



Coccolithophore ecology in the tropical and subtropical Atlantic Ocean: New perspectives from the Atlantic meridional transect (AMT) programme



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ABSTRACT

Coccolithophore species composition was determined in 199 samples collected from the upper 300 m of the Atlantic Ocean, spanning temperate, tropical and subtropical waters in both hemispheres during four Atlantic Meridional Transect (AMT) cruises over the period 2003–2005. Of the 171 taxa observed, 140 consistently represented <5% of total cell numbers, and were classed as rare. Multivariate statistical techniques were used on the common taxa to assess variability in community composition vertically in the water column, horizontally across hydrographic provinces (subtropical gyres, equatorial waters, temperate waters), and temporally between cruises. Sharper gradients of statistical dissimilarity in species composition occurred vertically over a few tens of metres than horizontally over hundreds of kilometres. Three floral groups were identified from analysis of the depth of normalised abundance maxima in the subtropical gyres and equatorial waters: the upper euphotic zone (UEZ, >10% surface irradiance); the lower euphotic zone (LEZ, 10–1% surface irradiance); and the sub-euphotic zone (SEZ, <1% surface irradiance). The LEZ includes the deep chlorophyll maximum (DCM) and nutricline, and was characterised by species such as *Emiliana huxleyi* and *Gephyrocapsa ericsonii* which were also abundant at higher latitudes. It is suggested that this pattern reflects similarities in the light (and inorganic nutrient) conditions between the LEZ and temperate waters. The SEZ is below the depth where light is thought to be sufficient to support photosynthesis, suggesting that deep-dwelling species such as *Florisphaera profunda* and *Gladiolithus* spp. may be mixotrophic or phagotrophic, although conclusive proof will need to be gained experimentally. Mixotrophy could also be an important nutritional strategy for species abundant (*Umbellosphaera* spp., holococcolithophores) in the UEZ where inorganic nutrient concentrations are depleted and limiting to growth, although other nutritional strategies, such as the use of organic nutrients, are also possible. Statistical differences were also found in the species composition between the different cruises, with high levels of similarity for similar timed cruises (May or September–October). Few individual taxa showed significant variability in abundance over the time-span of sampling, except species such as *E. huxleyi* and *G. ericsonii* at higher latitudes. In subtropical and equatorial waters, high levels of species richness and low levels of species dominance remained throughout the sampling period indicating that seasonal fluctuations reflected differences in the whole coccolithophore community rather than in just one or a few species. Multivariate analyses of the taxa classified as rare also indicated some level of temporal, as well as vertical, zonation. Such insights into coccolithophore ecology and community composition provide important new perspectives that require innovative research to fully understand their impact on ocean biogeochemistry.

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1. Introduction

Coccolithophores are unicellular marine algae, belonging to the Class Prymnesiophyceae, that possess one or more external layers of calcite scales (coccoliths). Coccoliths are formed intracellularly via calcification and are then extruded onto the external surface

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of the cell to form a coccosphere. Around 200 species of coccolithophore are extant in the modern ocean (Young et al., 2003), with considerable diversity in the shape of the cell, the shape, construction and crystallography of the coccoliths, and their number, diversity and geometry around the cell (Monteiro et al., 2016). Variability in the crystallography of the coccoliths is based around two forms which represent linked species' life stages and reproductive strategies; in their diploid form many species produce heterococcoliths (HET), composed of complex radially arranged inter-grown calcite crystals; in contrast, in the haploid form many species produce holococcoliths (HOL) formed of small equidimensional calcite crystals (see Monteiro et al., 2016 and references therein). Transitions between HOL and HET life forms are now recognised as being a key component of how coccolithophore species can adapt to changing environmental conditions (Houdan et al., 2004, 2006), although the biogeography of many HOL (and HET) forms has still not been fully identified (e.g., Dimiza et al., 2008; Cros and Estrada, 2013).

Although global production of calcite by coccolithophores is still not well constrained (Berelson et al., 2007; Poulton et al., 2007), coccoliths make up the major fraction of carbonate in marine sediments (Broecker and Clark, 2009). Furthermore, extensive dissolution of coccoliths occurs both in the upper water column due to biological processes (Milliman et al., 1999; Poulton et al., 2006a) and in deep waters situated below the lysocline for calcite. Geochemical evidence suggests that low latitude calcite production is an important term in the global carbonate budget (Sarmiento et al., 2002; Berelson et al., 2007) and, in the context of climate change and ocean acidification, considerable attention is now being given to how the physiology and ecology of coccolithophores will respond to, and affect, air-sea exchanges of carbon dioxide (e.g., Bach et al., 2015; O'Brien et al., 2016). However, our knowledge of the distributions and nutritional strategies of coccolithophores across the extensive ocean gyres is still based on relatively few detailed studies (e.g., McIntyre and Be, 1967; Winter et al., 1994; Hagino et al., 2000; Haidar and Thierstein, 2001). In this paper the coccolithophore communities in equatorial and subtropical waters of the Atlantic Ocean are examined through sampling on the Atlantic Meridional Transect (AMT) programme (Robinson et al., 2006).

The growth environment of coccolithophores, and other phytoplankton, living in equatorial and subtropical waters is strongly regulated by opposing gradients in light and nutrient availability. The upper water column above the semi-permanent thermocline is characterised by low (nano-molar; $<100 \text{ nmol kg}^{-1}$) inorganic nutrient concentrations and high levels of irradiance, whilst below the thermocline nutrient concentrations are relatively high (micro-molar; $>1 \text{ } \mu\text{mol kg}^{-1}$) and light levels are low. Vertical profiles of chlorophyll fluorescence show a deep chlorophyll-*a* maximum (DCM) associated with the deep increase in nutrients (a nutricline), at depths ranging from $\sim 60 \text{ m}$ around the tropical Equator to as much as 150 m in the subtropical gyres. The DCM is generally regarded as neither a maximum in phytoplankton biomass or in primary production (Marañón et al., 2000; Perez et al., 2005, 2006), but marks a depth where the relative availability of surface irradiance has decreased to 1% of its surface value and where photo-acclimation results in increased cellular chlorophyll levels. Phytoplankton associated with the DCM may be considered to be light-limited rather than nutrient-limited (Venrick, 1982; Poulton et al., 2006b), whereas the situation is reversed in the upper ocean.

The phytoplankton community in the warm, stratified waters of the Atlantic are dominated in terms of abundance, biomass and primary production by small ($<2 \text{ } \mu\text{m}$) picoplankton, mainly prokaryotes such as *Prochlorococcus* and *Synechococcus*, as well as small eukaryotes (Tarran et al., 2006; Zubkov, 2014). The nanoplankton ($2\text{--}20 \text{ } \mu\text{m}$), which includes most if not all species

of coccolithophore as well non-calcifying haptophytes (Liu et al., 2009), generally contribute 10–20% of biomass (chlorophyll-*a*), with higher values in equatorial waters and higher latitude waters during spring (Marañón et al., 2001; Poulton et al., 2006b). Simple estimates of coccolithophore contributions to primary production, based on calcification rates and cellular ratios of inorganic to organic production, imply that coccolithophores account for only $\sim 1\text{--}10\%$ (maximum 20%) of total phytoplankton primary production in equatorial and subtropical waters (Poulton et al., 2006a, 2007). Such low coccolithophore contributions match with similar estimates made in temperate and sub-polar environments, with significant contributions ($>30\text{--}40\%$) limited to coccolithophore blooms (Poulton et al., 2007, 2010, 2013, 2014).

Although coccolithophores are only a minor component of the phytoplankton community in the oligotrophic ocean they play an important role in the exchange of both carbon and sulphur between the atmosphere and surface waters and in the downward transport of biogenic material and its' accumulation in marine sediments (e.g., Holligan et al., 1993; Malin and Steinke, 2004; de Vargas et al., 2007; Ziveri et al., 2007). It is appropriate, therefore, to ask how the growth of coccolithophores in oligotrophic waters is regulated and how it might be affected by future changes in the oceanic environment (e.g., Bach et al., 2015). Despite numerous experimental studies of photosynthesis and calcification by coccolithophores (e.g., Bach et al., 2013), as well as examination of coccoliths in sediments across glacial-interglacial transitions (Beaufort et al., 2011; Meier et al., 2014), the effects on coccolithophore ecology of future changes in ocean carbonate chemistry due to rising CO_2 levels remain uncertain (Bach et al., 2015). This problem stems partly from the difficulties of growing most oceanic taxa in the laboratory and an incomplete understanding of their ecology and nutritional strategies in the open ocean.

Both latitudinal biogeographical and floral depth zones have been recognised on the basis of characteristic coccolithophore species assemblages (e.g., McIntyre and Be, 1967; Okada and McIntyre, 1977; Winter et al., 1994); with further work in the Atlantic (Kinkel et al., 2000; Haidar and Thierstein, 2001; Boeckel and Baumann, 2008) and Pacific (Hagino et al., 2000; Cortes et al., 2001) confirming the generality of earlier observations. Relative to the high cell density ($>1000 \text{ cells mL}^{-1}$) blooms that may occur in sub-polar waters, the reported abundances for surface subtropical waters of the Atlantic are in the range of $<1\text{--}300$ coccospheres (cells mL^{-1}) (the higher values being generally dominated by *Emiliana huxleyi*), with ~ 25 coccospheres mL^{-1} being typical of low-latitude gyre waters (e.g., Boeckel and Baumann, 2008). Seasonal variations in abundance are most pronounced in higher latitude waters ($>30^\circ\text{N/S}$) (e.g., Knappertsbusch and Brummer, 1995; Boeckel and Baumann, 2008; Baumann et al., 2008) and in equatorial upwelling regions (e.g., Kinkel et al., 2000), as a result of changes in water column stratification and nutrient distributions.

In this paper the results of SEM analyses of coccolithophore samples collected at depths of $5\text{--}300 \text{ m}$ between 47°N and 43°S on four AMT cruises during the period 2003–2005 are reported. The AMT programme provides an excellent platform to examine coccolithophore dynamics as it provides a wealth of ancillary environmental and ecological information to provide valuable contextual perspectives to view coccolithophore ecology in the wide expanses of the oligotrophic Atlantic Ocean. Our analysis of the coccolithophore database resulting from SEM analyses has four main aims: (1) to describe the general species composition of equatorial and subtropical communities in terms of species composition, relative abundance and diversity; (2) to examine the variability in vertical structure of the community in relation to light and nutrient availability; (3) to compare and contrast vertical biogeography with latitudinal variability in species composition; and (4) to investigate whether any temporal (inter-cruise) variability in

species composition is evident across the short time period of sampling (2003–2005).

2. Materials and methods

2.1. Sampling

The cruise tracks and dates for the cruises on which samples were collected for analyses of the coccolithophore communities are shown in Fig. 1. Coccolithophore data are analysed in relation to several hydrographic provinces across the Atlantic Ocean: Northern Gyre waters (NG, 35°N–10°N), equatorial waters (EQ, 10°N–10°S), and Southern Gyre waters (SG, 10°S–30°S), with temperate (TMP) waters >35°N and >30°S (see Robinson et al., 2006; Poulton et al., 2006b; Fig. 1). Seawater samples were collected during daily pre-dawn (02:00–04:00 h, local time) and mid-morning (11:00–12:00 h) deployments of a rosette sampler fitted with 24 water bottles (20 L), a Sea-Bird 9/11 CTD, and a Chelsea MKIII Aquatrack fluorometer. Chlorophyll-*a* and nitrate concentrations were measured at 10–15 depths from each CTD cast, whilst samples for coccolithophore enumeration were collected from 5 to 13 water depths over the upper 300 m. The standard sampling depths for the AMT programme during 2003–2005 (Poulton et al., 2006b; Robinson et al., 2006) were the depths of 55, 33, 14, 1 and 0.1% of surface irradiance (with optical depths of 0.6, 1.1, 2.0, 4.6 and 6.9, respectively); additional samples were collected at selected stations during AMT 14 to provide a more complete description of properties across the DCM down to 300 m.

Light depths were determined from the previous days light measurements, or assuming that the deep fluorescence maximum (DCM) approximates the depth (Z_{DCM} ; m) of 1% surface irradiance (optical depth = 4.6). The vertical attenuation coefficient of Photosynthetically Active Radiation (PAR; m^{-1}) was determined for each station as $4.6/Z_{DCM}$ and subsequent light depths determined as optical depth divided by the vertical attenuation coefficient. Optical depths for samples not collected at the five standard light depths were calculated as depth (Z ; m) \times vertical attenuation coefficient (m^{-1}). Sea-surface temperature was determined from the CTD sensors during daily deployments. Chlorophyll-*a* measurements were made on 250 mL water samples filtered onto Whatman GF/F filters, extracted in 90% acetone for 18–20 h at 4 °C, and measured on a TD700 Turner Designs fluorometer calibrated with a pure chlorophyll-*a* standard (Sigma-Aldrich, UK) (see Poulton et al., 2006a, 2006b for further details). Micromolar ($\mu mol kg^{-1}$) concentrations of nitrate were measured with a 5-channel Technicon, segmented flow colourimetric auto-analyser (Bran + Luebbe AAIL) (see Poulton et al., 2006a, 2006b for further details).

2.2. Coccolithophore enumeration and identification

For coccolithophore enumeration, 1–2 L of seawater was taken from 20 L Niskin bottles mounted on a CTD carousel, and filtered under gentle suction onto 25 mm 0.45 μm polycarbonate filters which were then dried at room temperature and stored in sealed Perspex boxes. The latitudes and depths for which samples were collected on the AMT 12 (May 2003) and AMT 14 (May 2004)

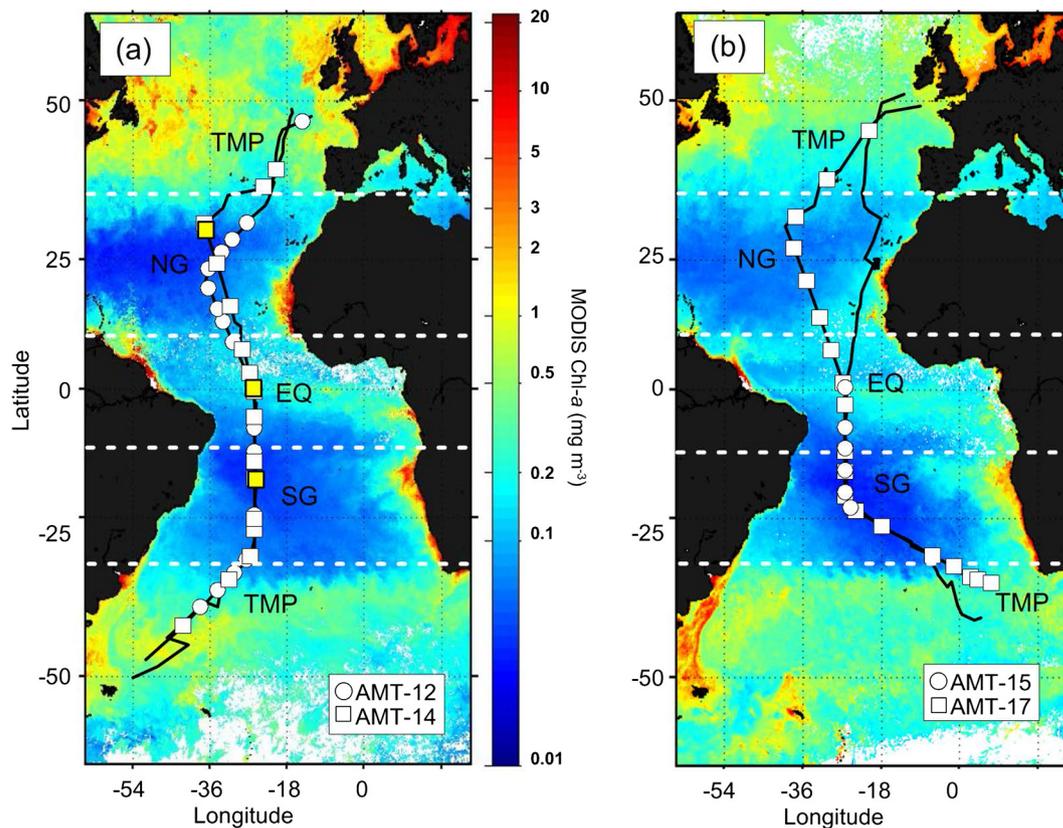


Fig. 1. Station positions (symbols) and cruise tracks (solid black lines) for the four AMT cruises sampled for coccolithophores in this study. (a) AMT 12 (12 May–15 June 2003) and AMT 14 (26 April–2 June 2004) superimposed on a spring 2014 (March 21st–June 20th) composite of MODIS-Aqua derived chlorophyll. (b) AMT 15 (19 September–29 October 2004) and AMT 17 (15 October–28 November 2005) superimposed on an autumn 2014 (September 21st–December 20th) composite of MODIS-Aqua derived chlorophyll. Yellow filled squares indicate the relative positions of the three high-resolution profiles presented in Figs. 4 and 5 (see Section 3.1). Horizontal dashed lines indicate the major hydrographic provinces used in the meridional analysis (see Sections 3.3 and 3.4) of coccolithophore taxa distribution (TMP, temperate waters; NG, Northern Gyre waters; EQ, equatorial waters; SG, Southern Gyre waters).

cruises are shown in Fig. 2, together with data on the vertical distributions of chlorophyll-*a* and nitrate for AMT 14, one of the AMT cruises with the highest depth resolution of chlorophyll-*a* and nitrate sampling. For a comparison between AMT cruises as to variability in the depth distribution of these properties see Robinson et al. (2006) (their Figs. 2 and 4). For the AMT 15 (September–October 2004) and AMT 17 (September–October 2005) cruises (see Fig. 1b), only surface samples (97% or 55% surface irradiance) were analysed: AMT 15, 6 samples from between 0.1°N (EQ) and 20.6°S (SG); AMT 17, 21 samples from between 44.3°N and 35°S.

