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1 **A comparative experimental approach to ecotoxicology in shallow-water and deep-sea**
2 **holothurians suggests similar behavioural responses**

3

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11

12 **Highlights**

- 13 • Shallow-water and deep-sea holothurians avoided copper-contaminated sediment.
- 14 • Shallow-water taxa may be suitable ecotoxicological proxies for deep-sea taxa.
- 15 • Avoidance behaviour may have bioenergetic consequences.

16

17 **Abstract**

18 Exploration of deep-sea mineral resources is burgeoning, raising concerns regarding
19 ecotoxicological impacts on deep-sea fauna. Assessing toxicity in deep-sea species is
20 technologically challenging, which promotes interest in establishing shallow-water
21 ecotoxicological proxy species. However, the effects of temperature and hydrostatic pressure
22 on toxicity, and how adaptation to deep-sea environmental conditions might moderate these

23 effects, are unknown. To address these uncertainties we assessed behavioural and
24 physiological (antioxidant enzyme activity) responses to exposure to copper-spiked artificial
25 sediments in a laboratory experiment using a shallow-water holothurian (*Holothuria*
26 *forskali*), and in an *in situ* experiment using a deep-sea holothurian (*Amperima* sp.). Both
27 species demonstrated sustained avoidance behaviour, evading contact with contaminated
28 artificial sediment. However, *A. sp.* demonstrated sustained avoidance of 5 mg l⁻¹ copper-
29 contaminated artificial sediment whereas *H. forskali* demonstrated only temporary avoidance
30 of 5 mg l⁻¹ copper-contaminated artificial sediment, suggesting that *H. forskali* may be more
31 tolerant of metal exposure over 96 h. Nonetheless, the acute behavioural response appears
32 consistent between the shallow-water species and the deep-sea species, suggesting that *H.*
33 *forskali* may be a suitable ecotoxicological proxy for *A. sp.* in acute (≤ 24 h) exposures which
34 may be representative of deep-sea mining impacts. No antioxidant response was observed in
35 either species, which was interpreted to be the consequence of avoiding copper exposure.
36 Although these data suggest that shallow-water taxa may be suitable ecotoxicological proxies
37 for deep-sea taxa, differences in methodological and analytical approaches, and in sex and
38 reproductive stage of experimental subjects, require caution in assessing the suitability of *H.*
39 *forskali* as an ecotoxicological proxy for *A. sp.* Nonetheless, avoidance behaviour may have
40 bioenergetic consequences that affect growth and/or reproductive output, potentially
41 impacting fecundity and/or offspring fitness, and thus influencing source-sink dynamics and
42 persistence of wider deep-sea populations.

43

44 **Keywords**

45 bathymetric; deep-sea mining; ecology; hydrostatic pressure; physiology; temperature;
46 toxicology

47

48 **Introduction**

49 Interest in exploiting deep-sea mineral resources is increasing, driven by rising metal prices
50 and the challenges of achieving environmentally and socio-economically acceptable solutions
51 for extraction, processing, and disposal of materials from low-grade rock deposits on land.

52 Economically important metals and rare-earth elements are present in deep-sea massive
53 polymetallic sulphides, polymetallic manganese nodules, and cobalt-rich ferromanganese
54 crusts (Petersen et al. 2016), and even muds (Kato et al. 2011). However, despite great
55 international attention the responses of targeted ecosystems to mineral extraction remain
56 largely unknown. The remoteness of deep-sea ecosystems makes experimental assessment of
57 deep-sea mining impacts challenging, and the urgency to exploit these resources may drive
58 researchers to use shallow-water ecosystems as proxies in assessments that influence policy-
59 making. The extensive data available for shallow-water taxa make this an attractive approach,
60 but the potential synergies that may arise from the interaction of stressors with environmental
61 factors, such as temperature, hydrostatic pressure, and oxygen concentration (Pörtner 2010;
62 Sokolova 2013; Brown and Thatje 2015), are typically ignored (Mestre et al. 2013).

63 Nevertheless, these synergies may be crucial to accurately project the impacts of deep-sea
64 mineral exploitation throughout species' thermal and baric distributions (Brown et al. in
65 revision). Further, whether shallow-water taxa are suitable proxies for deep-sea taxa remains
66 to be revealed; adaptation to environmental conditions in the deep sea may invalidate such an
67 approach by affecting sensitivity to potential stressors.

