1 Title: Characterising the nutritional status of children with primary ciliary dyskinesia 2 Authors: Marino LV<sup>1,2</sup>, Harris A<sup>3</sup>, Johnstone C<sup>1</sup>, Friend A<sup>3</sup>, Newell C<sup>2</sup>, Miles EA<sup>4</sup>, Lucas JS<sup>2,3,5</sup>, 3 Calder PC<sup>2,4</sup>, Walker WT<sup>2,3,5</sup> 4 5 6 Affiliations: Department of Dietetics/Speech and Language Therapy, University Hospital Southampton NHS Foundation Trust, Southampton, UK<sup>1</sup>, NIHR Southampton Biomedical 7 8 Research Centre, University Hospital Southampton NHS Foundation Trust and University of Southampton, Southampton, UK<sup>2</sup>, Primary Ciliary Dyskinesia Centre, University Hospital 9 Southampton NHS Foundation Trust, Southampton, UK<sup>3</sup>, Human Development & Health 10 Academic Unit, Faculty of Medicine, University of Southampton, Southampton, UK<sup>4</sup>, Clinical and 11 12 Experimental Sciences Academic Unit, Faculty of Medicine, University of Southampton, Southampton, UK<sup>5</sup>. 13 14 15 Keywords: nutrition, primary ciliary dyskinesia, bioelectrical impendence, micronutrients 16 17 Corresponding author: Luise Marino, University Hospital Southampton NHS Foundation Trust, Southampton, UK S016 6YD Tel: + 44 (0) 23 8079 6000 Email: Luise.Marino@uhs.nhs.uk 18 19

#### Abstract

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21 Introduction: Primary ciliary dyskinesia (PCD) is a rare, heterogeneous genetic disorder where 22 impaired mucociliary clearance is caused by dysfunctional motile cilia leading to bronchiectasis. 23 There is limited evidence characterising the nutritional status of children with PCD, although 24 lower body mass index (BMI) z-score has been associated with worse lung function (FEV<sub>1</sub>). 25 Methods: All children (n=43) with PCD, aged <16 years, from a single tertiary centre were 26 prospectively enrolled. Information on clinical phenotype and nutritional status including 27 bioelectrical impedance spectroscopy (BIS) phase-angle was collected. 28 Results: There was a weak positive association between height-for-age z-score (HAZ) and FEV<sub>1</sub> z-29 score (n=28, r=0.4, p=0.049). Those with a low fat free mass index (<-2 z scores) had a lower BMI z score (-1.3±1.2 vs. 0.8±0.7, p=0.0002). BIS phase angle identified more patients at nutritional 30 31 risk than using moderate malnutrition cut-offs of either HAZ or BMI ≤-2 z scores alone (21% vs. 32 4.6% vs. 6.9% respectively). PCD patients had a higher incidence of vitamin D insufficiency (<50 33 nmol/L) (54%) and deficiency (<30 nmol/L) (26%) than healthy children. 34 Conclusions We have characterised the nutritional phenotype of a cohort of children with PCD. 35 Monitoring vitamin D levels is important in PCD patients. There is a weak association between 36 lung function and nutritional status, and measures of BIS phase-angle. The use of BIS phase-37 angle may allow for early identification of at risk children and may therefore be of benefit for 38 nutritional assessments in the clinical setting. These findings will help inform a future nutritional 39 intervention strategy in children with PCD.

#### What we know:

- There is a positive association between lung function and body mass index in PCD.
- A low bioelectrical impendence spectroscopy phase angle is associated with poorer nutritional status, which may precede anthropometric changes and impact on clinical outcomes.

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# What this study adds:

- In children with PCD, there were weak associations between BIS phase angle, nutritional status and clinical outcomes
- In PCD, the routine use of BIS phase angle may add a sensitivity and specificity in identifying children with nutritional risk earlier than using anthropometry alone
- Vitamin D insufficiency may be more common amongst children with PCD

#### Introduction

Primary ciliary dyskinesia (PCD) is a rare, heterogeneous genetic disorder where impaired mucociliary clearance is caused by dysfunctional motile cilia. Infants with PCD usually have neonatal respiratory distress, and they go on to have persistent sinopulmonary infections, which leads on to bronchiectasis in over half of children and almost all adults [1]. Historically PCD has been considered a fairly mild respiratory condition; however, lung function (force expiratory volume on one second (FEV<sub>1</sub>)) has recently been shown to decline at a similar rate to that seen in patients with cystic fibrosis (CF) throughout childhood [2]. Despite their recurrent sinopulmonary infections, no biomarkers of chronic inflammation have been identified in PCD patients to date. Further to sinus and lung involvement, the majority of PCD patients have recurrent glue ear with conductive hearing loss, approximately half have *situs inversus totalis*, with a proportion having complex congenital heart disease; most men are infertile due to immotile sperm and gastrointestinal, renal and neurological sequelae have been described [1].

Until recently there has been a paucity of evidence characterising the nutritional status of children with PCD. However, one large international cross-sectional study has recently demonstrated lower height-for-age z-scores (HAZ) in PCD patients [3], with another showing a progressive longitudinal decline in linear growth throughout childhood resulting in a mean loss of body height of approximately 0.5–1 standard deviations by the age of 17 years [4]. The cause of this growth failure is likely to be multifactorial [4, 5] and may be as a result of increased respiratory effort, possible effects of chronic inflammation and suboptimal nutrition intake, although to our knowledge this has yet to be described. It is therefore important to use nutritional measurements that both identify accurately and, where possible, early those children at nutritional risk who would benefit from nutritional intervention.

In CF, it is well-established that patients with a lower body mass index z score (BMIZ) have poorer respiratory outcomes (specifically FEV<sub>1</sub>) [6]. This has led to intensive dietetic input into this cohort of patients and is a key factor in the improvements in life expectancy and clinical outcome measures seen in these patients over recent decades [7]. As a result, a body mass index (BMI)  $\leq$  0.67 z score ( $\leq$ 25<sup>th</sup> percentile) is generally used as a marker for children at nutritional risk [8]. However, in more recent years, the use of BMI as a stand-alone measure has been questioned as it does not distinguish between fat mass (i.e. adipose tissue) or fat free mass (i.e. lean body mass). As a result, children may have persistent chronic malnutrition (i.e. be short) but have a normal BMI as their weight to height ratio is normal. As such, utilising other methods of body composition analysis, such as bioelectrical impedance spectroscopy (BIS), may add a sensitivity and specificity to nutritional assessment over the use of BMI alone [9], identifying those children with declining nutritional status who may benefit from early nutritional support [10].

BIS measurements have been used in a variety of settings to quantify body composition [11]. BIS uses a weak, alternating electric current at a range of radiofrequencies to characterise the conductive and non-conductive tissue and fluid components of the body [12]. BIS phase angle depends on the opposition to the flow of electrical current (resistance) and the effect of the capacitive ability of cell membranes to impede the current (reactance) [13, 14]. A low BIS phase angle 50° has been shown to predict clinical outcomes relating to morbidity and mortality, in critical care, renal disease and HIV/AIDS [15-17]. More recently BIS phase angle 200/5° has been shown to be a marker of clinical outcomes and nutritional status in critically ill adults [18]. Furthermore, in this setting, BIS phase angle 200/5° also appears to be a measure of functional outcome, e.g. with lower BIS phase angle values seen in patients with poor handgrip strength [18, 19]. In our work in children with congenital heart disease we have demonstrated a

relationship between pre-operative BIS phase angle 200/5<sup>0</sup> measures and post-operative and intensive care unit length of stay [20, 21]. These results suggest that BIS phase angle 50<sup>0</sup> and 200/5<sup>0</sup> may be useful in identifying changes to cellular resistance, alterations in cell membrane integrity and total body water which occur independently of and precede anthropometric changes in children [21-23]. BIS phase angle is increasingly being considered as a useful surrogate measure of overall cellular health and resilience, in addition to being a marker of nutritional status [21].