Examination of filters by Scanning Electron Microscopy (SEM) generally followed the methodology of Charalampopoulou et al. (2011). Filter segments were examined with a LEO 1450 SEM at 5000× magnification and coccospheres were counted from up to 700 fields-of-view (FOV), equivalent to a maximum sample volume of ~15 mL. The total number of coccospheres counted per sample were ~300 for cruises AMT 12 and AMT 14, and ~150 per sample for cruises AMT 15 and AMT 17; when coccospheres were rare (mainly samples from below the DCM), <150 coccospheres were often encountered.

Species identification followed the nomenclature of Young et al. (2003), as well as Bollmann et al. (2006) for *Solisphaera* spp. and Kleijne and Cros (2009) for *Syracosphaera* spp. A full list of the coccolithophore taxa identified in this study is given in Appendix A. Counts of *Ophiaster formosus* and *O. hydroideus* were combined in order to remove uncertainty in distinguishing the two species when comparing coccosphere counts by different observers. No attempt was made to separate the different morphotypes of species such as *Emiliana huxleyi*, *Umbellosphaera tenuis* and *Calcidiscus leptoporus* (see Boeckel and Baumann, 2008), or *Florisphaera profunda* (see Quinn et al., 2005), although such information could be obtained from the digitally-stored SEM images. Counts of *Rhab-*

dosphaera xiphos are likely to have been underestimated due to the occurrence of aggregates of coccospheres on the SEM filters; also this species could not always be clearly separated from *Palusphaera vandellii*. The coccoliths of some species are readily detached on filters prepared for SEM analysis and in such cases aggregates of identical coccoliths were counted as a coccosphere. The full database of counts used in this study is available from the authors and via the British Oceanographic Data Centre (www.bodc.ac.uk). A total of 171 coccolithophore taxa (Appendix A) were identified in the 199 samples examined from the four AMT cruises. These taxa include recognised alternate life history phases (HET and HOL), three unnamed species (Young et al., 2003) and a few life history combinations for which two species names are still used (see Cros and Fortuno, 2002).

The low cell numbers and high diversity characteristics of subtropical coccolithophore communities (i.e., <30 cells mL⁻¹) lead to concern over the accuracy of SEM analyses on an examination volume (15 mL) which is a relatively small fraction of the filtered volume (1–2 L) to fully quantify the cell abundance and species diversity. One method to address such issues is to investigate how similar repeat counts of a single sample or of replicated samples are of one another in terms of cell counts and species composition. In this study, four replicate counts were performed on representative samples from below (120 m, 1% surface irradiance; optical depth = 4.6) and above (31 m, 55% surface irradiance; optical depth = 0.6) the DCM (Table 1; Fig. 3).

In general, there was good reproducibility of total numbers of taxa (species richness) and of relatively abundant cells (i.e., >1 mL⁻¹). However, for both sampling depths, many taxa were recorded in only one or two of the replicates, although these made a relatively small contribution to total cell counts; for the deep sample 60% of such taxa comprised only 31% of the total cell count,

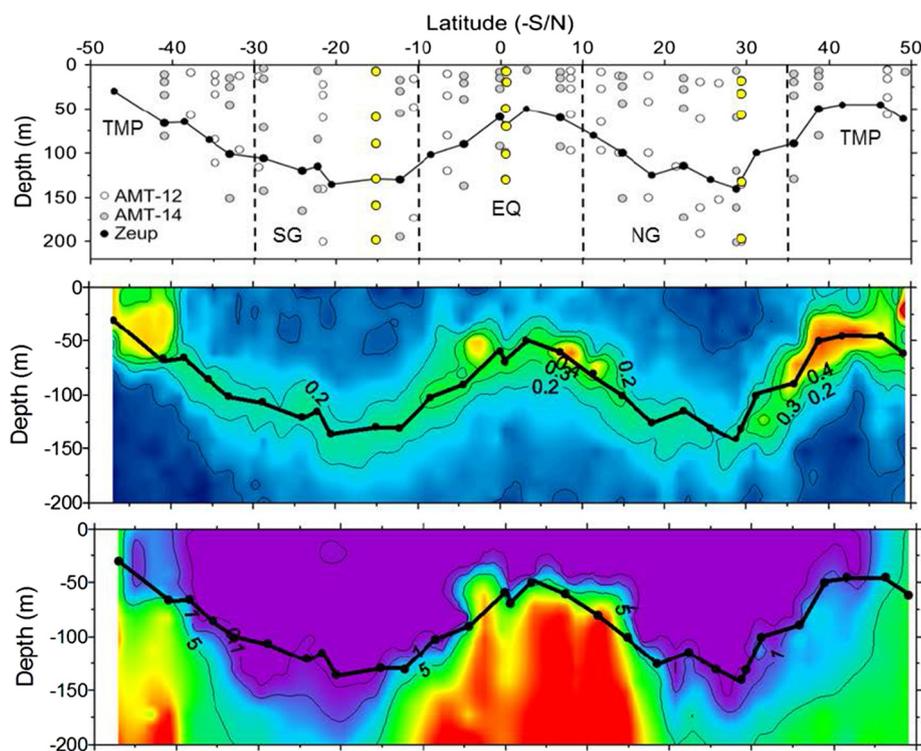


Fig. 2. Latitudinal sections showing (a) the depths at which coccolithophore samples were collected on the AMT 12 and AMT 14 cruises in relation to the depth of 1% surface irradiance (euphotic zone, $Z_{1\%}$) for AMT 14, (b) the distribution of fluorometrically-determined chlorophyll *a* (mg m^{-3}) in relation to the depth of 1% surface irradiance (solid black line) for AMT 14, and (c) the distribution of nitrate ($\mu\text{mol kg}^{-1}$) in relation to the depth of 1% surface irradiance (solid black line) for AMT 14. Vertical dashed lines in (a) indicate the major hydrographic provinces used in the meridional analysis (see Sections 3.3 and 3.4) of coccolithophore taxa distribution (TMP, temperate waters; NG, Northern Gyre waters; EQ, equatorial waters; SG, Southern Gyre waters).

Table 1
Replicate counts of coccospheres for samples collected on AMT 12 (Deep Chlorophyll Maximum, chlorophyll $a = 0.19 \text{ mg m}^{-3}$) and AMT 14 (surface layer, chlorophyll $a = 0.05 \text{ mg m}^{-3}$). AMT 12 sample: 15 taxa were recorded from all 4 Scanning Electron Microscopy (SEM) stubs, with the 3 most abundant ($>1 \text{ mL}^{-1}$) taxa (*F. profunda*, (average \pm standard deviation) $4.4 \pm 0.1 \text{ cells mL}^{-1}$; *Ophiaster* spp., $1.3 \pm 0.1 \text{ cells mL}^{-1}$; *Palusphaera vandellii*, $1.2 \pm 0.2 \text{ cells mL}^{-1}$) making up 57% of the total cell numbers; mean Relative Standard Deviation (RSD) for the other 12 taxa present was 11%; a further 4 taxa were recorded in 3 out of the 4 stubs. AMT 14 sample: 16 taxa were recorded from all 4 stubs, with the 4 most abundant ($>1 \text{ mL}^{-1}$) taxa (*E. huxleyi*, $1.1 \pm 0.3 \text{ cells mL}^{-1}$; *G. ericsonii*, $2.4 \pm 0.2 \text{ cells mL}^{-1}$; *Syracosphaera delicata*, $1.4 \pm 0.4 \text{ cells mL}^{-1}$; *U. tenuis*, $4.9 \pm 0.7 \text{ cells mL}^{-1}$) making up 48% of the total number of coccospheres; mean RSD for the other 11 taxa was 7%; a further 6 taxa were recorded in 3 out of the 4 stubs.

	AMT 12, CTD-26, 120 m			AMT 14, CTD-71, 31 m		
	Count volume (mL)	No. of taxa	Cells (mL^{-1})	Count volume (mL)	No. of taxa	Cells (mL^{-1})
Stub 1	15.1	27	12.9	6.6	40	22.6
Stub 2	15.1	29	12.3	7.3	32	19.2
Stub 3	15.0	28	10.7	7.2	36	20.2
Stub 4	15.0	32	12.6	7.2	37	19.7
Cumulative	60.2	49		28.3	66	
Mean (\pm S.D.)			12.1 (\pm 1.0)			20.4 (\pm 1.5)

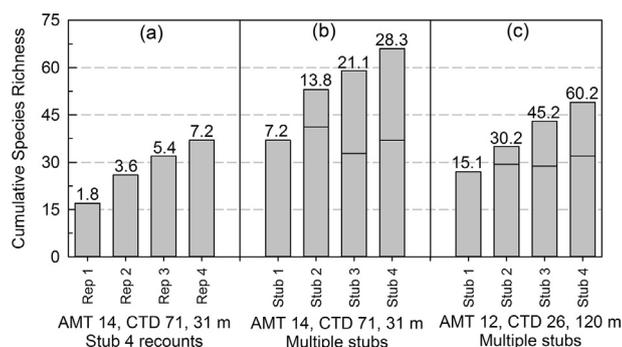


Fig. 3. Histograms of cumulative species richness in selected samples from AMT 14 (CTD 71, 31 m; (a, b) and AMT 12 (CTD 26, 120 m; c). Replicate counts from the same Scanning Electron Microscopy stub (a) and repeat counts from the same sample but different stubs (b, c) are shown. The cumulative volume (mL) counted for each sample is given as a value above each histogram. Horizontal bars on (b) and (c) indicate the number of species identified in each sample.

and for the surface the corresponding figures were 67% and 18%. The numbers of taxa recorded as a function of volume examined are shown in Fig. 3. For both types of sub-sampling it appears that a complete description of the coccolithophore flora can only be achieved by examining relatively large ($>100 \text{ mL}$) volumes of seawater. Thus SEM counts based on small seawater volumes (e.g., $<15 \text{ mL}$ as used for this study) give a measure of coccolithophore species richness whilst the abundance of rarer coccolithophore taxa can be described in relative terms only. To overcome such issues it is necessary to transform and pre-treat the resulting species counts before statistical analysis; including elimination of rare species which occur at low frequencies, consideration of standardised count data (i.e., percentage abundances), and a logarithmic transformation ($\log(X + 1)$), which will all remove the influence of low abundance species and focus the analysis on compositional changes rather than numerical changes (Clark and Warwick, 2001).

2.3. Statistical analysis

Multivariate statistical techniques were used to assess the vertical and horizontal (dis)similarity in species composition of samples, as well as to examine whether any inter-cruise differences in species composition existed, using PRIMER-6 (v 6.1.6, PRIMER-E Ltd.) (Clark and Warwick, 2001). Bray-Curtis Similarity was calculated from standardised count data, with the exclusion of certain taxa considered to be in such low occurrences as to be rare (herein defined as consistently contributing $<5\%$ to total cell numbers), and with a $\log(X + 1)$ transformation. Cluster analysis and non-metric Multi-Dimensional Scaling (nMDS) ordinations were then performed, with related ANOSIM (Analysis of Similarity) and SIMPER

(Similarity Percentages) performed on the resulting patterns, where appropriate (Clark and Warwick, 2001). ANOSIM allows statistical comparison of the variation in species abundance and composition between groups of samples. SIMPER allows statistical identification of which species are primarily responsible for differences between groups of samples, with ANOSIM assessing the significance of the difference between groups. PRIMER was also used to calculate Pielou's Evenness (J') which is a measure of the spread of cell abundances between the species present (Clark and Warwick, 2001). In this study, species richness was simply a measure of the number of taxa in each sample. Subsequent statistical analysis of the rare species was also performed on standardised data, with a $\log(X + 1)$ transformation.

The approach to the statistical examination of vertical, latitudinal and temporal patterns in species composition followed in this study follows a logical order of starting the analysis on a small set of samples (or species) and expanding our interpretation and analysis on this dataset by including other parts of the AMT database in steps. The main analysis focuses initially on the common ($>95\%$ total cell numbers) taxa, although an analysis of the rare species in the same manner to identify any potential patterns is also performed. The analysis follows the order: (a) firstly an examination of three high-resolution profiles from AMT 14 to identify key floral groups and vertical patterns in species composition (Section 3.1); (b) this vertical analysis is then widened to include the full cruise datasets from AMT 12 and AMT 14 (Section 3.2); (c) then consideration of latitudinal patterns in species composition based on just AMT 14 surface samples to identify key floral groups and patterns in species composition (Section 3.3); (d) this latitudinal analysis is then widened to include the full database (AMTs 12, 14, 15 and 17) to identify any inter-cruise differences or similarities in surface waters (Section 3.4); and (e) finally, an analysis of the rare species distribution, in terms of both vertical and latitudinal patterns (Section 3.5), using similar methods as used in the previous sections. The focus on AMT 14 is driven by this cruise representing the best sampled (vertically and latitudinally) AMT cruise, whilst only surface samples are included in the AMT database for AMT 15 and AMT 17.

