. van den Ouweland, M.J. Hooning, C. Seynaeve, C.H.M. van Deurzen, I.M. Obdeijn; Leiden University Medical Center, NL: C.J. van Asperen, J.T. Wijnen, R.A.E.M. Tollenaar, P. Devilee, T.C.T.E.F. van

Cronenburg; Radboud University Nijmegen Medical Center, NL: C.M. Kets, A.R. Mensenkamp; University Medical Center Utrecht, NL: M.G.E.M. Ausems, R.B. van der Luijt, C.C. van der Pol; Amsterdam Medical Center, NL: C.M. Aalfs, T.A.M. van Os; VU University Medical Center, Amsterdam, NL: J.J.P. Gille, Q. Waisfisz, H.E.J. Meijers-Heijboer; University Hospital Maastricht, NL: E.B. Gómez-García, M.J. Blok; University Medical Center Groningen, NL: J.C. Oosterwijk, A.H. van der Hout, M.J. Mourits, G.H. de Bock; The Netherlands Foundation for the detection of hereditary tumours, Leiden, NL: H.F. Vasen; The Netherlands Comprehensive Cancer Organization (IKNL): S. Siesling, J.Verloop; The Dutch Pathology Registry (PALGA): L.I.H. Overbeek. The HE BON study is supported by the Dutch Cancer Society grants NKI1998-1854, NKI2004-3088, NKI2007-3756, the Netherlands Organization of Scientific Research grant NWO 91109024, the Pink Ribbon grants 110005 and 2014-187.WO76, the BBMRI grant NWO 184.021.007/CP46 and the Transcan grant JTC 2012 Cancer 12-054. HE BON thanks the registration teams of IKNL and PALGA for part of the data collection. We wish to thank Hong Kong Sanatorium and Hospital for their continued support. We wish to thank the Hungarian Breast and Ovarian Cancer Study Group members (Janos Papp, Tibor Vaszko, Aniko Bozsik, Timea Pocza, Judit Franko, Maria Balogh, Gabriella Domokos, Judit Ferenczi, Department of Molecular Genetics, National Institute of Oncology, Budapest, Hungary) and the clinicians and patients for their contributions to this study. We wish to thank the Oncogenetics Group (VHIO), and the High Risk and Cancer Prevention Unit of the University Hospital Vall d'Hebron. We wish to thank the ICO Hereditary Cancer Program team led by Dr. Gabriel Capella. We would like to thank Dr Martine Dumont, Martine Tranchant for sample management and skillful technical assistance. J.S. is Chairholder of the

Canada Research Chair in Oncogenetics. J.S. and P.S. were part of the QC and Genotyping coordinating group of iCOGS (BCAC and CIMBA). We wish to thank Drs. Ana Peixoto, Catarina Santos, Patrícia Rocha and Pedro Pinto for their skillful contribution to the study. We wish to thank Heather Thorne, Eveline Niedermayr, all the kConFab research nurses and staff, the heads and staff of the Family Cancer Clinics, and the Clinical Follow Up Study (which has received funding from the NHMRC, the National Breast Cancer Foundation, Cancer Australia, and the National Institute of Health (USA)) for their contributions to this resource, and the many families who contribute to kConFab. Modifier Study of Quantitative Effects on Disease (MODSQUAD): MODSQUAD acknowledges ModSQuaD members Csilla Szabo (National Human Genome Research Institute, National Institutes of Health, Bethesda, MD, USA); Lenka Foretova and Eva Machackova (Department of Cancer Epidemiology and Genetics, Masaryk Memorial Cancer Institute and MF MU, Brno, Czech Republic); and Michal Zikan, Petr Pohlreich and Zdenek Kleibl (Oncogynecologic Center and Department of Biochemistry and Experimental Oncology, First Faculty of Medicine, Charles University, Prague, Czech Republic). Anne Lincoln, Lauren Jacobs. We wish to thank the NICCC National Familial Cancer Consultation Service team led by Sara Dishon, the lab team led by Dr. Flavio Lejbkowitz, and the research field operations team led by Dr. Mila Pinchev. We thank the investigators of the Australia New Zealand NRG Oncology group. We wish to thank members and participants in the Ontario Cancer Genetics Network for their contributions to the study. Leigha Senter, Kevin Sweet, Julia Cooper, Caroline Craven, and Michelle O'Connor were instrumental in accrual of study participants, ascertainment of medical records and database management. Samples were

