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Beetroot Juice versus Chard Gel: A Pharmacokinetic and Pharmacodynamic

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### Highlights

- When matched for nitrate content both beetroot juice and chard gels, known to
  be rich in nitrate, increased plasma nitrate and nitrite concentrations and
  reduced blood pressure to a similar extent.
  - Inter-individual variability to reach maximal plasma nitrite levels was considerable and should be taken into account when utilizing acute dietary nitrate supplementation.
    - Plasma concentrations of total nitrosated products were higher with beetroot juice than with chard gel despite comparable nitrate content.

#### **Abstract**

Dietary supplementation with inorganic nitrate (NO<sub>3</sub><sup>-</sup>) has been shown to induce a multitude of advantageous cardiovascular and metabolic responses during rest and exercise. While there is some suggestion that pharmacokinetics may differ depending on the NO<sub>3</sub><sup>-</sup> source ingested, to the best of our knowledge this has yet to be determined experimentally. Here, we compare the plasma pharmacokinetics of NO<sub>3</sub><sup>-</sup>, nitrite (NO<sub>2</sub><sup>-</sup>), and total nitroso species (RXNO) following oral ingestion of either NO<sub>3</sub><sup>-</sup> rich beetroot juice (BR) or chard gels (GEL) with the associated changes in blood pressure (BP). Repeated samples of venous blood and measurements of BP were collected from nine healthy human volunteers before and after ingestion of the supplements using a cross-over design. Plasma concentrations of RXNO and NO<sub>2</sub><sup>-</sup> were quantified using reductive gas-phase chemiluminescence and NO<sub>3</sub><sup>-</sup> using high pressure liquid ion chromatography. We report that, [NO<sub>3</sub><sup>-</sup>] and [NO<sub>2</sub><sup>-</sup>] were increased and systolic BP reduced to a similar extent in each experimental arm, with considerable inter-individual variation. Intriguingly, there was a greater increase in

[RXNO] following ingestion of BR in comparison to GEL, which may be a consequence of its higher polyphenol content. In conclusion, our data suggests that while differences in circulating  $NO_2^-$  and  $NO_3^-$  concentrations after oral administration of distinct  $NO_3^-$ -rich supplementation sources are moderate, concentrations of metabolic by-products may show greater-than-expected variability; the significance of the latter observation for the biological effects under study remains to be investigated.

Key Words: nitrite, nitric oxide, dietary supplementation, blood pressure

### 1. Introduction

Dietary nitrate ( $NO_3$ ) supplementation has been demonstrated to positively influence parameters of exercise performance (2, 25, 36) and vascular health (26, 27, 50, 54). These effects have been achieved utilizing a variety of different vehicles for  $NO_3$  delivery, including simple sodium (28) or potassium salts (23),  $NO_3$ -rich foods (44), concentrated beetroot juice (BR) (58), and chard gel (GEL) (37, 38). These studies have consistently shown that circulating plasma [ $NO_3$ ] and nitrite ([ $NO_2$ ]) concentrations are increased following ingestion of  $NO_3$  supplements. Whilst the biological consequences of dietary  $NO_3$  administration are not fully understood at present, it is known that  $NO_3$  can be reduced to  $NO_2$ , which is believed to be subsequently further converted to bioactive nitric oxide (NO) (1, 31). The enterosalivary circulation plays a vital role in NO homeostasis with ~25% of all circulating  $NO_3$  taken up by the salivary glands and concentrated in the saliva (51). The reduction of  $NO_3$  to  $NO_2$  takes place in the oral cavity where commensal facultative anaerobic bacteria on the surface of the tongue reduce  $NO_3$  to  $NO_2$  via  $NO_3$ 

reductase enzymes (12, 29). Once swallowed,  $NO_2$  reaches the stomach where a proportion is then converted to NO, with the remainder being absorbed into circulation via the intestinal tract (3, 32, 33).

It is well-established that increases in plasma [NO<sub>3</sub>] and [NO<sub>2</sub>] following dietary NO<sub>3</sub> supplementation occur in a dose-dependent manner (4, 19, 21, 23, 58, 59), however the influence of the vehicle, if any, is less certain. Several studies have reported that plasma [NO<sub>3</sub>] and [NO<sub>2</sub>] are reaches maximal quantities at ~ 1–1.5 h and 2.5–3h, respectively, after ingestion of BR (23, 35, 54, 58). Recent work from our laboratory has shown that consuming GEL results in similar plasma NO<sub>3</sub> pharmacokinetics but plasma [NO<sub>2</sub>] reaches maximal levels more quickly (~1.5 h) after ingestion (37). It is currently unclear whether the variance in NO<sub>2</sub> pharmacokinetics between BR and GEL is simply due to the vehicle of administration or profoundly influenced by inter-cohort differences in the response to NO<sub>3</sub> supplementation. Understanding if the vehicle of NO<sub>3</sub> supplementation affects the fate of NO-related metabolites may allow for the optimization of dosing strategies for sports performance and other contexts. Therefore, the purpose of this study was to compare the effects of ingesting BR and GEL on plasma NO metabolite pharmacokinetics and blood pressure (BP) pharmacodynamics in healthy individuals.

