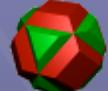


Smart Lab 2.5

The digital future of laboratory Chemistry

Jeremy G.Frey
School of Chemistry
University of Southampton, UK

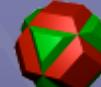




Talk

- Laboratory Research
- Laboratory Notebooks
- Blogs and Blogjects
- Publication@Source





Research Space UCB Guidelines

- Research space is a more complex problem. Changes over the past decade have no doubt been even more profound for research space than office space: but those changes are unique to each discipline, and to identify and characterize them would be a significant project in itself.....



UCB Space Study



29 Jan 2007



Jeremy G. Frey
University of Southampton

Space for model
building.....





The tea room is the ‘heart’ of the department - plans to create a more interdisciplinary equivalent for the Life Science Interface ut still only serves a local function



Growing need for the global (virtual) equivalent of the “Tea Room”

Social Space? Space for Discussions?

29 Jan 2007



Jeremy G. Frey
University of Southampton

Web 2.0 Workshop OGF 19



The CombeChem Project

- End to End linking of data and information
 - Publication@Source
- So collect data with regard to how it could eventually be used
 - Make sure the metadata is of high quality
 - Record properly at source in Digital Form
- The Chemistry Lab
 - People & Machines working together



Combechem

Smart Lab

Instruments on the Grid

Statistics

R4L

BioSimGrid

E-Malaria

e-Bank



If only I knew exactly
how she did this
experiments

I wish I had
recorded things at
the start the way I
do now.....

I wish I could get
the numbers from
this graph - the pdf
is not much use.

I know all this supplementary
information could be useful but
will people really remember the
format? Is it worth all the
hassle?

Typical Laboratory



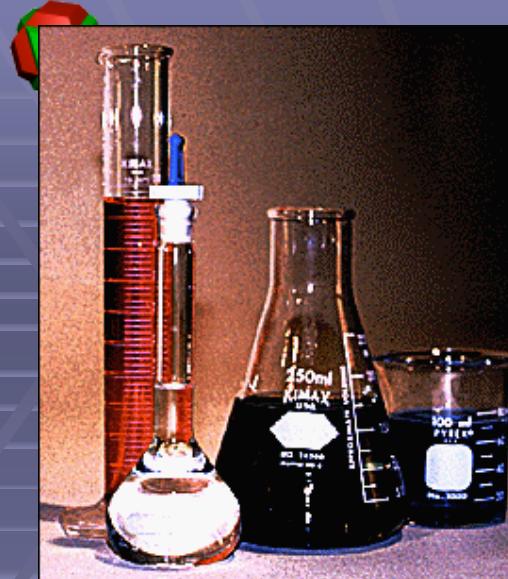
I am sure we collected that information a few years ago...

The details should be in her thesis....

Can you read what he says here....?

Can you find the file of data that were used to make the plot?

Some of these problems are due to the lack of information recorded at the time. Others are due to loss of information over time.



ChemLab

The Chemistry 3/5 & 6 Laboratories

- ▶ General Information
- ▶ Instruments & Techniques
- ▶ Chemistry 3/5 Experiments
- ▶ Chemistry 6 Experiments

DARTMOUTH COLLEGE

Safety

- [General Rules](#)
- [Safety Equipment](#)
- [Safety Hazards](#)
- [Emergency Procedures](#)

Resources

- [Applets](#)
- [General FAQ](#)
- [Uncertainty](#)

[ChemLab Home](#)

Permanent,
documented
and primary
record of
laboratory
observations

29 Jan 2007



| [Info](#) | [Techniques](#) | [Chem 3/5](#) | [Chem 6](#)

How to Keep a Notebook

One of the most useful skills you will acquire in the laboratory is the proper use of a laboratory notebook. Notebooks, or other formally kept records, are an essential tool in many careers, ranging from that of the research scientist to that of the practicing physician. The effort invested in developing good habits of notebook use will be amply repaid for students who pursue a future in the basic or applied sciences. Experience has indicated that skillful notebook use is developed by most students only through continued special effort--it does not come naturally. Some of the main principles of sound notebook use are outlined below.

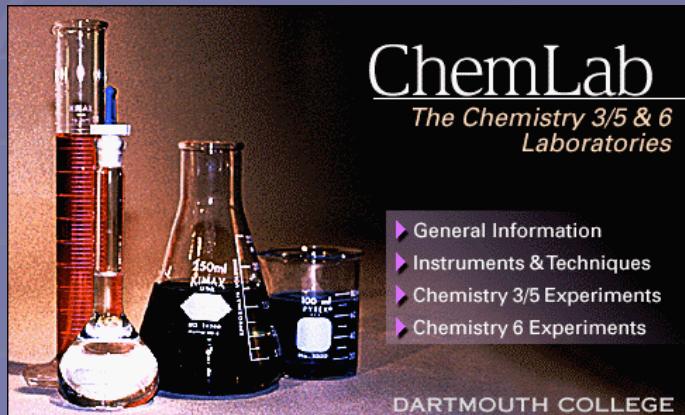
The laboratory notebook is a permanent, documented, and primary record of laboratory observations. Therefore, your notebook will be a bound journal with pages that should be numbered in advance and never torn out. A notebook will be supplied to you before the first laboratory period. Write your name, the name of your TA, and your lab section on the cover of your notebook. All notebook entries must be in ink and clearly dated. No entry is ever erased or obliterated by pen or "white out". Changes are made by drawing a single line through an entry in such a way that it can still be read and placing the new entry nearby. If it is a primary datum that is changed, a brief explanation of the change should be entered (e.g. "balance drifted" or "reading error"). No explanation is necessary if a calculation or discussion is changed; the section to be deleted is simply removed by drawing a neat "x" through it.



necessary if a calculation or discussion is changed; the section to be deleted is simply removed by drawing a neat "x" through it.

In view of the fact that a notebook is a primary record, data are not copied into it from other sources (such as this manual or a lab partner's notebook, in a joint experiment) without clear acknowledgment of the source. Observations are never collected on note pads, filter paper, or other temporary paper for later transfer into a notebook. If you are caught using the "scrap of paper" technique, your improperly recorded data may be confiscated by your TA or instructor at any time. It is important to develop a standard approach to using a notebook routinely as the primary receptacle of observations.

Each week at the beginning of lab lecture, you will turn in your prelab problems from the manual for grading. Prelab problems not turned in at the beginning of lab lecture will be



Observations are never collected on note pads, filter paper or other temporary paper for later transfer into a notebook

If you are caught using the "scrap of paper" technique, your improperly recorded data may be confiscated by your TA



Need to make
the data
available

Need to be
able to find it

But how to
expose it?



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From The New Yorker Book of Technology Cartoons.

First, they do an online search

29 Jan 2007



Jeremy G. Frey
University of Southampton

Web 2.0 Workshop OGF 19



COSHH

Leverage off things we already have to do – “We have a cunning plan”

COSHH ASSESSMENT FORM				Record No.
Substance Name	Physical Form	Quantity	Nature of Hazard	
Water	Liquid	1000ml	None	
Dextrose	Solid	<20g	possible irritation to eyes and skin	
Caffeine	Solid (tea)	<1g	Harmful if swallowed, induce vomiting.	
Milk	liquid	<100ml	No particular hazards	
Nature of Process liquid extraction of caffeine, followed by combination with dextrose to produce a sweet drink				
Is there a less hazardous substance? No If so, why not use it?				
Control Measures Required (No specific measure required) (Local exhaust ventilation, personal protection, etc.)				



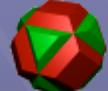
The fully semantic ELN - everything is linked!

