

Hyperpolarized Fumarate via Parahydrogen

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We produce hyperpolarized [1-¹³C]fumarate in the proton nuclear spin singlet state by pairwise *trans*-addition of parahydrogen to a molecular precursor using a ruthenium-based catalyst in water. The proton singlet state is transformed into observable carbon magnetization by radiofrequency pulses to enhance the ¹³C signal by a factor of 1000 using 50% para-enriched hydrogen gas.

NMR methods suffer from intrinsically weak signals, which makes *in vivo* imaging of metabolites at physiological concentrations very challenging. To overcome this barrier, hyperpolarization of the nuclear spins is a necessity, and can lead to signal enhancements of $\sim 10^5$ – 10^6 in a typical MRI scanner [1–3]. For example, hyperpolarized magnetic resonance imaging may be used to study the enzymatic conversion of fumarate (*trans*-butenedioic acid) into malate (2-hydroxybutanedioic acid), which acts as a marker for dying or diseased cells [4–6].

Currently, dissolution-dynamic nuclear polarization (dDNP) [7,8] is the technique used to hyperpolarize fumarate. Despite significant advances in this field over the last decade, dDNP still has some limitations: expensive equipment is required, the hyperpolarized material is delivered in batch mode, and the polarization build-up time per batch is in the order of one hour. An alternative hyperpolarization method is PHIP (Para-Hydrogen Induced Polarization) [9–11], which uses the nuclear spin singlet form of hydrogen gas as a polarization source. In hydrogenative PHIP a molecule of parahydrogen is chemically reacted with a molecular precursor to produce a ¹H-hyperpolarized product. The proton polarization can be observed directly, or transferred to a heteronuclear spin (i.e. ¹³C

or ¹⁵N) which allows for the generation of hyperpolarized metabolites in a cheap, continuous manner [12–16].

Here we show that PHIP can be used to prepare hyperpolarized fumarate by pairwise hydrogenation of an acetylene dicarboxylate precursor in water (Figure 1), using a commercially available ruthenium-based catalyst to achieve *trans*-hydrogenation [17–21]. This is in contrast to the vast majority of parahydrogenation reactions reported in the literature, which lead to the para-H₂ protons occupying *cis*-positions on the product molecule. For example, maleate, the *cis* isomer of butenedioic acid, is readily formed after

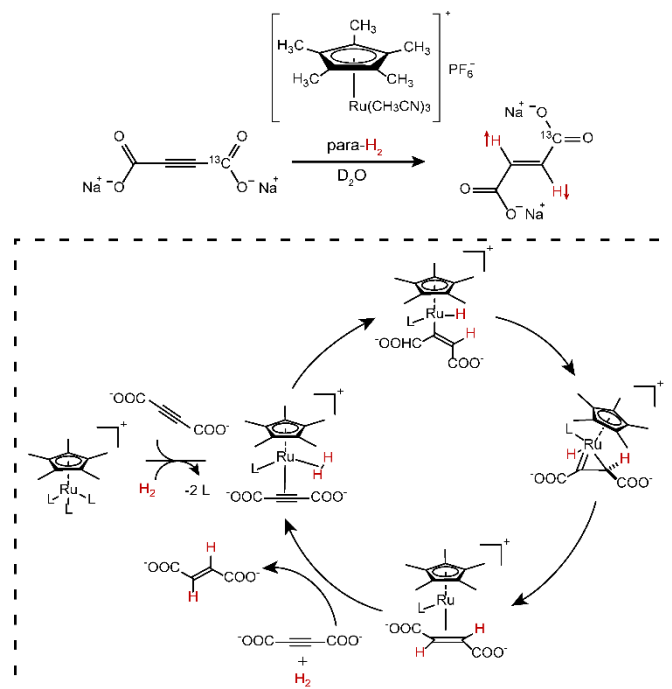


Fig 1. Acetylene [1-¹³C]dicarboxylate reacts with parahydrogen in the presence of [Cp^{*}Ru(CH₃CN)₃]PF₆ to form [1-¹³C]fumarate. The proton nuclear spin singlet state is indicated by opposing red arrows. The expected catalyst mechanism, modified from reference [19], is shown in the box.