68 Extraction of deep-sea mineral deposits may have significant ecotoxicological impacts on the
69 deep-sea fauna. For example, the polymetallic manganese nodules which form on abyssal
70 plains comprise high concentrations of potentially toxic metals such as copper (Ramirez-
71 Llodra et al. 2011) which may be released during extraction processes (Levin et al. 2016;

72 Gollner et al. 2017). Few studies have employed a comparative approach to explore the
73 effects of deep-sea conditions on sensitivity to toxicants, and these have predominantly
74 focused on adaptation to toxic challenges presented by hydrothermal vent environment.
75 However, these studies have assessed *in situ* biomarker expression (metallothioneins,
76 antioxidant enzymes, lipid peroxidation) and/or metal accumulation in hydrothermal vent and
77 coastal shrimp (Geret et al. 2002; Kadar et al. 2006; Gonzalez-Rey et al. 2007, 2008). Whilst
78 this approach identifies *in situ* physiology at ambient toxicant concentrations, it has yielded
79 limited information regarding similarities and/or differences in responses and sensitivity to
80 elevated toxicant concentrations among species from different environments. Such
81 information may only be revealed through a comparative experimental approach, where
82 individuals from different phylogenetically related shallow-water and deep-sea species are
83 exposed to similar challenges under native environmental conditions (e.g. hydrostatic
84 pressure, temperature).

85 Recent experimental assessment of the effects of hydrostatic pressure and temperature on
86 sublethal responses to copper exposure in the deep-sea hydrothermal vent shrimp *Rimicaris*
87 *exoculata* at 10°C and 30.0 MPa (Auguste et al. 2016) and the shallow-water shrimp
88 *Palaemon varians* provided the first opportunity to develop significant insight into caridean
89 adaptation to toxicity in a hydrothermal vent environment (Brown et al. in revision).
90 Comparisons suggest that sensitivity and responses to toxicants may not differ between these
91 species at a common temperature at native hydrostatic pressure, suggesting that shallow-
92 water taxa may be suitable ecotoxicological proxies for deep-sea taxa (Brown et al. in
93 revision). However, *P. varians* is significantly more tolerant of both copper and cadmium
94 than other shallow-water caridean shrimp species, perhaps as a result of adaptations to highly
95 variable environmental temperature, salinity, and oxygen concentration in estuarine, salt
96 marsh, and saline lagoon habitat (Brown et al. in revision). Consequently, matching the

97 environmental variability experienced by deep-sea species may be a key criterion for the
98 selection of shallow-water ecotoxicological proxy species (Brown et al. in revision).

99 Whilst bathyal hydrothermal vent taxa may be affected by the exploitation of seafloor
100 massive sulphides, extraction of polymetallic manganese nodules will impact abyssal fauna
101 (Levin et al. 2016; Gollner et al. 2017). Adaptations to the environmental conditions in the
102 abyssal deep-sea may be significantly different from adaptations to the bathyal hydrothermal
103 vent environment (Childress and Fisher 1992; Brown and Thatje 2014). For example,
104 temperature- and mass-normalised metabolic rate declines significantly with depth in
105 holothurians (Brown et al. under review). Consequently, ecotoxicological comparisons
106 between shallow-water species and bathyal hydrothermal vent species may not be
107 representative of other deep-sea taxa. To address this uncertainty, we assessed responses to
108 toxicant exposure in the temperate shallow-water holothurian *Holothuria forskali* and the
109 abyssal holothurian *Amperima* sp..

110 *H. forskali* occurs along the Atlantic coasts of northwest Europe and in the Mediterranean
111 Sea, from the intertidal to 100 m depth (OBIS, 2015). *H. forskali* can grow to 49 cm length
112 (Tuwo and Conand 1992). *H. forskali* is benthic and has a tough body wall, typically
113 occurring on boulders and rocks and moving using locomotory podia located on the ventral
114 sole (personal observation). In contrast, *Amperima* species occur widely in the abyssal deep
115 sea (>4000 m depth) and have been reported in the North and South Atlantic, Southern,
116 Indian, and South Pacific (OBIS, 2015). *A. sp.* can grow to 13 cm and is capable of
117 swimming by undulating its body, typically make brief excursions few metres into the water
118 column before drifting back to the seabed (personal observation). Both *H. forskali* and *A. sp.*
119 are detritivores (personal observation).

120 Copper was selected as the experimental toxicant based on representation in deep-sea mineral
121 deposits (see Ramirez-Llodra et al. 2011), and the demonstration that copper has the greatest
122 potential for rapid release during seafloor massive sulphide extraction (Knight and Roberts
123 2016).

124

125 **Materials and Methods**

126 *Shallow-water exposures*

127 Specimens of *Holothuria forskali* were sampled by divers (depth range 16 m to 19 m) on 27th
128 May 2015 from ~50 m² of rocky reef with sand gullies immediately south of Plymouth
129 Mewstone (50°18.158 N, 004°06.555 W) (indicative salinity 35.3, temperature 11.2°C,
130 dissolved oxygen 260.9 µmol l⁻¹ from 16 m to 19 m at the Western Channel Observatory L4
131 station at 50°25.00 N, 004°21.70 W on the 27th May 2015). Holothurians were transported to
132 the Marine Biology and Ecology Research Centre at the University of Plymouth, and
133 maintained for 1 week in a recirculating aquarium (salinity ~32, temperature 10°C, dissolved
134 oxygen concentration >245 µmol l⁻¹). Subsequently, holothurians were transported to the
135 National Oceanography Centre Southampton (NOCS) by temperature-controlled vehicle
136 (10°C, <6 hours), and transferred to a recirculating aquarium (salinity ~32, temperature 9.8°C
137 decreased by 1°C weekly to 4°C, pH 8.0, ammonia <0.1 mg l⁻¹, light:dark cycling following
138 seasonal changes). Holothurians were fed approximately 15 g of Marine Cuisine (Tropical
139 Marine Centre) three times each week. Holothurians were maintained for ≥1 weeks at 4°C
140 prior to experimental treatments. Treatments were performed concurrently to minimise
141 potential temporal influences on responses, but were logistically limited to 1 replicate of each
142 treatment per week by the availability of incubators. Experimental exposures were conducted