The aims of this study were to consider whether there were any associations between nutritional status and lung function in a cohort of children with PCD; to consider whether other measures, such as BIS phase angle 50° and 200/5°, might increase the sensitivity and specificity of identifying those patients at nutritional risk; to look for biomarkers of chronic inflammation in a PCD cohort; and finally, to measure a wide range of micro-nutrients, fatty acids and vitamins, in order to identify specific deficiencies that would need to be corrected in subsequent stratified nutritional interventions in this cohort of children.

#### **Materials and Methods**

The study was approved by the National Research Ethics Service (South Central (A) Committee 07/Q1702/109 ) and all participants provided written informed consent.

#### **Participants**

All children with PCD, aged between 0 and 16 years, from a single tertiary centre,
University Hospital Southampton (UHS) NHS Foundation Trust (Southampton, UK), seen in
outpatients between September 2016 and April 2017 were prospectively enrolled. The diagnosis
of PCD had been confirmed according to the ERS consensus guidelines [24]. Data on clinical
phenotype, anthropometry, BIS phase angle, and nutritional intake and blood samples were
collected during a single study visit.

#### **Spirometry measurement**

Spirometry measurements were performed using CareFusion Microlab 3500 Mk8 (CareFusion, California, USA) following calibration, according to the manufacturers' instructions. All measurements were obtained according to ERS/ATS guidance [25]. Global lung initiative (GLI) equations were used to estimate z scores for FEV<sub>1</sub> and forced vital capacity (FVC); ethnicity specific equations were used where available [26].

#### **Anthropometry**

Anthropometric measurements were performed and recorded in accordance with World Health Organisation (WHO) guidelines [27]. Infants aged ≤1 year were weighed naked and children aged ≥1 year with minimal clothing; weight was measured to the nearest 0.1 kg using a digital scale. Recumbent length was measured to the nearest 0.1 cm for all children aged ≤2 years using an infantometer (Seca 416; Birmingham, UK) and standing in older children under a stadiometer (Seca 213: Birmingham, UK). Reference values for fat free mass index (FFMI) were used to quantify z scores for children over the age of 2 years [28]. Z-scores were calculated using WHO Anthro software version 3.3.3 2011 [29] for participants aged ≤5 years and WHO AnthroPlus 3.2 [30] for those aged ≥5 years. WHO growth reference interpretation of cut offs for malnutrition were used. Moderate malnutrition was defined as a height-for-age, weight for height, BMI or FFMI of ≤-2 z-scores below the mean of the WHO child growth standards [27].

### Bioelectrical impedance spectroscopy measurements

BIS measurements were made using ImpediMedSFB7 (Pinkenba, QLD 4008 Australia), a single-channel tetra-polar device able to measure resistance and reactance across 256 frequencies. The machine was calibrated before use with a circuit of known impedance provided by the manufacturer. Measurements were conducted using a standard tetrapolar electrodes distribution; the inner arm electrode (sensor) was placed on the dorsal surface of the right wrist

and the leg electrode was placed on the anterior surface of the right ankle. Measurements were completed in unfasted subjects. Data files were processed using specialist software (Bioimp, ImpediMed), with data points rejected if they met any of the following criteria; i) positive X centre (Xc) values, ii) negative resistance values. Measurements were taken in triplicate and the mean used. BIS phase angle at a current frequency of 5°, 50°, 200° and 200/5° were analysed [31].

#### **Nutritional intake data collection**

A 3-day estimated food diary (2 weekdays and 1 weekend day) was recorded by carers/
children following the clinical appointment. Carers were provided with detailed instructions on
how to complete the diary accurately and were requested to return the diary via a prepaid
postal envelope. Nutritional intake data were assessed using CompEat Pro (Visual Informatics
Systems Ltd., Oxon, UK). Dietary intake for energy and protein were compared to the UK Dietary
Reference Values using the reference nutrient intake (RNI) for protein and estimated average
requirements (EARs) for energy [14]. As recommended by the Scientific Advisory Committee on
Nutrition in the United Kingdom (SACN), insufficient protein was defined as an intake <100 % of
the lower reference nutrient intake (LRNI—meeting nutrient requirements for 2.5 % of
population), sufficient intake was between the LRNI 100% and ≤200 % of the RNI and excessive
intake ≥200 % of the RNI [14]. For energy intake, the RNI was not used because it signifies an
excess energy intake for the majority of the population, as highlighted by the SACN [14]. As
advised by SACN, the EARs were used with children consuming ≤ 67 % classified as low energy
intake, between 67% EAR and 110 % as sufficient intake and excessive intake ≥ 110 % of the EAR
[15].

#### **Blood analysis**

Using standard laboratory techniques, routine clinical variables of interest were measured including C-reactive protein (CRP), selenium, zinc, copper, folate, vitamin B12 and vitamin D. Vitamin B6 concentrations were measured by high performance liquid chromatography with fluorescence detection as previously described [32]. Erythrocyte fatty acid levels were measured using a Hewlett Packard 6890 gas chromatograph (Hewlett-Packard; Avondale, PA, USA), as previously described [33]. Fatty acids are expressed as weight % of total fatty acids present. Plasma concentrations of interleukin 1 beta (IL-1 $\beta$ ), interleukin 2 (IL-2), interleukin 6 (IL-6), interleukin 8 (IL-8), tumour necrosis factor alpha (TNF- $\alpha$ ), and vascular endothelial growth factor 1 (VEGF-1) were measured using the FCSTM09-06 high sensitivity magnetic bead cytokine panel kit (Bio-Techne, R&D Systems Luminex Performance, Abingdon, UK). Plasma concentrations of prostaglandin  $E_2$  (PGE<sub>2</sub>), 6-keto prostaglandin  $F_1$  alpha (6-KPGF<sub>1 $\alpha$ </sub>) and 8-isoprostane were measured by ELISA (Cayman Chemical, Cambridge Biosciences, Cambridge UK). For all kits the manufacturer's instructions were followed and the plates were read on a Bioplex 200 (Bio-Rad, Watford, UK).

# Statistical analysis

An a priori statistical analysis plan was determined. SPSS version 24 (Chicago, IL) was used for conducting statistical analysis. Associations between BIS phase angle, anthropometry and clinical outcomes of  $FEV_1$  z scores and FVC z scores were investigated. Relationships between inflammatory mediators, markers of oxidative stress, nutritional intake and plasma/serum levels were also investigated. Data was tested for normality. For data that was normally distributed parametric tests were used, otherwise non-parametric tests were chosen. T-tests and one-way ANOVA with Dunn post test comparison were used to investigate differences between continuous variables of anthropometry, phase angle and clinical outcomes  $FEV_1$  and FVC z scores. Relationships between variables of interest were further investigated

using Spearman's correlations considering anthropometry, BIS phase angle clinical outcomes  $FEV_1$  and FVC z scores. A p value of <0.05 was used to define statistical significance. Where values were found to be normally distributed they are shown as mean and standard deviation or otherwise as median and inter-quartile range.