29 Jan 2007



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University of Southampton

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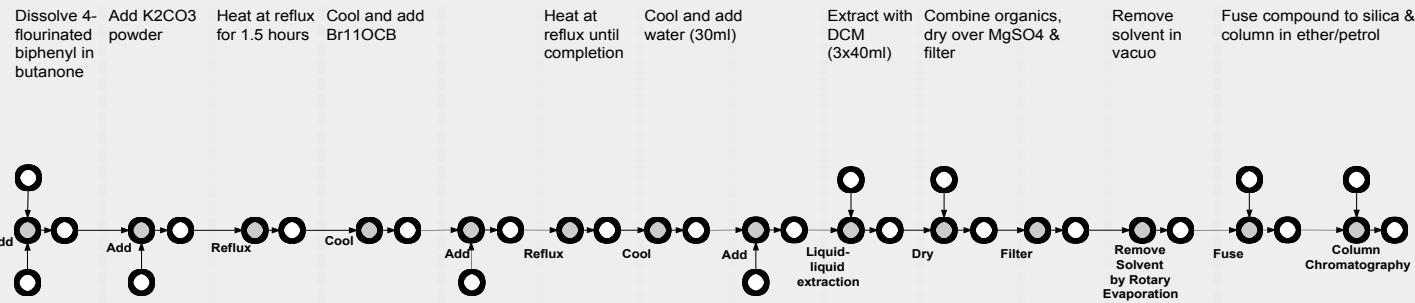


To [
List

TO DO LIST

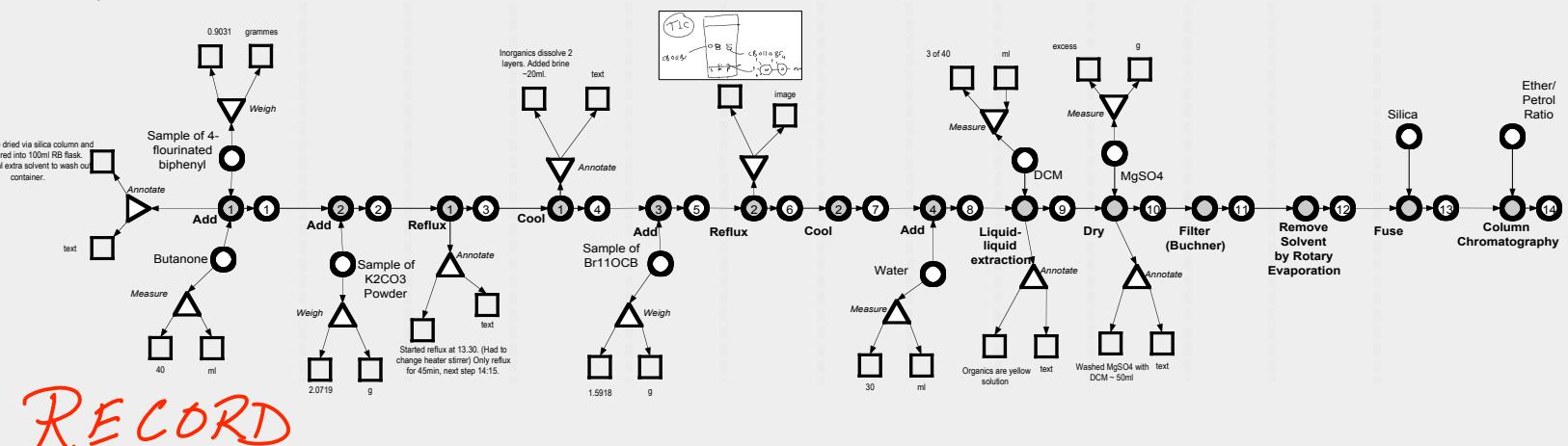
Ingredient List

Fluorinated biphenyl	0.9 g
Br11OCB	1.59 g
Potassium Carbonate	2.07 g
Butanone	40 ml

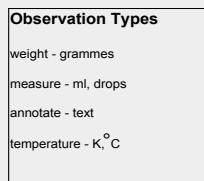
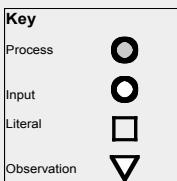


PLAN

Process
Record



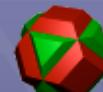
RECORD



Future Questions

Whether to have many subclasses of processes or fewer with annotations
 How to depict destructive processes
 How to depict taking lots of samples
 What is the observation/process boundary? e.g. MRI scan

Combechem
 30 January 2004
 gvh, hrm, gms



Name	Planned	Actual
Fluorinated biphenyl	0.9000 g	0.9031 g
Br11OCB	1.5900 g	1.5918 g
Potassium Carbonate	2.0700 g	2.0719 g
Butanone	40.0 ml	

Simple Interface

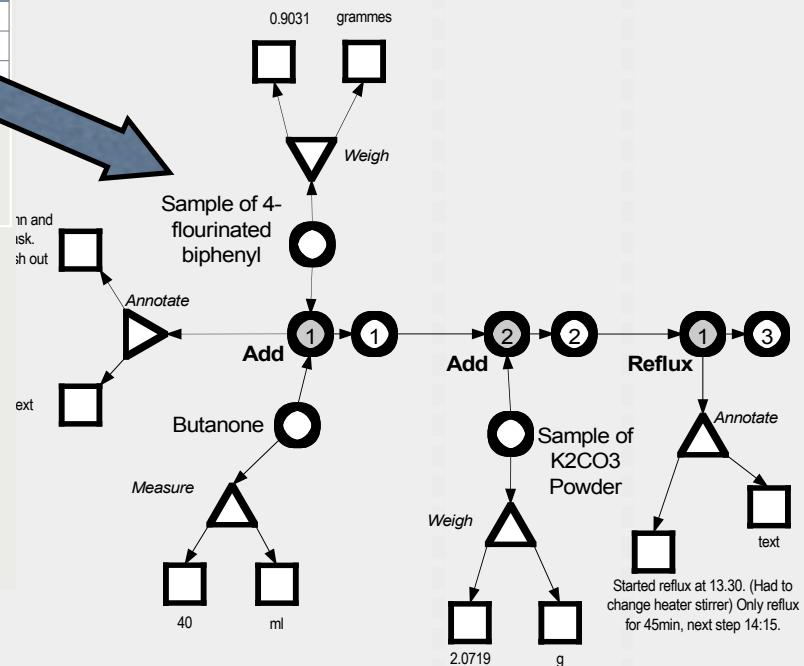
7	8	9
4	5	6
1	2	3
0	.	

Enter Del

Ingredient List

Fluorinated biphenyl	0.9 g
Br11OCB	1.59 g
Potassium Carbonate	2.07 g
Butanone	40 ml

Dissolve 4-flourinated biphenyl in butanone
Add K₂CO₃ powder
Heat at reflux for 1.5 hours



Smart Tea Project - User Centred Design, Design by Analogy to ensure the correct information is captured simply and easily.