143 between 17th July and 28st August 2015 to reduce potential variability in biochemical
144 responses resulting from annual reproductive cycles (Santos et al., 2016).

145 Static 96-hour experimental exposures were conducted in plastic aquaria (32.5 × 20 × 20 cm)
146 filled with 10 l aquarium water (salinity 32, dissolved oxygen concentration >245 µmol l⁻¹,
147 pH 8.0, ammonia <0.1 mg l⁻¹) at 4°C, covered with vented plastic lids allowing air
148 circulation, with 12:12 photoperiod. The experimental temperature was selected to represent
149 the lowest temperature occurring within the species' biogeographic range (Locarnini et al.
150 2013; OBIS, 2016). Adult holothurians were used in all treatments (size range 18 cm to 24
151 cm). Identification of *H. forskali* sex requires dissection. Post-treatment dissection indicated
152 that both male and female holothurians were used in all treatments (1 or 2 males in each
153 treatment), and that both male and female holothurians were in stage II (developing oocytes
154 or sperm) or stage III (mature oocytes or sperm) in the reproductive cycle (determined
155 according to Santos et al., 2016). Each holothurian was assigned to either a control, artificial
156 sediment addition, or copper-spiked artificial sediment addition treatments (n = 5). 500 g of
157 artificial sediment was prepared for each addition, with composition designed to reflect
158 particle-size distribution at the DISturbance and reCOLonisation (DISCOL) experimental
159 area in the abyssal Peru Basin (Grupe 2001), whilst incorporating OECD guidance (OECD
160 2004). Artificial sediment comprised 425 g white quartz sand (99% < 90 µm), 70 g kaolin
161 (96% < 45 µm), and 5 g peat (100% < 500 µm). Artificial sediment was spiked with copper
162 by 72 h incubation at 20°C in 250 ml of deionised water spiked with copper by the addition
163 of stock solution of CuSO₄·5H₂O, according to OECD guidance (OECD 2004). The stock
164 solution was prepared using deionised water and analytical grade CuSO₄·5H₂O, and
165 incubation concentrations were 0, 0.1, 1, 5, 10, and 20 mg l⁻¹. Artificial sediment and copper-
166 spiked artificial sediment was drained and released into the aquaria 24 hours prior to
167 introduction of the holothurian. Holothurians were gently detached from the maintenance

168 aquarium and transferred to the floor of the experimental aquarium with minimal aerial
169 exposure. Position within the aquarium (i.e. anterior and posterior height above aquarium
170 floor) and behaviour (e.g. contraction, ejection of cuvierian tubules, movement, feeding) were
171 monitored for 15 minutes immediately following introduction to experimental conditions, and
172 at 24-h intervals thereafter.

173 After 96 hours of experimental exposure, holothurians were removed from experimental
174 aquaria and dissected immediately. Individuals were not anaesthetised prior to dissection to
175 minimise potential for biochemical artefacts (Arafa et al. 2008). Feeding tentacle, body wall,
176 longitudinal muscle, gonad, respiratory tree, rete mirabile, digestive tract, and digestive tract
177 contents were sampled. Tissue was transferred to a 1.5 ml centrifuge tube, and snap frozen in
178 liquid nitrogen within 10 minutes of removal from experimental aquarium. Once frozen,
179 tissue was preserved at -80°C until analysed. Holothurians typically uptake heavy metals and
180 demonstrate antioxidant responses to heavy metals in the body wall, respiratory tree, and
181 digestive tracts (e.g. Wang et al. 2015, Li et al. 2016, Wang et al. 2016). *Holothuria forskali*
182 digestive tract contents did not include the artificial sediment and therefore biochemical
183 responses were only examined in respiratory tree and body wall tissues.

184

185 *Deep-sea exposures*

186 The *in situ* toxicology experiment was deployed from the RV *Sonne* during cruise SO242-2
187 of the Joint Program Initiative Oceans – Ecological Aspects of Deep-Sea Mining.
188 Deployments were made at the southern reference site ($007^{\circ}07.5\text{ S}$, $088^{\circ}27.0\text{ W}$) outside the
189 DISCOL experimental area in the abyssal Peru Basin (see Borowski 2001) (salinity 35.96,
190 temperature 1.85°C , dissolved oxygen $120.83\ \mu\text{mol l}^{-1}$). Experimental exposures were
191 conducted between 12th September and 16th September 2015. Reproductive cycles in other

192 *Amperima* species appear linked to changes in the flux of organic matter to the seafloor
193 (Wigham et al. 2003). However, the DISCOL region is a site with low organic matter flux
194 (Weber et al. 2000) suggesting that populations here may be sink populations rather than
195 source populations.