3. Results

3.1. Vertical community structure: AMT 14 analysis

An in-depth analysis of vertical variability in coccolithophore species composition is firstly carried out using three high-resolution profiles from the AMT 14 cruise. These three stations represent the southern gyre (SG), equatorial waters (EQ) and northern gyre (NG) regions of the Atlantic (Figs. 1, 4 and 5). At each of these stations the numbers of taxa and cells were highest in the surface layer above the depth of the DCM (1% surface irradiance;

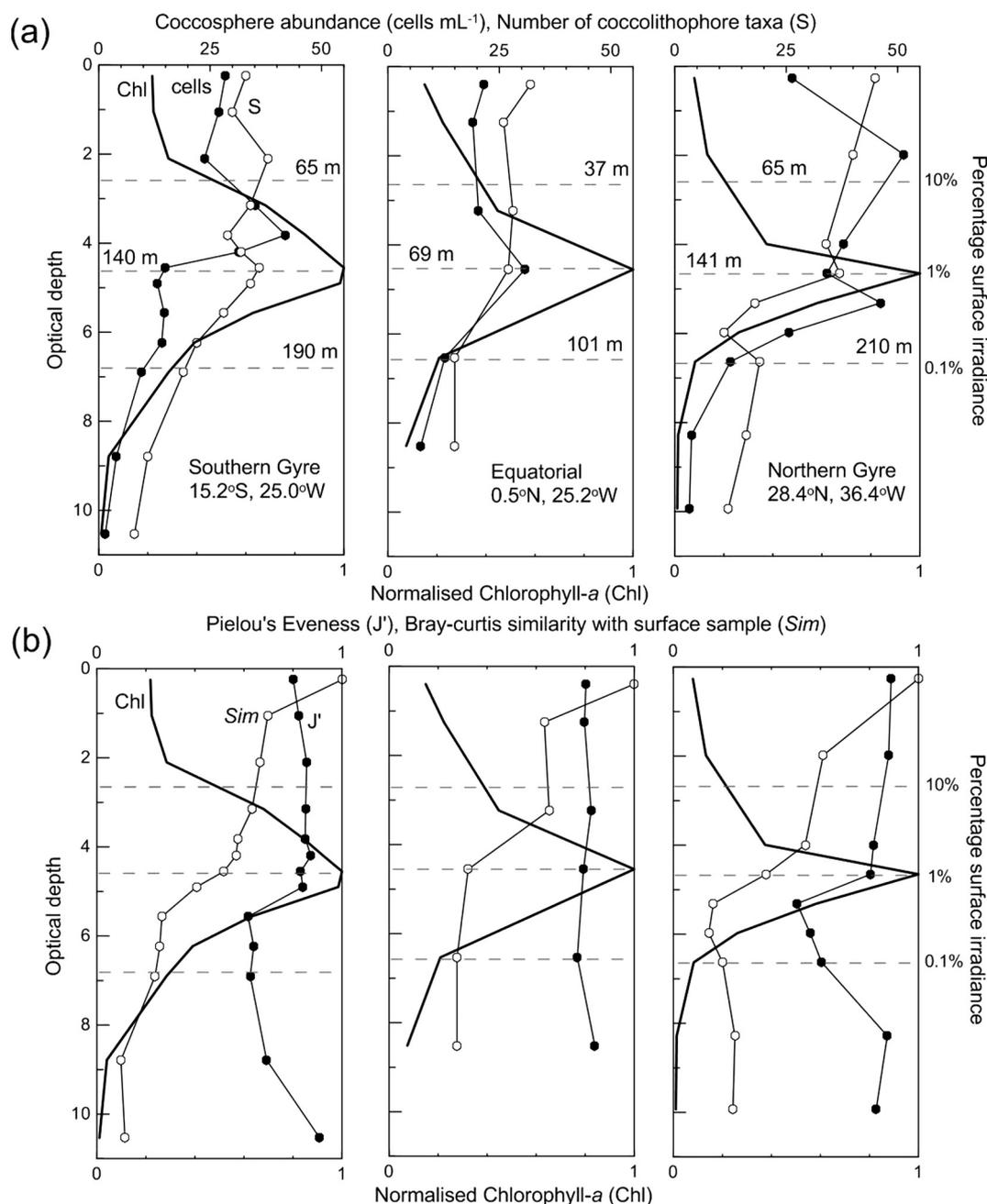


Fig. 4. Vertical profiles of coccolithophore community characteristics against optical and percentage surface irradiance depths for three AMT 14 stations taken to be representative of the Southern Gyre, equatorial waters, and the Northern Gyre. (a) Normalised chlorophyll *a* (Chl), coccosphere abundance (cells mL⁻¹), and the number of coccolithophore taxa (species richness) (*S*). (b) Normalised chlorophyll *a* (Chl), Pielou's Evenness (*J'*), and Bray-Curtis Similarity (*Sim*) referenced to the near-surface sample. The depth (m) of indicative levels of percentage surface irradiance are given against the horizontal dashed lines indicating the 10%, 1% and 0.1% surface irradiance depths.

optical depth = 4.6), and then declined to minimum values below the depth of 0.1% surface irradiance (optical depth = 6.9) (Fig. 4a). The relative abundance of cells and taxa within the DCM appears to vary between the two gyres, which may potentially be linked to seasonality, both being relatively low in the autumn (SG) when surface light is declining compared to the spring (NG) when surface light is increasing. In the EQ region, where the DCM and nutricline are shallow compared to the subtropical gyres (e.g., Fig. 2 for AMT 14), the decline in numbers of cells and taxa with depth are less marked. All three profiles show a relatively greater drop in cell number than in species richness below the DCM (Fig. 4a).

Each of the three profiles from AMT 14 (Fig. 4b) were characterised by a change in community structure of the coccolithophore

population with depth as indicated by the fall in Bray-Curtis Similarity to <50% (with reference to the surface sample at each site) at or slightly above the depth of the DCM. In contrast, *J'* was relatively constant down to the DCM (Fig. 4b); since numbers of taxa for each profile were fairly uniform above the DCM it appears that changes in species composition with depth in the surface layer were not accompanied by changes in the relative abundance of the dominant species. In the two subtropical gyres the sharp drop in *J'* below the DCM suggests that the coccolithophore population below the depth of 1% surface irradiance (optical depth = 4.6) was distinct from that in surface waters in terms of both species composition and dominance by fewer species (Fig. 4a). However, *J'* increased again below the depth of 0.1% surface irradiance (at

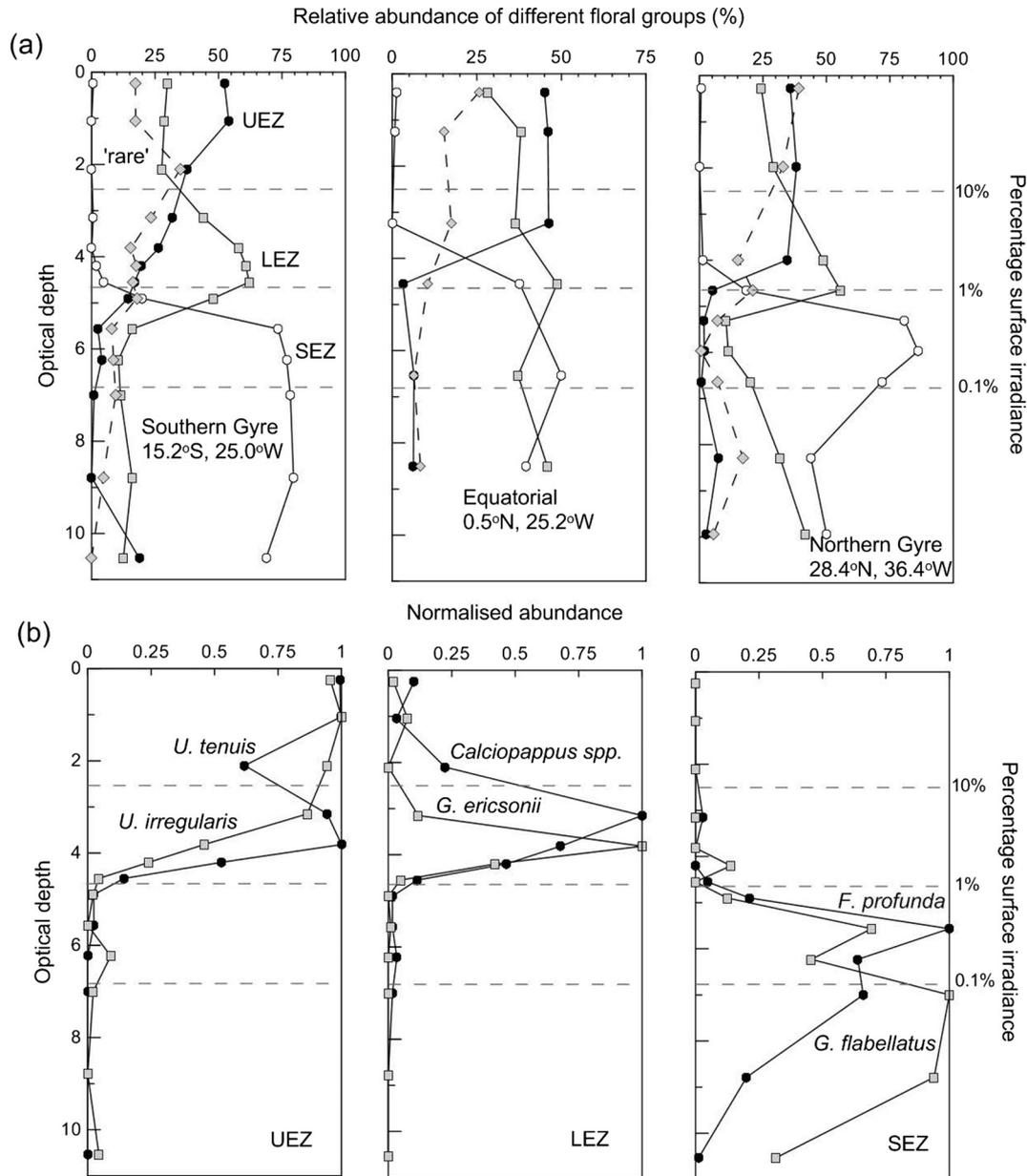


Fig. 5. Vertical profiles of abundance of the different coccolithophore floral groups (see Table 2; Section 3.1) and selected taxa from each group against optical and percentage surface irradiance depth for three AMT 14 stations taken to be representative of the Southern Gyre, equatorial waters, and the Northern Gyre. (a) Relative abundance of Upper Euphotic Zone (UEZ), Lower Euphotic Zone (LEZ), Sub-Euphotic Zone (SEZ) and rare (consistently <5% of total cell numbers) coccolithophore groups at the three stations. (b) Normalised abundance of taxa representative of the UEZ (*Umbellosphaera irregularis* and *U. tenuis*), LEZ (*Gephyrocapsa ericsonii* and *Calciopappus* spp.), and SEZ (*Florisphaera profunda* and *Gladiolithus flabellatus*) groups at the Southern Gyre station.

optical depths >8) and absolute depths deeper than 200 m, although these depths were characterised by very low cell numbers.

The depth distribution of the abundant coccolithophore taxa (i.e., >95% of total numbers) for the three stations in Fig. 4 (AMT 14) was further examined by plotting depth profiles of each species' cell abundance normalised to its maximum cell abundance observed in that profile (plots not shown). In our study, normalised abundance is the abundance in each sample relative to the maximum observed in the dataset; so a value of 0 would represent zero abundance and 1 would represent that species' maximum abundance. A species' depth habitat was then classified as the uppermost depth horizon where the maximum value (i.e., 1) and consistent values >0.75 were observed between the main relative surface irradiance depths presented in Fig. 4. Depth habitats were

compared across the three stations and where, in a few cases, species had high normalised abundances in more than one depth zone, the overall depth habitat was defined as the one common across all stations. This analysis of the depth distribution of the more abundant coccolithophore taxa recorded at the three stations identified three floral groups in our study (Table 2) which are taken to characterise the Upper Euphotic Zone (UEZ, >10% surface irradiance), Lower Euphotic Zone (LEZ, 1–10% surface irradiance), and Sub-Euphotic Zone (SEZ, <1% surface irradiance). As the DCM is generally considered to mark the base of the euphotic zone (i.e., 1% of surface irradiance), the deepest coccolithophore zone is referred to as the 'sub-euphotic' rather than 'lower photic' as in other studies (e.g., Winter et al., 1994).

The vertical distributions of each of these floral groups at the three AMT 14 stations analysed, and of the rare species

Table 2

Coccolithophore floral groups based on analysis of each species' cell abundance normalised to its maximum cell abundance observed in three stations taken as representative of Southern Gyre waters, equatorial waters, and Northern Gyre waters (see Fig. 1) during AMT 14. A species' depth habitat was classified as the uppermost depth horizon where the maximum value (i.e., 1) and consistent values >0.75 were observed between the 100%, 10%, 1% and 0.1% percentage surface irradiance depths (see Fig. 4). Each of the listed taxa consistently made up >5% of total cell numbers, and was assigned to the photic zone of overall maximum abundance. The number (n) of samples in each depth zone (across the three stations) in which species were analysed is given in parenthesis.

Upper Euphotic Zone (UEZ), >10% surface irradiance (n = 7)	Lower Euphotic Zone (LEZ), 10–1% surface irradiance (n = 8)	Sub-euphotic Zone (SEZ), <1% surface irradiance (n = 13)
<i>Corisphaera gracilis</i>	<i>Calciopappus</i> sp.	<i>Algirosphaera robusta</i>
<i>Palusphaera vandellii</i>	<i>Calciosolenia brasiliensis</i>	<i>Calciosolenia murrayi</i>
<i>Rhabdosphaera xiphos</i>	<i>Ceratolithus</i> spp.	<i>Florisphaera profunda</i>
<i>Syracosphaera bannockii</i>	<i>Discosphaera tubifera</i>	<i>Gladolithus flabellatus</i>
<i>S. bannockii</i> HOL	<i>Emiliana huxleyi</i>	<i>Hayaster perplexus</i>
<i>Syracosphaera delicata</i>	<i>Gephyrocapsa ericsonii</i>	<i>Reticulofenestra sessilis</i>
<i>Syracosphaera pulchra</i> HOL	<i>Michaelsarsia elegans</i>	<i>Tetralithoides quadrilaminata</i>
<i>Umbellosphaera irregularis</i>	<i>Oolithus antillarum</i>	
<i>Umbellosphaera tenuis</i>	<i>Ophiaster</i> spp.	
	<i>Rhabdosphaera clavigera</i>	
	<i>Solisphaera</i> spp.	
	<i>Syracosphaera anthos</i>	
	<i>Syracosphaera nana</i>	
	<i>Syracosphaera reniformis</i>	
	<i>Syracosphaera</i> sp.	

(consistently <5% of total cell number), are shown in Fig. 5a. In general the UEZ and rare taxa were most numerous above the depth of 10% surface irradiance but extended down to the depth of 1% surface irradiance, whereas SEZ taxa were largely confined to below the depth of 1% surface irradiance. In contrast, the LEZ taxa reached a maximum relative abundance close to the depth of 1% surface irradiance but also made significant contributions to coccolithophore populations of the UEZ at all three stations and as well as the SEZ at the equatorial station. Total cell numbers below the depth of 1% surface irradiance (see Fig. 4a) were low in all three profiles from AMT 14 so that the apparent importance of LEZ species below this depth at the EQ and NG stations reflects relatively low numbers of coccospheres. As an example of the depth distribution of characteristic taxa in each of the floral groups identified, Fig. 5b shows relative abundance profiles of two species from each of the UEZ, LEZ and SEZ groups (see Table 2) at the SG station. Fig. 5b also shows how two zones exist in the upper section of the water column, with the depth distribution of certain species peaking at intermediate depths below the depth of 10% surface irradiance but above the depth of the DCM and euphotic zone (e.g., *Calciopappus* spp. and *G. ericsonii*; Fig. 5b). The taxa within each floral group do not necessarily follow the same pattern; thus, for the UEZ, *Umbellosphaera tenuis* extended to a slightly greater depth than *U. irregularis* and, for the SEZ, the maximum abundance of *Gladolithus flabellatus* was well below that of *Florisphaera profunda*.

Using a comparative pair-wise statistical analysis of the similarity between species composition (ANOSIM) for the three selected AMT 14 stations it is clear that there are strong statistical differences in species composition between UEZ and SEZ (R-statistic 0.97, $p < 0.001$, average Bray-Curtis dissimilarity = 79.1%) and between LEZ and SEZ (R-statistic 0.74, $p < 0.001$, average Bray-Curtis dissimilarity = 70.7%), whereas the differences between UEZ and LEZ are relatively weaker (R-statistic 0.19, $p < 0.05$, dissimilarity = 55.2%; Table 3). The same pair-wise analysis for all

Table 3

Multivariate statistical analyses of differences in coccolithophore species composition for the three photic zones identified (UEZ, Upper Euphotic Zone; LEZ, Lower Euphotic Zone; SEZ, Sub-euphotic Zone). ANOSIM is a one way analysis of similarities; SIMPER is a one way analysis of (dis)similarity percentages species contributions to the differences between euphotic zone groups.