### 2. Methods

# 2.1 Participants

Nine healthy adult males (age  $28 \pm 4$  years, stature:  $181 \pm 8$  cm, body mass:  $83.4 \pm 101$  10.4 kg) volunteered to take part in the study, which was approved by the School of Science and Sport Ethics Committee of the University of the West of Scotland. All

participants provided written informed consent and a medical questionnaire before the study began. Healthy males between the ages of 18 and 45 who were physically active (taking part in recreational activity a minimum of 3 times per week) were eligible to participate in the study. Participants were excluded if they were currently taking dietary supplements or any medication, regularly used mouthwash, were smokers, had a current illness or virus within the previous month, had a known disorder or history of disorders of the hematopoietic system, were hypertensive (≥140/90 mmHg) or had a family history of premature cardiovascular disease. All procedures were conducted in accordance with the Declaration of Helsinki.

# 2.2 Experimental Design

Our study had a simple randomized cross-over design. Participants visited the laboratory on two separate occasions with a minimum 7-day washout period and a maximum of 14 days between visits. Participants consumed either concentrated BR (Beet It Organic Shot, James White Drinks, Ipswich, UK) or GEL (Science in Sport, GO+ Nitrates, Lancashire, UK) during each trial.

Participants were asked to refrain from the consumption of alcohol, caffeine, NO<sub>3</sub><sup>-</sup> rich foods as outlined by Hord and colleagues (22), and to avoid any strenuous exercise for 24 h before each trial. Participants were also asked to refrain from the use of anti-bacterial mouthwash and chewing gum for the duration of the study as they have been shown to disturb the oral bacterial flora required for the conversion of NO<sub>3</sub><sup>-</sup> to NO<sub>2</sub><sup>-</sup> in the saliva (17, 41). Compliance to these factors was determined at the start of each visit.

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Following a 12 h overnight fast, participants reported to the lab in the morning where they were asked to void the contents of their bladder and lie supine on a medical bed. After 15 min, BP was determined using an automated sphygmomanometer (Omron M10, Kyoto, Japan) three times, at 1 min intervals. A cannula was then inserted into the antecubital vein of the arm or a superficial vein on the dorsal surface of the hand and the line was kept patent by regular flushing with intravenous 0.9% saline solution. A sample of venous blood was then collected in a vacutainer containing EDTA and immediately centrifuged at 4000 rpm at 4°C for 10 min (Harrier 18/80, MSE, UK). The plasma was extracted carefully ensuring the cell layer was not disturbed and immediately frozen at -80°C for later analysis of plasma [NO<sub>3</sub>], [NO<sub>2</sub>], and total nitros ospecies [RXNO]. Participants then ingested either the BR or GEL supplements within 1 min of pre supplementation blood sampling. The GEL supplement comprised 120 ml of peach flavored sports gel containing 500 mg of NO<sub>3</sub> from natural chard and rhubarb sources. In the BR trial, participants ingested 117 ml of concentrated BR that also contained 500 mg of NO<sub>3</sub>. The NO<sub>3</sub> content of the supplements was later verified using high-pressure liquid ion chromatography (section 2.3).

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As outlined in Fig. 1 venous blood samples were collected simultaneously with measurements of BP pre-supplementation then at 1, 1.5, 2, 2.5, 3, 3.5 and 6 h postingestion of each supplement. The measurement of BP was carried out in triplicate, with the measurement being performed as close as possible to blood draw. The BP Cuff was placed on the opposite arm to the cannula. Participants remained supine

from the first blood sample until the 3.5 h sample, after which they were allowed to sit at a desk, returning 30 min before the final sample. During the experimental trials, participants were provided with standardized meals, which had a low NO<sub>3</sub><sup>-</sup> content. Specifically, participants consumed a cereal bar after 1.5 h and a cheese sandwich 3.5 h after ingestion of BR or GEL. Participants were provided with *ad libitum* access to tap water. The volume consumed in trial 1 was recorded and kept consistent for trial 2.

# 2.3 Additional Experimental Arm

The aforementioned procedures were conducted to address the primary objective of this experiment whereby doses of GEL and BR matched for NO<sub>3</sub><sup>-</sup> content were compared. Whereas the dose of GEL used in this experiment comprised two full gels as provided by the manufacturer (2 x 60g), 23 ml of BR was removed from one 70 ml bottle to ensure a matched NO<sub>3</sub><sup>-</sup> content. Given that both researchers and end-users are more likely to utilize the full 140 ml (e.g. (21, 58) the dose of BR used in this experiment was considered to be lacking in ecological validity. To this end, eight of the participants completed an additional experimental trial where they received 140 ml of BR (600 mg of NO<sub>3</sub><sup>-</sup>, H-BR) with the procedures repeated as previously described.

### 2.4 Analysis of Plasma NO Metabolites

High-pressure liquid ion chromatography was used to determine plasma  $[NO_3^-]$  and  $[NO_2^-]$ . Due to high variability in the  $NO_2^-$  measurements, which may relate to lack of specific sample processing without addition of N-ethylmaleimide prior to

centrifugation, the NO<sub>2</sub> data were re-analyzed using chemiluminescence and the latter was used in all calculations. Gas-phase chemiluminescence was used to determine plasma [RXNO]. Samples were thawed at room temperature in the presence of 5 mM N-ethylmale imide and subsequently analyzed using an automated NOx detection system (Eicom, ENO-20, Kyoto, Japan, combined with a Gilson auto-sampler for [NO<sub>3</sub>])(46) and a NO analyzer (Sievers NOA 280i, Analytix, UK for [NO<sub>2</sub>] and CLD 77AM sp, ECOphysicis, Durnten, Switzerland for [RXNO]) in conjunction with a custom-designed reaction chamber. NO<sub>2</sub> levels were determined using 1% potassium iodide in 5ml glacial acetic acid at room temperature for reduction of NO<sub>2</sub> to NO (42); RXNO levels were determined using the triiodide method (13). All samples were analyzed within 3 months of sample collection in order to minimize degradation of NO metabolites.