196 *In situ* treatments were conducted using corrals (Fig. 1). The internal dimensions of each
197 corral tapered from 40 cm × 40 cm to 30 cm × 30 cm, and each corral was covered by a fine
198 mesh. The corrals were deployed to the seafloor at 4196.5 m depth on the GEOMAR Remote
199 Operated Vehicle (ROV) elevator platform. Subsequently, the GEOMAR ROV *Kiel 6000*
200 collected each corral, located an *Amperima* sp. individual, and carefully placed the corral
201 over the holothurian. *A. sp.* was selected as the test species based on high abundances
202 observed during ROV dives on preceding days. Adult holothurians were used in all
203 treatments (size range 8 cm to 10 cm). Whilst post-treatment dissection did not resolve *A. sp.*
204 sex (all *A. sp.* were in stage I in the reproductive cycle without identifiable gonad), *A. spp.*
205 reach maturity at small size, e.g. 10 mm in *Amperima rosea* (Wigham et al. 2003). Each
206 holothurian was assigned to either a control, artificial sediment addition, or copper-spiked
207 artificial sediment addition treatment (n = 2). Control treatments remained undisturbed,
208 whereas artificial sediment or copper-spiked artificial sediment was released into each corral
209 using the ROV manipulator. Artificial sediment was transported to the seafloor aboard the
210 ROV, in 1 litre plastic bottles adapted to facilitate opening using the ROV manipulator. The
211 appropriate plastic bottle was inverted immediately above the mesh covering the corral, and
212 inserted through a flap in the mesh to release the artificial sediment. Preparation of artificial
213 sediments and copper-spiking were performed as described previously, but incubation
214 concentrations were limited to 0 and 5 mg l⁻¹ due to logistical constraints of operating in the
215 deep sea. Incubation concentrations were selected based on behavioural data from the
216 *Holothuria forskali* exposures. After ~94 hours holothurians were sampled using the ROV

217 suction pump, which was inserted through the flap in the mesh. Additional holothurians were
218 sampled to assess the effect of corral enclosure (n = 3). Holothurians were isolated in the
219 ROV suction pump chambers and recovered to the surface where they were dissected in a
220 temperature controlled laboratory (2°C). As with *Holothuria forskali*, individuals were not
221 anaesthetised prior to dissection. The anatomy of *Amperima* sp. differed from that of
222 *Holothuria forskali* and separation of different tissues was challenging. Consequently, only
223 bulk tissue samples (predominantly body wall) were taken from *Amperima* sp. individuals.
224 Tissue was transferred to a 1.5 ml centrifuge tube, and snap frozen in liquid nitrogen. Once
225 frozen, tissue was preserved at -80°C until analysed.

226

227 *Biochemical response*

228 Tissue samples were removed from -80°C and maintained on ice. A ~10 mg subsample was
229 dissected for biochemical analysis. Total protein and enzyme activities were determined by
230 absorbance using a microplate reader (FLUOstar OPTIMA, BMG Labtech). Total protein
231 was assessed at 584 nm absorbance using an assay kit employing the Lowry method with
232 Peterson's modification (Sigma-Aldrich TP0300). Superoxide dismutase (SOD) activity was
233 assessed at 450 nm absorbance using an assay kit employing inhibition of xanthine oxidase
234 activity (Sigma-Aldrich 19160). Glutathione peroxidase (GPx) activity was assessed at 340
235 nm absorbance using an assay kit employing NADPH oxidation in the presence of excess
236 glutathione reductase, reduced glutathione, and corresponding peroxide (Abcam ab102530).

237

238 *Statistical analysis*

239 *H. forskali* position and biochemical data were homoscedastic (Levene's test, $p > 0.05$).

240 Consequently, the effect of exposure to copper-spiked artificial sediment on *H. forskali*

241 position was assessed using two-way repeated measures analysis of variance (ANOVA), with
242 treatment (control, copper-spiked artificial sediment additions) and exposure duration as
243 fixed factors ($\alpha = 0.05$). The *post-hoc* Holm-Sidak multiple comparisons test was used to
244 determine significant differences from initial positions within treatments. Statistical
245 inferences were confirmed using the repeated measures ANOVA on ranks employing Dunn's
246 test to assess to assess differences in holothurian anterior and posterior position within each
247 treatment over exposure duration. The effect of exposure to copper-spiked artificial sediment
248 on *H. forskali* biochemistry was assessed using two-way ANOVA, with tissue (body wall,
249 respiratory tree) and treatment (control, copper-spiked artificial sediment additions) as fixed
250 factors ($\alpha = 0.05$). The effects of exposure to copper-spiked artificial sediment on *Amperima*
251 sp. position and biochemistry were not assessed statistically because of the low replication
252 (Vaux 2012).