29 Jan 2007



Jeremy G. Frey
University of Southampton

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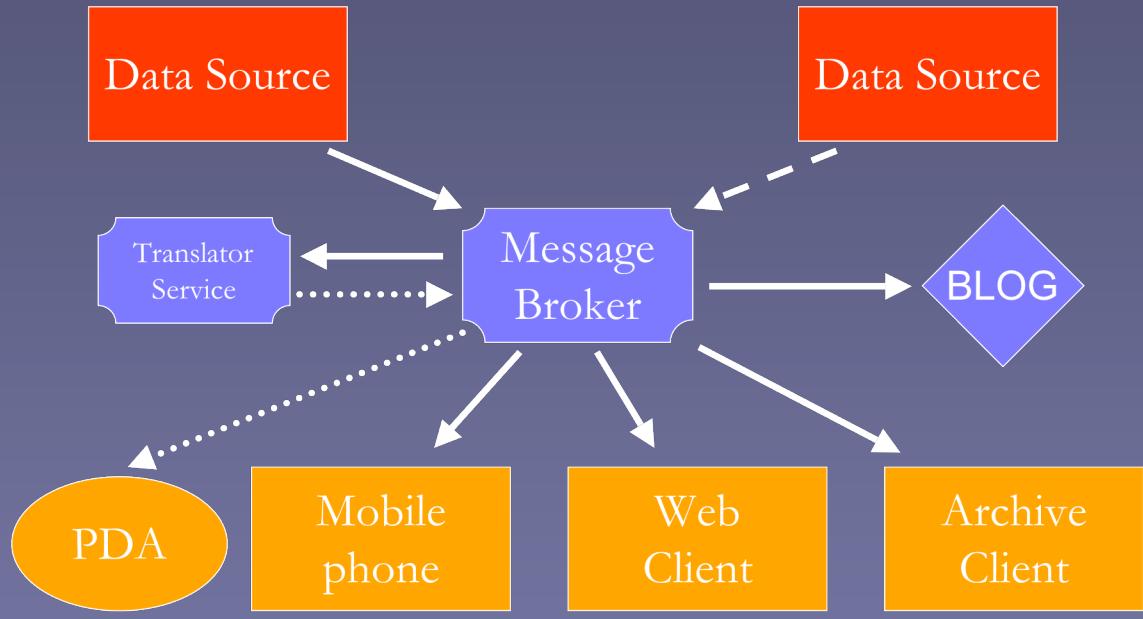


But what
about the
laboratory
environment?

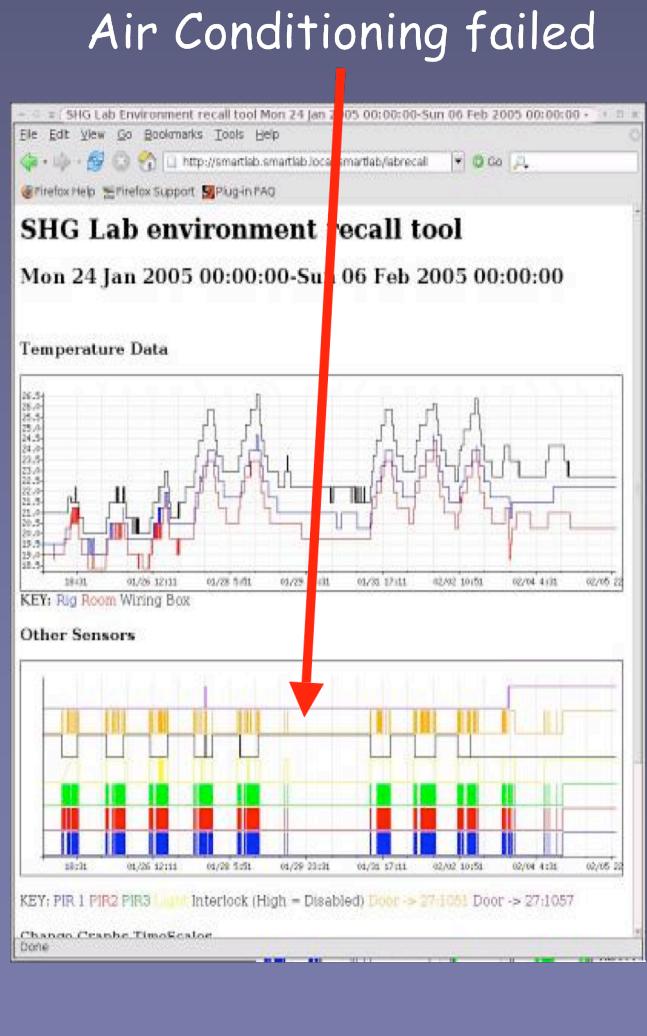


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From The New Yorker Book of Technology Cartoons.

Pub-Sub systems provide the flexible & extensible approach to distribution of real time laboratory monitoring & archiving



Smart Laboratory Spaces





Plans

- Plans in advance are useful
- This is the way things are supposed to be done
- The Plan provides a digital context so increases the value of planning
- Key to our ‘Smart Lab’ approach....
- But is it the best way?



He is charged with expressing contempt for meta-data

29 Jan 2007



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University of Southampton

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Laboratory “Blogs”

- Explore what is needed for a Blog to be the heart of an ELN
- Encourage and facilitate collaboration
- Need a data repository behind the Blog
 - R4L
 - E-Bank
- Building a VRE?



The 'Scientific Blog' is being tried in an attempt to combine laboratory notebooks and publication



Useful Chemistry - Microsoft Internet Explorer

File Edit View Favorites Tools Help

Address: <http://usefulchem.blogspot.com/>

The video ipod compatible podcast is available here.

Here it is on YouTube:

posted by Jean-Claude Bradley @ 4:30 PM 1 comments links to this post

WEDNESDAY, NOVEMBER 15, 2006

(11 items remaining)

start BBC NEWS | Politics | ... Document1 - Microsoft Internet Explorer Microsoft Internet Explorer

Note the use of
"YouTube"

usefulchem » Exp042 - Microsoft Internet Explorer

File Edit View Favorites Tools Help

Address: <http://usefulchem.wikispaces.com/Exp042>

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Actions

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Navigation

Home

Objective
To study formation of an imine from phenylacetaldehyde and t-butylamine

Procedure
CDCl₃ solutions of phenylacetaldehyde (240 μ L in 2 mL, 1 M) and t-butylamine (146 μ L in 2 mL, 0.7 M) are prepared in separate 1 dram vials. One mL of each solution are used to obtain initial H and C NMR spectra. The remaining 1 mL of each solution are mixed in a 1 dram vial and shaken vigorously. The resulting solution is transferred to an NMR tube and the reaction is monitored by H and C NMR.

Results

t-butylamine solution (BA)
H-NMR 1.27 (br s, NH₂), 1.15 (s, CH₃)

phenylacetaldehyde solution (PA)
H-NMR 9.71 (t, $J=2.3$ Hz, CHO, **0.65H**), 7.30 (m, 3H), 7.19 (d, $J=7.5$ Hz, 2H), 3.65 (d, $J=2.3$ Hz, CH₂, 2H)

42A 5 min
H-NMR new peaks 9.76 (t), 7.79 (d), 7.73 (d), 7.65 (t, $J=5.3$ Hz), 3.57 (d, $J=5.3$ Hz), 1.19 (s) and many other peaks in the 2-6 ppm region
From PA and BA: 9.73 (t), 7.1-7.4 (m, more than just PA), 3.67 (d), 1.18 (s)

42B 11 min
42C 28 min

Ads by Google
NMR
NMR Analytical Laboratories NMR Laboratory Network www.intertek-cb.com

Nmr Products
The UK Specialists in Stable Isotopes www.okgas.com

Mass:Spec MALDI Chips
On-target processing makes the most of your MALDI samples www.qiagen.com/protein

NMR Spectroscopy Info
On-Line and Analytical NMR Services Consulting and Analysis Process-NMR.Com

NIR Calibration On Demand
Continually built from your

An experiment that failed... Publishable?
Useful?