Groups	ANOSIM R-Statistic	p level	SIMPER Average dissimilarity (%)
<i>Comparison for selected stations in Fig. 4 (n = 28 samples)</i>			
UEZ (n = 7) v LEZ (n = 8)	0.19	$p < 0.05$	55.2
UEZ (n = 7) v SEZ (n = 13)	0.97	$p < 0.001$	79.1
LEZ (n = 8) v SEZ (n = 13)	0.74	$p < 0.001$	70.7
<i>Comparison for all AMT-14 stations (n = 85 samples)</i>			
UEZ (n = 41) v LEZ (n = 19)	0.49	$p < 0.001$	64.3
UEZ (n = 41) v SEZ (n = 25)	0.94	$p < 0.001$	79.2
LEZ (n = 19) v SEZ (n = 25)	0.54	$p < 0.001$	67.8

the samples from AMT 14, where samples were assigned to floral group zones by their relative irradiance depths, gives further support for the three floral groups (Table 3), with high levels of dissimilarity (>60%) in terms of species composition and significant ($p < 0.001$) R-statistics from the ANOSIM analyses. For both sets of analyses (selected AMT 14 stations and all AMT stations), the strongest statistical differences in terms of species composition were always seen between the UEZ and SEZ (R-Statistics > 0.9, $p < 0.001$) with average Bray-Curtis dissimilarity levels of almost 80%.

3.2. Vertical community structure: comparison between AMT 12 and 14

Vertical segregation of coccolithophore taxa during AMT 14 is clearly supported by analysis of normalised abundance (Figs. 4 and 5, Table 2) and multivariate comparison of species composition between the different sample groupings (Table 3). To test how widely these patterns exist in the fuller AMT database a multivariate statistical analysis of species composition in all AMT 12 and AMT 14 samples was performed. Coccolithophore species structure for all AMT 12 and AMT 14 samples (total 150) was investigated through calculation of Bray-Curtis Similarity and then ordination by non-metric Multi-Dimensional Scaling (nMDS). nMDS ordinations showing sample distribution with respect to cruise number, hydrographic region and optical depth are shown in Fig. 6a–c, respectively.

The nMDS ordination (Fig. 6) shows several interesting patterns in species composition, with the relative distance between samples indicative of their relative (dis)similarity. Samples with statistically similar species composition are closely aligned, whereas samples with low statistical similarity in terms of species composition are more widely spaced. Hence, the overlap in the distribution of samples from the two cruises (AMT 12 and 14) suggests strong similarities in species composition, especially for the gyre and equatorial regions (Fig. 6a and b). In terms of hydrographic region, the temperate (TMP) samples, mainly from shallow water depths, appear as a variable but relatively distinct group (Fig. 6b). In terms of irradiance levels, expressed as optical depth, the deep samples (from the depths of the 0.1% and, to a lesser extent, the 1% surface irradiance) appear as the most distinct groups (Fig. 6c), in contrast to the temperate samples (Fig. 6b) which show considerable overlap between optical depths (Fig. 6c).

Coccolithophores were found in all samples from AMT 12 and 14, with the highest cell concentrations in temperate waters (Fig. 6b–d) during both cruises. At gyre and equatorial stations (Fig. 6b and c), cell numbers below the DCM (1% surface irradiance; optical depth = 4.6) generally decreased with depth; for optical depths of 0.6–2, 4.6 and 6.9 (corresponding to the 55–14%, 1%

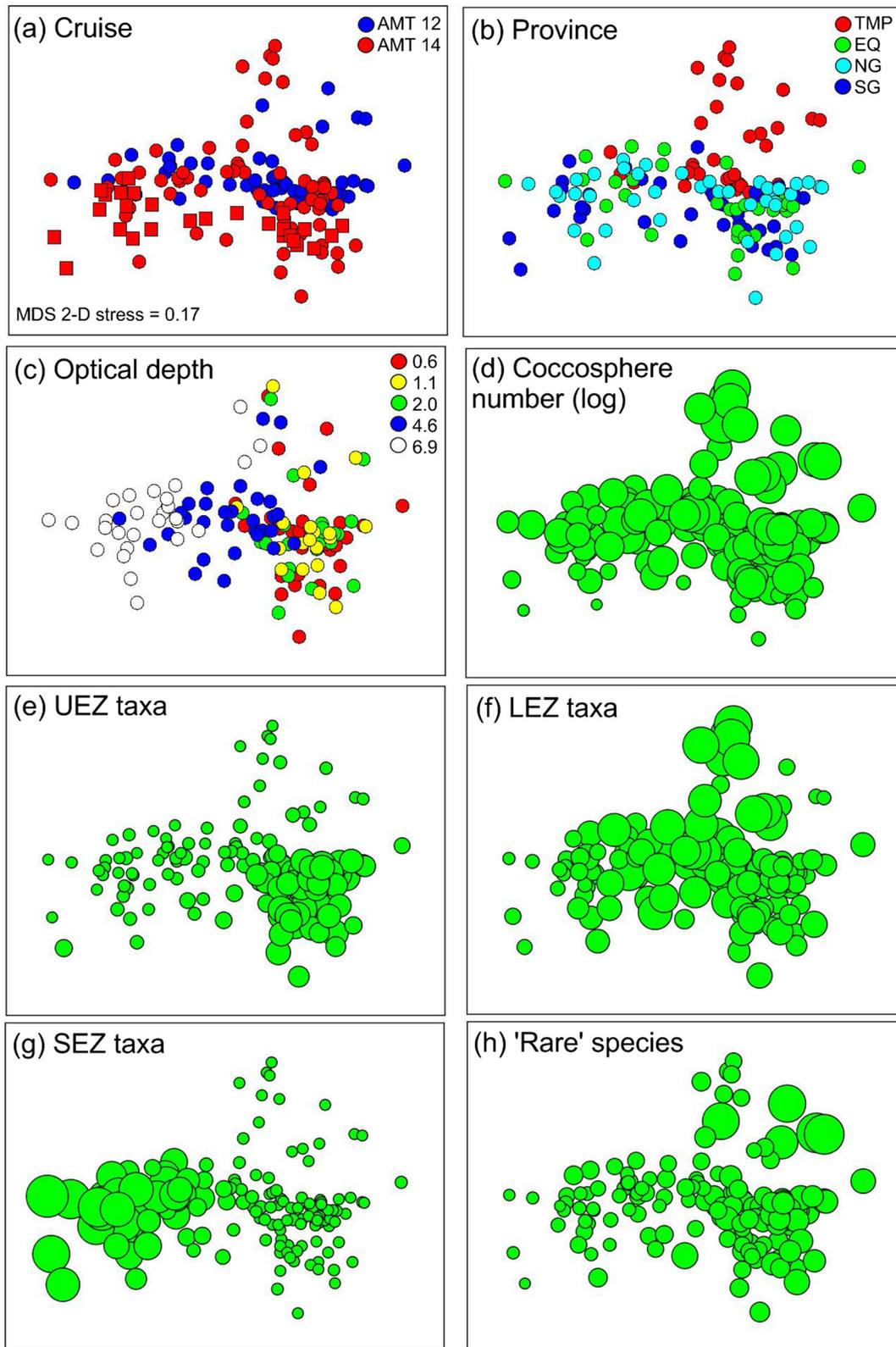


Fig. 6. Normalised Multi-Dimensional Scaling (nMDS) ordination and bubble plots showing latitudinal and depth variability in coccolithophore community structure for AMT 12 and AMT 14: (a) by cruise; (b) by latitudinal (TMP, Temperate waters; EQ, equatorial waters; NG, Northern Gyre waters; SG, Southern Gyre waters); (c) by optical depth; (d) by coccosphere abundance (log); (e) for UEZ taxa (relative abundance to other floral groups); (f) for LEZ taxa (relative abundance to other floral groups); (g) for SEZ taxa (relative abundance to other floral groups); and (h) for rare taxa (relative abundance to other floral groups). The samples from the three high-resolution depth profiles presented in Figs. 4 and 5 are identified in panel (a) as filled square symbols. Stress on the two-dimensional plot (see (a)) was 0.17 which indicates a 'good representation of the data in two-dimensional space' (Clark and Warwick, 2001).

and 0.1% surface irradiance levels, respectively) the mean cell counts were 24.4, 29.3 and 12.9 mL⁻¹, respectively.

The relative cell counts for the different floral groups identified in Table 2 are shown in Fig. 6e–h (i.e., the relative abundance of taxa from each floral group in each sample to total numbers) for the two AMT cruises. The upper optical depths (0.6–2.0; 55–14% surface irradiance; Fig. 6c) of the gyre (NG, SG) and equatorial (EQ) regions (Fig. 6b) are characterised by abundant UEZ taxa (Fig. 6e), and the lowest optical depth (6.9; 1% surface irradiance; Fig. 6c) of the same regions by SEZ taxa (Fig. 6g). In contrast, LEZ taxa are relatively widely distributed (Fig. 6f), extending into the temperate region (Fig. 6b) and from the surface down to as deep as the 1% surface irradiance level (optical depth = 6.9; Fig. 6c). Rare taxa (Fig. 6h) are relatively evenly distributed in terms of latitude (Fig. 6b) and depth (Fig. 6c), with maximum cell numbers in the UEZ (Fig. 6e) and temperate waters (Fig. 6b); however, they are not a homogenous group, with different species occurring at different depths and latitudes (data not shown) as has been found for the common species (see Section 3.1).

3.3. Horizontal (meridional) community structure: AMT 14 analysis

Based on routine sampling from 5 depths per station it is not possible to analyse objectively latitudinal variations in the LEZ and SEZ coccolithophore flora as samples collected at the depths of 1% and 0.1% surface irradiance might, or might not, correspond to the maximum cell abundances of characteristic species for these depths (see Fig. 5b). On the other hand the surface samples (55% of surface irradiance) may be considered as representative of the UEZ flora (see Fig. 5) and hence our horizontal (meridional) analysis of species composition focuses on only surface samples. Again, AMT 14 is chosen as the first cruise to base the analysis on, with the next section (Section 3.4) widening the analysis to include all four AMT cruises.

Results of a compositional statistical analysis of AMT 14 surface samples (55% relative surface irradiance only) are shown in Fig. 7. Generally, coccolithophore cell numbers in surface waters between 30°S and 35°N ranged from 20 to 40 cells mL⁻¹, and the number of taxa (species richness) in each sample ranged from 25 to 45 (Fig. 7a). The highest coccosphere numbers were recorded from temperate stations (>30°S) and the lowest from equatorial stations. The number of taxa were relatively high (>30) within surface waters of the subtropical gyres (NG, SG), and decreased at the northern temperate station and at some equatorial and Northern Gyre stations; for example, the surface (13 m) sample from the AMT 14 14.8°N station gave a coccosphere count of just 2.2 mL⁻¹ for just 10 taxa (Fig. 7a).

Species composition in surface samples across the full AMT 14 cruise was examined by comparing statistically one station in the NG (22.3°N) with all the other surface samples in terms of Bray-Curtis Similarity. This comparison showed that the coccolithophore communities were broadly comparable (i.e., ~50% similarity) in terms of species composition across a broad latitudinal range from 35°N to 32°S (Fig. 7b). A plot of Pielou's Evenness (Fig. 7b) for surface samples also suggests that the surface coccolithophore communities were comparable in terms of species diversity and dominance from 30°N to 30°S, except at higher latitudes where high cell densities (typically 10–100 cells mL⁻¹) of LEZ taxa such as *Emiliania huxleyi*, *Gephyrocapsa* spp., and *Calcidiscus leptoporus* were found. However, as shown in Fig. 7c, the relative abundance of individual taxa (e.g., *E. huxleyi*, *Umbellosphaera tenuis* and *U. irregularis*) between stations over 20 km apart was highly variable and latitudinal peaks in relative abundance were not always consecutive.

Using a similar approach for assessing the horizontal (latitudinal) distribution of coccolithophore species as used in Section 3.1

for vertical variability, the normalised species distribution for all species which represented over 5% of total numbers in surface samples was plotted again in order to determine whether each species' maximum relative abundance was consistently found (>0.75) within a specific hydrographic province (plots not shown). The results from this analysis are presented in Table 4, alongside the vertical floral group that each taxa (where appropriate) is aligned with. The nine UEZ taxa (see Table 2) are found mainly in the subtropical gyres (NG, SG), whilst LEZ taxa are found in both equatorial (EQ) and Temperate waters (Table 4). Of the SEZ taxa listed in Table 2, three species are found in surface temperate waters (*Algirosphaera robusta*, *Calciosolenia murrayi*, *Florisphaera profunda*), whereas the other four SEZ species were never observed in significant numbers in surface waters during AMT 14 (i.e., they were consistently <5% of total numbers and eliminated from the analysis). The relative abundances of these three biogeographic floras are plotted in Fig. 7d, along with the distribution of the rare (<5% of total numbers) surface water taxa. As expected, the gyre, equatorial and temperate floras reach maximum relative abundances in each of these regions, whilst the rare flora are found throughout the transect, apart from in temperate waters at either end of the transect (Fig. 7d).

A comparison of the Bray-Curtis Similarity between successive vertical samples at each station representing the UEZ, LEZ and SEZ (i.e., the statistical similarity between the 55% and 14%, 55% and 1%, 55% and 0.1% relative surface irradiance depths, respectively) again shows clear vertical gradients in species composition (Fig. 7e; see also Fig. 4b). Temperate stations are characterised by weak vertical gradients in Bray-Curtis Similarity, whilst relatively stronger vertical declines in similarity can be seen in both subtropical gyre and equatorial waters. Also, there is a marked contrast between the SG and NG samples, with high similarity (>70%) between the 55% and 14% species composition in the north which declines to ~50% in the south. A comparison of Fig. 7b and e shows that vertical gradients in species composition (in terms of Bray-Curtis Similarity) are much stronger than horizontal ones; similarity remains around 50% when comparing a sample from the NG with all samples from 30°N to 32°S (Fig. 7b), whereas similarity drops sharply below 50% when examining trends vertically over the upper 200 m (Fig. 7e). These trends highlight how there is sharper dissimilarity in coccolithophore species composition vertically over a few tens of metres than there is horizontally, over hundreds to thousands of kilometres.

3.4. Comparison between different cruises for surface communities

Analysis of meridional patterns of species composition in surface samples from AMT 14 shows several distinct features and trends in the level of statistical similarity between communities in different hydrographic provinces (Fig. 7). In this section this meridional analysis is widened to include UEZ samples (55% surface irradiance) from all four AMT cruises (AMTs 12, 14, 15 and 17), again using comparisons of Bray-Curtis Similarity in species composition in nMDS ordination (Fig. 8). In relation to cruise number, the northern spring/southern autumn (AMTs 12 and 14) and the northern autumn/southern spring (AMTs 15 and 17) samples appear to form distinct groups (Fig. 8a), a difference that may be attributable to seasonal changes in community composition rather than cellular abundances (due to the standardisation of the counts and pre-treatment of the data). When plotted in relation to hydrographic province the temperate samples form the most distinctive group and are the most scattered (Fig. 8b), again suggesting a potential seasonal signal. Variability between cruises is also evident in the equatorial and gyre waters of both hemispheres as samples from each temporal cruise pairing (i.e., AMT 12 and 14 versus AMT 15 and 17; Fig. 8a) are clearly distinct in the nMDS