253

254 **Results**

255 *Positional response*

256 *Holothuria forskali* responded to copper-spiked artificial sediment exposure by moving both
257 anterior and posterior away from ≥ 5 mg l⁻¹ copper-contaminated artificial sediment (Fig. 2)
258 (respectively, $F_{20,96} = 1.726$, $p = 0.042$ and $F_{20,96} = 3.391$, $p < 0.001$, with *post-hoc* test $p <$
259 0.05). However, only anterior position in the 5 mg l⁻¹ treatment differed significantly from
260 starting position and only after 24 hours exposure duration.

261 *Amperima* sp. in the control treatment remained on the seafloor. In contrast, individuals
262 exposed to uncontaminated or copper-spiked artificial sediment immediately engaged in
263 swimming. After ~94 hours, individuals in both the control treatment and the uncontaminated
264 artificial sediment addition treatment were on the seafloor. However, individuals exposed to

265 copper-spiked artificial sediment were attached inverted to the mesh covering the corral (Fig.
266 1).

267

268 *Biochemical response*

269 Exposure to copper-spiked artificial sediment had no effect on body wall and respiratory tree
270 superoxide dismutase (SOD) and glutathione peroxidase (GPx) activity in *H. forskali* (Fig. 3)
271 (respectively, $F_{5,48} = 0.541$, $p = 0.744$ and $F_{5,48} = 1.188$, $p = 0.329$). Exposure to copper-
272 spiked artificial sediment appeared to have no significant effect on bulk tissue (predominantly
273 body wall) SOD or GPx activity in *Amperima* sp. (Fig. 4).

274

275 **Discussion**

276 Differences in methodological (static laboratory exposures compared with *in situ* exposures)
277 and analytical approaches (tissue-specific analysis compared with bulk tissue analysis), and
278 in sex (male and female compared to undetermined) and reproductive stage (II or III
279 compared with I) of experimental subjects, require caution in assessing the suitability of
280 *Holothuria forskali* (a shallow-water species) as an ecotoxicological proxy for *Amperima* sp.
281 (a deep-sea species): these factors may affect results. Further, the low replication achieved
282 during *in situ* experimentation allows only extremely limited confidence in inferences.
283 Nonetheless, *A. sp.* demonstrated sustained avoidance of 5 mg l⁻¹ copper-contaminated
284 artificial sediment whereas *H. forskali* demonstrated only temporary avoidance of 5 mg l⁻¹
285 copper-contaminated artificial sediment, suggesting that *H. forskali* may be more tolerant of
286 metal exposure over 96 h. However, the acute behavioural response appears consistent
287 between the shallow-water species and the deep-sea species, suggesting that *Holothuria*

288 *forskali* may be a suitable ecotoxicological proxy for *A. sp.* in acute (≤ 24 h) exposures, which
289 may be representative of deep-sea mining impacts.

290 The positional response to exposure to copper-spiked artificial sediment displayed by both
291 the shallow-water and deep-sea holothurians likely represents an adaptive behavioural
292 response, contributing directly to individual survival (Dill 1987). Holothurians putatively
293 possess chemosensing capability, detecting sediment patches with higher organic carbon
294 content (Slater 2010) and selecting organic carbon-rich particles (Paltzat et al. 2008). The
295 avoidance behaviour observed in both shallow-water and deep-sea species suggests that the
296 chemosensory ability extends to potential toxicants. The absence of any antioxidant response
297 in either species may be a consequence of avoiding sustained copper exposure through
298 behavioural modification: analysis of copper concentrations or metallothionein
299 concentrations would be required to confirm this. However, sediment plume avoidance or
300 avoidance of redeposited contaminated sediments in deep-sea holothurians may be ineffective
301 in the mining context. Individuals may engage in swimming, but will may drift with the same
302 prevailing current that will disperse the potentially toxic sediment plume (Levin et al. 2016;
303 Gollner et al., 2017). Individuals may subsequently settle in areas with contaminated
304 sediment redeposition, thus maintaining metal exposure and stimulating further avoidance
305 behaviour. Bioenergetic consequences of avoidance behaviour or responses to sustained
306 sublethal metal exposures may be critical (Sokolova et al. 2012). Avoidance behaviour
307 increases energy allocation to activity through energetic trade-offs: increased allocation of
308 energy to activity during stress diminishes energy available for other functions, and
309 consequently reduces scope for growth and reproductive output (Kooijman 2010; Sokolova et
310 al. 2012). Responses to sustained sublethal metal exposures may have similar energetic
311 impacts (Sokolova et al. 2012).