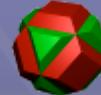


29 Jan 2007



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University of Southampton

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Southampton Experiment Blogs

- Example from a Bio-Organic Laboratory
 - Student (Jenny Hale)
 - Based in Southampton
 - Supervisor (Cameron Neylon)
 - Southampton only 1/3 time, RAL 2/3 time



Transformation of plasmid JRH4712/66 into BW25141 by electroporation

11th December 2006 @ 14:31

Transformations were set up according to the following protocol:

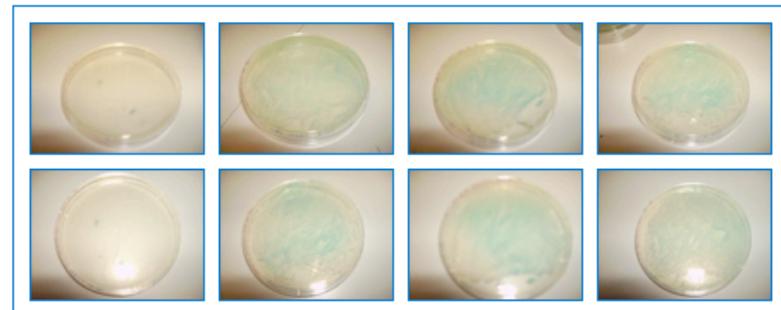
LB Ampicillin arabinose plates and SOC medium were warmed to 37 °C briefly before the arabinose plates were spread with X-glu (80 µL, 1:1 X-glu and LB) and allowed to continue warming.

BW25141 cells, plasmid JRH4712/66, p042, and electroporator cuvettes were cooled on ice. Items were added to the cuvettes as follows

-	1	+ve ctrl	-ve ctrl
BW25141	40 µL	40 µL	40 µL
plasmid 4712/66	4 µL	0 µL	0 µL
p042	0 µL	4 µL	0 µL

Cuvettes were electroporated at 1.75 kV, immediately had SOC medium (950 µL) added and the transformant transferred to eppendorf. The transformants were incubated at 37 °C for one hour with shaking. The transformants were diluted 1 in 20 with LB and 100 µL added to LB amp arabinose plates and incubated at 37 °C overnight.

Data



[Jennifer Hale](#) | [Beta-glucuronidase](#) | [Comments \(3\)](#)

Archives

[January 2007 \(24\)](#)
[December 2006 \(11\)](#)
[November 2006 \(5\)](#)

Sections

[beta-galactosidase preparation and assays \(18\)](#)
[Beta-glucuronidase \(18\)](#)
[Data \(Formatting\) \(1\)](#)
[Software discussions \(2\)](#)
[Starting materials and reagents \(1\)](#)

Lab Book Ref

[JRH4712-63 \(1\)](#)
[JRH4712-64 \(2\)](#)
[JRH4712-66 \(1\)](#)
[jrh4712-76 \(1\)](#)
[jrh4712-77 \(1\)](#)

Test digestions to check the activity of two batches of EcoRI and Ncol

22nd January 2007 @ 11:57

Lab Book Ref: [jrh4712-89](#)
 Sample Parent: [jrh4712-80_blue](#)
 Sample Parent2: [jrh4712-80_white](#)
 Digestions were set up as follows:

-	1	2	3	4	5	6	7	8	9	10	11
4712/80 blue	8 µL	-	-	8	-	-	-	8 µL	-	-	-
4712/80 white	-	8 µL	-	-	8 µL	-	-	-	8 µL	-	-
p042	-	-	5 µL	-	-	5 µL	5 µL	-	-	5 µL	5 µL
water	7.5 µL	7.5 µL	10.5 µL	7.5 µL	7.5 µL	10.5 µL	10 µL	7.5 µL	7.5 µL	10.5 µL	10 µL
EcoRI buffer 4	2 µL	2 µL	2 µL	-	-	-	2 µL	2 µL	2 µL	2 µL	2 µL
NEB buffer 2	-	-	-	2 µL	2 µL	2 µL	-	-	-	-	-
BSA	2 µL	2 µL	2 µL	2 µL	2 µL	2 µL	2 µL	2 µL	2 µL	2 µL	2 µL
EcoRI (a)	0.5 µL	0.5 µL	0.5 µL	-	-	-	0.5 µL	-	-	-	-
Ncol	-	-	-	0.5 µL	0.5 µL	0.5 µL	0.5 µL	-	-	-	0.5 µL
EcoRI (b)	-	-	-	-	-	-	-	0.5 µL	0.5 µL	0.5 µL	0.5 µL

*EcoRI (a) assay date 2/05
 EcoRI (b) assay date 7/05*

Digestions were incubated in a waterbath at 37 °C for 3 hours.

Archives

[January 2007 \(24\)](#)
[December 2006 \(11\)](#)
[November 2006 \(5\)](#)

Sections

[beta-galactosidase preparation and assays \(18\)](#)
[Beta-glucuronidase \(18\)](#)
[Data \(Formatting\) \(1\)](#)
[Software discussions \(2\)](#)
[Starting materials and reagents \(1\)](#)

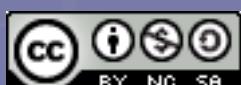
Lab Book Ref

[JRH4712-63 \(1\)](#)
[JRH4712-64 \(2\)](#)
[JRH4712-66 \(1\)](#)
[jrh4712-76 \(1\)](#)
[jrh4712-77 \(1\)](#)
[jrh4712-78 \(1\)](#)
[jrh4712-80 \(1\)](#)
[jrh4712-81 \(1\)](#)
[jrh4712-83 \(1\)](#)
[jrh4712-82 \(1\)](#)
[jrh4712-84 \(1\)](#)
[jrh4712-85 \(1\)](#)
[4712-88 \(1\)](#)
[jrh4712-89 \(1\)](#)
[4712-86 \(1\)](#)
[jrh4712-87 \(1\)](#)
[4712-90a \(1\)](#)

Product

[jrh4712-74 \(1\)](#)
[jrh4712-76 \(1\)](#)
[jrh4712-76a \(1\)](#)

29 Jan 2007



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 University of Southampton

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Mutagenesis of plasmid p042 via Taq and Genemorph1

9th January 2007 @ 14:34

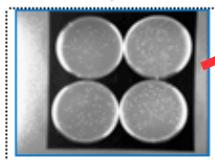
Lab Book Ref: jrh4712-78

Mutagenic PCR reactions were set up according to the following:

	Taq	Genemorph1	+ve ctrl	-ve ctrl
p042	2.5 µL	1 µL	5 µL	0 µL
Water	10 µL	38 µL	20.5 µL	25.5 µL
5 x GoTaq buffer	10 µL	-	10 µL	10 µL
mutazyme buffer	-	5 µL	-	-
Ordinary dNTPs	-	-	5 µL	5 µL
mutagenic dNTPs	5 µL	-	-	-
Mutazyme dNTP mix	-	1 µL	-	-
MgCl2	2.5 µL	-	3.5 µL	3.5 µL
MnCl2	12.5 µL	-	-	-
Primer fwd	2.5 µL	2 µL	2.5 µL	2.5 µL
Primer rev	2.5 µL	2 µL	2.5 µL	2.5 µL
GoTaq*	1 µL	-	1 µL	1 µL
Mutazyme	-	1 µL	-	-

*GoTaq=1 µL stock + 3 µL water.