#### **Results**

43 children were included in this study; of these 51% (n=22) were male. The average age at diagnosis was 2.7±3.8 years and at the time of study was 7.0±3.6 years. Patient demographics, nutritional status, spirometry values and blood results are described in Table 1.

# **Nutritional status and lung function**

FEV $_1$  z scores were available in children older than 6 years of age (n=28). There was a weak positive correlation between HAZ and FEV $_1$  z score (n=28, r=0.4, p=0.049). There was also an association between BMIZ and FEV $_1$  but this was not statistically significant (Figure 1a and b). There was a positive association with HAZ and FEV $_1$  z score; children children with a low FEV $_1$  z score of <-2 were also shorter compared to >-2 (mean HAZ -0.49±1.1 vs 0.2±0.7, p=0.05).

#### Early detection of nutrition depletion by BIS phase angle

Only 4.6% (n=2) of children had HAZ <-2 z scores and 6.9% (n=3) of children had a BMI <-2 z scores. There were 29 children, older than 2 years of age, for whom BIS phase angle data were available to calculate FFMI; of these, 21% (n=6) had a FFMI <-2 z scores (Table 3). Children with a FFMI of < -2 z scores had a lower phase angle  $5^{\circ}$  (p=0.03) and phase angle  $50^{\circ}$  (p=0.0002) compared to those with a FFMI of > -2 z scores (n=23) (Figure 2). Those with a low FFMI (<-2 z scores) had a significantly lower FVC z score (-1.5±1.0 vs. 0.3±1.3 (p=0.01)) and a lower BMI z score (-1.3±1.2 vs. 0.8±0.7 (p=0.0002)) (Table 3).

#### Biomarkers of chronic inflammation

All inflammatory and oxidative stress markers were detectable in all samples available (n=35; Figure 3). Mean plasma concentrations of the pro-inflammatory cytokines assessed (IL- $1\beta$ , IL-2, IL-6, IL-8 and TNF- $\alpha$ ) were not raised compared to available normative data [34, 35] (Figure 3). There were weak positive associations between VEGF-1 and FEV<sub>1</sub> z score (n=27, r=0.41, p=0.035), VEGF-1 and BMI z score (n=35, r=0.48, p=0.001), IL-2 and BIS phase angle 50° (n=35, r=0.38, p=-0.026), and IL-2 and BIS phase angle 200/5° (n=35, r=0.37, p=-0.031).

Red blood cells contained a mean of 14.2% linoleic acid (an essential fatty acid), 13.3% arachidonic acid, 0.5% eicosapentaenoic acid and 2.8% docosahexaenoic acid (Table 4). There were weak positive correlations between 6-keto PGF<sub>1 $\alpha$ </sub> and erythrocyte 18:1n-7 (n=35, r= 0.26, p=0.03) and 18:2n-6 (n=35, r=0.39, p=0.02). There were weak positive associations between PGE<sub>2</sub> and erythrocyte 14:0 (n=22, r=0.58, p=0.005), 16:0 (n=22, r=0.58, p=0.005), 18:1n-7 (n=22, r=0.4, p=0.02), 20:0 (n=22, r=0.39, p=0.03), 20:1n-9 (n=22, r=0.36, p=0.04), 22: 6n-3 (n=22, r=0.4, p=0.02) and 24:0 (n=22, r=0.37, p=0.04).

# Dietary intake, fatty acids, vitamins and micronutrients

None of the children studied had a low energy intake (defined as consuming less than 67 % of EAR) but 63% (n=14) had excessive intake (defined as consuming greater than 110% of EAR). 6% of children consumed inadequate amounts of protein ( $\leq$ 100 % of RNI); 22% of children consumed adequate amounts of protein (LRNI to  $\leq$ 200 % of the RNI) and 72% had an intake of protein  $\geq$ 200 % of the RNI [15]. There were no associations between energy and protein intake, with respect to BMI, height for age or BIS phase angle (Table 5). Whilst there were no statistically significant differences between the groups, FEV<sub>1</sub> z score was higher in those with a protein intake  $\geq$ 200% of RNI compared to those children with an adequate intake of protein (LRNI to  $\leq$ 200% RNI) (-2.5±1.4 vs -1.4±1.1, p=0.2).

Vitamin D insufficiency (<50 nmol/l) was present in 54% (19/35) of those in the cohort where plasma concentrations were available, of which 26% (n=9/35) were deficient (<30 nmol/l), requiring therapeutic supplementation. A weakly significant association was seen between low vitamin D status and FFMI z score (n=29, r=0.4, p=0.02). All other vitamin and micronutrient levels assessed were considered normal.

#### Discussion

We found a weak correlation between low HAZ and BMIZ and FEV<sub>1</sub> z score. Children with a lower FEV<sub>1</sub> z score were also more likely to be shorter, which was consistent with findings published in a large international cohort of children with PCD [24], and as such may be clinically meaningful. BIS phase angle  $50^{\circ}$  was able to identify children with declining nutritional status earlier than using standard measures of anthropometry alone e.g. HAZ and BMIZ  $\leq$ -2. Children in our cohort commonly had insufficient vitamin D levels but all other micronutrient levels were within the normal range. However, despite recruiting all children seen in the National PCD Management Service at our centre over a 6-month recruitment window, due to the rarity of PCD, only 43 children were enrolled, and, as such, our results should be interpreted with caution.

The observation that poorer nutritional status is associated with poorer lung function in PCD patients, although weak, is similar to that seen in patients with CF [7, 36], with a relationship described between FEV $_1$  and BMI [3, 37]. However in CF the main nutritional issues are thought to be due to malabsorption secondary to pancreatic insufficiency and PCD patients have normal pancreatic function. This may imply that having chronic suppurative lung disease, in isolation from pancreatic involvement, may have a negative impact on nutritional status. However, although this does not imply causality as it is possible that other clinical and

nutritional factors may impact on linear growth [38], it would be an important relationship to explore in a future larger study.

Improving nutritional status is one of the cornerstones of the management of children with CF and is widely thought to have contributed significantly to the improvements in mortality seen over the past decades [7, 36]. We, therefore, hypothesise that similar benefits may be seen in children with PCD and postulate that if stratified nutritional interventions were developed to provide appropriate amounts of both macro- and micronutrients for those patients displaying early nutritional decline, longer term outcomes may be improved [39].

Although not statistically significantly different, FEV<sub>1</sub> z score was higher in those with a protein intake ≥200% of RNI compared to those with an adequate intake of protein (LRNI to ≤200% RNI). Whilst nutritional interventions have been successful in improving some outcomes for children with chronic diseases such as CF [40], strategies have focused on increasing energy intake at the expense of considering the need for type II nutrients, which support linear growth such as zinc, sulphur (protein) and magnesium [41-43]. Children with chronic diseases are surviving longer. As such, it is essential that excess adiposity, through targeting energy intake alone, is not promoted with the aim of achieving a target BMI, without considering what is happening to linear growth and lean body mass deposition [10].