312 The potential ecological implications of holothurian avoidance behaviour extend beyond
313 individual holothurian species. Holothurians are keystone deep-sea species (Gage & Tyler
314 1991; Kuhnz et al. 2014), deepening the oxic sediment horizon through bioturbation and
315 therefore influencing the distribution of infaunal species (Iken et al. 2001; Soto et al. 2010).
316 Further, bioturbation by holothurians can moderate toxicity to both epi- and infauna by
317 distributing toxicants within sediments (Storelli et al. 2001; Yuan et al. 2016). Reduced
318 bioturbation resulting from behavioural avoidance may therefore also affect the persistence of
319 both epi- and infaunal species (Mermillod-Blondin 2011) in areas impacted by sediment
320 plumes incorporating toxicants. Sediment re-deposition may occur over scales of 10-100 km
321 from mining sites (see Wedding et al. 2013), therefore the behavioural avoidance reported
322 here may have significant farfield ecosystem scale effects that require consideration in
323 environmental management of deep-sea mineral exploitation.

324

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340

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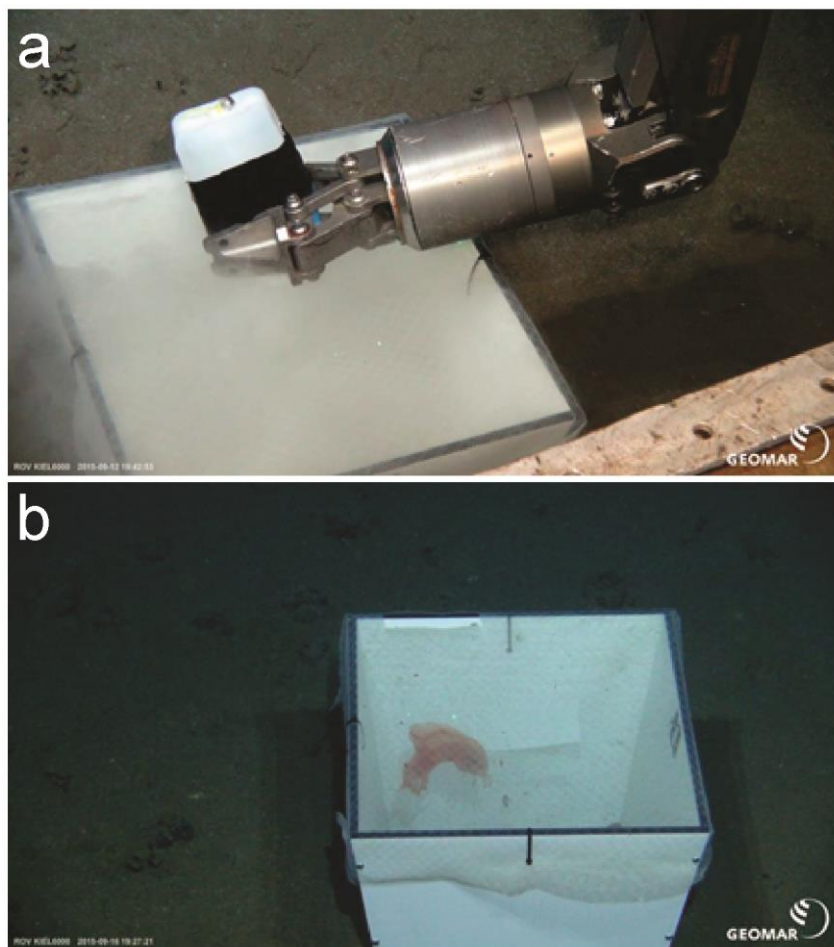
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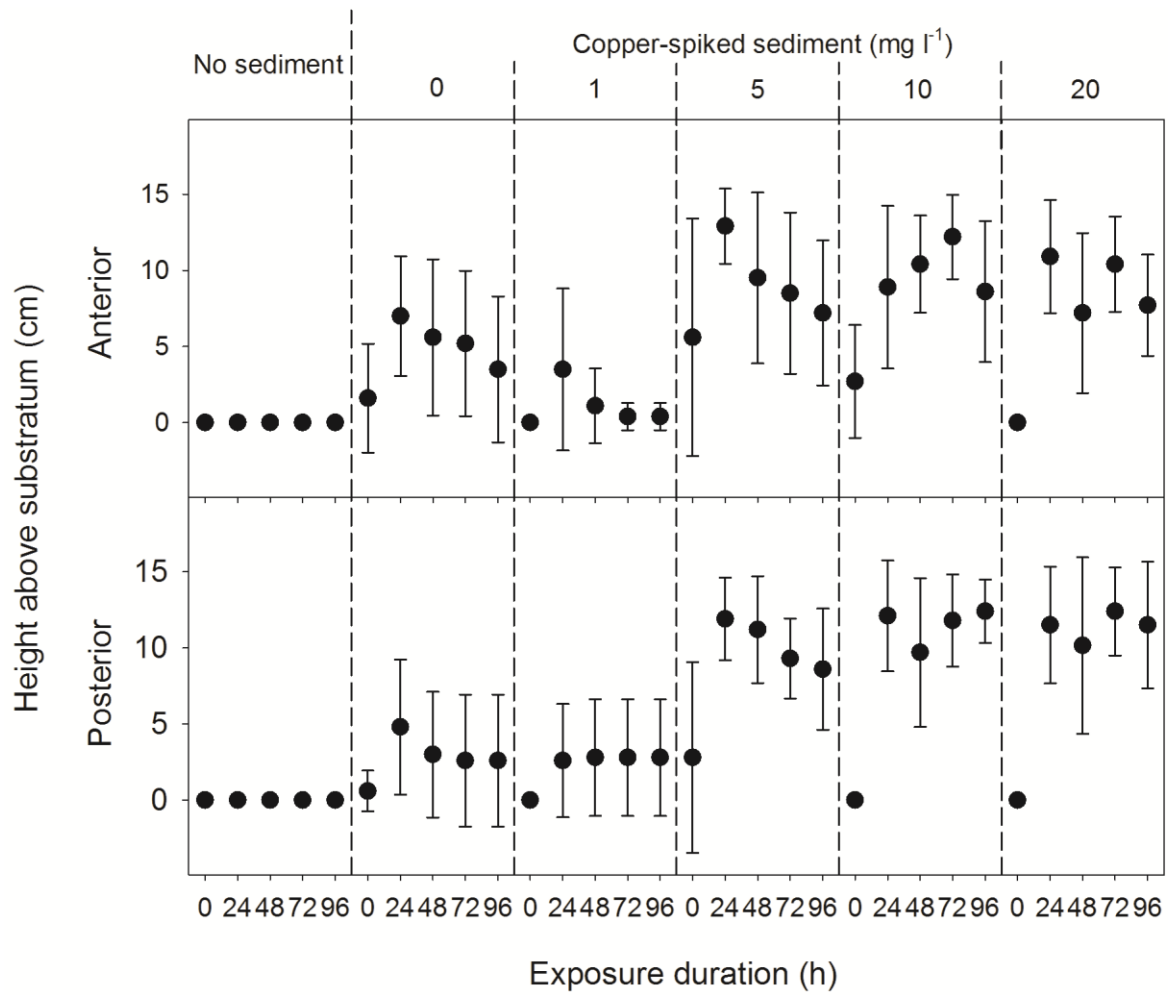
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- 475