## 2.5 Data Analysis

All analyses were carried out using the Statistical Package for the Social Sciences, Version 22 (SPSS Inc., Chicago, IL, USA) or GraphPad Prism version 6 (GraphPad Software Inc., San Diego, USA) for kinetic analyses. For brevity, data from the additional H-BR trial are not displayed in figures. The sample size was determined *a priori* using a power calculation which revealed that a minimum of eight participants was required to detect differences in the time taken for NO<sub>2</sub> to peak between GEL and BR conditions. To establish the time to reach maximal [NO<sub>2</sub>] and [NO<sub>3</sub>] a log (Gaussian) non-linear regression model was applied to the data using the following equation:

 $Y=Amplitude*exp(-0.5*(ln(X/Center)/Width)^2).$ 

Data are expressed as the change in the mean ( $\Delta$ )  $\pm$  standard error of the mean (S.E.M) as compared to baseline or the mean and 95% confidence interval (CI) for time to reach maximal values. The distribution of the data was tested using the Shapiro-Wilk test. A two-way repeated-measures ANOVA was used to examine the differences between condition and over time for plasma NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup>, RXNO, and BP. *Post-hoc* analysis to determine the difference from the baseline was conducted using a paired samples t-tests with Bonferroni correction. Statistical significance was declared when P < 0.05.

# 3. Results and Discussion

Plasma [NO<sub>3</sub>] and [NO<sub>2</sub>] at baseline amounted to  $26 \pm 5.7 \,\mu\text{M} \,\text{NO}_3^-$ ,  $95 \pm 31.9 \,\text{nM}$  $NO_2$  for BR and 33  $\pm$  3.4  $\mu$ M  $NO_3$  and 25  $\pm$  6.7 nM  $NO_2$  for GEL. As expected, oral NO<sub>3</sub> supplementation significantly increased plasma [NO<sub>3</sub>] and [NO<sub>2</sub>] in each experimental arm (P < 0.001) ( $\Delta [NO_3]$  with BR: 319.4 ± 32.1  $\mu$ M, with GEL: 383.9  $\pm$  35.7 µM, Fig. 2;  $\Delta$  [NO<sub>2</sub>] with BR: 205.4  $\pm$  51.9 nM, with GEL: 207.4  $\pm$  58.1 nM, Fig. 3). The magnitude of the increase, however, was not different between BR and GEL (P > 0.10). In the H-BR arm,  $[NO_2]$  and  $[NO_3]$  increased to a greater extent than BR and GEL ( $\Delta$  [NO<sub>2</sub>] 277 ± 161 nM,  $\Delta$  [NO<sub>3</sub>] 457 ± 22  $\mu$ M, both P < 0.01). Following ingestion of BR, [NO<sub>2</sub>] reached maximal values at 3 h (95% CI 2.1 – 3.9 h), which was not different to GEL (2.8 h, 95% CI 2.3 - 3.2 h, P = 0.739). Likewise, the time taken for plasma [NO<sub>3</sub>] to reach maximal concentrations was not different between BR and GEL (BR: 1.4 h 95% CI 0.8 – 1.9 h, GEL: 1.4 h 95% CI 0.7 – 2.1 h, P = 0.737). In the H-BR arm,  $[NO_2]$  and  $[NO_3]$  reached maximal concentration in the plasma after 3.2 h (95%CI 2.1 – 4.2 h) and 1.5 h (95%CI 0.9 – 2.1 h), respectively. 

These data collectively suggest that the vehicle of delivery, be it liquid or gel, does not impact the kinetics of the reduction of  $NO_3^-$  to  $NO_2^-$  or the maximal plasma concentrations of these metabolites. Nevertheless, it remains to be established whether  $NO_3^-$  supplementation in solid forms, such as whole vegetables or concentrated BR flapjacks, results in different  $NO_x$  pharmacokinetics.

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In the present study, plasma [NO<sub>2</sub>] and [NO<sub>3</sub>] reached maximal quantities within a similar timeframe to previous research with BR (19, 29, 40, 43). However, on this occasion [NO<sub>2</sub>] took substantially longer after GEL (2.8 h) compared with our own previous work (1.5 h) (37). Given that descriptive and anthropometric variables were similar between the two study cohorts, it seems likely that physiological variations between individuals may account for these differences in time. Although plasma [NO<sub>2</sub>] is likely to be substantially elevated in most individuals 2.5 h after ingestion of either BR or GEL, the peak may reasonably occur anywhere between 2.1 and 3.9 h. To further highlight this Figure 4 displays the individual variability in the plasma NO<sub>2</sub> response to both vehicles of supplementation. Another important factor to acknowledge when comparing different studies is the methods of analysis for NO metabolites. The sensitivity of chemiluminescence and HPLC has been highlighted with factors such as sample preparation, type of analyzer used, and duration of sample storage, all potentially influencing the result acquired (8, 42). Whilst the precise mechanisms explaining the disparity in plasma [NO<sub>2</sub>] pharmacokinetics between these studies are unclear, we speculate that this may at least be partially explained by variances in the gut microbiota (14), pH of oral cavity and stomach (18, 43), and differences in the composition of the oral bacterial flora required for NO<sub>3</sub> reduction (11, 18). The importance of the oral microbiome for NO<sub>3</sub> reduction has been clearly