processed by the OSU Human Genetics Sample Bank. We would like to thank Yip Cheng Har, Nur Aishah Mohd Taib, Phuah Sze Yee, Norhashimah Hassan and all the research nurses, research assistants and doctors involved in the MyBrCa Study for assistance in patient recruitment, data collection and sample preparation. In addition, we thank Philip Iau, Sng Jen-Hwei and Sharifah Nor Akmal for contributing samples from the Singapore Breast Cancer Study and the HUKM-HKL Study respectively. The Malaysian Breast Cancer Genetic Study is funded by research grants from the Malaysian Ministry of Science, Technology and Innovation, Ministry of Higher Education (UM.C/HIR/MOHE/06) and charitable funding from Cancer Research Initiatives Foundation. SMC team wishes to acknowledge the assistance of the Meirav Comprehensive breast cancer center team at the Sheba Medical Center for assistance in this study. Swedish scientists participating as SWE-BRCA collaborators are: from Lund University and University Hospital: Åke Borg, Håkan Olsson, Helena Jernström, Karin Henriksson, Katja Harbst, Maria Soller, Ulf Kristoffersson; from Gothenburg Sahlgrenska University Hospital: Anna Öfverholm, Margareta Nordling, Per Karlsson, Zakaria Einbeigi; from Stockholm and Karolinska University Hospital: Anna von Wachenfeldt, Annelie Liljegren, Annika Lindblom, Brita Arver, Gisela Barbany Bustinza, Johanna Rantala; from Umeå University Hospital: Beatrice Melin, Christina Edwindsdotter Ardnor, Monica Emanuelsson; from Uppsala University: Hans Ehrencrona, Maritta Hellström Pigg, Richard Rosenquist; from Linköping University Hospital: Marie Stenmark-Askmal, Sigrun Liedgren. We wish to thank Cecilia Zvocec, Qun Niu, physicians, genetic counselors, research nurses and staff of the Cancer Risk Clinic for their contributions to this resource, and the many families who contribute to our program. We thank Joyce Seldon MSGC and Lorna

Kwan, MPH for assembling the data for this study. We would like to thank Dr. Robert Nussbaum and the following genetic counselors for participant recruitment: Beth Crawford, Kate Loranger, Julie Mak, Nicola Stewart, Robin Lee, Amie Blanco and Peggy Conrad. And thanks to Ms. Salina Chan for her data management. We thank Simon Gayther, Carole Pye, Patricia Harrington and Eva Wozniak for their contributions towards the UKFOCR. Geoffrey Lindeman, Marion Harris, Martin Delatycki of the Victorian Familial Cancer Trials Group. We thank Sarah Sawyer and Rebecca Driessen for assembling this data and Ella Thompson for performing all DNA amplification.