Anthropometrical measures of height and weight are used to determine nutrition risk, with WHO cut offs <-2 used for moderate malnutrition [27]. In this cohort using <-2 HAZ and BMIZ scores 4.6% vs. 6.9% cases respectively would have been identified as being at nutritional risk. FFMI and FMI z scores are increasingly being used to quantify body composition, which when used in conjunction with BIS phase angle 50°, might offer a method for early identification of patients with declining nutritional status, as described in children with CF [44, 45]. In this study 21% of children had a FFMI z score of <-2. This group of children also had a lower BIS

phase angle 50° of 4.3±0.4 compared to children with a FFMI z score of > -2 (n=23) whose BIS phase angle 50° was 4.9±0.8 (p=0.0002). However, their BMI z score was -1.3±1.2 suggesting they were normally nourished. As such, had nutritional cut offs of <-2 z scores only been used to identify nutritional risk, these children may not have been recognised as being at risk. Therefore it may be that BIS phase angle 50° may increase the sensitivity and specificity of identifying those children at risk of nutritional decline earlier than routine measures of anthropometry, such as height and weight currently afford and described amongst children with congenital heart disease [21]. Children with lower BIS phase angle 50° had lower lung function, suggesting this measure may potentially be associated with other independent and clinically important outcomes, as seen amongst other children with lung disease [44, 45]. However, we acknowledge the numbers in this study are small and further work amongst a larger cohort of children with PCD will be required to confirm this observation as well as determining the causality as it may be an incidental finding within this cohort.

In this study BIS phase angle 50° was measured using an ImpediMedSFB7 device. BIS technology is easy to use in a clinical setting as it is non-invasive. Interest is increasing in trying to develop BIS phase angle reference values, which would allow for the comparison of derived BIS phase angle 50° measures amongst children who vary in age, gender and clinical condition. This has the potential to allow for the earlier identification of those with declining nutritional status, presenting an opportunity for earlier intervention [21]. Limitations for all studies, including ours, conducted in this area are the use of differing machines, varying populations and the various methods and approaches used to determining a value for phase angle [46]. This makes it challenging to apply a single defined cut-off for phase angle, which may have been defined for a specific study. Therefore, like others, our values should only be considered in the context of the condition and population studied. In order to strengthen the application of this

technology, the utility of phase angle should be studied longitudinally, in a considerably larger cohort across multiple centres, with the aim of developing cut-off values and creating reference z scores for phase angle to be used in the context of PCD [47].

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A wide array of micronutrients and vitamins were measured in order to identify any specific deficiencies that might need to be considered when planning future nutritional intervention strategies. The only noteworthy finding seen was that 54% of the cohort in whom plasma concentrations were available, had insufficient vitamin D levels (<50 nmol/L), with 26% being deficient (<30 nnol/L), needing treatment. This is three times the prevalence seen in an Italian study assessing 22 PCD patients, which found 18% had vitamin D levels <50 nmol/L [48]. Furthermore, it is higher than the prevalence of vitamin D insufficiency seen in UK children, shown to be 35% in one large population-based study [49]. This is particularly evident once geographical location is considered as the reported mean vitamin D levels of UK children living in Southern England was 62.3 nmol/L [49] whereas in our cohort it was 52.5 nmol/L. There was also a weak association between low vitamin D status and FFMI. Vitamin D has previously been shown to be essential for calcium transport, lean body mass development and protein metabolism. In vitamin D deficient children and adults with a classical Fontan circulation lean body mass is significantly lower than in those with normal levels [50]. The relationship between vitamin D status and body composition should be further investigated in future nutritional studies.

Patients with PCD have persistent and recurrent sinopulmonary infections. However, to our knowledge, there have been no studies to date considering blood biomarkers of inflammation in PCD patients. Many chronic conditions have an inflammatory component and low-grade inflammation can affect metabolism and body composition. Cytokines have a pivotal role in orchestrating the inflammatory response to infection. One group studied levels of IL-8 in

the sputum of children with PCD, finding it to be three times greater than in the sputum of children with CF [51]. Furthermore, it has been shown that monocytes from PCD patients produce higher levels of pro-inflammatory cytokines (IL-1 $\beta$  and TNF- $\alpha$ ) in response to lipopolysaccharide than those from healthy individuals [52]. It was interesting, therefore, that mean plasma levels of the pro-inflammatory cytokines assessed were not raised in our cohort compared to the available, albeit limited, normative data [34, 35]. We had hypothesised that those patients with evidence of inflammation may have poorer nutrition outcomes. It was therefore noteworthy that we only observed a weak correlation between the pro-inflammatory cytokine IL-2 and BIS phase angle. The function of IL-2 is not completely understood but elevated levels are associated with cancer cachexia and weight loss [53]. This finding is hypothesis generating and further research into IL-2 and its association with nutritional status would be of interest.

We also considered relationships between markers of oxidative stress (6-keto-PGF $_{1\alpha}$  and 8-isoprostane), various fatty acids and BIS phase angle. Red blood cells are considered stable markers of status of polyunsaturated fatty acids and are reflective of dietary intake patterns. The composition reported here is similar to compositions reported in children in other settings [54]. N-3 fatty acids, especially EPA and DHA are anti-inflammatory and capable of inhibiting inflammatory processes including leukocyte chemotaxis, adhesion molecule expression, leukocyte-endothelial adhesive interactions and production of inflammatory cytokines and lipid mediators. Dietary fat intake in the UK is high in n-6 fatty acids which facilitate the production of eicosanoids like prostaglandins and leukotrienes from arachidonic acid (n-6 fatty acid cascade) and subsequently are considered to promote inflammation and oxidative stress [55]. We found a weak positive relationship between linoleic acid, an n-6 fatty acid, in red blood cells and

plasma 6-keto  $PGF_{1\alpha}$ , which would be of interest to investigate further in future nutrition studies.

There are a number of limitations of this study. Firstly, there are limitations to the accuracy of dietary intake methods, which are highlighted in many studies [56, 57]. In our study we chose to use a 3-day semi quantitative food diary as oppose to a 7-day diary in order to reduce the fatigue effect of recording dietary information for such a long time. However, future studies in children with PCD may benefit form adding a second dietary intake method to ensure that dietary intake recorded is accurate [56]. In addition, the observation regarding the potential benefit of BIS phase angle 50° in early identification of children with nutritional problems will need further assessment in larger cohorts before stronger conclusions can be drawn. In order to assess the benefit of this and any subsequent nutritional intervention, it is likely that a multicentre design will be required.

### Conclusion

We have characterised the nutritional phenotype of a cohort of children with PCD. There appears to be a weak relationship between lung function and nutritional status, in addition to measures of BIS phase angle 50°. The use of BIS phase angle 50° may be better at identifying early nutritional decline in children with PCD and may therefore be of benefit for nutritional assessments in the clinical setting. These findings may help inform a future nutritional intervention strategy in children with PCD.

#### Acknowledgements

The authors wish to thank the children and their families for agreeing to be part of this study. Vitamin B6 assays completed by Christine Glenn, NIHR Southampton Biomedical Research Centre, Wessex Investigational Sciences Hub Laboratory (WISH Lab). The PCD team included physiotherapists Victoria Keenan and Hannah Wilkins.

#### **Funding**

This work was supported by a Team Fellowship Award from NIHR Health Education England (Wessex). ALH was also supported by an Individual Fellowship from NIHR Health Education (Wessex). This work is also part of independent research completed by LVM arising from a Health Education England/NIHR Clinical Lectureship (ICA-CL-2016-02-001)) supported by the National Institute for Health Research. The views expressed in this publication are those of the author(s) and not necessarily those of the NHS, the National Institute for Health Research, Health Education England or the Department of Health. The National PCD Service at UHS is commissioned and funded by NHS England. PCD research in Southampton is supported by NIHR Southampton Respiratory Biomedical Research Unit and NIHR Wellcome Trust Clinical Research Facility.

