**PFTs affect drought response in peatland**

**Plant functional types define magnitude of drought response in peatland CO2 exchange**

**Jan J Kuipera,b, Wolf M Mooija,c, Luca Bragazzad,e,f, Bjorn JM Robroekb,d,e\***

*aDepartment of Aquatic Ecology, Netherlands Institute of Ecology (NIOO-KNAW), Wageningen, The Netherlands; bEcology and Biodiversity Group, Institute of Environmental Biology, Utrecht University, Utrecht, The Netherlands; cAquatic Ecology and Water Quality Management Group, Wageningen University, Wageningen, The Netherlands; dÉcole Polytechnique Fédérale de Lausanne (EPFL), School of Architecture, Civil and Environmental Engineering (ENAC), Laboratory of Ecological Systems (ECOS), station 2, CH-1015 Lausanne, Switzerland; eWSL - Swiss Federal Institute for Forest, Snow and Landscape Research, Research Unit Community Ecology, Site Lausanne, station 2, CH-1015 Lausanne, Switzerland; fUniversity of Ferrara, Department of Life Science and Biotechnologies, Corso Ercole I d’Este 32, Ferrara 44121, Italy.*

\* Corresponding author: b.j.m.robroek@uu.nl

*Abstract.* Peatlands are important sinks for atmospheric carbon (C), yet the role of plant functional types (PFTs) for C sequestration under climatic perturbations is still unclear. A plant removal experiment was used to study the importance of vascular PFTs for the net ecosystem CO2 exchange (NEE) during (i.e. resistance) and after (i.e. recovery) an experimental drought. The removal of PFTs caused a decrease of NEE, but the rate differed between microhabitats (i.e. hummocks and lawns) and the type of PFTs. Ericoid removal had a large effect on NEE in hummocks, while in the lawns the graminoids played a major role. The removal of PFTs did not affect the resistance or the recovery after the experimental drought. We argue that the response of *Sphagnum* mosses (the only PFT present in all treatments) to drought is dominant over that of coexisting PFTs. However, we observed that the moment in time in which the system switched from C sink to C source during the drought was controlled by the vascular PFTs. In the light of climate change, the shifts in species composition or even the loss of certain PFTs are expected to strongly affect the future C dynamics in response to environmental stress.

*Key words:* *drought; net ecosystem CO2 exchange; peatlands; plant functional types; recovery; resistance; Sphagnum*

INTRODUCTION

The extensive amount of organic carbon (C) stored in northern peatlands has been shown to be highly vulnerable to increasing soil temperature (Freeman et al. 2001a, Piao et al. 2008, Dorrepaal et al. 2009) and decreasing soil moisture (Freeman et al. 2001b, Fenner and Freeman 2011). In particular, prolonged periods of drought can significantly impact the structure and the functioning of peatlands (Weltzin et al. 2003, Bragazza 2008) which then can (temporarily) shift from C sinks to C sources (Lafleur et al. 2003, Lund et al. 2012). The adverse effects of drought on the peatland C balance may, however, strongly depend on plant community composition (Johnson et al. 2011), and the co-occurrence of species or plant functional types may help maintaining ecosystem processes in response to disturbance (Mulder et al. 2001, Craine et al. 2012).

The role of plant species richness and diversity for the stability of ecosystems in response to environmental changes is heavily debated (Loreau and de Mazancourt 2013). Generally, high species diversity and high diversity in traits promotes non-additive effects on ecosystem services due to complementarity, facilitation and competition between species or species groups (Hooper et al. 2005, Dias et al. 2013). More evidence is provided that species richness promotes the stability of ecosystem processes (Hector et al. 2010, Isbell et al. 2011, Bloor and Bardgett 2012). Alternatively, the stability of ecosystems is suggested to be mainly driven by the most dominant species, or species group, and to be proportionate to the primary production of these species (Grime 1998).

Mosses of the genus *Sphagnum*, generally dominate ombrotrophic peatlands, or bogs, and their decay-resistant litter makes the bulk of the stored peat (Hájek et al. 2011). Due to their ability to modify their environment, *Sphagnum* mosses are considered typical *ecosystem engineers* (Rochefort 2000). Consequently, *Sphagnum* mosses reduce the potential success of competing plant species (van Breemen 1995, Rydin and Jeglum 2006). Additionally, through their high polyphenol content *Sphagnum* mosses inhibit microbial activity by which they control decomposition processes and thus the export of C from peatlands (Bragazza and Freeman 2007). Vascular plant species, with different functional traits, are typical inhabitants of bogs (Chapin et al. 1996, Dorrepaal et al. 2007), where plant communities are reported to be remarkably stable in terms of species composition for long periods (Backéus 1972, Rydin and Barber 2001). Studies at species level (Robroek et al. 2007, Otieno et al. 2009, Robroek et al. 2009), community level (Alm et al. 1999, Bubier et al. 2003, Laine et al. 2007, Ward et al. 2009) and ecosystem level (Deppe et al. 2010) revealed that biota play a decisive role in controlling peatland C dynamics. The apparent protective role of vascular plant species on moss growth and C uptake has previously been described. Photo inhibition of photosynthesis, for example, under high irradiance has been reported to decrease under shading due to vascular plant cover (Murray et al. 1989, 1993), and shading by vascular plants may additionally decrease moss evaporation and protect the moss canopy from drying (Heijmans et al. 2004, McNeil and Waddingon 2003). Few studies, however, have explicitly studied the role and function of different plant functional groups on peatland C uptake and its robustness during summer drought.