established, with the oral reductase capacity substantially interrupted when using antibacterial mouthwash (5, 41, 55) or spitting of saliva following NO<sub>3</sub> supplementation (30, 54). Equally, physical fitness has been suggested to affect the individual response to NO<sub>3</sub> supplementation (18). In contrast to the direct association between endothelial NO production (as measured by plasma  $NO_2$ ) and exercise performance (47, 53). Porcelli and colleagues (45) demonstrated that there was a negative association between aerobic capacity (VO<sub>2peak</sub>) and the increase in plasma [NO<sub>2</sub>] following ingestion of a NO<sub>3</sub> supplement. Although not measured in either the present study or our previous work on NO<sub>3</sub> pharmacokinetics (37), it is conceivable that individual differences in physical fitness, diet, or other lifestyle habits may contribute to the between-group variation reported here and elsewhere within the literature (18). Although it has not been thoroughly investigated, it is also conceivable that oral (and gut) microbial flora changes as a result of frequent NO<sub>3</sub> supplementation. It has been recently demonstrated following 2 weeks of NO<sub>3</sub> supplementation via BR there is an increase in salivary pH suggesting a role of NO<sub>3</sub> supplementation in altering composition of the oral microbiome (20).

Whilst the NO<sub>3</sub> and NO<sub>2</sub> responses were similar between experimental arms, an unexpected finding was that ingestion of BR tended to increase plasma [RXNO] to a greater extent in comparison to GEL ( $\Delta$  in BR: 408.1  $\pm$  127.9 nM vs.  $\Delta$  in GEL: 148.1  $\pm$  35.1 nM, P = 0.08, Fig. 5.). Plasma [RXNO] at baseline amounted to 79.5  $\pm$  13.1 nM for BR and 71.9  $\pm$  10.9 nM for GEL. There was, however, a high degree of variability in the change in [RXNO] between individuals and the small sample size likely explains why this finding was not statistically significant. The increase in [RXNO] was even greater in the H-BR trial ( $\Delta$ 563.8  $\pm$  116.7 nM) at 2 h post ingestion

than in GEL (P = 0.004) and BR (P = 0.03). Although plasma [RXNO] is not measured routinely in NO<sub>3</sub><sup>-</sup> supplementation studies, the magnitude by which [RXNO] increased following BR in the present study is greater than what has been previously reported [6]. Equally surprising was that the rise in [RXNO] exceeded that of [NO<sub>2</sub><sup>-</sup>] following ingestion of BR. The explanation for this is presently uncertain and while differences in supplementation regimen, NO<sub>3</sub><sup>-</sup> dose, and study participants may explain the disparity with previous research, further work is required to explore the changes in [RXNO] and [NO<sub>2</sub><sup>-</sup>] following ingestion of BR.

What is also unclear is why ingestion of BR increases [RXNO] to a greater extent (at least in the H-BR trial) compared to GEL. Although care was taken to match the supplements for total NO<sub>3</sub> content, differences in the polyphenol content between beetroot and chard may account for this outcome (24, 57). Furthermore, alongside the primary sources of NO<sub>3</sub> the BR supplement contained additional ingredients including lemon juice and the GEL contained rhubarb juice, gelling agents, preservatives, and flavorings. While the total antioxidant and polyphenol content of BR has been defined (56, 57) there is no comparable data on GEL. The total polyphenol content of each supplement may be important for overall NO bioavailability. Ingestion of flavonoid rich apples, for example, has been shown to increase [RXNO] in healthy adults (6), and nitrated polyphenols are formed from acidified NO<sub>2</sub> under simulated stomach conditions (40). Moreover, it has been shown that polyphenols augment the reduction of NO<sub>2</sub> to NO in the gut (48, 49). Given that S-nitrosothiols (RSNO), a component of RXNO, act as a carrier and store of NO in the blood, a polyphenol-induced increase in the bioavailability of NO may reasonably be exhibited by an increase in total nitroso products following BR. The importance of

the polyphenol content of NO<sub>3</sub> supplements and the role of RXNO in the translation to consequent physiological outcomes has yet to be established. However, the high polyphenol content of BR (56, 57), may explain the greater reduction in oxygen consumption following BR compared to sodium NO<sub>3</sub> (15). RXNOs are protected from direct NO scavenging by reactive oxygen species allowing NO to be transported by e.g. serum albumin and red blood cells (7, 52). This establishes an NO reservoir for the sustained release of NO from these biological storage forms (9, 16, 34). Potentially allowing for the targeted delivery of NO to where it is required such as sites of ischemia during exercise.