### Statement of authorship

All authors have made substantial contributions to the following areas of this manuscript: LVM and WW designed the study. EAM completed the cytokine, pro-oxidant and fatty acid analyses. WW, ALH, LVM and CJ carried out the data collection. ALH, LVM, BL, WW and PCC completed the data and statistical analyses and drafted the manuscript. All authors edited, read and approved the final manuscript.

## **Conflict of interest**

None of the authors has any conflict of interest to declare in relation to this research.

#### 394 References

- Goutaki M, Meier AB, Halbeisen FS, Lucas JS, Dell SD, Maurer E, Casaulta C, Jurca M,
   Spycher BD, Kuehni CE: Clinical manifestations in primary ciliary dyskinesia: systematic
   review and meta-analysis. The European respiratory journal 2016, 48(4):1081-1095.
- Davis SD, Ferkol TW, Rosenfeld M, Lee HS, Dell SD, Sagel SD, Milla C, Zariwala MA,
   Pittman JE, Shapiro AJ et al: Clinical features of childhood primary ciliary dyskinesia by
   genotype and ultrastructural phenotype. American journal of respiratory and critical
   care medicine 2015, 191(3):316-324.
- 402 3. Goutaki M, Halbeisen FS, Spycher BD, Maurer E, Belle F, Amirav I: **Growth and**403 **nutritional status, and their association with lung function: a study from the**404 **international Primary Ciliary Dyskinesia Cohort**. 2017, **50**(6).
- Svobodova T, Djakow J, Zemkova D, Cipra A, Pohunek P, Lebl J: Impaired Growth during
   Childhood in Patients with Primary Ciliary Dyskinesia. International journal of
   endocrinology 2013, 2013:731423.
- Maglione M, Bush A, Nielsen KG, Hogg C, Montella S, Marthin JK, Di Giorgio A,
   Santamaria F: Multicenter analysis of body mass index, lung function, and sputum
   microbiology in primary ciliary dyskinesia. Pediatric pulmonology 2014, 49(12):1243-1250.
- 412 6. Engelen MP, Schroder R, Van der Hoorn K, Deutz NE, Com G: **Use of body mass index**413 **percentile to identify fat-free mass depletion in children with cystic fibrosis**. *Clinical*414 *nutrition (Edinburgh, Scotland)* 2012, **31**(6):927-933.
- 7. Dodge JA, Turck D: Cystic fibrosis: nutritional consequences and management. Best
   practice & research Clinical gastroenterology 2006, 20(3):531-546.
- 417 8. Foundation CT (ed.): **Nutritional Management of Cystic Fibrosis: A consensus guideline**; 418 2016.
- Alvarez JA, Ziegler TR, Millson EC, Stecenko AA: Body composition and lung function in cystic fibrosis and their association with adiposity and normal-weight obesity.
   Nutrition (Burbank, Los Angeles County, Calif) 2016, 32(4):447-452.
- 422 10. Konstan MW, Pasta DJ, Wagener JS, VanDevanter DR, Morgan WJ: **BMI fails to identify**423 **poor nutritional status in stunted children with CF**. *Journal of cystic fibrosis : official*424 *journal of the European Cystic Fibrosis Society* 2017, **16**(1):158-160.
- 425 11. Kyle UG, Earthman CP, Pichard C, Coss-Bu JA: **Body composition during growth in**426 **children: limitations and perspectives of bioelectrical impedance analysis**. *European*427 *journal of clinical nutrition* 2015, **69**(12):1298-1305.
- Mulasi U, Kuchnia AJ, Cole AJ, Earthman CP: Bioimpedance at the bedside: current
   applications, limitations, and opportunities. Nutrition in clinical practice: official
   publication of the American Society for Parenteral and Enteral Nutrition 2015, 30(2):180-193.
- 432 13. Kyle UG, Bosaeus I, De Lorenzo AD, Deurenberg P, Elia M, Manuel Gomez J, Lilienthal
   433 Heitmann B, Kent-Smith L, Melchior JC, Pirlich M et al: Bioelectrical impedance analysis 434 part II: utilization in clinical practice. Clinical nutrition (Edinburgh, Scotland) 2004,
   435 23(6):1430-1453.
- 436 14. Kyle UG, Bosaeus I, De Lorenzo AD, Deurenberg P, Elia M, Gomez JM, Heitmann BL,
   437 Kent-Smith L, Melchior JC, Pirlich M et al: Bioelectrical impedance analysis--part I:
   438 review of principles and methods. Clinical nutrition (Edinburgh, Scotland) 2004,
   439 23(5):1226-1243.

- Kyle UG, Soundar EP, Genton L, Pichard C: Can phase angle determined by bioelectrical impedance analysis assess nutritional risk? A comparison between healthy and hospitalized subjects. Clinical nutrition (Edinburgh, Scotland) 2012, 31(6):875-881.
- da Silva TK, Berbigier MC, Rubin Bde A, Moraes RB, Correa Souza G, Schweigert Perry ID:
   Phase angle as a prognostic marker in patients with critical illness. Nutrition in clinical practice: official publication of the American Society for Parenteral and Enteral Nutrition 2015, 30(2):261-265.
- Lee Y, Kwon O, Shin CS, Lee SM: Use of bioelectrical impedance analysis for the
   assessment of nutritional status in critically ill patients. Clinical nutrition research 2015,
   4(1):32-40.
- 450 18. Kuchnia A, Earthman C, Teigen L, Cole A, Mourtzakis M, Paris M, Looijaard W, Weijs P,
   451 Oudemans-van Straaten H, Beilman G et al: Evaluation of Bioelectrical Impedance
   452 Analysis in Critically III Patients: Results of a Multicenter Prospective Study. JPEN
   453 Journal of parenteral and enteral nutrition 2017, 41(7):1131-1138.
- Thibault R, Makhlouf AM, Mulliez A, Cristina Gonzalez M, Kekstas G, Kozjek NR, Preiser JC, Rozalen IC, Dadet S, Krznaric Z et al: Fat-free mass at admission predicts 28-day mortality in intensive care unit patients: the international prospective observational study Phase Angle Project. Intensive care medicine 2016, 42(9):1445-1453.
- 458 20. Marino LV GM, Pappachan JV **Pre-operative bioelectrical impedance predicts intensive**459 **care length of stay in children following cardiac surgery**. *Cardiololgy in the Young* 2018,
  460 **epub ahead of print**.
- 461 21. Marino LV, Meyer R, Johnson M, Newell C, Johnstone C, Magee A, Sykes K, Wootton SA,
   462 Pappachan JV: Bioimpedance spectroscopy measurements of phase angle and height
   463 for age are predictive of outcome in children following surgery for congenital heart
   464 disease. Clinical nutrition (Edinburgh, Scotland) 2017.
- Farias CL, Campos DJ, Bonfin CM, Vilela RM: Phase angle from BIA as a prognostic and nutritional status tool for children and adolescents undergoing hematopoietic stem cell transplantation. Clinical nutrition (Edinburgh, Scotland) 2013, 32(3):420-425.
- Pileggi VN, Monteiro JP, Margutti AV, Camelo JS, Jr.: Prevalence of child malnutrition at a university hospital using the World Health Organization criteria and bioelectrical impedance data. Brazilian journal of medical and biological research = Revista brasileira de pesquisas medicas e biologicas 2016, 49(3).
- 472 24. Lucas JS, Barbato A, Collins SA: **European Respiratory Society guidelines for the**473 **diagnosis of primary ciliary dyskinesia**. 2017, **49**(1).
- 474 25. Miller MR, Hankinson J, Brusasco V, Burgos F, Casaburi R, Coates A, Crapo R, Enright P,
   475 van der Grinten CP, Gustafsson P et al: Standardisation of spirometry. The European
   476 respiratory journal 2005, 26(2):319-338.
- 477 26. Quanjer PH, Stanojevic S, Cole TJ, Baur X, Hall GL, Culver BH, Enright PL, Hankinson JL, Ip
  478 MS, Zheng J *et al*: **Multi-ethnic reference values for spirometry for the 3-95-yr age**479 **range: the global lung function 2012 equations**. *The European respiratory journal* 2012,
  480 **40**(6):1324-1343.
- 481 27. **Growth reference 5-19 years: BMI-for-age (5-19years)**482 [http://www.who.int/growthref/who2007 bmi for age/en/]
- Nagy P, Kovacs E, Moreno LA, Veidebaum T, Tornaritis M, Kourides Y, Siani A, Lauria F,
   Sioen I, Claessens M et al: Percentile reference values for anthropometric body
   composition indices in European children from the IDEFICS study. International journal
   of obesity (2005) 2014, 38 Suppl 2:S15-25.
- 487 29. WHO WHO: WHO Anthro (version 3.2.2, January 2011). In.; 2015.