In this study, we aim to elucidate the role of plant functional types in affecting C fluxes in a peatland during a drought event. Specifically, we performed a plant removal experiment in order to explore how two dominant plant functional types, i.e., ericoid dwarf-shrubs and graminoids, can affect the resistance and the recovery of CO2 uptake during and after a drought event.

MATERIALS AND METHODS

*Field sampling and experimental set-up*

In November 2009, thirty-two intact peat cores (diameter 22 cm, depth 16 cm) were collected from Tofte mose, Lille Vildmose Natural Park, Denmark (56°50’N, 10°15’E). Half of the cores were taken from wet lawns, and half were taken from relatively dry hummocks in order to include the two dominant microhabitats. All cores were representative for the site and encompassed equal (i.e., within natural variation) amounts of three plant functional types (PFTs): *Sphagnum* mosses, graminoids and evergreen ericoid shrubs (Chapin et al. 1996). In the lawns, *Sphagnum cuspidatum* Ehrh. Ex Hoffm. was the dominant moss species, while the vascular plant community consisted of the graminoids *Eriophorum angustifolium* Honck. and *Rhynchospora alba* (L.), and the ericoids *Erica tetralix* L.*, Vaccinium oxycoccus* L. and *Andromeda polifolia* L. In the hummocks, *Sphagnum magellanicum* Brid. and *S. rubellum* Wils. dominate the peat moss layer, while the vascular plant community consisted of the graminoid *Eriophorum vaginatum* L., and the ericoids *E. tetralix, V. oxycoccus, A. polifolia, Empetrum nigrum* L. and *Calluna vulgaris* (L.). The vascular plant cover ranged between 25-50% in both microhabitats.

After extraction, the cores were carefully placed in plastic containers (hereafter referred to as mesocosms), transported to Utrecht University, The Netherlands, and kept in a growth room (20/18 °C [12/12 h; day/night], 70% RH, 400 ppm CO2, 200 µmol m-2 s-1 light intensity) for seven weeks to acclimate (*acclimation period*). Mesocosms were watered twice a week, using artificial rainwater (Garrels and Christ 1965), which was sprinkled over the vegetation to mimic an average precipitation of approximately 2 mm day-1 (730 mm year-1; *cf.* the region of Lille Vildmose NP; www.dmi.dk). Additionally, water levels were kept constant at 2 cm and 10 cm below the *Sphagnum* capitula for lawns and hummocks, respectively, by funneling artificial rainwater to the mesocosms. One mesocosm became visibly infected with fungi and was removed from the experiment, resulting in 31 mesocosms (16 hummocks, 15 lawns).

After the *acclimation period*, vascular plant functional type (PFT) composition was manipulated by removing ericoids (Erem), graminoids (Grem), or both the PFTs so to leave a pure *Sphagnum* stand (n = 4 for each treatments and microhabitat). Removal was realized by clipping the above ground biomass flush to the moss layer(McLellan et al. 1995). An extra set of mesocosms was used a control treatment, but in order to take into account the potential effects of the clipping procedure, about 20% of the ericoid and graminoid cover was removed from the control mesocosms (Table 1). The period between PFT removal and the drought event lasted four weeks (*pre-drought period*), and was used to recover from the removal procedure. The second experimental period entailed the actual drought event (*drought period*), during which the mesocosms were allowed to dry out ‘naturally’. After four weeks of drought, all mesocosms were rewetted and allowed to recover (*recovery period*). Rainwater was added to completely submerge the mesocosms over a one-night period, after which the water tables were brought back to their initial levels and the original pre-drought watering regime was resumed. The process of recovery was monitored for ten weeks. Throughout all the experimental periods, the treatments were maintained by regular removal of regrown biomass.

*Carbon exchange measurements*

We used net ecosystem CO2 exchange (NEE) measurements to approximate net ecosystem productivity. Ecosystem respiration (Re; autotrophic and heterotrophic) was measured by darkening the chambers. Closed flux chambers (diameter 20 cm, height 29 cm, fitted with a circulating fan) were placed over the mesocosms to measure CO2 fluxes using an Innova Bruel Kjær BK 1302 photoacoustic multi gas analyzer (Bruel and Kjær, type 1302, Denmark), connected to a multipoint sampler (CBISS MK2, 4-channel, CBISS Ltd. England). During chamber closure, increment in air pressure in the chamber headspace was avoided by a 10 mm opening at the top of the plastic container, which was closed before any records were taken. The chambers made no contact with the vegetation wherefore disturbance of pressure gradients across the soil-atmosphere interface was avoided. Chamber measurements comprised five succeeding sampling points with an 8-minute interval. NEE was measured weekly during the *pre-drought period* and every two days during the *drought period*. During the *recovery period*, measurements were initially performed every two days, but intensity was reduced at a later stage. Re was measured just before and after plant removal, and throughout the *drought period* and *recovery period*. The ecological sign convention was used for the NEE and Re data, so that positive fluxes indicate CO2 uptake and negative fluxes indicate CO2 loss to the atmosphere.

*Calculations and Statistical Analyses*

NEE was calculated from the change in CO2 concentration in the chamber headspace with time, using an exponential non-linear function as proposed by Kutzbach et al. (2007). We used the nls function in the *stats* package in R to fit the models to the first four CO2 readings of the individual time series. In general, the coefficients of determination of the fitted models were very convenient (91% of the regressions showing an r2 > 0.995, n=940). The net CO2 flux was then calculated for the time zero, representing the flux at the start of the enclosure (see Appendix A for specifics). Re was determined by a linear regression over the five succeeding sampling points after darkening the chambers.