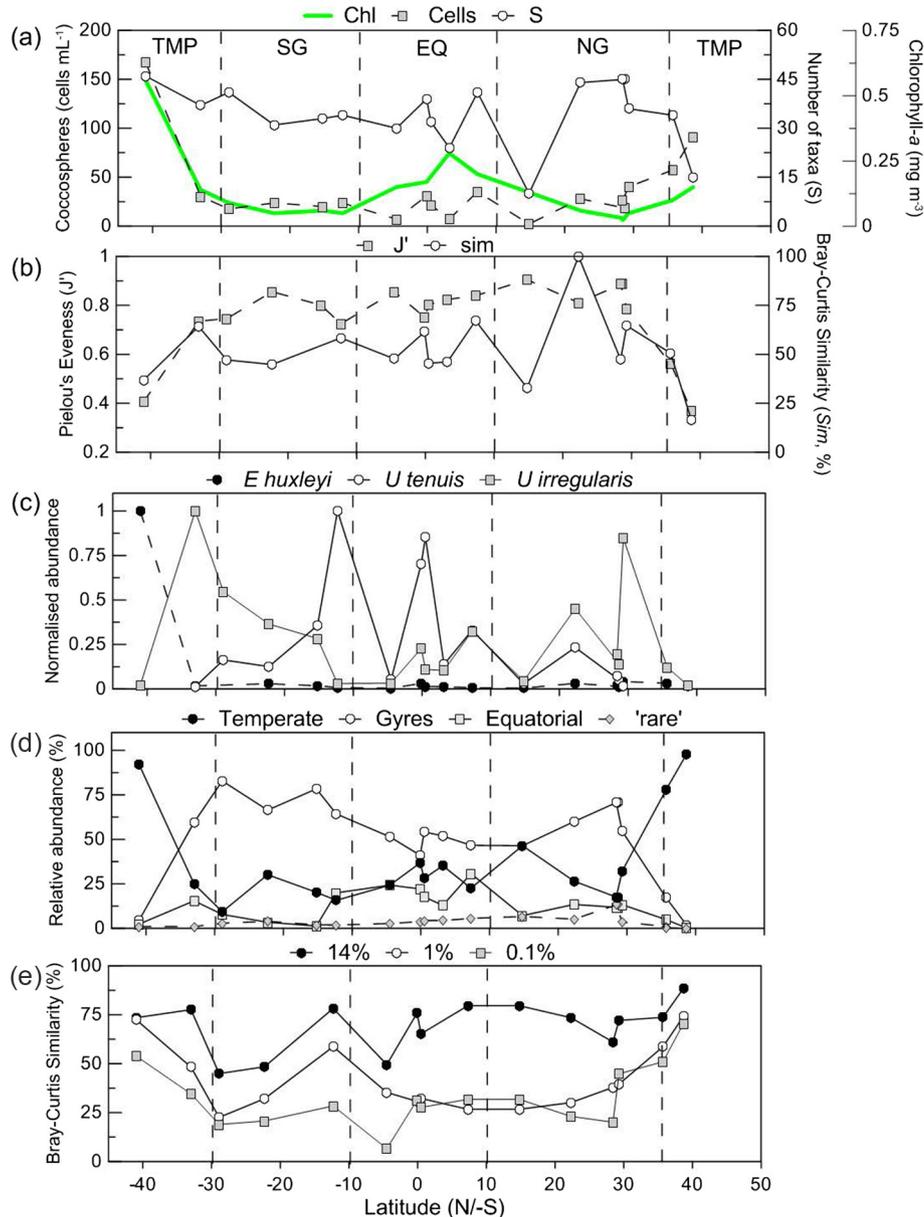


Fig. 7. Latitudinal trends of coccolithophore composition in samples from the near-surface (optical depth = 0.6 (55% surface irradiance)) during AMT 14. (a) Chlorophyll *a* (Chl, mg m^{-3}), cell abundance (cells, mL^{-1}) and the number of coccolithophore taxa (species richness) (*S*). (b) Pielou's Evenness (*J'*) and Bray-Curtis Similarity (*Sim*) referenced to a Northern Gyre station at 22.3°N. (c) Normalised abundance of *Umbellosphaera tenuis*, *U. irregularis* and *E. huxleyi*. (d) Relative abundance of coccolithophore flora (see Table 4) representative of Temperate waters, Gyre waters (northern and southern), and equatorial waters, as well as the rare group (consistently <5% of total cell numbers). (e) The degree of similarity of species composition from the 14%, 1% and 0.1% surface irradiance depths compared to the 55% sample (see a) expressed as percentage Bray-Curtis Similarity (*Sim*). Vertical dashed lines in panel (a) indicate the positions of the major hydrographic provinces (TMP, temperate waters; NG, Northern Gyre waters; EQ, equatorial waters; SG, Southern Gyre waters).

(Fig. 8b). Hence, the species composition of cruises sampled in the same season but different years appears to be more similar than between successive cruises in different years; i.e., implying that seasonal differences in coccolithophore species composition are stronger than inter-annual differences.

To further test the statistical significance of the sample segregation shown in Fig. 8 an identical analysis used on spatial trends in species composition (Fig. 6) is applied to the dataset. Table 5 shows the ANOSIM and SIMPER analyses on the data presented in Fig. 8, whilst species level variability is further examined (Table 6) in the context of changes in sea-surface temperature and chlorophyll-*a* concentration. The ANOSIM analysis supports the trends identified, with statistically significant ($p < 0.05$) differences

in surface coccolithophore communities between AMT 12 and AMT 14 (May) versus AMT 15 and AMT 17 (September–October) for all four biogeographic provinces. Although the highest dissimilarity between seasonal samples was found for Temperate waters (69.1%), the other regions still had dissimilarity >55% (57.5–62.6%) and statistically significant ($p < 0.01$) R-statistics (Table 5). These results support the perspective that seasonal differences in coccolithophore species composition are stronger than inter-annual differences and the multivariate statistics are identifying seasonal variability (within the limited time-series of sampling).

Examining the raw count data further elucidates the compositional changes identified in Fig. 8, although it should be noted that due to the data treatment (standardisation, $\log(X + 1)$

Table 4

Coccolithophore floral groups based on the latitudinal range of their maximum abundance relative to hydrographic provinces (see Fig. 1; TMP, Temperate waters; EQ, equatorial waters; Northern and Southern Gyre waters, NG and SG). The list is based on taxa found in 16 surface (55% surface irradiance) samples from AMT 14 and which represented >5% of total cell numbers in at least one surface sample. The affiliation of taxa in terms of the vertical floral zones (see Table 2) are also indicated (UEZ, Upper Euphotic Zone; LEZ, Lower Euphotic Zone; SEZ, Sub-euphotic Zone).

Temperate (TMP), >35°N or >30°S	Gyres (SG, NG), 30°S–10°S or 10°N–35°N	Equator (EQ), <10°S to <10°N
<i>Algirosphaera robusta</i> (SEZ)	<i>Acanthoica quattrosolina</i>	<i>Calciosolenia brasiliensis</i> (LEZ)
<i>Alisphaera ordinata</i>	<i>Anthosphaera fragaria</i>	<i>Corisphaera tyrreniensis</i>
<i>Alisphaera pinnigera</i>	<i>Anthosphaera periperforata</i>	<i>Gephyrocapsa oceanica</i>
<i>Alisphaera quadrilatera</i>	<i>Calcidiscus leptopus</i>	<i>Gephyrocapsa spinosa</i>
<i>Calcioappus</i> sp. (LEZ)	<i>Calyptrolithina multipora</i>	<i>Homozygospaera papillifera</i>
<i>Calciosolenia murrayi</i> (SEZ)	<i>Calyptrolithophora papillifera</i>	<i>Michaelsarsia elegans</i> (LEZ)
<i>Calyptrolithina divergens</i>	<i>Ceratolithus</i> spp. (LEZ)	<i>Palusphaera vandellii</i> (UEZ)
<i>Emiliania huxleyi</i> (LEZ)	<i>Corisphaera gracilis</i> (UEZ)	<i>Papposphaera lepida</i>
<i>Florisphaera profunda</i> (SEZ)	<i>Coronosphaera mediterranea</i> HOL	<i>Poricalyptra magnaghii</i>
<i>Gephyrocapsa ericsonii</i> (LEZ)	<i>Cyrtoisphaera</i> sp.	<i>Syracosphaera ampliora</i>
<i>Gephyrocapsa muelleriae</i>	<i>Discosphaera tubifera</i> (LEZ)	<i>Syracosphaera marginaporata</i>
<i>Gephyrocapsa ornata</i>	<i>Helicosphaera carteri</i>	<i>Syracosphaera pirus</i>
<i>Helicosphaera pavimentum</i>	<i>Helicosphaera carteri</i>	<i>Syracosphaera rotula</i>
<i>Oolithotus antillarum</i> (LEZ)	HOL	
<i>Oolithotus fragilis</i>	<i>Helladosphaera cornifera</i>	
<i>Ophiaster</i> spp. (LEZ)	<i>Homozygospaera arethusae</i>	
<i>Reticulofenestra parvula</i>	<i>Pappomonas</i> sp. Type 4	
<i>Syracosphaera anthos</i> (LEZ)	<i>Pappomonas</i> sp.	
<i>Syracosphaera corolla</i>	<i>Polycrater galapagensis</i>	
<i>Syracosphaera delicate</i> (UEZ)	<i>Poricalyptra aurisinae</i>	
<i>Syracosphaera halldalii</i>	<i>Rhabdosphaera clavigera</i> (LEZ)	
<i>Syracosphaera histrica</i>	<i>Rhabdosphaera xiphos</i> (UEZ)	
<i>Syracosphaera molischii</i>	<i>Solisphaera</i> spp. (LEZ)	
<i>Syracosphaera protrudens</i>	<i>Sphaerocalyptra</i> sp. 1	
<i>Syracosphaera squamosa</i>	<i>Syracosphaera anthos</i> HOL	
<i>Syracosphaera sp.</i> (LEZ)	<i>Syracosphaera bannockii</i> (UEZ)	
<i>Umbilicosphaera sibogae</i>	<i>S. bannockii</i> HOL (UEZ)	
	<i>Syracosphaera lamina</i>	
	<i>Syracosphaera nana</i> (LEZ)	
	<i>Syracosphaera noroitica</i>	
	<i>Syracosphaera prolongata</i>	
	<i>Syracosphaera pulchra</i>	
	<i>Syracosphaera pulchra</i> HOL (UEZ)	
	<i>Syracosphaera hastata</i>	
	<i>Syracosphaera reniformis</i> (LEZ)	
	<i>Umbellosphaera irregularis</i> (UEZ)	
	<i>Umbellosphaera tenuis</i> (UEZ)	
	<i>Umbilicosphaera hulburtiana</i>	
	Unknown holococcolith	

transformation) actual cell numbers have limited impact on the nMDS. Average coccolithophore abundances were slightly higher during northern spring in the subtropical gyres and equatorial waters than during southern spring (Table 6), although the differences were not statistically significant. As expected, Temperate waters showed the highest absolute cell abundances and highest

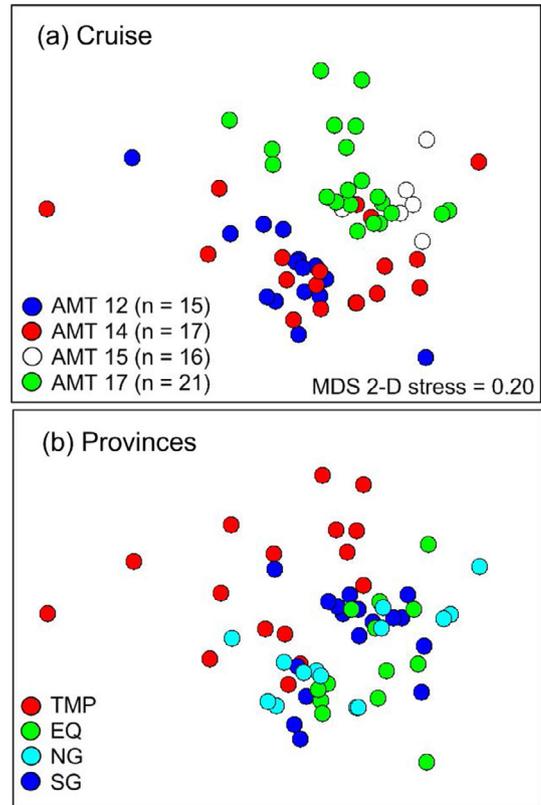


Fig. 8. Normalised Multi-Dimensional Scaling (nMDS) ordination showing inter-cruise and latitudinal differences in the UEZ coccolithophore community composition for AMTs 12, 14, 15 and 17: (a) by cruise; and (b) by hydrographic provinces (TMP, temperate waters; NG, Northern Gyre waters; EQ, equatorial waters; SG, Southern Gyre waters). Stress on the two-dimensional plot (see (a)) was 0.20 which indicates a 'good representation of the data in two-dimensional space' (Clark and Warwick, 2001).

Table 5

Analysis of inter-cruise differences between surface coccolithophore communities during cruises in May (M) (AMT 12, n = 15; AMT 14, n = 17) and September (S) (AMT 15, n = 6; AMT 17, n = 21) for the four hydrographic provinces (see Fig. 1; TMP, Temperate waters; EQ, equatorial waters; NG, Northern Gyre waters; SG, Southern Gyre waters). ANOSIM is a one way analysis of similarities; SIMPER is a one way analysis on (dis)similarity percentages species contributions to the statistical differences between communities. The number of samples (n) in each province in each season is given in parenthesis.

Groups	ANOSIM R-Statistic	p level	SIMPER Average Dissimilarity (%)
M-TMP (n = 9) v S-TMP (n = 8)	0.44	p < 0.001	69.1
M-SG (n = 7) v S-SG (n = 11)	0.54	p < 0.001	57.5
M-EQ (n = 8) v S-EQ (n = 5)	0.41	p < 0.01	57.6
M-NG (n = 10) v S-NG (n = 4)	0.53	p < 0.01	62.6

variability. The pattern for species richness is similar, with average species numbers being higher (>30) in Temperate, subtropical gyre and equatorial waters in northern spring than in southern spring (Table 6). Differences in J' between northern and southern spring cruises in gyre and equatorial waters were minor (Table 6). These trends imply that although the size and composition of the coccolithophore community may change seasonally, evenness (J') was consistently high (>0.8 in gyre and equatorial waters) indicating no overall dominance of the community by one or a few species.

Overall for the four cruises, values of J' were lowest in Temperate waters in both northern and southern spring (average 0.5–0.6, Table 6) which may be due to the high relative abundances of E.

Table 6
Comparison of mean values (\pm Standard Deviation) of surface coccolithophore community composition, sea-surface temperature (SST) and chlorophyll *a* concentrations. Hydrographic provinces (see Fig. 1) are: TMP, Temperate waters; NG, Northern Gyre waters; SG, Southern Gyre waters; EQ, equatorial waters. Species were selected from one-way SIMPER analysis of species contributing to the upper 15% of dissimilarity between flora in different hydrographic provinces grouped by season. The number of samples (n) included in the analysis in each season and each province are also indicated.

Characteristic	Northern spring (AMT 12 and 14, n = 35)				Southern spring (AMT 15 and 17, (n = 27)			
	TMP (n = 9)	NG (n = 10)	SG (n = 7)	EQ (n = 8)	TMP (n = 8)	NG (n = 4)	SG (n = 5)	EQ (n = 11)
Total cells (cells mL ⁻¹)	51.2 (52.4)	24.6 (11.5)	23.7 (4.3)	19.5 (10.0)	166.3 (103.3)	18.2 (4.8)	13.8 (3.5)	16.6 (6.9)
Species richness	30.6 (14.4)	38.9 (11.1)	36.7 (4.2)	32.4 (5.4)	18.7 (6.7)	30.5 (4.1)	29.8 (3.3)	25.6 (4.3)
Pielou's Evenness (J')	0.6 (0.2)	0.8 (0.1)	0.8 (0.1)	0.8 (0.0)	0.5 (0.2)	0.8 (0.1)	0.8 (0.0)	0.8 (0.0)
Shannon-Weiner diversity (H')	2.1 (0.7)	3.0 (0.4)	2.8 (0.2)	2.8 (0.1)	1.6 (0.4)	2.6 (0.3)	2.6 (2.1)	2.6 (0.2)
<i>Emiliania huxleyi</i>	0.30 (0.74)	0.10 (0.06)	0.07 (0.05)	0.08 (0.06)	0.64 (0.55)	0.18 (0.13)	0.12 (0.06)	0.24 (0.11)
<i>Gephyrocapsa ericsonii</i>	0.26 (0.44)	0.05 (0.06)	0.02 (0.03)	–	0.15 (0.19)	0.22 (0.16)	0.27 (0.06)	0.03 (–)
<i>Umbellosphaera irregularis</i>	–	0.03 (0.04)	0.18 (0.17)	0.14 (0.14)	–	0.06 (0.04)	0.08 (0.03)	0.11 (0.10)
<i>Umbellosphaera tenuis</i>	0.05 (0.07)	0.10 (0.10)	0.09 (0.09)	0.06 (0.03)	0.02 (0.01)	0.14 (0.07)	0.21 (0.13)	0.09 (0.06)
<i>Rhabdosphaera xiphos</i>	0.01 (0.01)	0.03 (0.03)	0.08 (0.11)	0.01 (0.01)	0.01 (0.01)	0.08 (0.05)	0.07 (0.03)	0.06 (0.04)
<i>Palusphaera vandellii</i>	0.06 (0.03)	0.11 (0.05)	0.12 (0.05)	0.13 (0.11)	0.01 (0.00)	0.01 (–)	–	0.01 (–)
SST (°C)	17.8 (3.9)	22.9 (1.5)	25.3 (2.2)	27.5 (0.8)	17.9 (0.8)	24.0 (1.4)	24.3 (0.9)	26.9 (0.9)
Chlorophyll <i>a</i> (mg m ⁻³)	0.18 (0.15)	0.04 (0.01)	0.05 (0.02)	0.16 (0.07)	0.26 (0.14)	0.05 (0.02)	0.04 (0.02)	0.16 (0.06)

huxleyi (30–64%) and *G. ericsonii* (15–26%). In subtropical gyre and equatorial waters, *Umbellosphaera irregularis*, *U. tenuis*, *R. xiphos*, and *P. vandellii* (Table 6) had the highest relative abundances in the upper-ocean coccolithophore community. Although these four species contributed to the upper 15% of the dissimilarity between flora in the different hydrographic provinces in different seasons (according to a one-way SIMPER analysis), it is clear from their average relative abundances and high standard deviations in Table 6 that there is little consistent pattern in species level changes. Hence, the whole community (i.e., the species contributing to the other 85% of the dissimilarity) also varied and caused the seasonal differentiation seen in Fig. 8 and Table 5. High values of *J'* further support this notion of whole community compositional variability rather than variability by a few species. A notable exception to this was *P. vandellii*, which seems to have been completely absent in the subtropical gyre and equatorial waters during southern spring (Table 6) although, as already noted, distinguishing this species from *R. xiphos* can be difficult.