- 488 30. WHO WHO: WHO AnthroPlus. In.; 2015.
- 489 31. Azevedo ZM, Moore DC, de Matos FA, Fonseca VM, Peixoto MV, Gaspar-Elsas MI,
  490 Santinoni E, Dos Anjos LA, Ramos EG: **Bioelectrical impedance parameters in critically ill**491 **children: importance of reactance and resistance**. *Clinical nutrition (Edinburgh,*

492 *Scotland*) 2013, **32**(5):824-829.

- 493 32. Talwar D, Quasim T, McMillan DC, Kinsella J, Williamson C, O'Reilly DS: **Optimisation and**494 validation of a sensitive high-performance liquid chromatography assay for routine
  495 measurement of pyridoxal 5-phosphate in human plasma and red cells using pre496 column semicarbazide derivatisation. *Journal of chromatography B, Analytical*497 technologies in the biomedical and life sciences 2003, **792**(2):333-343.
- 498 33. Eltweri AM, Thomas AL, Fisk HL, Arshad A, Calder PC, Dennison AR, Bowrey DJ: **Plasma**499 and erythrocyte uptake of omega-3 fatty acids from an intravenous fish oil based lipid
  500 emulsion in patients with advanced oesophagogastric cancer. *Clinical nutrition*501 (Edinburgh, Scotland) 2017, **36**(3):768-774.
- Kleiner G, Zanin V, Monasta L, Crovella S, Caruso L, Milani D, Marcuzzi A: Pediatric
   patients with inflammatory bowel disease exhibit increased serum levels of
   proinflammatory cytokines and chemokines, but decreased circulating levels of
   macrophage inhibitory protein-1beta, interleukin-2 and interleukin-17. Experimental
   and therapeutic medicine 2015, 9(6):2047-2052.
- Kim HO, Kim H-S, Youn J-C, Shin E-C, Park S: Serum cytokine profiles in healthy young
   and elderly population assessed using multiplexed bead-based immunoassays. *Journal of Translational Medicine* 2011, 9:113-113.
- 510 36. Konstan MW, Butler SM, Wohl ME, Stoddard M, Matousek R, Wagener JS, Johnson CA,
   511 Morgan WJ: Growth and nutritional indexes in early life predict pulmonary function in
   512 cystic fibrosis. The Journal of pediatrics 2003, 142(6):624-630.
- 513 37. Cohen-Cymberknoh M, Simanovsky N, Hiller N, Hillel AG, Shoseyov D, Kerem E:
   514 Differences in disease expression between primary ciliary dyskinesia and cystic fibrosis
   515 with and without pancreatic insufficiency. Chest 2014, 145(4):738-744.
- Ripa P, Robertson I, Cowley D, Harris M, Masters IB, Cotterill AM: The relationship
   between insulin secretion, the insulin-like growth factor axis and growth in children
   with cystic fibrosis. Clinical endocrinology 2002, 56(3):383-389.
- Sanders DB, Zhang Z, Farrell PM, Lai HJ: Early life growth patterns persist for 12years
   and impact pulmonary outcomes in cystic fibrosis. Journal of cystic fibrosis: official
   journal of the European Cystic Fibrosis Society 2018.
- Turck D, Braegger CP, Colombo C, Declercq D, Morton A, Pancheva R, Robberecht E,
   Stern M, Strandvik B, Wolfe S et al: ESPEN-ESPGHAN-ECFS guidelines on nutrition care
   for infants, children, and adults with cystic fibrosis. Clinical nutrition (Edinburgh,
   Scotland) 2016, 35(3):557-577.
- 526 41. Engelen MP, Com G, Deutz NE: Protein is an important but undervalued macronutrient
   527 in the nutritional care of patients with cystic fibrosis. Current opinion in clinical
   528 nutrition and metabolic care 2014, 17(6):515-520.
- Jonker R, Engelen MP, Deutz NE: **Role of specific dietary amino acids in clinical conditions**. *The British journal of nutrition* 2012, **108 Suppl 2**:S139-148.
- 531 43. Golden MH: **Proposed recommended nutrient densities for moderately malnourished children**. *Food and nutrition bulletin* 2009, **30**(3 Suppl):S267-342.
- Groeneweg M, Tan S, Boot AM, de Jongste JC, Bouquet J, Sinaasappel M: **Assessment of** nutritional status in children with cystic fibrosis: conventional anthropometry and