The resistance to drought is the ability of the system to reduce the decrease of NEE throughout the *drought period*. Here, the resistance was calculated, for each individual mesocosm, as the slope of the linear regression through the NEE data-points over the drought period. These regression parameters were also used to calculate the time when NEE switched from positive to negative value (i.e., NEE = 0) for each individual mesocosm. Similar to our measure of resistance, the recovery was calculated using the slope through the NEE data-points over the *recovery period*. Since NEE values decreased abruptly immediately after the recovery (data not shown), the use of a linear regression would not be appropriate. To circumvent this problem, recovery was calculated using the data collected one week after the start of the *recovery period*. Resilience was calculated as the ratio of post-recovery to pre-drought NEE. As such, resilience combines resistance and recovery and provides a measure of the extent to which the NEE has recovered to the pre-drought level.

To test the influence of our treatments on net ecosystem exchange (NEE) and ecosystem respiration (Re), we used the generalized linear models (GLM) assuming a Gaussian distribution of the data. We coded microhabitat (MH), ericoid removal (Erem) and graminoid removal (Grem) as binary factors and initially we tested the treatment effect on NEE and Re at three crucial moments during the experiment, i.e., just before the initiation of the *drought period* (Pre-drought), at the end of the *drought period* (Post-drought), and at the end of the *recovery period* (Post-rewetting). Similarly, we tested the treatment effect on the resistance, recovery and resilience of the net ecosystem exchange.

As the amount of removed biomass can be considered as an explanatory factor, we initially examined two models. The first model contained the factors MH, Erem, and Grem, while the second model considered the amount of ‘biomass removed’ as co-variable. The goodness-of-fit in relation to the model complexity was evaluated by comparing values of the corrected Akaike information criterion (AICc), using the selMod function in *pgirmess* package in R (Giraudoux 2013). The model with the smallest AICc was selected, and further analyses were performed using the ANOVA function and the F-statistic in the *stats* package of R. The *lme* package in R (Pinheiro et al. 2011) was used to perform repeated measures-ANOVA to determine the effects of Erem, Grem and MH on NEE through the drought period and the recovery period. As microhabitats differed significantly in their CO2 exchange dynamics (significant MH effects, or MH × Erem and MH × Grem interactions, see also Appendix C and D), these analyses were also performed separately for the two microhabitats. All analyses were performed with the software R 2.15.2 (R Core Team 2012).

RESULTS

*Pre-drought period: the effect of vascular plant functional type removal*

To test the potential role of the amount of removed biomass in affecting net ecosystem exchange (NEE) and ecosystem respiration (Re), we examined the explanatory power of a set of models with different complexities (see Appendix B). The amount of removed biomass was rather important for the change in NEE and Re before and after plant functional type (PFT) removal. Therefore, the co-variable ‘biomass removed’ was always taking into the potential models when testing for the effects on PFT removal in further analyses.

Plant functional type (PFT) removal caused a decrease in pre-drought NEE, which differed between the microhabitats in relation to the PFTs (Fig. 1A,B; Appendix C). Indeed, in the lawns, graminoid removal reduced NEE (F1, 12 = 16.9*,* *p* ≤ 0.01). Ericoids removal caused a reduction of NEE only in combination with graminoid removal (Ericoid × Graminoid (F1, 11 = 3.5*,* *p* = 0.09). In the hummocks, ericoid removal resulted in a reduction of NEE (F1, 14 = 27.3*,* *p* ≤ 0.001), whereas graminoid removal did not affect the NEE. Including removed biomass in these GLM model did not enhance the explanatory power, i.e. higher AICc (Appendix C).

*Drought period: the resistance to drought*

Net ecosystem exchange (NEE) and ecosystem respiration (Re) decreased immediately after the start of the experimental drought in all the mesocosms (Fig. 1 and 2). The resistance to drought differed between microhabitats (Fig 3A,B, Table 2). The decrease of NEE was faster in the lawns, resulting in lower post-drought NEE (Fig. 1; Appendix C,D). In fact, lawns switched from CO2 sinks (NEE > 0) to CO2 sources (NEE < 0) earlier than hummocks (Fig. 1 and. 3C,D, Table 2). The decrease of Re was faster in the hummocks (Fig. 2; Appendix D).

PFT removal did not significantly affect the resistance in both microhabitats (Table 2), though in the lawn microhabitats graminoid removal seemed to slightly slow down the decrease in NEE (F1, 12 = 3.4, *p* = 0.09), and though not significant the removal of ericoids seemed to slightly speed up the decrease of NEE. Nevertheless, PFT removal affected the moment of the switch from CO2 sink to CO2 source in the two microhabitats (Fig. 3C,D, Table 2). When analyzed separately it appears that in the lawn microhabitats, graminoid removal caused an earlier switch of the ecosystem from CO2 sink to CO2 source (F1, 12 = 14.4*,* *p* ≤ 0.01), while in the hummock microhabitats ericoid removal enhanced such switch, although not significantly (F1, 14 = 3.9*,* *p* = 0.07).