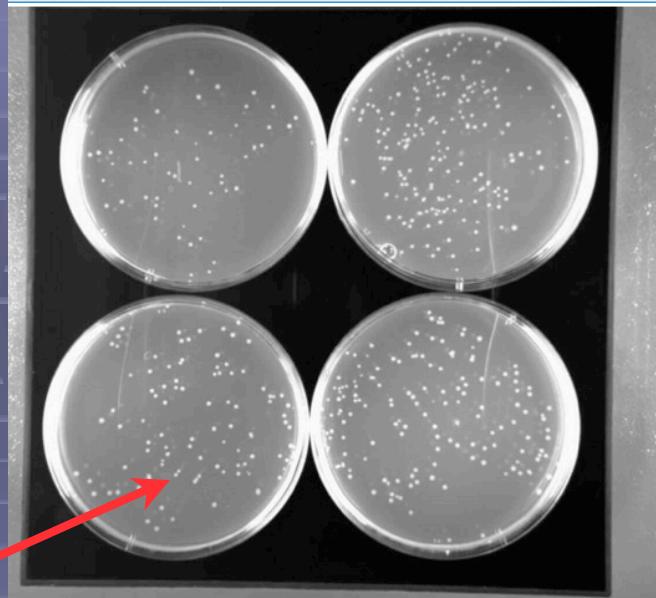
The reactions were run on program MUTAGB for 30 cycles. 5 µL product was run on a 1% normal agarose analytical gel. Taq mutagenesis had failed, Genemorph and +ve ctrl had worked. -ve control was correct.



Jennifer Hale | Beta-glucuronidase | Comments (3)

<< Previous Page

Next Page >>





Re: PCR of beta-galactosidase third attempt by Jennifer Hale

14th December 2006 @ 11:10

Unfortunately the purification appears not to have gone well. Though I also can't get any consistency from the figures given by the nano-drop. These are the results I got:

-	reading 1	reading 2	reading 3	reading 4	reading 5	reading 6	average
PCR product before*	282.3 ng/µL	283.4 ng/µL	281.1 ng/µL	N/A	N/A	N/A	282.3 ng/µL
PCR product after*	7.8 ng/µL	12.9 ng/µL	17.6 ng/µL	85.4 ng/µL	22.4 ng/µL	12.8 ng/µL	?

*Both reactions combined together after PCR

I'm going to do another PCR again. That step is working really well. I'm just not sure what to do about purifying it. The only other thing I can try is eluting in TE buffer rather than water (which it says you can also elute into)

In this purification I used preheated water and followed the instructions closely. Perhaps the DNA will elute into TE more effectively.

Re: PCR of beta-galactosidase third attempt by David Neylon

14th December 2006 @ 18:32

I would definitely compare these on a gel so as to see whether it is just the nanodrop that is the problem. It might help also if you are explicit about how much solution you are trying to purify and what the final volume is.

jrh4712-80_blue (2)

4712-86 (1)

4712-84_beta-gal (1)

4712-80_blue (1)

4712-88 (1)

Sample Parent2

jrh4712-80_blue (1)

jrh4712-80_white (2)

4712-84_pBad (1)

4712-80_white (1)

Sample Parent 3

4712-74 (1)

Search

Links

Admin

[New Post](#)

Live Copy



29 Jan 2007



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University of Southampton

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Re: Software development by Jennifer Hale

1st December 2006 @ 15:03

I'm not sure that I trust this thing. Just now there was an error message saying there were syntax errors in the mySQL. my post had appeared twice and my comment had disappeared. I close the window and then log back in and it all seems to be okay: the post appears once and my comment has reappeared but the post I wrote in response to the posts apparently disappearing and doubling up has disappeared.

Re: Software development by Andrew Milsted

1st December 2006 @ 17:18

I am working on the session problem, I have looked at the long list and most of the points are on my todo list. I will let you know when things become available. If you are writing long posts (I know this sounds crazy) but use word and copy and paste into the blog, once I have fixed the sessions this won't be needed.

Re: Software development by Jennifer Hale

5th December 2006 @ 11:19

My gel pictures when seen in close up are awful quality - very pixelated. It doesn't show a clear picture at all. Should I try saving the files as a different file type as they are currently a jpeg?

The thing I don't understand the most though is that if I click on the file on my computer and it opens up in photo editor or viewer or whatever it is, the picture is fine even on a large scale - really clear.



Instrument Blog

[Login](#) [more blogs](#)

MQTT Lego Microscope

A highly advanced remote control microscope.

Data Collection

15th September 2006 @ 16:52

A data collection was made by Andrew Milsted (ajm3) with sample description: Paper Clip

Data

The image shows a close-up of a paper clip under a microscope. To the right is a graph with a red peak. The x-axis is labeled 'x=21.661538461538,y=117' and has values 0, 32, 64, 96, 128, 160, 192, 224, 256. The y-axis ranges from 0 to 700,000. A green vertical line is at x=21.661538461538.

ajm3 | [Data Collection](#) | [Comments \(0\)](#)

Archives
[September 2006 \(2\)](#)

Sections
[Data Collection \(2\)](#)

Search

Links

‘Blog-jects’

29 Jan 2007



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University of Southampton

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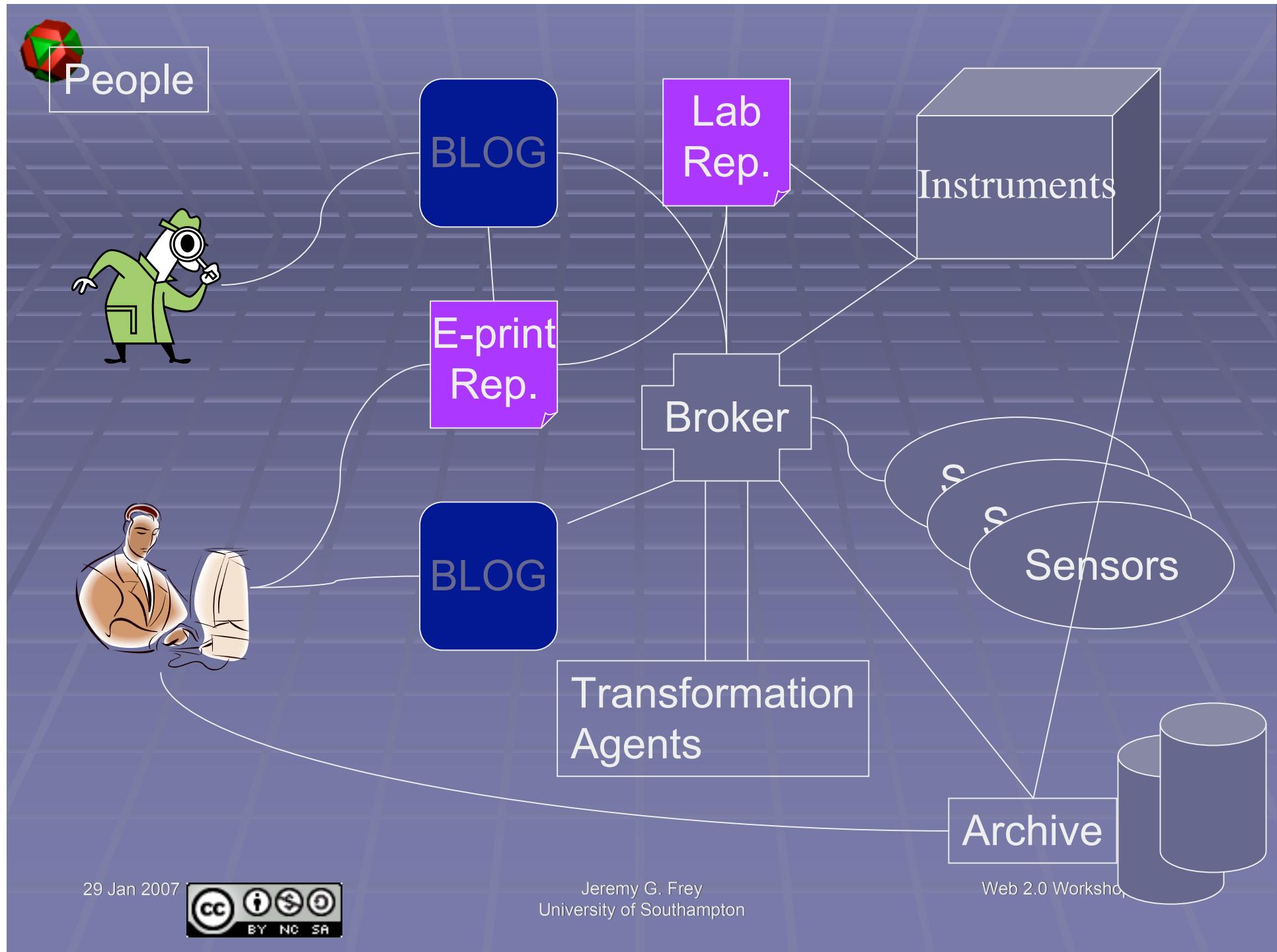
Pub-Sub