- bioelectrical impedance analysis. A cross-sectional study in Dutch patients. *Journal of cystic fibrosis : official journal of the European Cystic Fibrosis Society* 2002, **1**(4):276-280.
- Hauschild DB, Barbosa E, Moreira EA, Ludwig Neto N, Platt VB, Piacentini Filho E,
   Wazlawik E, Moreno YM: Nutrition Status Parameters and Hydration Status by
   Bioelectrical Impedance Vector Analysis Were Associated With Lung Function
   Impairment in Children and Adolescents With Cystic Fibrosis. Nutrition in clinical
   practice: official publication of the American Society for Parenteral and Enteral Nutrition
   2016, 31(3):378-386.
- Nichols J, Going S, Loftin M, Stewart D, Nowicki E, Pickrel J: Comparison of two
   bioelectrical impedance analysis instruments for determining body composition in
   adolescent girls. International journal of body composition research 2006, 4(4):153-160.
- Norman K, Stobaus N, Pirlich M, Bosy-Westphal A: Bioelectrical phase angle and impedance vector analysis--clinical relevance and applicability of impedance parameters. Clinical nutrition (Edinburgh, Scotland) 2012, 31(6):854-861.
- Mirra V, Caffarelli C, Maglione M, Valentino R, Perruolo G, Mazzarella C, Di Micco LL,
   Montella S, Santamaria F: Hypovitaminosis D: a novel finding in primary ciliary
   dyskinesia. Italian journal of pediatrics 2015, 41:14.
- 552 49. Absoud M, Cummins C, Lim MJ, Wassmer E, Shaw N: **Prevalence and predictors of** 553 **vitamin D insufficiency in children: a Great Britain population based study**. *PloS one* 554 2011, **6**(7):e22179.
- Avitabile CM, Leonard MB, Zemel BS, Brodsky JL, Lee D, Dodds K, Hayden-Rush C,
   Whitehead KK, Goldmuntz E, Paridon SM et al: Lean mass deficits, vitamin D status and
   exercise capacity in children and young adults after Fontan palliation. Heart (British
   Cardiac Society) 2014, 100(21):1702-1707.
- 559 51. Bush A, Payne D, Pike S, Jenkins G, Henke MO, Rubin BK: Mucus properties in children
   with primary ciliary dyskinesia: comparison with cystic fibrosis. Chest 2006,
   129(1):118-123.
- 562 52. Cockx M, Gouwy M, Ruytinx P, Lodewijckx I, Van Hout A, Knoops S, Portner N, Ronsse I,
   563 Vanbrabant L, Godding V et al: Monocytes from patients with Primary Ciliary
   564 Dyskinesia show enhanced inflammatory properties and produce higher levels of pro 565 inflammatory cytokines. Scientific reports 2017, 7(1):14657.
- 566 53. Gonda K, Shibata M, Shimura T, Machida T, Suzuki S, Nakamura I, Ohki S, Sakurai K, Ohto
   567 H, Tomita R et al: Serum Soluble Interleukin-2 Receptor is Increased in Malnourished
   568 and Immunosuppressed Patients With Gastric and Colorectal Cancer: Possible
   569 Influence of Myeloid-Derived Suppressor Cells. World journal of oncology 2012,
   570 3(4):158-164.
- 571 54. Brigandi SA, Shao H, Qian SY, Shen Y, Wu BL, Kang JX: Autistic children exhibit
   572 decreased levels of essential Fatty acids in red blood cells. International journal of
   573 molecular sciences 2015, 16(5):10061-10076.
- 574 55. Calder PC: **Omega-3 fatty acids and inflammatory processes: from molecules to man**. *Biochemical Society transactions* 2017, **45**(5):1105-1115.
- 576 56. Lanigan JA, Wells JC, Lawson MS, Cole TJ, Lucas A: **Number of days needed to assess**577 **energy and nutrient intake in infants and young children between 6 months and 2**578 **years of age**. *European journal of clinical nutrition* 2004, **58**(5):745-750.
- 579 57. Freisling H, Ocke MC, Casagrande C, Nicolas G, Crispim SP, Niekerk M, van der Laan J, de
   580 Boer E, Vandevijvere S, de Maeyer M et al: Comparison of two food record-based
   581 dietary assessment methods for a pan-European food consumption survey among

| 582<br>583 | infants, toddlers, and children using data quality indicators. European journal of nutrition 2015, <b>54</b> (3):437-445. |
|------------|---|
| 584        |   |
| 585        |   |

# **PCD**

| Variables  | n= | Mean ± SD              |  |  |  |  |
|--|----|------------------------|--|--|--|--|
| Age at diagnosis (years)                                 | 43 | 2.7±3.6                |  |  |  |  |
| Age at time of study (years)                             | 43 | 7.0±5.2                |  |  |  |  |
| Forced Expiratory Volume in 1 second (FEV <sub>1</sub> ) | 29 |                        |  |  |  |  |
| predicted (%)  | 29 | 79.3±20.4              |  |  |  |  |
| FEV <sub>1</sub> z score                                 | 28 | -1.7±1.7               |  |  |  |  |
| Forced Volume Capacity (FVC) (L)                         | 29 | 2.2±0.9                |  |  |  |  |
| FVC predicted (%)  | 29 | 90.9±19.1              |  |  |  |  |
| FVC z score  | 28 | -0.8±1.7               |  |  |  |  |
| Oral antibiotic courses per year (n)                     | 40 | 3.5±1.8                |  |  |  |  |
| Gender   | 43 | Male n=22; Female n=21 |  |  |  |  |
| Weight (kg)  | 43 | 30.3±19.8              |  |  |  |  |
| Height (m)   | 43 | 1.2±0.3                |  |  |  |  |
| BMI (kg/m <sup>2</sup> )                                 | 43 | 17.2±4.6               |  |  |  |  |
| Weight for age z score (WAZ)                             | 26 | 0.1±1.1                |  |  |  |  |
| Height for age z score (HAZ)                             | 43 | -0.2±1.1               |  |  |  |  |
| Mid upper arm circumference (MUAC) (cm)                  | 33 | 19.7±4.7               |  |  |  |  |
| Vitamin D (nmol/L)                                       | 35 | 52.5±29.2              |  |  |  |  |
| Selenium (μmol/L) (ref range 0.2- 0.9)                   | 35 | 1.4±2.3                |  |  |  |  |
| Zinc (μmol/L) (ref range 11 – 24)                        | 35 | 14.2±2.1               |  |  |  |  |
| Copper (µmol/L) (ref range 10-22)                        | 35 | 20.0±4.2               |  |  |  |  |
| Ferritin (μg/L) (ref range 20-200)                       | 36 | 24.0±13.4              |  |  |  |  |
| Folate (nmol/mL) (ref range 5-21)                        | 36 | 13.0±6.6               |  |  |  |  |
| Vitamin B12 (pg/mL) (ref range 180 – 1000)               | 36 | 623.5±271              |  |  |  |  |
| Vitamin B6 (μg/L) (ref range 5-50)                       | 19 | 75.7±42.0              |  |  |  |  |
| Iron (μmol/L) (ref range 5 – 31)                         | 36 | 12.0±5.3               |  |  |  |  |
| Transferrin (g/L) (ref range 1.8 - 3.3 g/L)              | 36 | 3.0±0.4                |  |  |  |  |
| Transferrin iron saturation (%) (ref range <16)          | 36 | 19.5±9.4               |  |  |  |  |
| Haemoglobin (g/L) (ref range 95-135)                     | 36 | 129.9±13.8             |  |  |  |  |
| Albumin (g/L) (ref range 35 – 45)                        | 36 | 41.4±3.5               |  |  |  |  |
| Calcium (mmol/L) (ref range 2.20-2.60)                   | 22 | 2.3±0.1                |  |  |  |  |
| C-reactive protein (g/L) (ref range 0 – 3.6)             | 36 | 5.9±17.4               |  |  |  |  |
| Phosphate (mmol/L) (ref range 1.25-2.10)                 | 36 | 1.5±0.3                |  |  |  |  |
| Magnesium (mmol/L) (ref range 0.70-1.00)                 | 36 | 0.9±0.8                |  |  |  |  |

| Alkaline Phosphatase (U/L) (ref range 145-420) | 35 | 260±96 |
|--|----|--------|
|--|----|--------|