*Recovery after, and resilience to drought*

Rewetting caused a sudden increase in CO­­2 emission in all the mesocosms (data not shown). After this initial ‘desaturation respiration’ (Gerdol et al. 1996), CO2 uptake recommenced relatively fast. On the whole, recovery was not influenced by the PFTs, although mesocosms without ericoids seemed to recover faster (Fig. 3E,F, Table 2), especially in the lawn microhabitats. Ecosystem resilience differed between microhabitats (Table 2), and was higher in the hummocks (45 ± 6.3%) than in the lawns (25 ± 4.6%). In both microhabitats, the PFT removal treatment did not affect the resilience of NEE (Table 2).

DISCUSSION

Using a plant removal experiment, we tested the role of vascular plant functional types on short-term CO2 fluxes in peatland in response to drought. Removal experiments are useful to study the relation between plant functional types and ecosystem processes (Dıaz et al. 2003), although we acknowledge they have their limitations (Bret-Harte et al. 2004, Ward et al. 2009, Gundale et al. 2010). Remaining plant roots, for example, may increase ecosystem respiration (Re), and concomitantly decrease net ecosystem exchange (NEE). We, however, observed decreased Re after vascular PFT removal, confirming the contribution of vascular plant biomass to ecosystem respiration in peatlands (Bubier et al. 2003). Interestingly the decrease in NEE upon PFT removal was trait dependent and different between lawns and hummocks. Indeed, decreased NEEin hummocks was mainly caused by the removal of ericoids, whereas in lawns it was mainly caused by graminoids removal (Fig. 1). The decrease in NEE due to plant removal can partly be explained by the amount of removed biomass (Appendix B), yet the effects of PFT removal and biomass removal are highly collinear, i.e. hummocks are dominated by ericoids, while lawns are dominated by graminoids. Contrasting to our results, Ward et al. (2009) did not find any change in NEE after ericoid or graminoid removal from a blanket bog, due to an increase of both the gross C uptake and respiration rate. We argue that the removal of the high cover (about 70%) of vascular plants in their experiment may have stimulated the photosynthetic rates of the underlying moss layer. This apparent “competitive release effect” (Wardle et al. 1999, Symstad and Tilman 2001) did not take place in our experiment where the initial cover of vascular plants was much lower (25-50%). Concurrently, negative effects of vascular plant cover on underlying mosses or soil processes have been reported to occur mainly at dense vascular plant cover (Chapin and Shaver 1985, Heijmans et al. 2002, Blok et al. 2011).

Our findings that NEE decreased with plant removal indicate an important role of vascular plants and their corresponding traits in affecting NEE (McNeil and Waddington 2003), yet the overall aim of this study was to elucidate the role of PFTs on the resistance of NEE to, and its recovery after an extreme summer drought. The decrease in NEE during the drought period in all treatments, indicates an intrinsic sensitivity of peatlands to drought (Cai et al. 2010, Fenner and Freeman 2011, Lund et al. 2012). Lawns were less resistant to drought than hummocks (Fig. 3), supporting earlier findings of greater resistance of hummock *Sphagnum* species to desiccation (Andrus 1986, Strack and Price 2009). While vascular plant species seem to be important in controlling levels of NEE, they seem not to play an important role in mitigating the negative effect of drought on NEE. This is surprising, as vascular plants can avoid desiccation during drought by actively regulating their water content through morphological adaptations (roots, stomata, vascular system). *Sphagnum* mosses are poikilohdyric and cannot actively control their water contents. They can only tolerate drought through physiological responses, like quenching its basal chlorophyll fluorescence (Hajek and Beckett 2008, Turetsky et al. 2012). Based on the absence of an effect of PFTs on the resistance to drought (Table 2), we reason that the decrease of NEE is primarily controlled by the drought-induced decrease of *Sphagnum* photosynthetic assimilation. Indeed, most mesocosms were visible desiccated toward the end of the drought period (Robroek et al. 2009). Would we, however, approach resistance as the ecosystems capacity to maintain positive CO2 uptake during drought (i.e. time to NEE ≤ 0), the presence of vascular plants becomes rather important (Fig. 3). According to previous hypotheses from grassland studies (Wang et al. 2007, van Ruijven and Berendse 2009), the initial productivity (or NEE) and not the plant functional type diversity *per se* then determines the resistance of the peatland to environmental perturbations.

The observed recovery after drought resembles that already presented by Robroek et al. (2009). They, however, report data from pure *Sphagnum* stands. Interestingly, *Sphagnum* mosses are generally described as a drought intolerant genus, being very sensitive to (repeated) desiccation (Gerdol et al. 1996, Schipperges and Rydin 1998) and slow in their recovery after drought (Gerdol et al. 1996, Bragazza 2008). The faster recovery observed in hummocks indicates a greater tolerance to desiccation of hummock *Sphagnum* mosses (Hájek and Beckett 2008). Our results, however, show that after a ten-week recovery period, CO2 uptake returned only to <45% of the pre-drought perturbation levels. Although our results indicate a relatively swift recovery of C uptake after a drought, these results also indicate that a single drought can have a substantial impact on the annual CO2 budget (Alm et al. 1999). In fact, any cessation in carbon uptake can have a strong effect on the annual C budget in peatlands (McNeil and Waddington 2003). The size of this effect depends on the timing, the severity and the duration of the drought (Lund et al. 2012).