3.5. Distribution of rare taxa

Apart from the 31 taxa listed in Table 2, the other species in the AMT database consistently contributed <5% to total cell numbers and are regarded as rare in our analysis. Due to these low cell counts, intermittent occurrences will have a large influence on any statistical analysis of composition differences for this group and hence any trends should be viewed with caution. However, there is still value in examining the compositional changes of the rare species whilst being aware of the potential pitfalls. Examination of the distribution of the rare species shows that this group are widely distributed with respect to both depth and latitude (Figs. 5b and 7d).

A more detailed analysis of the distribution of this large group of species in the AMT samples by nMDS ordination (Fig. 9) shows strongly overlapping distributions, although subgroups of sample composition are identifiable and appear characteristic of temperate (Fig. 9b) and deep euphotic zone (1% and 0.1% surface irradiance; optical depths of 4.6 and 6.9) waters (Fig. 9c). ANOSIM pairwise tests further support statistically significant differences in rare species composition between samples in each of the hydrographic provinces, although differences between temperate and gyre/equatorial waters are statistically strongest ($p < 0.001$) and between samples in the different floral zones, with the differences between UEZ and SEZ statistically strongest ($p < 0.001$) (Table 7). This is indicative of the existence of biogeographic and euphotic

zone floral groups within the rare species, although these species contribute little to the overall numerical abundances (i.e., consistently <5% of the total community).

4. Discussion

4.1. Coccolithophores in the equatorial and subtropical Atlantic Ocean

Total coccosphere (cell) counts of 150–300 per sample were sufficient to reliably quantify the abundance of species present at concentrations of >0.5 cell mL⁻¹ (see Table 1), who represented at least 95% of total cell numbers. The diversity (species richness) of coccolithophore communities is more difficult to determine due to the presence of a high number (140) of relatively rare species defined as consistently representing <5% of total numbers. The presence of a large number of rare coccolithophore species has also been described for other subtropical waters, such as the North Pacific gyre (Cortes et al., 2001; Thierstein et al., 2004). Species richness is dependent on the volume of sample examined which, for this study, was set by the number of coccospheres counted per sample and was generally in the range of 5–16 mL; consideration of Fig. 3 indicates that 50–70% of total species in any one sample were encountered depending on the volume examined. The high species richness, low evenness characteristics of phytoplankton communities, as discussed by Thierstein et al. (2004) for coccolithophores (see also O'Brien et al., 2016) and by Cermeno et al. (2014) for mixed phytoplankton populations, are likely to strongly influence ecological responses to both small and large scale variability in environmental conditions that affect cell growth and survival.

Setting the AMT dataset into the wider context of other open-ocean coccolithophore observations shows several interesting trends. The maximum observed counts for key coccolithophore taxa from the AMT dataset (199 samples) and from repeat sampling at the Bermuda Atlantic time-series (BATS) station (217 samples; Haidar and Thierstein, 2001) and the Hawaiian Oceanographic time-series (HOT) station (183 samples; Thierstein et al., 2004) in the subtropical Atlantic and Pacific oceans respectively are shown in Table 8. For the more numerous species, values are generally higher for BATS and lower for HOT compared to those from the AMT transect, reflecting relatively strong seasonality in surface stratification and nutrient levels at the subtropical gyre margin (BATS), which is poorly resolved within the AMT dataset, and persistent oligotrophic conditions

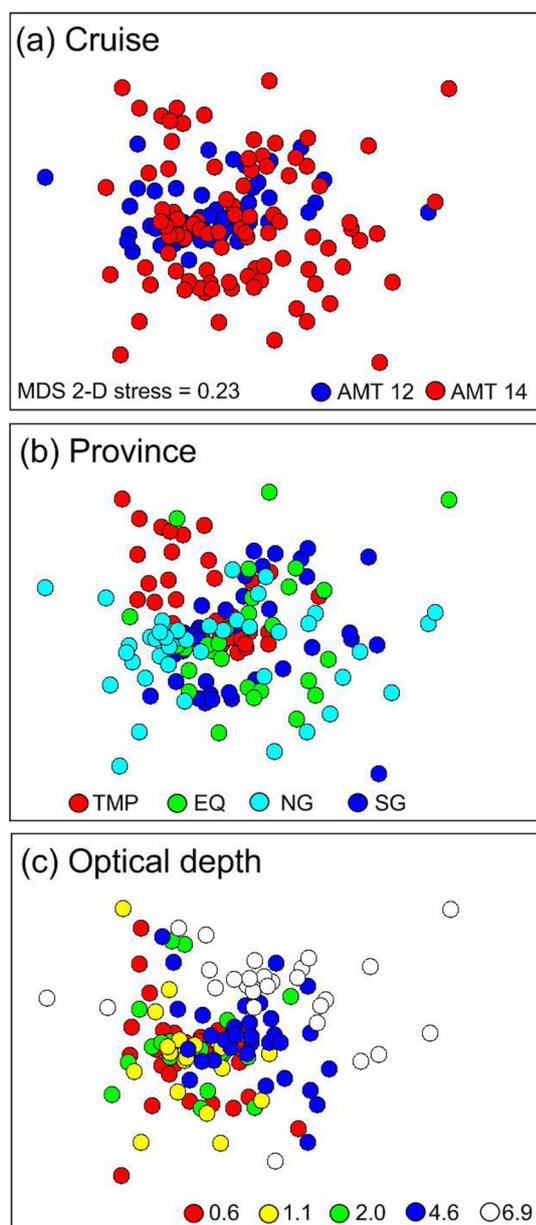


Fig. 9. Normalised Multi-Dimensional Scaling (nMDS) ordination showing relative distributions of rare coccolithophore taxa recorded for all AMT 12 and AMT 14 samples: (a) by cruise; (b) by hydrographic province (TMP, temperate waters; NG, Northern Gyre waters; EQ, equatorial waters; SG, Southern Gyre waters); and (c) by optical depth. Stress on the two-dimensional plot (see (a)) was 0.23 which indicates a 'good representation of the data in two-dimensional space' (Clark and Warwick, 2001).

throughout the year nearer to the gyre centre (HOT) compared to the more varied hydrography along the AMT transect (e.g., Fig. 2).

Another detailed taxonomic study of coccolithophores in the subtropical Atlantic is the work of Boeckel and Baumann (2008) who examined around 60 samples along a transect between 8°S and 45°S. Boeckel and Baumann (2008) did not provide coccolithophore counts for individual taxa, preferring instead to express their results as 'sphere units' in order to take account of detached coccoliths. However, the most numerous taxa found by these authors north of 34°S are all listed in Table 8. Similarly, analyses of coccolithophore assemblages in warm waters of the Pacific (Okada and Honjo, 1973; Honjo and Okada, 1974; Hagino et al., 2000; Hagino and Okada, 2004) and Indian Oceans (Takahashi and Okada, 2000) are also consistent with the AMT data (i.e., iden-

Table 7

Multivariate statistical analyses of rare coccolithophore distributions (i.e., species consistently contributing <5% of total numbers in all samples) for all stations on the AMT 12 and AMT 14 cruises. The species composition for each sample is examined in relation to: (a) cruise number (AMT 12 or AMT 14); (b) hydrographic province; and (c) photic zone floral group. Hydrographic provinces (see Fig. 1) are: Temperate waters (TMP), equatorial waters (EQ), Northern Gyre waters (NG), and Southern Gyre waters (SG). Photic zone floral groups are: Upper Euphotic Zone (UEZ), Lower Euphotic Zone (LEZ), and Sub-euphotic Zone (SEZ). ANOSIM is a one way analysis of similarities; SIMPER is a one way analysis of (dis)similarity percentages species contributions to the differences between euphotic zone groups. The number of samples (n) in each province in each season is given in parenthesis.

Groups	ANOSIM R-Statistic	p level	SIMPER Average dissimilarity (%)
<i>Comparison by Hydrographic province (n = 150)</i>			
TMP (n = 35) v SG (n = 38)	0.21	p < 0.001	80.7
TMP (n = 35) v EQ (n = 34)	0.18	p < 0.001	80.1
TMP (n = 35) v NG (n = 43)	0.15	p < 0.001	81.6
SG (n = 38) v EQ (n = 34)	0.14	p < 0.001	80.8
SG (n = 38) v NG (n = 43)	0.07	p < 0.005	81.0
EQ (n = 34) v NG (n = 43)	0.10	p < 0.01	81.8
<i>Comparison by Euphotic Zone Floral Groups (n = 85)</i>			
UEZ (n = 41) v LEZ (n = 19)	0.29	p < 0.001	79.3
UEZ (n = 41) v SEZ (n = 25)	0.58	p < 0.001	90.4
LEZ (n = 19) v SEZ (n = 25)	0.12	p < 0.01	86.8

tical species lists to those in Tables 2 and 4). The main differences within the equatorial and subtropical Atlantic, and between the Atlantic and other oceanic regions, are in the regional abundances of taxa such as *E. huxleyi*, *G. oceanica* and *C. leptopus*, which are all more abundant in areas of elevated nutrient levels, higher surface water temperatures and stronger water column stratification (Kinkel et al., 2000; Hagino et al., 2000; Franklin et al., 2009).

The species from each of the three floral depth groups and their ranges of maximum abundance listed in Table 8 can therefore be considered as representative of coccolithophore populations throughout the oligotrophic ocean. Regional increases in the strength of localised upwelling, both at the ocean margins and close to the Equator, and in seasonal mixing along the high latitude boundaries of the subtropical gyres are typically associated with a greater abundance of LEZ taxa. Comparative quantitative information on the less common coccolithophore taxa has yet to be collated, but all open ocean samples examined by SEM have shown species richness comparable, though slightly lower, to that found for the AMT transect (171 taxa): 112 taxa were reported from the South Atlantic (Boeckel and Baumann, 2008); 100–125 taxa from the Pacific Ocean (Hagino et al., 2000; Cortes et al., 2001; Hagino and Okada, 2004); and 92 taxa from the Indian Ocean (Takahashi and Okada, 2000).

From an ecological perspective, in terms of the response of communities to environmental variability, it is important to consider the significance of the high diversity (species richness) of coccolithophore communities in oligotrophic waters (Fig. 3). Such diversity is characteristic of many types of microbes in the ocean, including the related non-calcifying haptophytes (Liu et al., 2009), generally taking the form of a few abundant species and a large number of rare ones (Logares et al., 2014), and is thought to be fundamental to the maintenance of ecosystem function and associated biogeochemical processes (Caron and Countway, 2009). The high species richness of each of the coccolithophore floral zones (UEZ, LEZ, SEZ) in the water column (see Tables 2 and 4) supports the conclusion of Logares et al. (2014), that assemblages have fairly regular proportions of abundant and rare taxa but contrasting structuring patterns across space and time. Within the context of environmental change, diversity represents a genetic reservoir of community adaptability. However, for coccolithophores or other types of microbes, it will not be possible to

Table 8

A comparison of maximum coccospere counts (cells mL⁻¹) from equatorial and subtropical samples from AMT (cruises 12, 14, 15 and 17; this study) with those from the Bermuda Atlantic Time-Series (BATS; Haidar and Thierstein, 2001) and Hawaii Ocean Time-series (HOT; Cortes et al., 2001; Thierstein et al., 2004). LM indicates light microscope counts; SEM indicates Scanning Electron Microscope counts. The vertical floral group affiliations are also indicated (UEZ, Upper Euphotic Zone; LEZ, Lower Euphotic Zone; SEZ, Sub-euphotic Zone; R indicates species classified as rare in the vertical floral analysis, see Section 3.1). Values in parentheses are maximum counts from AMT temperate water samples.

Species	Floral Group	AMT 32°S–30°N SEM	BATS 32.2°N LM	HOT 22.7°N SEM
<i>Calcidiscus leptoporus</i>	R	1.5 (37.0)	2.3	0.2
<i>Discosphaera tubifera</i>	UEZ	3.7 (8.2)	2.3	na
<i>Emiliana huxleyi</i>	LEZ	35.4 (265.8)	92.7	19.9
<i>Florisphaera profunda</i>	SEZ	42.7	67.5	15.0
<i>Gephyrocapsa ericsonii</i>	LEZ	12.0 (224.1)	26.2 ^a	7.9
<i>G. oceanica</i>	R	4.0	1.4	3.1
<i>Gladiolithus flabellatus</i>	SEZ	9.6	5.2	na
<i>Helicosphaera</i> spp.	R	1.2	1.4	0.2
<i>Rhabdosphaera clavigera</i>	LEZ	6.2	4.7	na
<i>Syracosphaera pulchra</i>	UEZ	1.8	0.9	0.8
<i>Umbellosphaera irregularis</i>	UEZ	11.2	16.2	19.9
<i>U. tenuis</i>	UEZ	9.8	34.1	16.4
<i>Umbilicosphaera hulbertiana</i>	R	2.6 (8.8)	0.5	na
<i>U. sibogae</i>	R	1.2 (1.4)	1.6	0.2

na = not available.

^a Counted as small coccospere.

detect adaptation without initial detailed taxonomic or genomic descriptions of community structure, including the rare species.

Within the limits set by the AMT sampling programme (2 cruises per year, 2003–2005) inter-cruise differences in coccolithophore community composition were evident (Fig. 8a, Table 5). Such inter-cruise differences may relate to seasonal variability, though the timeframe of sampling is too limited to directly link these to seasonality. Clear and statistically significant (Table 5) inter-cruise differences were most obvious in Temperate waters (Fig. 8b), where large variations were observed in the abundance of common taxa such as *E. huxleyi*. Statistically significant differences in species composition were not limited to Temperate waters, with seasonal differences also detected in the subtropical gyres and equatorial waters (Table 5). Finally, it should be noted that, although large scale community distributional patterns can be generally related to large scale hydrographic and climatic factors, the causes for differences in the abundance of widespread taxa such as, for example, *Umbellosphaera* spp. over relatively small spatial scales (see Fig. 7c) remain unknown. High variability in rates of primary production in the oligotrophic Atlantic (e.g., Marañón et al., 2003) has been attributed to variations in rates of localised nutrient supply (see Johnson et al., 2010), and hence it appears that the structure, composition and dynamic properties of biological communities in the subtropical gyres is much less uniform than previously thought.