Systolic (SBP), diastolic (DBP), and mean arterial pressure (MAP) at baseline were as follows SBP:  $123 \pm 2$  mmHg, DBP:  $70 \pm 1$  mmHg, MAP:  $88 \pm 1$  mmHg for BR and SBP:  $124 \pm 2$  mmHg, DBP:  $73 \pm 2$  mmHg, MAP:  $90 \pm 2$  mmHg for GEL. In the present study, both BR and GEL reduced SBP and MAP ( $\Delta$  SBP with BR:  $-10 \pm 2$  mmHg, P < 0.001, vs. Baseline; with GEL:  $-12 \pm 2$  mmHg, P < 0.001;  $\Delta$  MAP with BR:  $-5 \pm 2$  mmHg, P = 0.012 vs Baseline; with GEL:  $-7 \pm 2$  mmHg, P = 0.010, Fig. 6). The magnitude of the reductions in SBP and MAP were not different between BR and GEL ( $P \ge 0.12$ ). Neither GEL nor BR significantly altered DBP (P = 0.18) nor was there any difference between experimental arms (P = 0.197). Likewise, SBP ( $\Delta - 11 \pm 2$  mmHg, P < 0.001) and MAP ( $\Delta - 8 \pm 3$  mmHg, P < 0.001) were reduced and DBP remained unchanged from baseline in the H-BR arm. It must be acknowledged that maintenance of the supine position for a prolonged period of time also likely contributed to a reduction in BP. Without a control condition, however, it is impossible to determine the extent of this effect. Nevertheless, these findings are consistent with previous literature demonstrating that ingestion of either BR or GEL

reduces SBP and MAP among healthy individuals (23, 37, 54, 58). The response in DBP appears to be more variable, however, although several previous studies have reported comparable data (2, 10, 23). Given the data presented here, it appears that the plasma [NO<sub>3</sub>] and [NO<sub>2</sub>] mirrors acute hemodynamic response to dietary NO<sub>3</sub> closely. Of notable interest, however, is that the changes in [RXNO] did not appear to be associated with the magnitude of the reduction in BP. This is in contrast to work by Oplander and colleagues (39) who demonstrated that reductions in BP were associated with an increased plasma availability of RXNO but not NO<sub>2</sub> following exposure of the skin to ultraviolet radiation. It is conceivable, therefore, that the method by which NO bioavailability is augmented will alter the mechanisms by which BP is reduced.

## 4. Conclusion

Our data suggests that dietary NO<sub>3</sub> supplementation via BR and GEL elicits similar plasma [NO<sub>2</sub>] and [NO<sub>3</sub>] pharmacokinetics when examined within the same participant cohort. Likewise, both BR and GEL are capable of reducing SBP and MAP with little difference in the magnitude of these effects. Nevertheless, we here present data demonstrating that the time course of ingesting the NO<sub>3</sub> supplements to maximal [NO<sub>2</sub>] in blood plasma is profoundly variable between individuals. This is of major relevance for researchers wishing to determine the same. We also report, for the first time, that ingesting BR leads to a greater availability of RXNO compared to GEL, which we speculate may be attributed to the higher polyphenol content of the BR supplement.

#### 347 **References**

- 348 1. Bailey JC, Feelisch M, Horowitz JD, Frenneaux MP, Madhani M.
  349 Pharmacology and therapeutic role of inorganic nitrite and nitrate in vasodilatation. *Pharmacol Ther* 2014:144(3):303–20.
- Bailey SJ, Winyard P, Vanhatalo A, et al. Dietary nitrate supplementation reduces the O2 cost of low-intensity exercise and enhances tolerance to high-intensity exercise in humans. *J Appl Physiol* 2009;107(4):1144–55.
- 354 3. Benjamin N, O'Driscoll F, Dougall H, et al. Stomach NO synthesis. *Nature* 1994;368(6471):502.
- 356 4. Bondonno CP, Croft KD, Puddey IB, et al. Nitrate causes a dose-dependent augmentation of nitric oxide status in healthy women. *Food Funct* 2012;3(5):522.
- 359 5. Bondonno CP, Liu AH, Croft KD, et al. Antibacterial mouthwash blunts oral nitrate reduction and increases blood pressure in treated hypertensive men and women. *Am J Hypertens* 2015;28(5):572–5.
- Bondonno CP, Yang X, Croft KD, et al. Flavonoid-rich apples and nitraterich spinach augment nitric oxide status and improve endothelial function in healthy men and women: A randomized controlled trial. *Free Radic Biol Med* 2012;52(1):95–102.
- 366 7. Bryan NS, Fernandez BO, Bauer SM, et al. Nitrite is a signaling molecule and regulator of gene expression in mammalian tissues. *Nat Chem Biol* 2005;1(5):290–7.
- Bryan NS, Grisham MB. Methods to detect nitric oxide and its metabolites in biological samples. *Free Radic Biol Med* 2007;43(5):645–57.
- Bryan NS, Rassaf T, Maloney RE, et al. Cellular targets and mechanisms of nitros(yl)ation: an insight into their nature and kinetics in vivo. *Proc Natl Acad Sci U S A* 2004;101(12):4308-13.
- 374 10. Coles LT, Clifton PM. Effect of beetroot juice on lowering blood pressure in free-living, disease-free adults: a randomized, placebo-controlled trial.
   376 Nutr J 2012;11(1):106.
- 377 11. Doel JJ, Benjamin N, Hector MP, Rogers M, Allaker RP. Evaluation of bacterial nitrate reduction in the human oral cavity. *Eur J Oral Sci* 2005;113(1):14–9.
- 380 12. Duncan C, Dougall H, Johnston P, et al. Chemical generation of nitric oxide 381 in the mouth from the enterosalivary circulation of dietary nitrate. *Nat* 382 *Med* 1995;1(6):546–51.
- 383 13. Feelisch M, Rassaf T, Mnaimneh S, et al. Concomitant S-, N-, and heme-384 nitros(yl)ation in biological tissues and fluids: implications for the fate of 385 NO in vivo. *FASEB J* 2002;16(13):1775–85.
- 386 14. Flint HJ, Scott KP, Louis P, Duncan SH. The role of the gut microbiota in