The apparent absence of a crucial role for PFT composition on the resistance to, and recovery after, a drought in peatlands, together with the response of net photosynthesis of monospecific *Sphagnum* stands to drought (Robroek et al. 2009, Adkinson and Humphreys 2011), indicate a dominant mechanistic role for the *Sphagnum* community. Raised bogs have a rather low diversity and are characterized by the presence of a clear ecosystem engineer, which may have a disproportionate influence on ecosystem processes (Brown 1995, Polley et al. 2007). Consequently, *Sphagnum* mosses are largely controlling the ecosystem C cycling. Such a strong effect displayed by a single group, has been earlier reported as a possible explanation for the absence of diversity–ecosystem functioning relationships (Hooper et al. 2005). As in peatlands, *Sphagnum* mosses have the largest share of the aboveground green biomass in all plant communities (Laine et al. 2012), our results support Grime’s biomass ratio hypothesis(Grime 1998), which states that the effect of each species or plant functional group is proportional to its relative biomass in the ecosystem.

Changes in short-term CO2 fluxes due to drought are of relevance for the long-term C budget of peatlands (Ward et al. 2009), especially if extreme drought events are expected to occur more often (Dai 2012). Our study shows that vascular PFTs do not affect the resistance of the peatland ecosystem to, nor the recovery after an experimental drought. Nevertheless, vascular PFTs are important in controlling the net ecosystem productivity, and thereby the moment the systems can switch from C sink to C source during a drought event. As evidenced by our data, in both lawns and hummocks, the presence of vascular plants secures the C sink function during drought events. In the light of climate change, shifts in the species community composition or even the loss of certain PFTs can thus have strong effects on future C dynamics in response to environmental stress.

ACKNOWLEDGMENTS

The Lille Vildmose area is owned and conserved by the Aage V. Jenssen Foundation (Denmark), and we appreciate their permission, arranged through Jacob Pålsgaard, to enter the reserve and collect samples. This paper greatly improved after the comments of two anonymous referees. We are also grateful to Alexandre Buttler, Mariet Hefting, Constant Signarbieux and Jasper van Ruijven for many valuable suggestions on the presentation of our results. The Division for Earth and Life Sciences (ALW) with financial aid from the Netherlands Organization for Scientific Research (NWO; Research Innovation Scheme grant nr. 863.10.014) supports this work.

LITERATURE CITED

Adkinson, A. C., and E. R. Humphreys. 2011. The response of carbon dioxide exchange to manipulations of *Sphagnum* water content in an ombrotrophic bog. Ecohydrology 4:733–743.

Alm, J., L. Schulman, J. Walden, H. Nykänen, P. J. Martikainen, and J. Silvola. 1999. Carbon balance of a boreal bog during a year with an exceptionally dry summer. Ecology 80:161–174.

Andrus, R. E. 1986. Some aspects of *Sphagnum* ecology. Canadian Journal of Botany 64:416–426.

Backéus, I. 1972. Bog vegetation re-mapped after sixty years, studies on Skagershultamossen, central Sweden. Oikos:384–393.

Blok, D., M. M. P. D. Heijmans, G. Schaepman-Strub, J. Ruijven, F. J. W. Parmentier, T. C. Maximov, and F. Berendse. 2011. The cooling capacity of mosses: controls on water and energy fluxes in a Siberian tundra site. Ecosystems 14:1055–1065.

Bloor, J. M. G., and R. D. Bardgett. 2012. Stability of above-ground and below-ground processes to extreme drought in model grassland ecosystems: Interactions with plant species diversity and soil nitrogen availability. Perspectives in Plant Ecology, Evolution and Systematics 14:193–204.

Bragazza, L. 2008. A climatic threshold triggers the die‐off of peat mosses during an extreme heat wave. Global Change Biology 14:2688–2695.

Bragazza, L., and C. Freeman. 2007. High nitrogen availability reduces polyphenol content in *Sphagnum* peat. The Science of the Total Environment 377:439–443.

Bret-Harte, M. S., E. A. García, V. M. Sacré, J. R. Whorley, J. L. Wagner, S. C. Lippert, and F. S. Chapin III. 2004. Plant and soil responses to neighbour removal and fertilization in Alaskan tussock tundra. Journal of Ecology 92:635–647.

Brown, J. H. 1995. Organisms as engineers: a useful framework for studying effects on ecosystems? Trends in Ecology and Evolution 10:51–52.

Bubier, J., P. Crill, A. Mosedale, S. Frolking, and E. Linder. 2003. Peatland responses to varying interannual moisture conditions as measured by automatic CO2 chambers. Global Biogeochemical Cycles 17:1066.

Cai, T., L. B. Flanagan, and K. H. Syed. 2010. Warmer and drier conditions stimulate respiration more than photosynthesis in a boreal peatland ecosystem: analysis of automatic chambers and eddy covariance measurements. Plant, Cell and Environment 33:394–407.

Chapin, F. S., III, and G. R. Shaver. 1985. Individualistic growth response of tundra plant species to environmental manipulations in the field. Ecology 66:564–576.

Chapin, F. S., III, M. S. Bret Harte, S. E. Hobbie, and H. Zhong. 1996. Plant functional types as predictors of transient responses of arctic vegetation to global change. Journal of Vegetation Science 7:347–358.

Craine, J. M., T. W. Ocheltree, J. B. Nippert, E. G. Towne, A. M. Skibbe, S. W. Kembel, and J. E. Fargione. 2012. Global diversity of drought tolerance and grassland climate-change resilience. Nature Climate Change 3:63–67.

Dai, A. 2012. Increasing drought under global warming in observations and models. Nature Climate Change 3:52–58.

Deppe, M., K. H. Knorr, D. M. McKnight, and C. Blodau. 2010. Effects of short-term drying and irrigation on CO2 and CH4 production and emission from mesocosms of a northern bog and an alpine fen. Biogeochemistry 100:89–103.