4.2. Coccolithophore biogeography: vertical and horizontal distributions

The characteristic vertical distribution of coccolithophore taxa observed at stations representative of the subtropical gyres and equatorial waters (Fig. 5) follows closely the general pattern observed for subtropical waters from earlier studies (e.g., McIntyre and Be, 1967; Winter et al., 1994). The terminology used in this study (UEZ, LEZ, SEZ) differs from that of other studies (e.g., Winter et al., 1994; Upper Photic Zone, Middle Photic Zone, Lower Photic Zone) due to a focus on the euphotic zone which supports upper ocean primary production. As the DCM is generally considered to mark the base of the euphotic zone (i.e., 1% of surface irradiance), the deepest coccolithophore zone is referred to as the 'Sub-euphotic Zone' rather than 'Lower Photic Zone', with two further zones in the upper euphotic zone (i.e., from the depth of 1% surface irradiance to the surface). The existence of the two upper

euphotic zones (UEZ, LEZ) is supported by the species' normalised abundance profiles, with certain species having peaks not in the upper portion of the water column (UEZ) but deeper and above the depth of 1% surface irradiance (e.g., *Calciopappus* spp. and *G. ericsonii*; Fig. 5b).

For stations where samples were taken from just the 5 standard light depths used for AMT primary production studies (e.g., Poulton et al., 2006a, 2006b), it is difficult to describe or compare in detail the coccolithophore communities of the three vertical zones; the DCM (1% surface irradiance) is located at the transition between the LEZ and SEZ, with each of these two lower zones represented by just one sample (1% and 0.1% surface irradiances, respectively) that may or may not correspond to the maximum abundance of the key representative species (see Fig. 5). In contrast, the UEZ is better represented by three samples (55%, 33% and 14% surface irradiances).

Similarly, the horizontal (meridional) spacing of stations at which coccolithophores were sampled on AMT cruises 12 and 14 (Fig. 2a) is too coarse to show how oceanographic boundaries (see Aiken et al., 2000) or mesoscale eddy structure might relate to coccolithophore abundance or composition (e.g., Jin et al., 2016). In general, statistically similar communities in terms of species composition were found at closely spaced stations (<20 km apart, e.g., at the Equator on AMT 14; Fig. 7c), but there were wide differences in the relative abundance of common taxa (Fig. 7c). However, comparison of Fig. 6b and c shows that, in general, coccolithophore communities differed to a much greater degree vertically in the water column (surface layer, DCM, sub-DCM) than horizontally across hydrographic provinces (gyres, equatorial waters). This is further supported through comparison of vertical profiles of (Bray-Curtis) similarity (Figs. 5a and 7e) to meridional changes in similarity with latitude (Fig. 7b).

The depth distribution of the main floral groups summarised in Table 2 is fully consistent with the conclusions of Winter et al. (1994) (see also McIntyre and Be, 1967), who for subtropical waters, identified upper, middle and lower photic zones using similar reference light levels (>10%, 1–10%, and <1% of surface irradiance, with the DCM at the 1% level). Winter et al. (1994) stated that the middle photic zone was 'not easily distinguished by a characteristic flora', as also shown by the nMDS ordination analysis (Fig. 6c) for the AMT samples, with LEZ species (optical depth = 4.6) showing considerable overlap with both the UEZ group (optical depths 0.6, 1.1 and 2; irradiance levels 55, 33 and 14%) and SEZ

group (optical depths 4.6 and 6.9; 1 and 0.1%). It is not surprising therefore that some species listed for the LEZ in Table 2 have been previously assigned to the layers above (e.g., *Ceratolithus* spp.) or below (e.g., *Syracosphaera anthos*). The conclusion of Winter et al. (1994) that *Umbellosphaera tenuis*, *Syracosphaera* spp., and placolith-bearing genera (such as *E. huxleyi* and *Gephyrocapsa* spp.) show no depth preference but tend to be most abundant in the middle photic zone is generally well supported by the AMT data, although some of the diverse *Syracosphaera* group (in particular *S. pulchra*) are found mainly in the UEZ (Table 2) and *U. tenuis* is classed as a UEZ taxon that extends to greater depths than *U. irregularis* (see Fig. 5a) in the AMT analysis.

Two general features of coccolithophore vertical distribution shown by the AMT data are firstly that the Bray-Curtis Similarity index declines with respect to the surface sample throughout the UEZ and LEZ (Fig. 4b; see also Fig. 7e), whereas the total numbers of both cells and species remain uniform between the surface and the depth of 1% surface irradiance (Fig. 4). Secondly, cell numbers for some characteristic SEZ taxa (in particular *Gladiolithus flabellatus*; see Fig. 5a) reach a maximum at or below the depth of 0.1% surface irradiance (Fig. 5b). The first observation implies that despite compositional changes in species composition with depth, the relative size of the community remains consistent, and that rare taxa as well as abundant ones (Table 2) change with depth; within the AMT dataset rare taxa that were largely confined to the LEZ include *Acanthoica quattrosolina*, *Alisphaera pinnigera*, *Corisphaera gracilis*, *Gephyrocapsa oceanica*, *G. ornata*, *Helicosphaera* spp., *Picicola margalefi*, *Syracosphaera reniformis* and *Umbellosphaera* spp. Peaks in species abundances well below the depth of 1% surface irradiance, where light can be assumed to be limiting to photosynthetic processes whilst nutrients are non-limiting (e.g., Poulton et al., 2006b), are likely indicative that most, if not all, SEZ species (e.g., *G. flabellatus*) have alternative nutritional strategies than autotrophy (i.e., mixotrophy or phagotrophy).

Regional anomalies in the vertical distribution of coccolithophores can be inferred from Fig. 7e. Firstly, greater similarity between samples from the depths of 14% and 0.1% of surface irradiance in temperate waters compared to subtropical waters reflects greater vertical mixing of the upper water column at higher latitudes and a more uniform environment; in autumn (i.e., at the southern stations on AMT 12 and 14), this trend can be attributed partly to populations of SEZ species, including *Florisphaera profunda*, being mixed towards the surface, whereas in spring (northern stations) UEZ and LEZ species occur throughout the water column before the DCM and associated flora becomes well established. Secondly, greater statistical similarity of species composition between samples from the 1% and 0.1% surface irradiance depths in subtropical waters of the northern hemisphere (spring) compared to those of the southern hemisphere (autumn) could be related to seasonal changes in the depth of the permanent DCM (see Letelier et al., 2004); in spring, when the depth of 1% surface irradiance deepens such that the DCM is displaced downwards into a layer where SEZ species, in particular, *F. profunda* are relatively abundant.

The UEZ and SEZ coccolithophore taxa (Table 2) are largely confined to the well-stratified upper waters (<100 m) of the equatorial and subtropical Atlantic (Fig. 7e), with their distributions extending polewards in summer months with seasonal strengthening of stratification (see Haidar and Thierstein, 2001). In contrast, LEZ taxa, and associated rare taxa (e.g., as listed above, Fig. 10b), are widely distributed with respect to both latitude and depth (Fig. 6f) and reach their maximum abundance in temperate waters (Figs. 6d and 7f) where high concentrations (>50 coccospheres mL⁻¹) of species such as *E. huxleyi* and *G. ericonii* are observed. Hence, the LEZ taxa often represent the dominant species in temperate and high latitude waters, a phenomenon poten-

tially linked to light availability in that light levels experienced in the LEZ (i.e., 10% of 2000 μmol photons m⁻² s⁻¹ on a 14–16 h day equates to 10.1–11.5 mol photons m⁻² d⁻¹) are similar to average mixed layer irradiances experienced by temperate coccolithophore communities (e.g., 10–15 mol photons m⁻² d⁻¹; see for e.g., Poulton et al., 2010, 2014).

4.3. Ecological and nutritional strategies of subtropical coccolithophores

A notable feature of the AMT data is a strong difference in the mean vertical distribution between HET and HOL forms of the same species (see Tables 2 and 4), with the latter consistently occurring at shallower depths as has been described for stratified waters of the Mediterranean by Dimiza et al. (2008) and Cros and Estrada (2013). The best example in the AMT dataset is provided by *Syracosphaera anthos* (Fig. 10a); with the distribution of the HET form which belongs to the LEZ floral group often extending well below the DCM, whereas the HOL form is characteristic of the UEZ group and typically confined to surface waters. Widening this analysis to all species that have recognised HOL-HET stages in the AMT database (9 species; *Acanthoica quattrosolina*, *Calcidicus leptoporus*, *Coccolithus pelagicus*, *Coronosphaera mediterranea*, *Heli-*

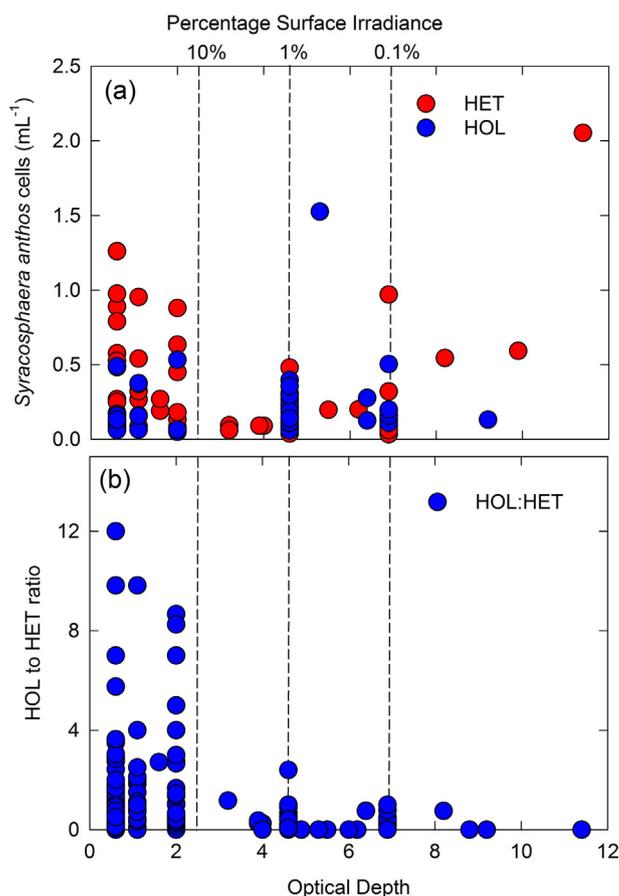


Fig. 10. Depth distribution of hetero- (HET) and holo-coccosphere (HOL) taxa against optical and percentage surface irradiance depth for all AMT 12 and AMT 14 samples from Southern Gyre waters, equatorial waters and Northern Gyre waters. (a) Depth distribution of *Syracosphaera anthos* cells in both HET and HOL form. (b) All coccolithophore taxa present as both HOL and HET forms (i.e., *Acanthoica quattrosolina*, *Calcidicus leptoporus*, *Coccolithus pelagicus*, *Coronosphaera mediterranea*, *Helicosphaera carteri*, *Syracosphaera anthos*, *S. bannockii*, *S. nana*, and *S. pulchra*). Vertical dashed lines indicate the percentage surface irradiance depths used to differentiate between floral depth zones (UEZ, >10%; LEZ, 10–1%; SEZ, <0.1%).

cosphaera carteri, *Syracosphaera anthos*, *S. bannockii*, *S. nana*, and *S. pulchra*) and plotting the ratio of HOL to HET cell counts (Fig. 10b) shows a clear decrease in the relative abundance of HOL forms with depth. However, there are also clearly high numbers of HET forms for some species (e.g., *Acanthoica quattrosipina*) in surface waters as indicated by the low HOL:HET ratios at light levels >10%.

A clear trend in the ecology of subtropical coccolithophores, in terms of floral groups, rare species and HOL-HET species combinations is that a relatively large number of cells and species are found deep in the water column (Tables 2 and 6; Figs. 4–6, 9 and 10). Such deep waters experience low light levels in both relative (<1% of surface irradiance) and absolute terms (<20 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ based on a surface irradiance of 2000 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ or <2 mol photons $\text{m}^{-2} \text{d}^{-1}$ on a 14–16 h day), which are likely to be limiting to autotrophic growth. The oligotrophic waters of the equatorial and subtropical Atlantic Ocean are characterised by a DCM which is situated on the nutricline (Fig. 2); autotrophs above the DCM are likely (inorganic) nutrient-limited, whilst below they are likely light-limited (e.g., Poulton et al., 2006b). Hence, most coccolithophore species face the problem of either insufficient inorganic (nitrate, phosphate) nutrients (UEZ and LEZ taxa) in the upper water column or insufficient light (SEZ taxa) at depth in order to compete with the dominant, small (<2 μm) photosynthetic picoplankton that have a high efficiency for the assimilation of nutrients and light energy (Zubkov, 2014).

The DCM is generally considered to be a pigment rather than a biomass maximum (i.e., composed of cells with a low carbon-to-chlorophyll ratio and hence high cellular pigmentation) across the oligotrophic Atlantic Ocean. Picoeukaryotes, autotrophic cells <2 μm in diameter, are relatively abundant in the DCM of the Atlantic subtropical gyres (<1000–5000 cells mL^{-1} ; Tarran et al., 2006) and, in the northern gyre, their distribution has been shown to be closely associated with deep peaks in uptake rates for nitrate (Painter et al., 2014). Tarran et al. (2006) also reported that the maximum biomass of autotrophic nanoeukaryotes (2–10 μm diameter) occurred immediately above the DCM, where growth is likely to be enhanced close to the nutricline by some upward diffusion of nutrients and by nutrient regeneration associated with biological activity at the DCM.

For the coccolithophores, the DCM represents a transitional zone between the LEZ and SEZ floral groups (Fig. 5a); the number of coccospheres in the DCM, although dominated by LEZ taxa, is lower than in the UEZ except at equatorial stations (Fig. 4a), and important LEZ species often show a clear maximum in abundance above, rather than at, the DCM (Fig. 5b). Thus, there appears to be no clear relationship between the vertical distribution of coccolithophores and the position of the nutricline, suggesting that irradiance at this depth is too low to support nitrate assimilation by these relatively large (5–20 μm) cells. However, the LEZ group includes taxa (*E. huxleyi*, *Gephyrocapsa* spp.) that increase in abundance in response to nutrient enrichment as well as taxa (*Calciolenia*, *Discosphaera*, *Michaelsarsia*) that do not (see Brand, 1994). The former probably have a similar nutritional status to the nano-eukaryotes mentioned above whereas the latter, together with UEZ taxa, are considered more characteristic of nutrient-depleted oceanic waters and, almost without exception, have not been grown successfully in culture. Little is known about the biology of SEZ taxa, even to the extent as to whether or not they all have functional chloroplasts and are autotrophic. At light levels <1% of surface irradiance it seems very unlikely that they could reach observed abundances (Table 8) by phototrophy (photosynthesis and reduction of inorganic nutrients) alone.

There are a number of references to possible mixotrophy by coccolithophores in connection with their survival in low-nutrient oceanic waters (Brand, 1994), the function of the haptonema (flagella) as a food gathering organelle (Kawachi and

Inouye, 1995), the suggested function of modified coccoliths as 'particle collectors' (Aubry, 2009), and their phylogenetic linkages to heterotrophic flagellates (de Vargas et al., 2007). There is still little observational or experimental evidence for mixotrophy by coccolithophores. However, from a consideration of (i) the physiological ecology of species inhabiting oligotrophic oceanic waters (Brand, 1994), (ii) mixotrophy by non-calcifying haptophytes (Liu et al., 2009; Unrein et al., 2014), (iii) the quantitative importance of mixotrophy in oligotrophic oceanic ecosystems (Hartmann et al., 2012), (iv) the existence of heterotrophic coccolithophores in polar waters (Thomsen et al., 1991), and (v) the recognition of the significance of mixotrophy for the oceanic biological carbon pump (Mittra et al., 2014) it is highly likely that many coccolithophores are actually mixotrophic, with possible exceptions being placolith-bearing species belonging to the LEZ group which form blooms in nutrient-enriched waters (e.g., *E. huxleyi*, *Gephyrocapsa* spp., *Calcidiscus* spp., *Umbilicosphaera* spp.).