References

1. Evans DG, Graham J, O'Connell S, *et al.* Familial breast cancer: summary of updated NICE guidance. *BMJ* 2013;346:f3829.
2. Clark AS, Domchek SM. Clinical management of hereditary breast cancer syndromes. *J Mammary Gland Biol Neoplasia* 2011;16(1):17-25.
3. Parker WH, Jacoby V, Shoupe D, *et al.* Effect of bilateral oophorectomy on women's long-term health. *Womens Health (Lond Engl)* 2009;5(5):565-76.
4. Rivera CM, Grossardt BR, Rhodes DJ, *et al.* Increased mortality for neurological and mental diseases following early bilateral oophorectomy. *Neuroepidemiology* 2009;33(1):32-40.
5. Rivera CM, Grossardt BR, Rhodes DJ, *et al.* Increased cardiovascular mortality after early bilateral oophorectomy. *Menopause* 2009;16(1):15-23.
6. Rocca WA, Bower JH, Maraganore DM, *et al.* Increased risk of cognitive impairment or dementia in women who underwent oophorectomy before menopause. *Neurology* 2007;69(11):1074-83.
7. Eccles DM. Identification of personal risk of breast cancer: genetics. *Breast Cancer Res* 2008;10 Suppl 4:S12.
8. Michailidou K, Hall P, Gonzalez-Neira A, *et al.* Large-scale genotyping identifies 41 new loci associated with breast cancer risk. *Nat Genet* 2013;45(4):353-61, 361e1-2.
9. Kuchenbaecker KB, Ramus SJ, Tyrer J, *et al.* Identification of six new susceptibility loci for invasive epithelial ovarian cancer. *Nat Genet* 2015;47(2):164-71.
10. Michailidou K, Beesley J, Lindstrom S, *et al.* Genome-wide association analysis of more than 120,000 individuals identifies 15 new susceptibility loci for breast cancer. *Nat Genet* 2015;47(4):373-80.
11. Kuchenbaecker KB, Neuhausen SL, Robson M, *et al.* Associations of common breast cancer susceptibility alleles with risk of breast cancer subtypes in BRCA1 and BRCA2 mutation carriers. *Breast Cancer Res* 2014;16(6):3416.
12. Couch FJ, Wang X, McGuffog L, *et al.* Genome-wide association study in BRCA1 mutation carriers identifies novel loci associated with breast and ovarian cancer risk. *PLoS Genet* 2013;9(3):e1003212.
13. Gaudet MM, Kuchenbaecker KB, Vijai J, *et al.* Identification of a BRCA2-specific modifier locus at 6p24 related to breast cancer risk. *PLoS Genet* 2013;9(3):e1003173.
14. Chowdhury S, Dent T, Pashayan N, *et al.* Incorporating genomics into breast and prostate cancer screening: assessing the implications. *Genet Med* 2013;15(6):423-32.
15. Antoniou AC, Beesley J, McGuffog L, *et al.* Common breast cancer susceptibility alleles and the risk of breast cancer for BRCA1 and BRCA2 mutation carriers: implications for risk prediction. *Cancer Res* 2010;70(23):9742-54.
16. Milne RL, Antoniou AC. Genetic modifiers of cancer risk for BRCA1 and BRCA2 mutation carriers. *Ann Oncol* 2011;22 Suppl 1:i11-7.
17. Antoniou AC, Beesley J, McGuffog L, *et al.* Common breast cancer susceptibility alleles and the risk of breast cancer for BRCA1 and BRCA2 mutation carriers: implications for risk prediction. *Cancer Res* 2010;70(23):9742-9754.