476 **Figures**

477

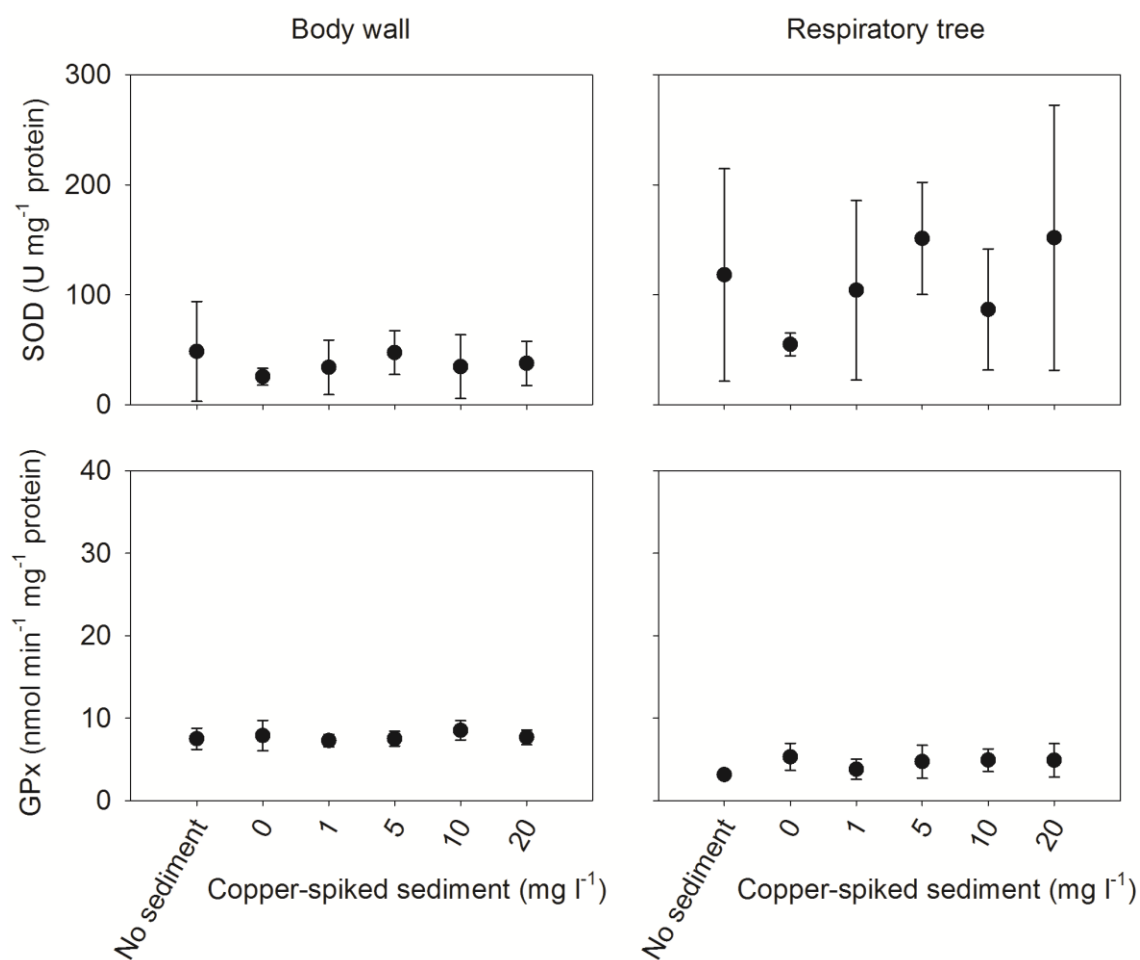
478 **Fig. 1.** *In situ* ecotoxicology experiments. (a) Introduction of artificial sediment to a corral
479 using the ROV *Kiel 6000* manipulator. (b) *Amperima* sp. position ~94 hours after exposure to
480 5 mg l⁻¹ copper-spiked artificial sediment. The internal dimensions of each corral tapered
481 from 40 cm × 40 cm to 30 cm × 30 cm.