Table 2: Body composition measure of fat mass, fat free mass and BIS phase angle 50° and plasma cytokine concentrations in children with PCD

|                           | N  | Mean <u>+</u> SD |
|---------------------------|----|------------------|
| BIS phase angle 50°       | 36 | 4.5 ± 0.9        |
| Fat mass (kg)             | 36 | 5.9 ± 6.7        |
| Fat free mass (kg)        | 36 | 22.8 ± 13.6      |
| Fat mass (%)              | 36 | 19.2 ± 20.7      |
| Fat free mass (%)         | 36 | 80.7 ± 79.2      |
| IL-1β (pg/ml)             | 41 | 0.6 ± 1.1        |
| IL-2 (pg/ml)              | 41 | 0.4 ± 0.8        |
| IL-6 (pg/ml)              | 41 | 2.5 ± 4.1        |
| IL-8 (pg/ml)              | 41 | 5.3 ± 4.5        |
| TNFα (pg/ml)              | 41 | 7.7 ± 4.6        |
| VEGF-1 (pg/ml)            | 41 | 30.7 ± 31.9      |
| C-reactive protein (mg/L) | 34 | 5.9±17.4         |

# Table 3: Comparison of children with PCD over the age of 2years with a fat free mass index <-2 z scores to those with a fat free mass index > -

# 2 z scores

|                                  | Age              | FEV1 z            | FVC z             | HAZ      | BMI z    | Fat      | Fat Mass  | Fat free | Fat Free  | Total body | Extra    | Intra    | R200/5  | Phase   | Phase    | Phase   |
|----------------------------------|------------------|-------------------|-------------------|----------|----------|----------|-----------|----------|-----------|------------|----------|----------|---------|---------|----------|---------|
|                                  | (years)          | score             | score             | score    | score    | Mass     | (%)       | mass     | Mass      | water (%)  | cellular | cellular | HZ2     | angle 5 | angle 50 | angle   |
|                                  |                  |                   |                   |          |          | Index    |           | index    | (%)       |            | Fluid %  | Fluid %  |         | Hz      | Hz       | 200 Hz  |
| Fat free mass                    | s index < 2 (r   | n=6)              | 1                 |          |          |          | 1         |          |           | ı          |          |          | 1       | 1       | 1        |         |
| Mean±SD                          | 5.9 <b>±</b> 3.5 | -2.5 <b>±</b> 1.0 | -1.5 <b>±</b> 1.0 | 0.0±1.4  | -1.3±1.2 | -2.7±0.5 | 12.4±10.6 | -3.0±0.0 | 87.6±10.6 | 64.1±7.8   | 47.7±2.1 | 52.3±2.1 | 0.9±0.0 | 1.5±0.1 | 4.3±0.4  | 5.2±0.3 |
| Fat free mass index > - 2 (n=23) |                  |                   |                   |          |          |          |           |          |           |            |          |          |         |         |          |         |
| Mean±SD                          | 9.3 <b>±</b> 4.5 | -1.2±1.4          | -0.3 <b>±</b> 1.3 | -0.2±0.9 | 0.8±0.7  | -0.3±2.2 | 19.4±10.5 | 1.5±0.8  | 80.6±10.5 | 59.0±7.7   | 43.7±4.4 | 56.3±4.4 | 0.8±0.0 | 1.8±0.3 | 4.9±0.8  | 5.2±0.8 |
| p values                         | 0.1              | 0.09              | 0.01              | 0.9      | 0.0002   | 0.04     | 0.2       | 0.0002   | 0,2       | 0.2        | 0.0002   | 0.03     | 0.07    | 0.03    | 0.0002   | 0.9     |

# stress (pg/ml) in children with PCD

| % erythrocyte fatty acids & markers of oxidative stress  | N= | Mean ±standard deviation |  |  |  |  |  |
|--|----|--------------------------|--|--|--|--|--|
| 14:0   | 22 | 0.4±0.2                  |  |  |  |  |  |
| 16:0   | 22 | 22.7±1.7                 |  |  |  |  |  |
| 16:1n-7  | 22 | 0.3±0.1                  |  |  |  |  |  |
| 18:0   | 22 | 15.7±1.3                 |  |  |  |  |  |
| 18:1n-9  | 22 | 17.2±1.4                 |  |  |  |  |  |
| 18:1n-7  | 22 | 1.5±0.5                  |  |  |  |  |  |
| 18:2n-6  | 22 | 14.2±1.7                 |  |  |  |  |  |
| 18:3n-6  | 22 | 0.06±0.5                 |  |  |  |  |  |
| 18:3n-3  | 22 | 0.2±0.1                  |  |  |  |  |  |
| 20:0   | 22 | 0.1±0.6                  |  |  |  |  |  |
| 20:1n-9  | 22 | 0.3±0.07                 |  |  |  |  |  |
| 20:2n-6  | 22 | 0.2±0.07                 |  |  |  |  |  |
| 20:3n-6  | 22 | 1.9±0.4                  |  |  |  |  |  |
| 20:4n-6  | 22 | 13.3±1.5                 |  |  |  |  |  |
| 22:0   | 22 | 0.09±0.04                |  |  |  |  |  |
| 20:4n-3  | 22 | 0.1±0.04                 |  |  |  |  |  |
| 20:5n-3  | 22 | 0.5±0.2                  |  |  |  |  |  |
| 24:0   | 22 | 2.9±0.6                  |  |  |  |  |  |
| 24:1n-9  | 22 | 0.3±0.06                 |  |  |  |  |  |
| 22:5n-3  | 22 | 1.7±0.3                  |  |  |  |  |  |
| 22:6n-3  | 22 | 2.8±0.8                  |  |  |  |  |  |
| Prostaglandin E <sub>2</sub> (PGE <sub>2</sub> ) (pg/ml) | 34 | 96.9±110                 |  |  |  |  |  |
| 8-isoprostane (pg/ml)                                    | 34 | 45.9±79.7                |  |  |  |  |  |
| 6-keto PGF <sub>1<math>\alpha</math></sub> (pg/ml)       | 36 | 95.1±120                 |  |  |  |  |  |

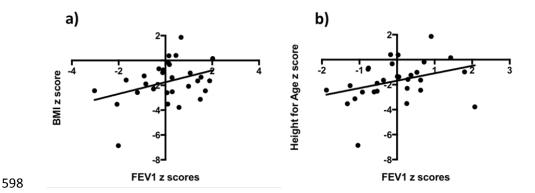
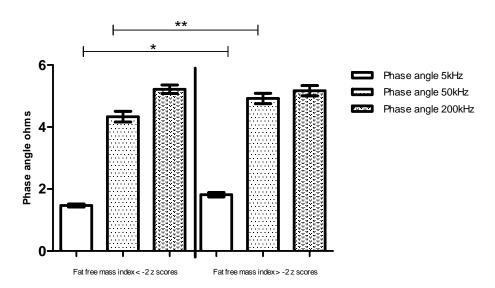
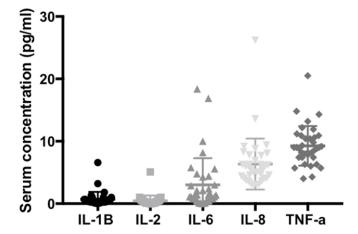


Figure 1: Relationship between FEV1 z score and a) BMI z score (n=29; r=0.36, p=0.055) and b)

height for age z score (n=29; r=0.4, p=0.049) in children with PCD.



**Figure 2:** Children with a fat free mass index of <-2 z scores (n=6) had a significantly lower phase angle  $5^{\circ}$  (p=0.03) and phase angle  $50^{\circ}$  (p=0.0002) compared to those with a fat free mass index of > - 2 scores (n=23)



**Figure 3** Concentrations of five pro-inflammatory cytokines (IL-1B, IL-2, IL-6, IL-8 & TNF- $\alpha$ ) in children with PCD (n=41) (Mean (SD), measured in pg/ml)