18. Couch FJ, Wang X, McGuffog L, *et al.* Genome-wide association study in BRCA1 mutation carriers identifies novel loci associated with breast and ovarian cancer risk. *PLoS Genet* 2013;9(3):e1003212.
19. Mavaddat N, Peock S, Frost D, *et al.* Cancer risks for BRCA1 and BRCA2 mutation carriers: results from prospective analysis of EMBRACE. *J Natl Cancer Inst* 2013;105(11):812-22.
20. Mavaddat N, Pharoah PD, Michailidou K, *et al.* Prediction of breast cancer risk based on profiling with common genetic variants. *J Natl Cancer Inst* 2015;107(5).
21. Lin WY, Camp NJ, Ghousaini M, *et al.* Identification and characterization of novel associations in the CASP8/ALS2CR12 region on chromosome 2 with breast cancer risk. *Hum Mol Genet* 2015;24(1):285-98.
22. Ghousaini M, Edwards SL, Michailidou K, *et al.* Evidence that breast cancer risk at the 2q35 locus is mediated through IGFBP5 regulation. *Nat Commun* 2014;4:4999.
23. Bojesen SE, Pooley KA, Johnatty SE, *et al.* Multiple independent variants at the TERT locus are associated with telomere length and risks of breast and ovarian cancer. *Nat Genet* 2013;45(4):371-84, 384e1-2.
24. French JD, Ghousaini M, Edwards SL, *et al.* Functional variants at the 11q13 risk locus for breast cancer regulate cyclin D1 expression through long-range enhancers. *Am J Hum Genet* 2013;92(4):489-503.
25. Meyer KB, O'Reilly M, Michailidou K, *et al.* Fine-scale mapping of the FGFR2 breast cancer risk locus: putative functional variants differentially bind FOXA1 and E2F1. *Am J Hum Genet* 2013;93(6):1046-60.
26. Dunning AM, Michailidou K, Kuchenbaecker KB, *et al.* Breast cancer risk variants at 6q25 display different phenotype associations and regulate ESR1, RMND1 and CCDC170. *Nat Genet* 2016;48(4):374-86.
27. Southey MC, Goldgar DE, Winqvist R, *et al.* PALB2, CHEK2 and ATM rare variants and cancer risk: data from COGS. *J Med Genet* 2016; 10.1136/jmedgenet-2016-103839.
28. BCAC. *iCOGS summary results*.
<http://bcac.ccge.medschl.cam.ac.uk/bcacdata/icogs-summary-results/>.
29. Antoniou AC, Goldgar DE, Andrieu N, *et al.* A weighted cohort approach for analysing factors modifying disease risks in carriers of high-risk susceptibility genes. *Genet Epidemiol* 2005;29(1):1-11.
30. Harrell FE, Califf RM, Pryor DB, *et al.* Evaluating the Yield of Medical Tests. *Jama-Journal of the American Medical Association* 1982;247(18):2543-2546.
31. Amin Al Olama A, Benlloch S, Antoniou AC, *et al.* Risk Analysis of Prostate Cancer in PRACTICAL, a Multinational Consortium, Using 25 Known Prostate Cancer Susceptibility Loci. *Cancer Epidemiol Biomarkers Prev* 2015;24(7):1121-9.
32. Aulchenko YS, Ripke S, Isaacs A, *et al.* GenABEL: an R library for genome-wide association analysis. *Bioinformatics* 2007;23(10):1294-6.
33. StataCorp. *Stata Statistical Software: Release 13*. In. College Station, TX: StataCorp LP; 2013.
34. Lakhani SR, Jacquemier J, Sloane JP, *et al.* Multifactorial analysis of differences between sporadic breast cancers and cancers involving BRCA1 and BRCA2 mutations. *J Natl Cancer Inst* 1998;90(15):1138-45.

35. Mavaddat N, Barrowdale D, Andrulis IL, *et al.* Pathology of breast and ovarian cancers among BRCA1 and BRCA2 mutation carriers: results from the Consortium of Investigators of Modifiers of BRCA1/2 (CIMBA). *Cancer Epidemiol Biomarkers Prev* 2012;21(1):134-47.
36. Symons MJ, Moore DT. Hazard rate ratio and prospective epidemiological studies. *J Clin Epidemiol* 2002;55(9):893-9.
37. Husing A, Canzian F, Beckmann L, *et al.* Prediction of breast cancer risk by genetic risk factors, overall and by hormone receptor status. *J Med Genet* 2012;49(9):601-8.
38. Rocca WA, Grossardt BR, de Andrade M, *et al.* Survival patterns after oophorectomy in premenopausal women: a population-based cohort study. *Lancet Oncol* 2006;7(10):821-8.
39. Rebbeck TR, Kauff ND, Domchek SM. Meta-analysis of risk reduction estimates associated with risk-reducing salpingo-oophorectomy in BRCA1 or BRCA2 mutation carriers. *J Natl Cancer Inst* 2009;101(2):80-7.
40. NCCN. *NCCN Clinical Practice Guidelines in Oncology: Genetic/Familial High-Risk Assessment: Breast and Ovarian. V1. 2016.*
http://www.nccn.org/professionals/physician_gls/pdf/genetics_screening.pdf.
41. Antoniou AC, Cunningham AP, Peto J, *et al.* The BOADICEA model of genetic susceptibility to breast and ovarian cancers: updates and extensions. *Br J Cancer* 2008;98(8):1457-66.
42. Tung N, Domchek SM, Stadler Z, *et al.* Counselling framework for moderate-penetrance cancer-susceptibility mutations. *Nat Rev Clin Oncol* 2016;13(9):581-8.
43. Anderson K, Jacobson JS, Heitjan DF, *et al.* Cost-effectiveness of preventive strategies for women with a BRCA1 or a BRCA2 mutation. *Ann Intern Med* 2006;144(6):397-406.
44. Evans DG, Lalloo F, Ashcroft L, *et al.* Uptake of risk-reducing surgery in unaffected women at high risk of breast and ovarian cancer is risk, age, and time dependent. *Cancer Epidemiol Biomarkers Prev* 2009;18(8):2318-24.