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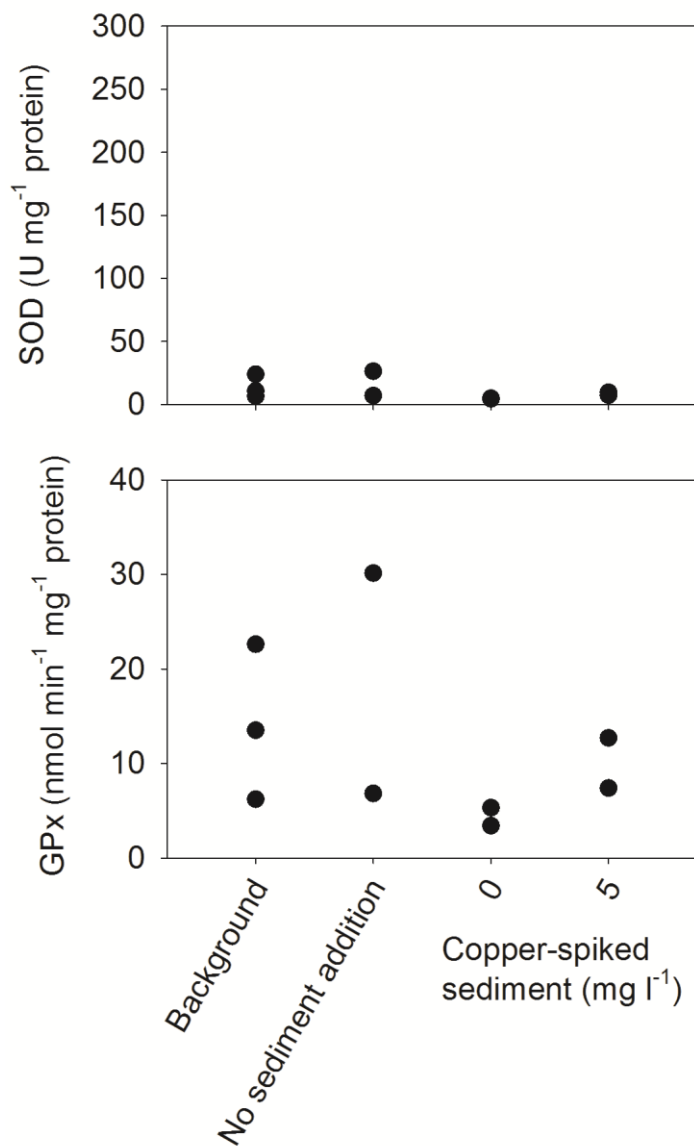
484 **Fig. 2.** Effect of exposure to copper-spiked sediment on *Holothuria forskali* position above
 485 the sediment surface (mean \pm SD; n = 5). Error bars are absent where all individuals were
 486 positioned on the floor of the aquarium.



487

488 **Fig. 3.** Effect of exposure to copper-spiked sediment on body wall and respiratory tree
 489 superoxide dismutase (SOD) and glutathione peroxidase (GPx) activity in *Holothuria forskali*
 490 (mean \pm SD; n = 5). Error bars appear absent where SD is smaller than the data point.

491



492

493 **Fig. 4.** Effect of exposure to copper-spiked sediment on bulk tissue (predominantly body
494 wall) superoxide dismutase (SOD) and glutathione peroxidase (GPx) activity in *Amperima*
495 sp. (individual data).