- nutrition and health. *Nat Rev Gastroenterol Hepatol* 2012;9(10):577–89.
- Flueck JL, Bogdanova A, Mettler S, Perret C. Is beetroot juice more effective than sodium nitrate? The effects of equimolar nitrate dosages of nitrate-rich beetroot juice and sodium nitrate on oxygen consumption during
- 391 exercise. *Appl Physiol Nutr Metab* 2016;41(4):421–9.
- 392 16. Ford PC, Wink DA, Stanbury DM. Autoxidation kinetics of aqueous nitric oxide. *FEBS Lett* 1993;326(1–3):1–3.
- 394 17. Govoni M, Jansson EA, Weitzberg E, Lundberg JO. The increase in plasma 395 nitrite after a dietary nitrate load is markedly attenuated by an 396 antibacterial mouthwash [Internet]. *Nitric Oxide* 2008;19(4):333–7.
- 397 18. Hezel MP, Weitzberg E. The oral microbiome and nitric oxide homoeostasis. *Oral Dis* 2015;21(1):7–16.
- Hobbs DA, Kaffa N, George TW, Methven L, Lovegrove JA. Blood pressurelowering effects of beetroot juice and novel beetroot-enriched breads in normotensive male subjects. *Br J Nutr* 2012;108(11):2066–74.
- 402 20. Hohensinn B, Haselgrübler R, Müller U, et al. Sustaining elevated levels of nitrite in the oral cavity through consumption of nitrate-rich beetroot juice in young healthy adults reduces salivary pH [Internet]. *Nitric Oxide* 2016; Ahead of Print
- Hoon MW, Jones AM, Johnson NA, et al. The effect of variable doses of inorganic nitrate-rich beetroot juice on simulated 2000-m rowing performance in trained athletes. *Int J Sports Physiol Perform* 2014;9(4):615–20.
- 410 22. Hord NG, Tang Y, Bryan NS. Food sources of nitrates and nitrites: the physiologic contact for potential health benefits. *Am J Clin Nutr* 2009;90(6):1–10.
- 413 23. Kapil V, Milsom AB, Okorie M, et al. Inorganic Nitrate Supplementation 414 Lowers Blood Pressure in Humans: Role for Nitrite-Derived NO. 415 *Hypertension* 2010;56(2):274–81.
- 416 24. Kazimierczak R, Hallmann E, Lipowski J, et al. Beetroot (Beta vulgaris L.)
  417 and naturally fermented beetroot juices from organic and conventional
  418 production: Metabolomics, antioxidant levels and anticancer activity. *J Sci*419 *Food Agric* 2014;94(13):2618–29.
- 420 25. Lansley KE, Winyard PG, Bailey SJ, et al. Acute dietary nitrate 421 supplementation improves cycling time trial performance. *Med Sci Sports* 422 *Exerc* 2011;43(6):1125–31.
- Lara J, Ashor AW, Oggioni C, Ahluwalia A, Mathers JC, Siervo M. Effects of inorganic nitrate and beetroot supplementation on endothelial function: a systematic review and meta-analysis. *Eur J Nutr* 2016;55(2):451–9.
- 426 27. Larsen FJ, Ekblom B, Sahlin K, Lundberg JO, Weitzberg E. Effects of Dietary
  427 Nitrate on Blood Pressure in Healthy Volunteers To the Editor: Nitric
  428 oxide, generated by nitric. N Engl J Med 2006;355(26):2792–3.