The distinct vertical profiles for the HOL and HET forms of the same species (Dimiza et al., 2008; Cros and Estrada, 2013), may therefore reflect differences in the degree or type of mixotrophy across the group; HOLs in shallower, more oligotrophic water could supplement phototrophy with heterotrophy whereas HETs, in the absence of evidence for an ability to utilise nitrate under low light at depth, could be largely or entirely heterotrophic. A shift from one life-cycle stage to another may also represent an ecological strategy (Houdan et al., 2004, 2006) in response to changes in environmental conditions. In the case of the HET form of *Syracosphaera anthos* (Fig. 10), nano-eukaryotes in the DCM (Painter et al., 2014) are potentially an important food source. Alternative nutritional strategies for coccolithophores in low (inorganic) nutrient conditions may also include the use of dissolved organic carbon and nutrient sources to supplement or replace photosynthetic growth, though the composition, lability and bioavailability of this material is unclear. Clearly, innovative future research on the nutritional strategies of (subtropical) coccolithophores is key to fully understanding the ecology and biogeography of such species, as well as gaining a better perspective on coccolithophore impacts on the biological carbon pump in the oligotrophic ocean.

A better understanding of the nutrition of coccolithophores will also open the way to improved interpretation of data on the distribution of coccoliths in marine sediments in relation to environmental conditions. In particular for the important palaeo-indicator species *Florisphaera profunda*, SEZ taxa are mainly or entirely found below the DCM and if they are partly or wholly heterotrophic then their abundance is more likely to reflect the availability of living and non-living particulate material rather than of irradiance or inorganic nutrients (e.g., Molfino and McIntyre, 1990) or of a well-developed DCM (e.g., Grelaud et al., 2012). Furthermore, the depths of the nutricline and the SEZ are probably not independent variables as envisaged by Molfino and McIntyre (1990), in the sense that the depth of the nutricline approximates the 1% surface irradiance level, whether it is deep or shallow, and varies with the degree of biological attenuation of light in the overlying water (and the uptake of nitrate as light becomes available; Letelier et al., 2004). Hence, a shallowing of the depth of the 1% surface irradiance will be accompanied by a shallower nutricline as nitrate uptake follows light availability.

Furthermore, oceanic ecosystems associated with a shallow thermocline are more productive, and also generally show a greater abundance of *F. profunda* in the water column; for example, Fig. 2 shows how the nutricline (and DCM) shallows in the equatorial region and northern gyre in spring during AMT 14 (see also Figs. 2 and 4 in Robinson et al., 2006), also at gyre margins (BATS) compared to gyre centres (HOT) (see Table 8). Therefore, it appears that changes in the relative abundance of *F. profunda* coccoliths in

ocean sediments may reflect changes in the production and/or dissolution of coccoliths of UEZ and LEZ taxa. In the oligotrophic subtropical ocean with a deep nutricline, phytoplankton are relatively scarce and will be efficiently consumed by herbivores within surface waters (e.g., Poulton et al., 2006a), leading to high losses of coccoliths due to dissolution (Milliman et al., 1999), whereas at gyre boundaries and around upwelling regions, increases in coccolithophore abundance in the euphotic layer will lead to sudden downward fluxes of UEZ and LEZ coccoliths, a proportion of which may escape dissolution. However, at ocean boundaries and within enclosed seas, other factors may control the relative abundance of different types of coccoliths in the sediments (Ahagon et al., 1993); organic matter derived from continental shelf and terrestrial sources may be a significant source of food for heterotrophic coccolithophores, whilst freshwater inputs can affect the growth conditions for autotrophic taxa either positively or negatively through effects on nutrient levels, on light penetration (turbidity), and on depth of the thermocline. For example, in the western subtropical Atlantic, sediments in which *G. flabellatus* is more abundant than *F. profunda* are found in offshore regions affected by the outflow of the Amazon River, close to the continental slope affected by the Brazil Current and over the mid-Atlantic ridge (Boeckel et al., 2006); it would be of great interest to know why this happens in such contrasting situations.

4.4. Future recommendations

Though sample collection was limited to a depth resolution of only five standard light depths across much of the AMT transect, these light depths still reflected the general pattern of the vertical distribution of species composition in the equatorial and subtropical Atlantic Ocean. Ideally, higher resolution sampling (e.g., 15–20 sampling levels) would lend further support to the vertical patterns identified in this study, and if combined with increased meridional sampling frequency would also greatly expand the species level insights gained. The difficulty in reaching an examination volume which fully accounts for the incredible level of species diversity represents a limitation to gaining further insights into the biogeography and ecology of the rare species. Furthermore, the point where the cumulative species richness plateaus against examination volume (i.e., where the rarefaction curve plateaus) will likely be sample specific, also providing a limitation. However, examination of 150–300 coccospores per sample did account for the dominant species in terms of cell numbers: with 31 out of the 171 taxa representing 95% of total cell numbers, and hence though it is not possible to fully describe the full species richness of the community it is possible to reliably analyse the vertical and meridional distribution of the numerically dominant species. In order to confidently identify seasonal and interannual variability in species composition in the subtropical ocean, a longer time-frame is required to be able to differentiate random variability from temporal trends. Such an expanded dataset, if geographically aliased in a suitable way, would also provide further insights into the biogeography of rare coccolithophore species.

The new perspectives gained from statistical exploration of coccolithophore distribution in the equatorial and subtropical Atlantic Ocean warrant further examination and confirmation. Such research will require innovative methodology and experimentation to confirm the nutritional strategies of upper-ocean and deep-dwelling coccolithophore species. For example, tracing particulate uptake through the use of fluorescently labelled prey items, high-resolution microscopy to examine the presence/absence of chloroplasts, and tracing the cellular uptake of labelled compounds. Heterotrophic or mixotrophic nutrition of coccolithophores has significant implications for their biogeochemical roles; for example, the impact on dissolved inorganic carbon and

alkalinity through calcification, photosynthesis, respiration and dissolution will be very different to a cell which lacks photosynthesis and has high ratios of calcification to respiration. Indeed, the cellular coupling of photosynthesis and calcification, in terms of internal (carbon) substrate competition, will be very different in a cell which attains its energetic needs through respiration rather than autotrophy. How this impacts on cellular rates of calcification and growth, and hence the fitness and competition between autotrophic and heterotrophic (partly or fully) coccolithophores is completely unknown, but a key issue to be addressed when considering the species composition of oceanic communities. Moreover, communities dominated by autotrophic, heterotrophic or mixotrophic coccolithophores will have different influences on air-sea CO₂ fluxes through variable roles in organic and inorganic production, and the biological carbon pump and carbonate-counter pump.

5. Conclusions

Across 199 samples collected in the equatorial and subtropical Atlantic Ocean, 171 coccolithophore taxa were identified (Appendix A); of which 140 were classified as rare, consistently representing <5% of total cell numbers in all samples, whilst 31 were observed in enough abundance to allow us to analyse their vertical and meridional distribution. From multivariate statistical analysis, strong vertical gradients in species composition were identified in equatorial and subtropical waters, which lessened in Temperate waters. Three vertical zones in terms of floral composition were identified: an upper euphotic zone (UEZ), a lower euphotic zone (LEZ), and a sub-euphotic zone (SEZ). This vertical zonation is closely tied to the light availability through the water column, and the species involved in each zone are broadly similar to those identified by Winter et al. (1994) (see also McIntyre and Be, 1967). Light levels in the SEZ are likely to be well below those required to support photosynthesis and hence it is suggested that ingestion of other plankton (mixotrophy or full phagotrophy) or dissolved organic compounds may support the growth (and hence calcification) of species at these depths.

Coccolithophore cell numbers were highest in temperate waters, whilst species richness was highest in the subtropical gyres and equatorial waters; comparison of gyre samples with other surface samples highlights relatively high similarity (>50%) across large horizontal distances, which contrasts with the sharp declines in similarity with depth seen in the gyres and equatorial waters. UEZ taxa were largely restricted to the subtropical gyres, whilst the LEZ taxa were found in both equatorial (EQ) and Temperate regions and some SEZ species were found in surface Temperate waters. LEZ species often represent the dominant species in Temperate and high latitude waters, which is potentially linked to similar light (and inorganic nutrient) climates between the LEZ in the subtropics and mixed layers in Temperate waters. Seasonal differences in coccolithophore species composition were stronger than inter-annual differences. The strongest seasonal variability seen was in Temperate waters, with elevated abundances of species such as *E. huxleyi* and *G. ericonii* (both LEZ). Clearly the coccolithophore communities in low-latitude, low (inorganic) nutrient waters respond to temporal changes in environmental conditions and these environments are not quiescent regions of the global ocean.

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Appendix A

List of taxa identified by Scanning Electron Microscopy. Holococcolithophores (HOL) indicated in bold italics.

Species name	Species name
<i>Acanthoica maxima</i>	<i>Cyrtosphaera lecaliae</i>
<i>Acanthoica quattrosolina</i>	<i>Cyrtosphaera</i> sp.
<i>Acanthoica quattrosolina</i> HOL	<i>Discosphaera tubifera</i>
<i>Acanthoica</i> sp.	<i>Emiliana huxleyi</i>
<i>Algirosphaera cucullata</i>	<i>Ericolus?</i> sp.
<i>Algirosphaera robusta</i>	<i>Florisphaera profunda</i>
<i>Alisphaera capulata</i>	<i>Flosculosphaera calceolariopsis</i>
<i>Alisphaera extenta</i>	<i>Gephyrocapsa ericsonii</i>
<i>Alisphaera gaudii</i>	<i>Gephyrocapsa muelleriae</i>
<i>Alisphaera ordinata</i>	<i>Gephyrocapsa oceanica</i>
<i>Alisphaera pinnigera</i>	<i>Gephyrocapsa ornata</i>
<i>Alisphaera quadrilatera</i>	<i>Gephyrocapsa</i> sp.
<i>Alisphaera unicornis</i>	<i>Gladiolithus flabellatus</i>
<i>Alisphaera</i> sp.	<i>Gliscolithus amitakareniae</i>
<i>Alveosphaera bimurata</i>	<i>Hayaster perplexus</i>
<i>Anacanthoica acanthos</i>	<i>Helicosphaera carteri</i>
<i>Anacanthoica cidaris</i>	<i>Helicosphaera carteri</i> HOL
<i>Anthosphaera fragaria</i>	<i>Helicosphaera hyalina</i>
<i>Anthosphaera lafourcadii</i>	<i>Helicosphaera pavementum</i>
<i>Anthosphaera periperforata</i>	<i>Helicosphaera wallichii</i>
<i>Calcidiscus leptoporus</i>	<i>Helicosphaera</i> sp.
<i>Calcidiscus leptoporus</i> HOL	<i>Helladosphaera cornifera</i>
<i>Calciopappus</i> sp.	<i>Helladosphaera pienaarii</i>
<i>Calciosolenia brasiliensis</i>	<i>Helladosphaera vavilovii</i>
<i>Calciosolenia murrayi</i>	<i>Homozygosphaera arethusae</i>
<i>Calicasphaera diconstricta</i>	<i>Homozygosphaera spinosa</i>
<i>Calyptrolithina divergens</i>	<i>Homozygosphaera triarcha</i>
<i>Calyptrolithina multipora</i>	<i>Homozygosphaera vercellii</i>
<i>Calyptrolithophora papillifera</i>	<i>Hymenomonas lacunae</i>
<i>Calyptrosphaera dentata</i>	<i>Jomonolithus</i> sp.
<i>Calyptrosphaera heimdaliae</i>	<i>Michaelsarsia adriaticus</i>
<i>Calyptrosphaera sphaeroidea</i>	<i>Michaelsarsia elegans</i>
<i>Canistrolithus</i> sp. 1	<i>Ochrosphaera neapolitana</i>
<i>Ceratolithus</i> spp.	<i>Oolithotus antillarum</i>
<i>Coccolithus pelagicus</i>	<i>Oolithotus fragilis</i>
<i>Coccolithus pelagicus</i> HOL	<i>Ophiaster</i> spp.
<i>Corisphaera gracilis</i>	<i>Palusphaera vandellii</i>
<i>Corisphaera strigilis</i>	<i>Palusphaera</i> sp.1
<i>Corisphaera tyrrheniensis</i>	<i>Pappomonas</i> sp. Type 2

Appendix A (continued)

Species name	Species name
<i>Coronosphaera binodata</i>	<i>Pappomonas</i> sp. Type 3
<i>Coronosphaera maxima</i>	<i>Pappomonas</i> sp. Type 4
<i>Coronosphaera mediterranea</i>	<i>Pappomonas</i> sp.
<i>Coronosphaera mediterranea</i> HOL hellenica	<i>Papposphaera arctica</i>
<i>Coronosphaera mediterranea</i> HOL wettsteinii	<i>Papposphaera borealis</i>
<i>Coronosphaera</i> sp.	<i>Papposphaera lepida</i>
<i>Cyrtosphaera aculeata</i>	<i>Papposphaera thomsenii</i>
	<i>Papposphaera</i> sp. Type 1
Species name	Species name
<i>Papposphaera</i> sp. Type 2	<i>Syracosphaera pirus</i>
<i>Papposphaera</i> sp. Type 4	<i>Syracosphaera prolongata</i>
<i>Papposphaera</i> sp.	<i>Syracosphaera protrudens</i>
<i>Picarola margalefii</i>	<i>Syracosphaera pulchra</i>
<i>Placorhombus ziveriae</i>	<i>Syracosphaera pulchra</i> HOL pirus
	<i>Syracosphaera pulchra</i> HOL oblonga
<i>Pleurochrysis carterae</i> var. <i>carterae</i>	<i>Syracosphaera rotula</i>
<i>Pleurochrysis roscoffensis</i>	<i>Syracosphaera tumularis</i>
<i>Polycrater galapagensis</i>	<i>Syracosphaera hastata</i>
<i>Polycrater</i> sp. 'ladle like'	<i>Syracosphaera didyma</i>
<i>Polycrater</i> sp.	<i>Syracosphaera castellata</i>
<i>Pontosphaera syracusana</i>	<i>Syracosphaera reniformis</i>
<i>Poricalyptra aurisinae</i>	<i>Syracosphaera squamosa</i>
<i>Poricalyptra gaarderiae</i>	<i>Syracosphaera leptolepis</i>
<i>Poricalyptra isselii</i>	<i>Syracosphaera</i> sp.
<i>Poricalyptra magnaghii</i>	<i>Tetralithoides quadrilaminata</i>
<i>Poritectolithus maximus</i>	<i>Tetralithoides</i>
	<i>Turrilithus latericiooides</i>
<i>Poritectolithus poritectum</i>	<i>Umbellosphaera irregularis</i>
<i>Reticulofenestra parvula</i>	<i>Umbellosphaera tenuis</i>
<i>Reticulofenestra sessilis</i>	<i>Umbilicosphaera anulus</i>
<i>Rhabdosphaera clavigera</i>	<i>Umbilicosphaera hurburtiana</i>
<i>Rhabdosphaera xiphos</i>	<i>Umbilicosphaera sibogae</i>
	<i>Zygosphaera amoena</i>
<i>Solisphaera</i> spp.	Undescribed
<i>Scyphosphaera apsteinii</i>	heterococcolithophore A
<i>Sphaerocalyptra quadridentata</i>	Undescribed
	coccolithophore
<i>Sphaerocalyptra</i> sp. 1	Undescribed
<i>Sphaerocalyptra</i> sp. 3	holococcolithophore
<i>Sphaerocalyptra</i> sp.	
<i>Syracolithus ponticuliferus</i>	
<i>Syracolithus schilleri</i>	
<i>Syracolithus</i> sp. A	
<i>Syracosphaera ampliora</i>	
<i>Syracosphaera anthos</i>	
<i>Syracosphaera anthos</i> HOL	
<i>Syracosphaera bannockii</i>	
<i>Syracosphaera bannockii</i> HOL	
<i>Syracosphaera borealis</i>	
<i>Syracosphaera corolla</i>	
<i>Syracosphaera delicata</i>	
<i>Syracosphaera dilatata</i>	
<i>Syracosphaera epigrosa</i>	
<i>Syracosphaera exigua</i>	
<i>Syracosphaera florida</i>	
<i>Syracosphaera halldalii</i>	

Appendix A (continued)

Species name	Species name
<i>Syracosphaera histrica</i>	
<i>Syracosphaera lamina</i>	
<i>Syracosphaera marginaporata</i>	
<i>Syracosphaera molischii</i>	
<i>Syracosphaera nana</i>	
<i>Syracosphaera nana</i> HOL	
<i>Syracosphaera nodosa</i>	
<i>Syracosphaera noroitica</i>	
<i>Syracosphaera ossa</i>	

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