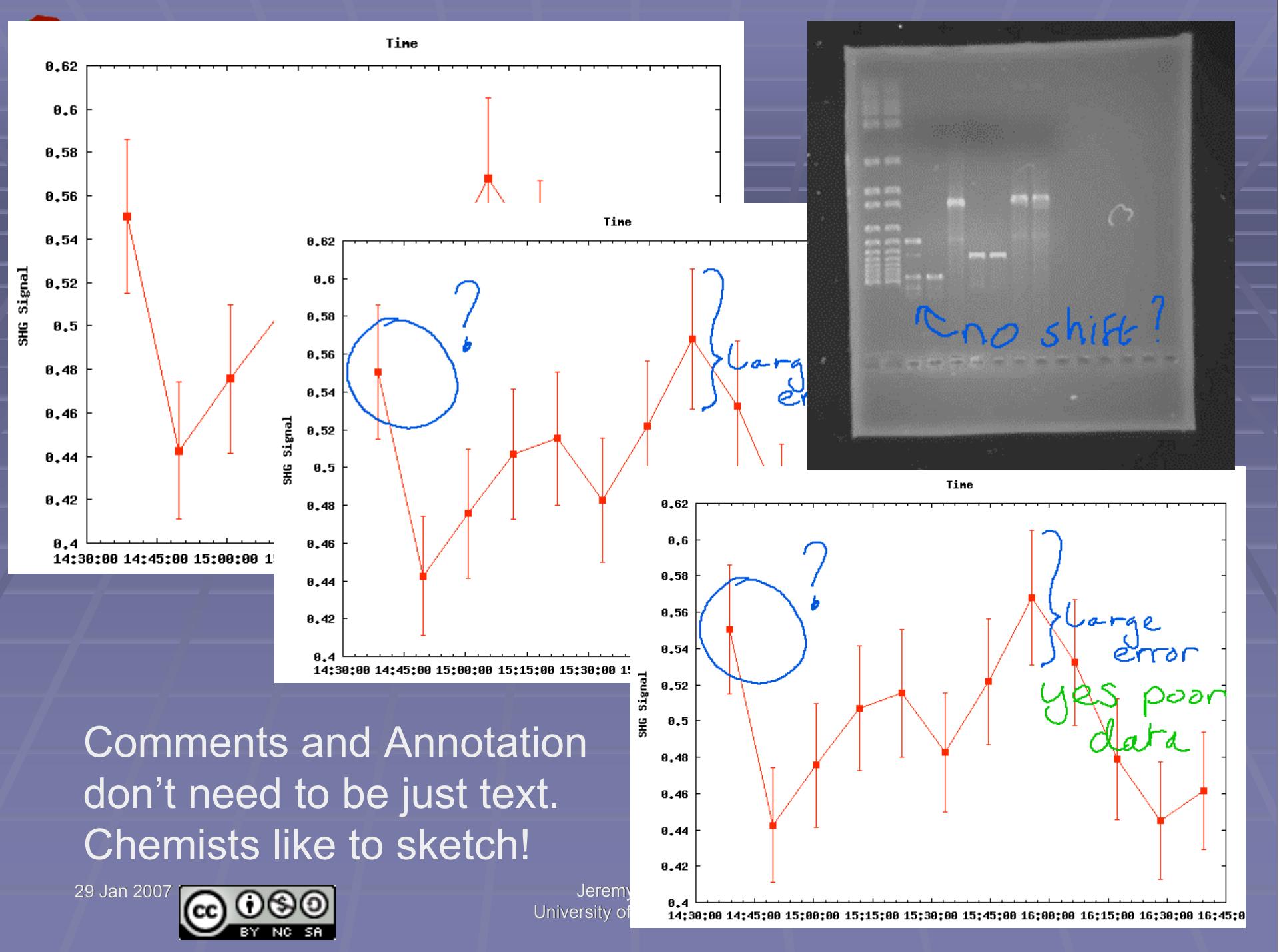
- Pub Sub & Data Brokers
 - Important way to include laboratory environmental & people data in a flexible manner
 - Backed by archive database service
 - Correlate with experiments via time & place



Blog-jects

- Equipment become first class members of the web
- Interacts well with Pub-Sub as items are attached to topics, topics relate the Bog items
- With automation this evolves to a two-way communication
- Live Copy essential







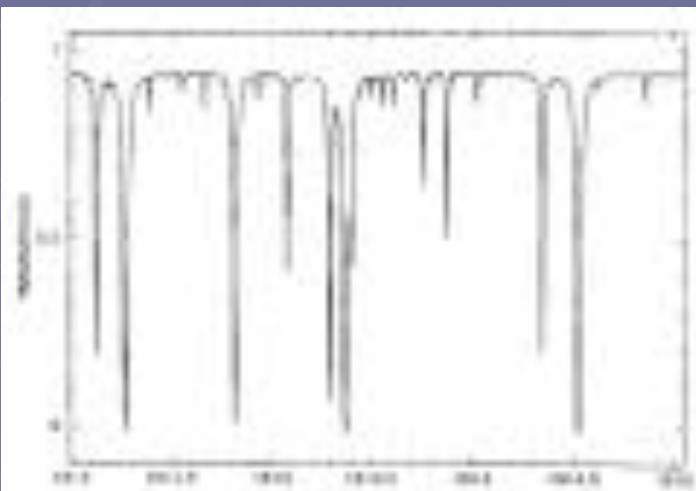
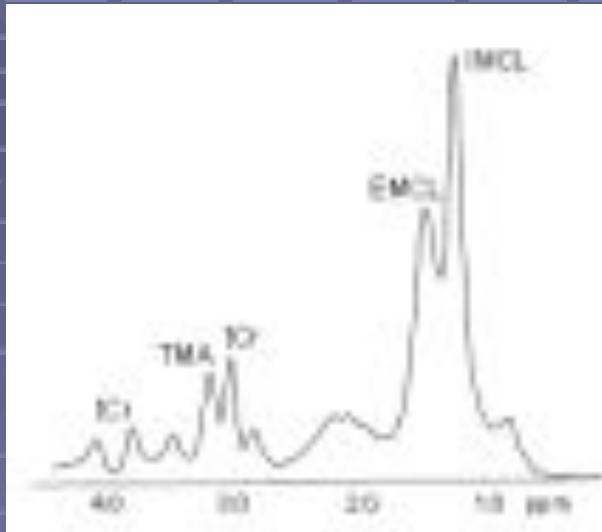
Validation

- Increasing the value of data
- How to bring all the necessary information together to enable appropriate validation
- Increasingly difficult & expensive to achieve
- Need provenance and context
- Essential step otherwise just a collection of items



Why?