- 429 28. Larsen FJ, Weitzberg E, Lundberg JO, Ekblom B. Effects of dietary nitrate on oxygen cost during exercise. *Acta Physiol* 2007;191(1):59–66.
- 431 29. Li H, Duncan C, Townend J, et al. Nitrate-reducing bacteria on rat tongues. *Appl Environ Microbiol* 1997;63(3):924–30.
- 433 30. Lundberg JO, Govoni M. Inorganic nitrate is a possible source for systemic generation of nitric oxide. *Free Radic Biol Med* 2004;37(3):395–400.
- 435 31. Lundberg JO, Weitzberg E, Gladwin MT. The nitrate-nitrite oxide pathway in physiology and therapeutics. *Nat Rev Drug Discov* 2008;7(2):156–67.
- 438 32. Lundberg JO, Weitzberg E, Lundberg JM, Alving K. Intragastric nitric oxide 439 production in humans: measurements in expelled air. *Gut* 440 1994;35(11):1543–6.
- 441 33. McKnight GM, Smith LM, Drummond RS, Duncan CW, Golden M, Benjamin N. Chemical synthesis of nitric oxide in the stomach from dietary nitrate in humans. *Gut* 1997;40(2):211–4.
- 444 34. Miersch S, Mutus B. Protein S-nitrosation: Biochemistry and characterization of protein thiol-NO interactions as cellular signals. *Clin Biochem* 2005;38(9):777–91.
- 447 35. Miller GD, Marsh AP, Dove RW, et al. Plasma nitrate and nitrite are increased by a high-nitrate supplement but not by high-nitrate foods in older adults. *Nutr Res* 2012;32(3):160–8.
- 450 36. Muggeridge DJ, Howe CCF, Spendiff O, Pedlar C, James PE, Easton C. A 451 single dose of beetroot juice enhances cycling performance in simulated 452 altitude. *Med Sci Sports Exerc* 2014;46(1):143–50.
- 453 37. Muggeridge DJ, Sculthorpe N, Grace FM, et al. Acute whole body UVA 454 irradiation combined with nitrate ingestion enhances time trial 455 performance in trained cyclists. *Nitric Oxide - Biol Chem* 2015;48:3–9.
- 456 38. Muggeridge DJ, Sculthorpe N, James PE, Easton C. The effects of dietary 457 nitrate supplementation on the adaptations to sprint interval training in 458 previously untrained males. *J Sci Med Sport* 2016; Ahead of Print
- 39. Oplander C, Volkmar CM, Paunel-go A, et al. Whole Body UVA Irradiation
   Lowers Systemic Blood Pressure by Release of Nitric Oxide From
   Intracutaneous Photolabile Nitric Oxide Derivates. *Circ Res* 2009;105(10):1031-40.
- 463 40. Peri L, Pietraforte D, Scorza G, Napolitano A, Fogliano V, Minetti M. Apples 464 increase nitric oxide production by human saliva at the acidic pH of the 465 stomach: A new biological function for polyphenols with a catechol group? 466 Free Radic Biol Med 2005;39(5):668–81.
- 41. Petersson J, Carlström M, Schreiber O, et al. Gastroprotective and blood 468 pressure lowering effects of dietary nitrate are abolished by an antiseptic 469 mouthwash. *Free Radic Biol Med* 2009;46(8):1068–75.

- 470 42. Pinder AG, Rogers SC, Khalatbari A, Ingram TE, James PE. The
- 471 Measurement of Nitric Oxide and Its Metabolites in Biological Samples by
- 472 Ozone-Based Chemiluminescence. In: *Redox-Mediated Signal Transduction:*
- 473 *Methods and Protocols.* NJ: Humana Press; 2008 p. 11–28.
- 474 43. Pinheiro LC, Amaral JH, Ferreira GC, et al. Gastric S-nitrosothiol formation
- drives the antihypertensive effects of oral sodium nitrite and nitrate in a
- rat model of renovascular hypertension. Free Radic Biol Med 2015;87:252–
- 477 62.
- 478 44. Porcelli S, Pugliese L, Rejc E, et al. Effects of a Short-Term High-Nitrate Diet
- on Exercise Performance. *Nutrients* 2016;8(9):534. 5
- 480 45. Porcelli S, Ramaglia M, Bellistri G, et al. Aerobic Fitness Affects the Exercise
- Performance Responses to Nitrate Supplementation. *Med Sci Sports Exerc*
- 482 2014;47(8); 1643-1651.
- 483 46. Rassaf T, Bryan NS, Kelm M, Feelisch M. Concomitant presence of N-
- nitroso and S-nitroso proteins in human plasma. *Free Radic Biol Med*
- 485 2002;33(11):1590-6.
- 486 47. Rassaf T, Lauer T, Heiss C, et al. Nitric oxide synthase-derived plasma
- nitrite predicts exercise capacity. *Br J Sport Med* 2007;41(2):669–73;
- discussion 673.
- 489 48. Rocha BS, Gago B, Barbosa RM, Laranjinha J. Dietary polyphenols generate
- 490 nitric oxide from nitrite in the stomach and induce smooth muscle
- 491 relaxation. *Toxicology* 2009;265(1–2):41–8.
- 492 49. Rocha BS, Nunes C, Pereira C, Barbosa RM, Laranjinha J. A shortcut to
- 493 wide-ranging biological actions of dietary polyphenols: modulation of the
- 494 nitrate-nitrite oxide pathway in the gut. *Food Funct*
- 495 2014;5(8):1646-52.
- 496 50. Siervo M, Lara J. Inorganic nitrate and beetroot juice supplementation
- reduces blood pressure in adults: a systematic review and meta-analysis.
- 498 *The Journal of Nutrition* 2013;143(6):818–26.
- 499 51. Spiegelhalder B, Eisenbrand G, Preussmann R. Influence of dietary nitrate
- on nitrite content of human saliva: Possible relevance to in vivo formation
- of N-nitroso compounds. *Food Cosmet Toxicol* 1976;14(6):545–8.
- 502 52. Stamler JS, Jaraki O, Osborne J, et al. Nitric oxide circulates in mammalian
- plasma primarily as an S-nitroso adduct of serum albumin. *Proc Natl Acad*
- *Sci U S A* 1992;89(16):7674–7.
- 505 53. Totzeck M, Hendgen-Cotta UB, Rammos C, et al. Higher endogenous nitrite
- levels are associated with superior exercise capacity in highly trained
- 507 athletes. *Nitric Oxide Biol Chem* 2012;27(2):75–81.
- 508 54. Webb AJ, Patel N, Loukogeorgakis S, et al. Acute blood pressure lowering,
- vasoprotective, and antiplatelet properties of dietary nitrate via
- bioconversion to nitrite. *Hypertension* 2008;51(3):784–90.
- 55. Woessner M, Smoliga JM, Tarzia B, Stabler T, Van Bruggen M, Allen JD. A