Dias, A., M. P. Berg, F. Bello, A. R. van Oosten, K. Bílá, and M. Moretti. 2013. An experimental framework to identify community functional components driving ecosystem processes and services delivery. Journal of Ecology 101: 29-37.

Dorrepaal, E., S. Toet, R. S. P. Van logtestijn, E. Swart, M. J. Van De Weg, T. V. Callaghan, and R. Aerts. 2009. Carbon respiration from subsurface peat accelerated by climate warming in the subarctic. Nature 460:616–619.

Dıaz, S., A. J. Symstad, F. Stuart Chapin, D. A. Wardle, and L. F. Huenneke. 2003. Functional diversity revealed by removal experiments. Trends in Ecology and Evolution 18:140–146.

Fenner, N., and C. Freeman. 2011. Drought-induced carbon loss in peatlands. Nature Geoscience 4:895-900.

Freeman, C., C. D. Evans, D. T. Monteith, B. Reynolds, and N. Fenner. 2001a. Export of organic carbon from peat soils. Nature 412:785.

Freeman, C., N. Ostle, and H. Kang. 2001b. An enzymic “latch” on a global carbon store. Nature 409:149.

Garrels, R. M., and C. L. Christ. 1965. Solutions, minerals and equilibria. Jones and Bartlett Publishers, Boston, United States.

Gerdol, R., A. Bonora, R. Gualandri, and S. Pancaldi. 1996. CO2 exchange, photosynthetic pigment composition, and cell ultrastructure of *Sphagnum* mosses during dehydration and subsequent rehydration. Canadian Journal of Botany 74:726–734.

Giraudoux, P. 2013. pgirmess: Data analysis in ecology. R package version 1.5.7. http://CRAN.R-project.org/package=pgirmess.

Grime, J. P. 1998. Benefits of plant diversity to ecosystems: immediate, filter and founder effects. Journal of Ecology 86:902-910.

Gundale, M. J., D. A. Wardle, and M.-C. Nilsson. 2010. Vascular plant removal effects on biological N fixation vary across a boreal forest island gradient. Ecology 91:1704–1714.

Hájek, T., and R. P. Beckett. 2008. Effect of water content components on desiccation and recovery in *Sphagnum* mosses. Annals of Botany 101:165–173.

Hájek, T., S. Ballance, J. Limpens, M. Zijlstra, and J. T. A. Verhoeven. 2011. Cell-wall polysaccharides play an important role in decay resistance of Sphagnum and actively depressed decomposition in vitro. Biogeochemistry 103:45–57.

Hector, A., Y. Hautier, P. Saner, L. Wacker, R. Bagchi, J. Joshi, M. Scherer-Lorenzen, E. M. Spehn, E. Bazeley-White, M. Weilenmann, M. C. Caldeira, P. G. Dimitrakopoulos, J. A. Finn, K. Huss-Danell, A. Jumpponen, C. P. H. Mulder, C. Palmborg, J. S. Pereira, A.-S. D. Siamantziouras, A. C. Terry, A. Y. Troumbis, B. Schmid, and M. Loreau. 2010. General stabilizing effects of plant diversity on grassland productivity through population asynchrony and overyielding. Ecology 91:2213–2220.

Heijmans, M. M. P. D., H. Klees, and F. Berendse. 2002. Competition between *Sphagnum magellanicum* and *Eriophorum angustifolium* as affected by raised CO2 and increased N deposition. Oikos 97:415–425.

Heijmans, M. M. P. D., W. J. Arp, and F. S. Chapin III. 2004. Controls on moss evaporation in a boreal black spruce forest. Global Biogeochemical Cycles 18:GB2004.

Hooper, D. U., F. S. Chapin III, J. J. Ewel, A. Hector, P. Inchausti, S. Lavorel, J. H. Lawton, D. M. Lodge, M. Loreau, and S. Naeem. 2005. Effects of biodiversity on ecosystem functioning: a consensus of current knowledge. Ecological Monographs 75:3–35.

Isbell, F., V. Calcagno, A. Hector, J. Connolly, W. S. Harpole, P. B. Reich, M. Scherer-Lorenzen, B. Schmid, D. Tilman, J. van Ruijven, A. Weigelt, B. J. Wilsey, E. S. Zavaleta, and M. Loreau. 2011. High plant diversity is needed to maintain ecosystem services. Nature 477:199–202.

Johnson, D., J. Vachon, A. J. Britton, and R. C. Helliwell. 2011. Drought alters carbon fluxes in alpine snowbed ecosystems through contrasting impacts on graminoids and forbs. New Phytologist 190:740–749.

Kutzbach, L., J. Schneider, T. Sachs, M. Giebels, H. Nykänen, N. J. Shurpali, P. J. Martikainen, J. Alm, and M. Wilmking. 2007. CO2 flux determination by closed-chamber methods can be seriously biased by inappropriate application of linear regression. Biogeosciences 4:1005–1025.

Lafleur, P. M., N. T. Roulet, J. L. Bubier, S. Frolking, and T. R. Moore. 2003. Interannual variability in the peatland-atmosphere carbon dioxide exchange at an ombrotrophic bog. Global Biogeochemical Cycles 17:1036.

Laine, A., D. Wilson, G. Kiely, and K. A. Byrne. 2007. Methane flux dynamics in an Irish lowland blanket bog. Plant and Soil 299:181–193.