Publishing Data and Information Loss



Jeremy G. Frey
University of Southampton

Web 2.0 Workshop OGF 19

Preparation of $\text{CPh}_3[\text{C}_6\text{F}_5]_3\text{BCNPBB}$ (2)

$\text{Me}_3\text{SiNCB}(\text{C}_6\text{F}_5)_3$ (0.51 g, 0.84 mmol) and Ph_3CCl (0.23 g, 0.84 mmol) were stirred in 20 mL of dichloromethane for 0.5 h to give a yellow solution. After removal of volatiles *in vacuo*, the residue was washed with pentane (30 mL), PBB (0.81 g, 0.84 mmol) and dichloromethane (30 mL) were added, and the mixture was stirred for 2 h. The solvent was then removed. The product was washed again with 30 mL of pentane and dried *in vacuo* to yield a yellow-orange powder (yield 1.01 g, 5.8 mmol, 69%). Attempts to crystallise the product from dichloromethane were not successful. IR (nujol): 2284 cm^{-1} ($\nu_{\text{C=C}}$). $^1\text{H NMR}$ (CD_2Cl_2 , 20 °C, 300.13 MHz): δ 8.56 (t, 3, $J = 8.0\text{ Hz}$ *p-Ph*), 7.90 (t, 6 H, $J = 7.5\text{ Hz}$, *m-Ph*), 7.70 (d, 6 H, $J = 7.2\text{ Hz}$, *o-Ph*). $^{13}\text{C NMR}$ (CD_2Cl_2 , 20 °C, 75.48 MHz): δ 211.0 (CPh_3), 144.1 (*p-C*), 143.0 (*m-C*), 140.1 (*ipso-C*), 130.9 (*o-C*). $^{11}\text{B NMR}$ (CD_2Cl_2 , 20 °C, 96.3 MHz): δ -4.35 (s, br, 1 B, $\text{N}(\text{C}_6\text{F}_5)_3$). $^{19}\text{F NMR}$ (CD_2Cl_2 , 20 °C, 282.4 MHz): δ -18.72 (br, s, 1 F), -120.22 (br, s, 1 F), -121.99 (br, s, 1 F), -122.50 (s, 1 F), -132.20 (s, 1 F), -133.94 (br, 6 F, *o-F* on $\text{B}(\text{C}_6\text{F}_5)_3$), overlapping signals (-134.15, -134.39, -134.95, -135.27, -135.64), 136.89 (br, 1 F), -137.81 (br, 3 F), -138.79 (d, 1 F), -144.73 (t, 1 F), -149.78 (t, 1 F), -151.11 (t, 1 F), -154.65 (t, 1 F), -154.93 (t, 1 F), -155.32 (t, 1 F), -156.86 (t, 1 F), -157.24 (t, 1 F), -157.55 (t, 1 F), -158.29 (t, 1 F), -158.90 (t, 1 F), -159.57 (t, 3 F, $J = 20\text{ Hz}$, *p-F* on $\text{B}(\text{C}_6\text{F}_5)_3$), -159.98 (t, 1 F), -161.44 (br, 2 F), -164.0 to -164.4 (overlapping signals, 3 F), -165.33 (br, 2 F), -166.12 (t, 3 F, $J = 20\text{ Hz}$, *m-F* on $\text{B}(\text{C}_6\text{F}_5)_3$). Anal. Calcd for $\text{C}_{24}\text{H}_{15}\text{B}_2\text{F}_{12}$:

Done

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Separating Data from Interpretations: A crystallography example

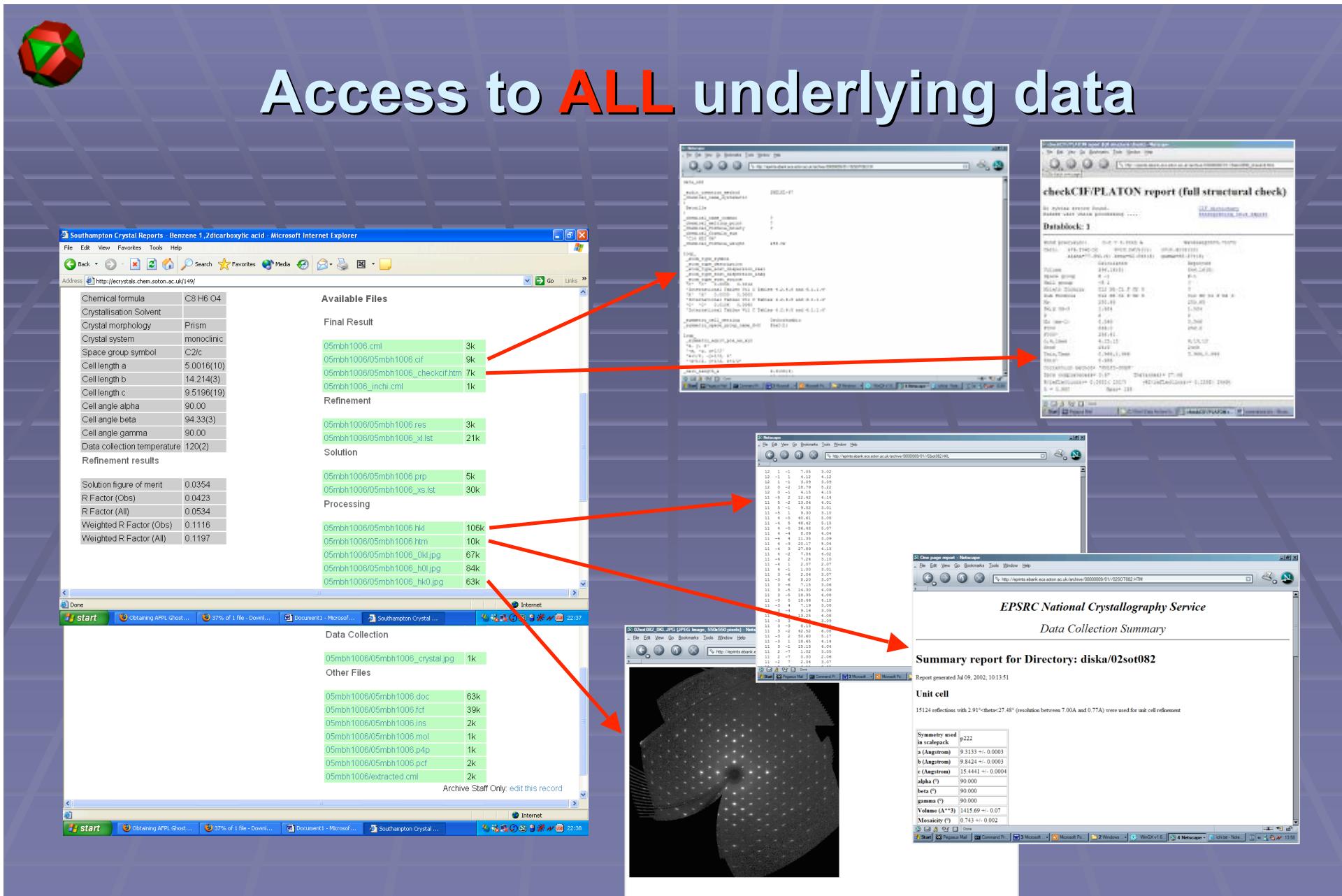
Underlying data

Intellect & Interpretation

Jeremy G. Frey
University of Southampton

<img alt="Screenshot of a computer screen showing a software interface for crystallography data entry. The window is titled 'e-Prints Soton' and shows a table of experimental details for a crystal structure. The table includes columns for crystal data, chemical formulas, cell dimensions, temperature, and other parameters. The data is