Tables

Table 1. Per-standard-deviation hazard ratios (HR) and 95% confidence intervals (CI) for the associations of polygenic risk scores (PRS) with breast (BC) and ovarian cancer (OC) risk in *BRCA1* and *BRCA2* carriers*

PRS	No. of SNPs	<i>BRCA1</i> carriers		<i>BRCA2</i> carriers	
		HR (95%CI)	p†	HR (95%CI)	p†
Outcome: breast cancer					
Overall BC PRS	88	1.14 (1.11-1.17)	1.8x10 ⁻¹⁸	1.22 (1.17-1.28)	7.2x10 ⁻²⁰
ER-positive ^{&} BC PRS	87	1.11 (1.08-1.15)	3.5x10 ⁻¹³	1.22 (1.16-1.27)	4.0x10 ⁻¹⁹
ER-negative ^{&} BC PRS	53	1.27 (1.23-1.31)	8.2x10 ⁻⁵³	1.15 (1.10-1.20)	6.8x10 ⁻¹⁰
Outcome: ovarian cancer					
OC PRS	17	1.28 (1.22-1.34)	2.5x10 ⁻²⁶	1.49 (1.34-1.65)	8.5x10 ⁻¹⁴

*The PRS created from the latest reported population-based study results were used.

†P-value for a two-sided test using a weighted cohort Cox-regression with time to breast or ovarian cancer diagnosis, respectively, as the outcome

[&]oestrogen-receptor-positive and -negative

Table 2. Proportion of samples and number of events in percentile categories of polygenic risk scores (PRS) and their associations with breast and ovarian cancer risks*

Percentile category in %	<i>BRCA1</i> carriers			<i>BRCA2</i> carriers		
	% samples in percentile category	No. of events	HR (95%CI)†	% samples in percentile category	No. of events	HR (95%CI)†
Outcome: Breast cancer						
0-5	3.6	222	0.76 (0.64-0.91)	4.0	138	0.80 (0.63-1.02)
5-10	4.1	250	0.70 (0.59-0.82)	4.2	142	0.68 (0.54-0.87)
10-20	8.7	551	0.77 (0.68-0.87)	8.9	340	0.92 (0.77-1.09)
20-40	18.7	1377	0.98 (0.89-1.07)	18.8	764	1.00 (0.87-1.15)
40-60	20.4	1534	1 (reference)	19.1	793	1 (reference)
60-80	21.0	1729	1.21 (1.11-1.33)	21.2	950	1.16 (1.02-1.32)
80-90	11.0	950	1.32 (1.19-1.47)	11.4	557	1.37 (1.17-1.60)
90-95	5.8	519	1.50 (1.31-1.72)	5.8	309	1.76 (1.43-2.17)
95-100	6.7	665	1.82 (1.61-2.07)	6.7	337	1.51 (1.25-1.82)
Outcome: Ovarian cancer						
0-5	4.7	85	0.66 (0.51-0.86)	4.8	20	0.76 (0.39-1.47)
5-10	5.3	110	0.81 (0.64-1.02)	5.3	18	0.67 (0.34-1.32)
10-20	10.5	215	0.80 (0.66-0.96)	10.4	39	0.87 (0.54-1.39)
20-40	20.9	478	0.95 (0.82-1.10)	20.4	104	1.02 (0.71-1.46)
40-60	19.9	468	1 (reference)	20.4	107	1 (reference)
60-80	19.5	519	1.19 (1.03-1.38)	19.5	159	1.73 (1.25-2.40)
80-90	9.3	267	1.43 (1.20-1.70)	9.1	76	1.84 (1.24-2.72)
90-95	4.9	155	1.54 (1.24-1.91)	4.8	45	1.87 (1.16-3.02)
95-100	5.1	165	1.86 (1.51-2.29)	5.4	63	3.04 (2.00-4.61)

* The PRS created from reported population-based study results were used. The percentile boundaries were derived assuming a normally-distributed PRS. The oestrogen receptor-negative breast cancer PRS was used for the associations with breast cancer risk in *BRCA1* carriers and overall breast cancer PRS in *BRCA2* carriers. CI=confidence interval;