512 513 514		of mouthwash following a dietary nitrate load. <i>Nitric Oxide</i> 2016;54(16):1-7.
515 516 517 518 519	56.	Wootton-Beard PC, Moran A, Ryan L. Stability of the total antioxidant capacity and total polyphenol content of 23 commercially available vegetable juices before and after in vitro digestion measured by FRAP, DPPH, ABTS and Folin-Ciocalteu methods. <i>Food Res Int</i> 2011;44(1):217–24.
520 521 522	57.	Wootton-Beard PC, Ryan L. A beetroot juice shot is a significant and convenient source of bioaccessible antioxidants. <i>J Funct Foods</i> 2011;3(4):329–34.
523 524 525	58.	Wylie LJ, Kelly J, Bailey SJ, et al. Beetroot juice and exercise: pharmacodynamic and dose-response relationships. <i>J Appl Physiol</i> 2013;115(3):325–36.
526 527 528	59.	Wylie LJ, Ortiz de Zevallos J, Isidore T, et al. Dose-dependent effects of dietary nitrate on the oxygen cost of moderate-intensity exercise: Acute vs chronic supplementation. <i>Nitric Oxide</i> 2016;
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533 **Figure Captions** 534 535 Figure 1: Study overview: time-points for beetroot juice/chard gel administration, 536 venous blood sampling, blood pressure measurements and food intake. 537 Figure 2: Changes in plasma nitrate concentrations following supplementation with BR and GEL ( $\Delta$  Mean  $\pm$  S.E.M). \* Significant difference from baseline (pre-538 539 supplementation) (P < 0.001). 540 Figure 3: Changes in plasma nitrite concentrations following supplementation with BR and GEL ( $\Delta$  Mean  $\pm$  S.E.M). \* Significant difference from baseline (pre-541 542 supplementation) 543 Figure 4: Individual plasma nitrite pharmacokinetics and Systolic BP for BR and GEL. Each participant is represented by the same different colour in each figure. 544 545 Figure 5: Changes in total nitroso species concentrations following supplementation 546 with BR and GEL ( $\Delta$  Mean  $\pm$  S.E.M). \* Significant difference from baseline (pre-547 supplementation) Figure 6: Systolic (A), diastolic (B) and mean arterial pressure (C) changes following 548 549 supplementation with BR and GEL (Δ Mean ± S.E.M). \* Significant difference from 550 baseline (pre-supplementation)

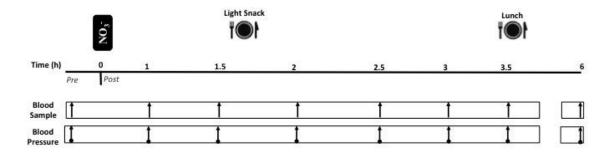
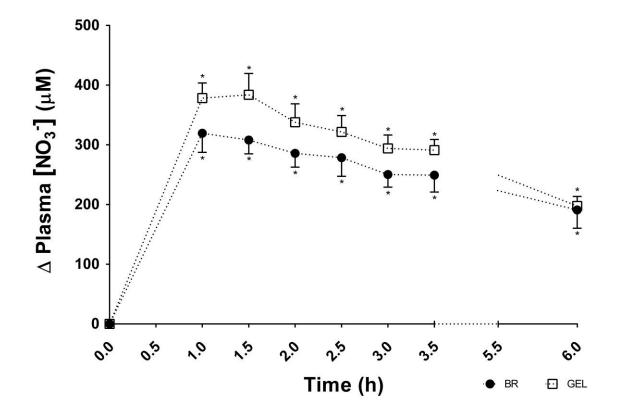


Fig. 2



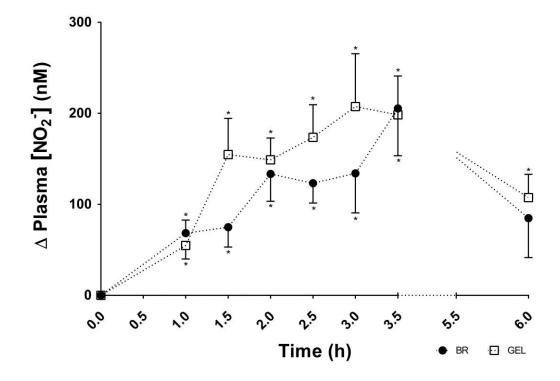


Fig. 4

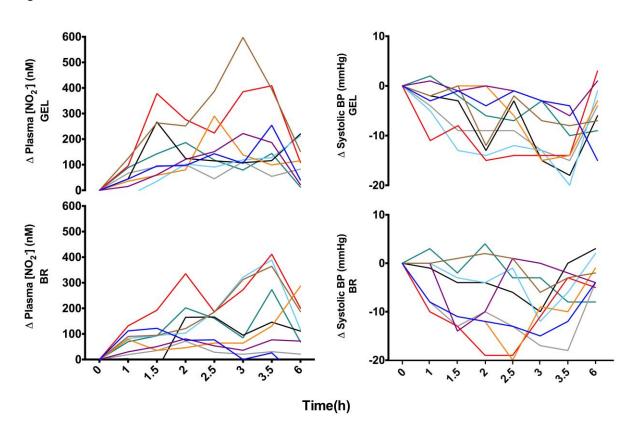


Fig. 5