Access to **ALL** underlying data



29 Jan 2007

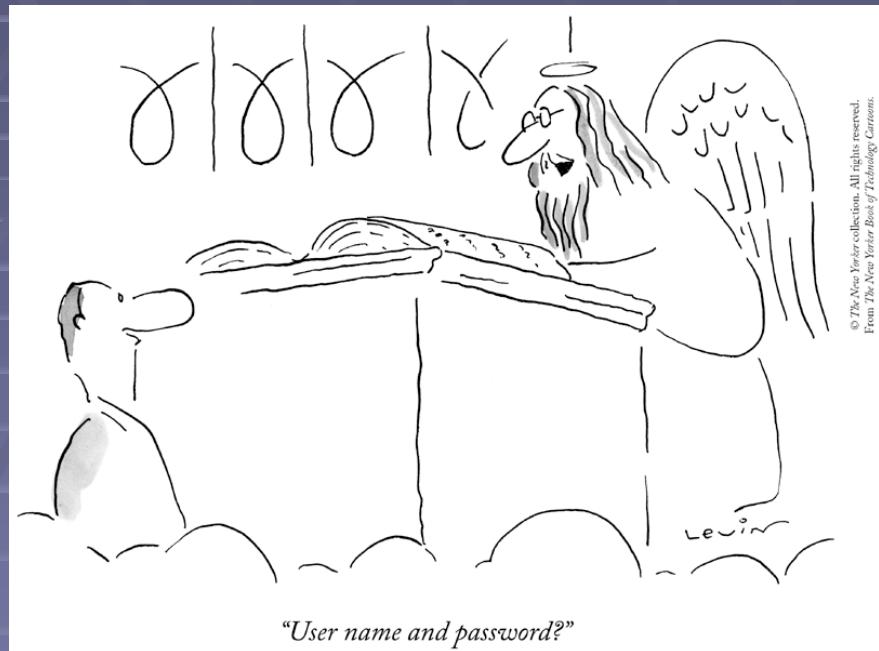


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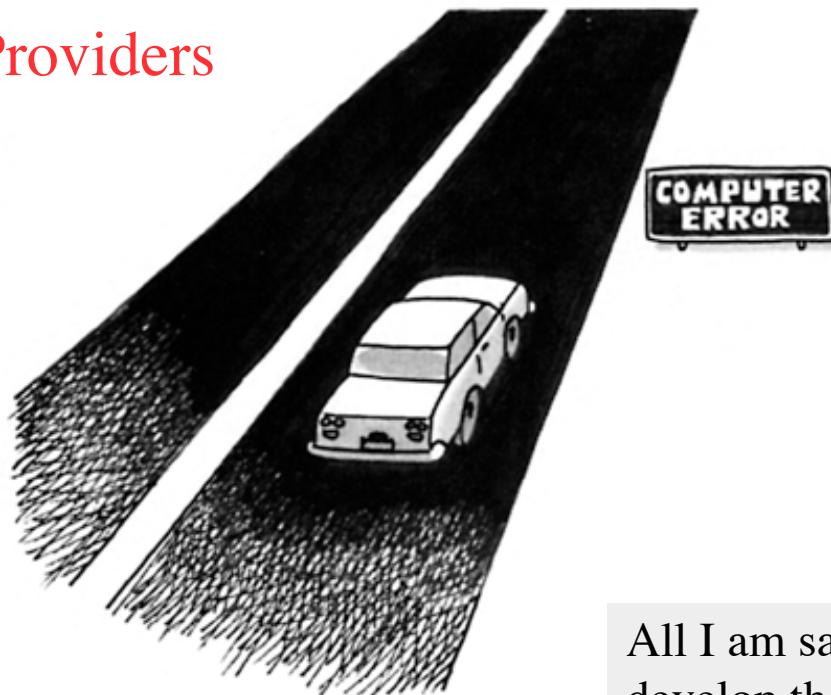
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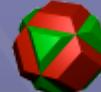


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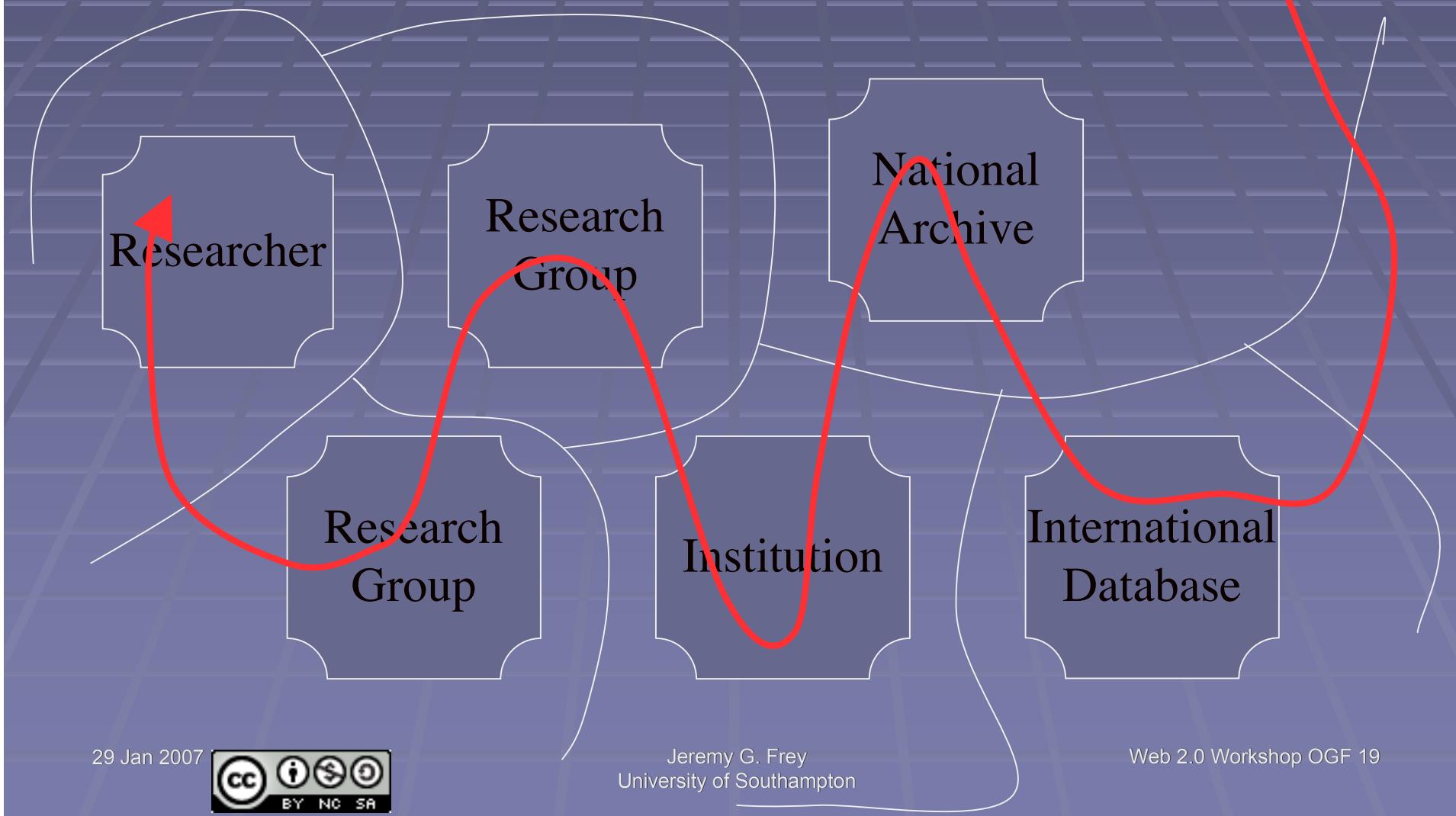


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All I am saying is that now is the time to develop the technology to deflect an asteroid



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