† HR=hazard ratio from a weighted cohort Cox-regression with time to breast or ovarian cancer diagnosis, respectively, as the outcome

Table 3. Age-specific hazard ratio (HR) estimates for the PRS associations and HR estimates for a PRS x age interaction*

Age category	<i>BRCA1</i> carriers			<i>BRCA2</i> carriers		
	No. of events	HR per unit SD increase in the ER-PRS	P†	No. of events	HR per unit SD increase in the overall BC PRS	P†
Outcome: Breast cancer						
18-39	4125	1.63 (1.52-1.74)	-	1731	1.65 (1.44-1.88)	-
40-49	2557	1.18 (1.13-1.23)	4.2x10 ⁻¹⁵	1587	1.22 (1.14-1.31)	8.5x10 ⁻⁵
50-59	846	1.14 (1.09-1.21)	0.40	718	1.10 (1.02-1.19)	0.05
≥60	269	1.20 (1.11-1.29)	0.33	294	1.12 (1.03-1.23)	0.75
Interaction HR		0.993 (0.990-0.996)	3.3x10 ⁻⁶		0.995 (0.991-0.999)	0.01
Main effect PRS		1.69 (1.50-1.91)			1.55 (1.29-1.87)	
Outcome: Ovarian cancer						
18-49	1258	1.55 (1.42-1.69)		172	3.05 (2.35-3.97)	
50-59	808	1.11 (1.05-1.18)	1.1x10 ⁻⁹	227	1.52 (1.26-1.84)	8.2x10 ⁻⁶
≥60	396	1.14 (1.06-1.21)	0.67	232	1.21 (1.12-1.30)	0.03
Interaction HR		0.992 (0.988-0.998)	0.003		0.991 (0.979-1.00)	0.11
Main effect PRS		1.83 (1.43-2.34)			2.48 (1.34-4.58)	

* The population-derived PRS for oestrogen receptor-negative breast cancer was used for the associations with breast cancer in *BRCA1* carriers and the overall breast cancer PRS in *BRCA2* carriers. P-value relate to the difference in PRS association between each age group from the preceding younger group and to the interaction term.

† P-value for a two-sided test using a weighted cohort Cox-regression with time to breast or ovarian cancer diagnosis, respectively, as the outcome

Table 4. Discrimination of population-derived polygenic risk scores (PRS) for breast (BC) and ovarian cancer (OC) in *BRCA1* and *BRCA2* carriers

PRS	Harrell's c statistic (95%confidence interval)	
	<i>BRCA1</i> carriers	<i>BRCA2</i> carriers
Discrimination for breast cancer		
Overall BC PRS	0.541 (0.530-0.551)	0.566 (0.551-0.581)
ER-positive BC PRS	0.532 (0.522-0.543)	0.566 (0.551-0.581)
ER-negative BC PRS	0.581 (0.571-0.592)	0.538 (0.523-0.553)
Discrimination for ovarian cancer		
OC PRS	0.579 (0.559-0.600)	0.628 (0.592-0.665)

Figure legends

Figure 1. Hazard ratios (HR) and 95% confidence intervals (error bars) for percentiles of the polygenic risk score (PRS) relative to the middle quintile. The oestrogen receptor-negative breast cancer (BC) PRS (A) and the overall-BC PRS (C) were used for breast cancer in *BRCA1* and *BRCA2* carriers, respectively, and the ovarian cancer (OC) PRS for the OC associations (B, D). Lines denote the theoretical estimates under a multiplicative polygenic model with means and standard deviations of $\bar{x}=0.10$ and $SD=0.41$ for the ER-negative BC PRS, $\bar{x}=0.41$ and $SD=0.50$ for the overall BC PRS, $\bar{x}=0.47$ and $SD=0.37$ for the OC PRS.

Figure 2. Predicted breast cancer risks by percentile of the polygenic risk scores (PRS). The oestrogen receptor-negative breast cancer PRS was used for *BRCA1* carriers (A) and the overall breast cancer PRS for *BRCA2* carriers (C). Ovarian cancer risks are given by percentile of the ovarian cancer PRS in *BRCA1* (B) and *BRCA2* (D) carriers. Age-specific PRS associations were used to calculate these cumulative risks.