Laine, A. M., J. Bubier, T. Riutta, M. B. Nilsson, T. R. Moore, H. Vasander, and E. S. Tuittila. 2012. Abundance and composition of plant biomass as potential controls for mire net ecosystem CO2 exchange. Botany 90:63–74.

Loreau, M., and C. de Mazancourt. 2013. Biodiversity and ecosystem stability: a synthesis of underlying mechanisms. Ecology letters.

Lund, M., T. R. Christensen, and A. Lindroth. 2012. Effects of drought conditions on the carbon dioxide dynamics in a temperate peatland. Environmental Research Letters 7:045704.

McLellan, A. J., A. H. Fitter, and R. Law. 1995. On decaying roots, mycorrhizal colonization and the design of removal experiments. The Journal of Ecology:225–230.

McNeil, P., and J. M. Waddington. 2003. Moisture controls on *Sphagnum* growth and CO2 exchange on a cutover bog. Journal of Applied Ecology 40:354–367.

Murray, K. J., J. D. Tenhunen, and J. Kummerow. 1989. Limitations on *Sphagnum* growth and net primary production in the foothills of the Philip Smith Mountains, Alaska. Oecologia 80:256–262.

Murray, K. J., J. D. Tenhunen, and R. S. Nowak. 1993. Photoinhibition as a control on photosynthesis and production of *Sphagnum* mosses. Oecologia 96:200–207.

Mulder, C. P., D. D. Uliassi, and D. F. Doak. 2001. Physical stress and diversity-productivity relationships: the role of positive interactions. Proceedings of the National Academy of Sciences of the United States of America 98:6704–6708.

Otieno, D. O., M. Wartinger, A. Nishiwaki, M. Z. Hussain, J. Muhr, W. Borken, and G. Lischeid. 2009. Responses of CO2 exchange and primary production of the ecosystem components to environmental changes in a mountain peatland. Ecosystems 12:590–603.

Piao, S., P. Ciais, P. Friedlingstein, P. Peylin, M. Reichstein, S. Luyssaert, H. Margolis, J. Fang, A. Barr, A. Chen, A. Grelle, D. Y. Hollinger, T. Laurila, A. Lindroth, A. D. Richardson, and T. Vesala. 2008. Net carbon dioxide losses of northern ecosystems in response to autumn warming. Nature 451:49–52.

Pinheiro, J., D. Bates, S. DebRoy, D. Sarkar, and T. R. D. C. Team. 2011. *nlme*: Linear and nonlinear mixed effects models. R package version 3.1-102.

Polley, H. W., B. J. Wilsey, and J. D. Derner. 2007. Dominant species constrain effects of species diversity on temporal variability in biomass production of tallgrass prairie. Oikos 116:2044–2052.

R Core Team. 2012. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org/.

Robroek, B. J. M., J. Limpens, A. Breeuwer, and M. G. C. Schouten. 2007. Effects of water level and temperature on performance of four *Sphagnum* mosses. Plant Ecology 190:97–107.

Robroek, B. J. M., M. G. C. Schouten, J. Limpens, F. Berendse, and H. Poorter. 2009. Interactive effects of water table and precipitation on net CO2 assimilation of three co‐occurring *Sphagnum* mosses differing in distribution above the water table. Global Change Biology 15:680–691.

Rochefort, L. 2000. *Sphagnum* - A keystone genus in habitat restoration. The Bryologist 103:503–508.

Rydin, H., and J. K. Jeglum. 2006. The Biology of Peatlands. Oxford University Press, USA.

Rydin, H., and K. E. Barber. 2001. Long-term and fine-scale coexistence of closely related species. Folia Geobotanica 36:53–61.

Schipperges, B., and H. Rydin. 1998. Response of photosynthesis of *Sphagnum* species from contrasting microhabitats to tissue water content and repeated desiccation. New Phytologist 140:677–684.

Strack, M., and J. S. Price. 2009. Moisture controls on carbon dioxide dynamics of peat‐Sphagnum monoliths. Ecohydrology 2:34–41.

Symstad, A. J., and D. Tilman. 2001. Diversity loss, recruitment limitation, and ecosystem functioning: lessons learned from a removal experiment. Oikos 92:424–435.

Turetsky, M. R., B. Bond-Lamberty, E. Euskirchen, J. Talbot, S. Frolking, A. D. McGuire, and E. S. Tuittila. 2012. The resilience and functional role of moss in boreal and arctic ecosystems. New Phytologist 196:49–67.

van Breemen, N. 1995. How *Sphagnum* bogs down other plants. Trends in Ecology and Evolution 10:270–275.

van Ruijven, J., and F. Berendse. 2009. Diversity enhances community recovery, but not resistance, after drought. Journal of Ecology 98:81–86.

Wang, Y., S. Yu, and J. Wang. 2007. Biomass-dependent susceptibility to drought in experimental grassland communities. Ecology letters 10:401–410.

Ward, S. E., R. D. Bardgett, N. P. McNamara, and N. J. Ostle. 2009. Plant functional group identity influences short‐term peatland ecosystem carbon flux: evidence from a plant removal experiment. Functional Ecology 23:454–462.

Wardle, D. A., K. I. Bonner, G. M. Barker, G. W. Yeates, K. S. Nicholson, R. D. Bardgett, R. N. Watson, and A. Ghani. 1999. Plant removals in perennial grassland: vegetation dynamics, decomposers, soil biodiversity, and ecosystem properties. Ecological Monographs 69:535–568.

Weltzin, J. F., S. D. Bridgham, J. Pastor, J. Chen, and C. Harth. 2003. Potential effects of warming and drying on peatland plant community composition. Global Change Biology 9:141–151.

SUPPLEMENTAL MATERIAL

APPENDIX A.Description on the exponential non-linear model used for calculating CO2 fluxes from the mesocosms.

APPENDIX B.The effects of biomass removal on net ecosystem CO2 exchange (NEE) and ecosystem respiration (Re).

APPENDIX C.Model test results for two models, one with and one without ‘removed biomass’ as a factor.

APPENDIX D. Test results of the repeated measures-ANOVA on net ecosystem CO2 exchange during the drought period.

**Table 1** Amounts of removed vascular plant aboveground biomass (g dwt), and cover of the different plant functional types (PFT) after biomass removal for the PFT removal treatments on the lawn and hummock microhabitats. Note that in the control treatment part of the ericoid and graminoid aboveground biomass has been removed. Total evapotranspiration rates have been calculated over the drought period; significant differences (Tukey’s *post-hoc* test; *p* ≤ 0.05) are indicated by different letters.

|  |  |  |  |
| --- | --- | --- | --- |
|  | Biomass removed (g dwt) | PFT cover after removal (%) | Total Evapotranspiration during the drought period (ml m-2) |
| Treatment |  | *Sphagnum* | Ericoids | Graminoids | Vascular plants |  |
| **Lawn** |  |  |  |  |  |  |
| Control | 0.95 ± 0.3 | 100 ± 0 | 15.0 ± 3.5 | 15.0 ± 2.9 | 30.0 ± 3.5 | 67.53 ± 4.3a |
| Erem | 3.35 ± 0.9 | 100 ± 0 | - | 27.5 ± 3.2 | 27.5 ± 3.2 | 67.53 ± 4.3a |
| Grem | 0.54 ± 0.2 | 100 ± 0 | 17.5 ± 7.2 | - | 17.5 ± 7.2 | 59.30 ± 2.6a |
| Erem + Grem | 3.91 ± 0.3 | 100 ± 0 | - | - | - | 60.58 ± 4.3a |
| **Hummock** |  |  |  |  |  |  |
| Control | 1.93 ± 0.7 | 100 ± 0 | 20.8 ± 2.9 | 9.2 ± 2.5 | 30.0 ± 1.6 | 69.89 ± 2.5ab |
| Erem | 4.50 ± 0.7 | 100 ± 0 | - | 17.1 ± 3.0 | 17.1 ± 3.0 | 61.36 ± 1.7a |
| Grem | 0.12 ± 0.1 | 100 ± 0 | 21.3 ± 3.5 | - | 21.3 ± 3.5 | 72.64 ± 1.8b |
| Erem + Grem | 5.26 ± 0.5 | 100 ± 0 | - | - | - | 65.24 ± 3.2ab |

**Table 2** ANOVA results (F and *p*-values) for the interactive effects of microhabitat, ericoids removal and graminoid removal on the resistance, time to NEE = 0, and recovery of net ecosystem CO2 exchange. *p-*values in bold indicate significant values.

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  |  |  | Resistance NEE |  | NEE = 0 |  | Recovery NEE |  | Resilience NEE |
|  | *Df1* | *Df2* | F |  | *p* |  | F |  | *p* |  | F |  | *p* |  | F |  | *p* |
| Microhabitat (MH) | 1 | 29 | 7.1 |  | **≤ 0.05** |  | 11.6 |  | **≤ 0.01** |  | 0.5 |  | 0.48 |  | 0.6 |  | 0.44 |
| Erem | 1 | 28 | 1.4 |  | 0.24 |  | 6.1 |  | **≤ 0.05** |  | 2.2 |  | 0.15 |  | 0.7 |  | 0.41 |
| Grem | 1 | 27 | 0.0 |  | 1.00 |  | 4.5 |  | **≤ 0.05** |  | 0.4 |  | 0.51 |  | 0.9 |  | 0.34 |
| MH : Erem | 1 | 26 | 0.5 |  | 0.49 |  | 1.3 |  | 0.26 |  | 0.0 |  | 0.9 |  | 0.0 |  | 0.91 |
| MH : Grem | 1 | 25 | 4.9 |  | **≤ 0.05** |  | 0.1 |  | 0.78 |  | 0.0 |  | 0.94 |  | 0.5 |  | 0.47 |
| Erem : Grem | 1 | 24 | 2.1 |  | 0.16 |  | 0.1 |  | 0.77 |  | 0.3 |  | 0.59 |  | 0.5 |  | 0.48 |
| MH : Erem : Grem | 1 | 23 | 0.9 |  | 0.35 |  | 0.0 |  | 0.99 |  | 1.0 |  | 0.32 |  | 0.2 |  | 0.66 |

 FIGURE LEGENDS

**Figure 1** Comparison of net ecosystem exchange (± SEM) rates between the different plant functional type removal at the pre-drought, post-drought, and post-recovery period (A: lawns; B hummocks), as well as during the drought period (C: lawns; D: hummocks). Negative values indicate net CO2 loss. For statistics, see Appendix C and D.

**Figure 2** The effect of plant functional type removal on ecosystem respiration during the drought period (A: lawns; B: hummocks). Negative values indicate net CO2 loss. For statistics, see Appendix D.

**Figure 3** The effect of plant functional type removal on the resistance (A, B), the time to NEE = 0 (C, D), and the recovery (E, F) in the two microhabitats. For definitions, see material and methods section.