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hydrogenation of acetylene dicarboxylate with parahydrogen using the most widely used hydrogenative PHIP catalyst, i.e. [1,4-bis(diphenylphosphino)butane](1,5-cyclooctadiene)rhodium(I) tetrafluoroborate [22–24]. Maleate itself is a toxic compound, so for *in vivo* application a second hydrogenative step is used to produce the metabolite succinate [24].

If ^{13}C nuclei are absent, the protons in fumarate are chemically and magnetically equivalent, so the hyperpolarized proton singlet order remains in an NMR-silent form, and enhanced NMR signals cannot be directly observed. The hyperpolarized ^1H NMR signals can be released by enzymatic conversion to malate [25], or chemical desymmetrization [26]. In the cases of maleate and succinate, magnetic inequivalence caused by a difference in J_{CH} couplings to a ^{13}C spin has been used to convert the ^1H singlet order into observable ^{13}C magnetisation [23,24]. Here we use the same principle: ^{13}C labelling of one of the carboxylate moieties breaks the magnetic equivalence of the protons, which have different ^1H - ^{13}C J -couplings to the ^{13}C label. Since the protons remain close to magnetic equivalence, the application of a simple $\pi/2$ pulse on ^{13}C leads to relatively weak hyperpolarized ^{13}C NMR signals, which are due to singlet-triplet mixing [27]. A specific pulse sequence such as S2hM (singlet-to-heteronuclear magnetization) [23][28], on the other hand, leads to full conversion of proton singlet order into ^{13}C magnetization, leading to strongly enhanced ^{13}C NMR signals.

The S2hM pulse sequence, shown in Figure 2, exploits the molecular J -couplings (and not chemical shifts) to achieve polarization transfer, and is optimised for the case of small heteronuclear coupling to the parahydrogen proton pair. The sequence is designed to work independent of magnetic field homogeneity [23], which is an improvement on similar sequences [29,30]. It can additionally be made very robust to RF field inhomogeneity by the use of composite pulses, and can therefore be readily implemented on large sample volumes at low magnetic field, as is commonly the case in a clinical setting.

It should be noted that although *trans*-hydrogenation has been used before in combination with parahydrogen [17–20], the product molecules formed in those cases have chemically inequivalent proton pairs, giving rise to so-called PASADENA-enhanced NMR signals. This approach is ineffective for releasing the proton singlet order as observable magnetization for the chemically equivalent protons of fumarate.

To generate hydrogen gas at 50% para enrichment, hydrogen gas (purity 99.995%) was passed through a home-built parahydrogen generator containing an iron(III) oxide

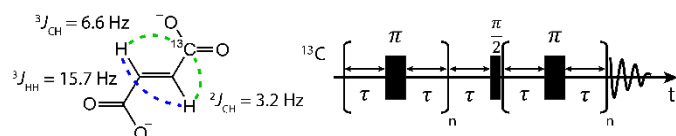


Fig 2. The S2hM pulse sequence used to convert proton singlet order into ^{13}C magnetization. The delay is given by $\tau = 1 / (4\sqrt{J_{\text{HH}}^2 + (\Delta J_{\text{CH}}/2)^2})$ and the number of loops is $n = \text{round}(\pi / (4\arctan(\Delta J_{\text{CH}}/2J_{\text{HH}})))$. The fumarate J -couplings are shown. The values of τ and n used in experiments were $\tau = 15.8$ ms and $n = 7$.

catalyst cooled to 77 K. The solution prior to hydrogenation reactions was 6 mM $[\text{Cp}^*\text{Ru}(\text{CH}_3\text{CN})_3]\text{PF}_6$, 100 mM disodium acetylene $[1-^{13}\text{C}]$ dicarboxylate and 100 mM sodium sulphite in D_2O . For sample hydrogenation, parahydrogen gas was bubbled at 5 bar and 50 °C for 30 s through 1/16 inch PEEK tubing that extended to the bottom of a pressurizable 5 mm NMR tube. The bubbling was performed inside the magnet and was controlled manually by hand-operated valves. NMR experiments were performed at 11.7 T in a 5 mm BBO probe using a Bruker AVANCE III console.

To demonstrate hyperpolarization of the ^{13}C spins, we hydrogenated the sample as described, and then applied the S2hM pulse sequence optimised for the J -couplings in fumarate on the ^{13}C channel before ^{13}C detection. For comparison, the hyperpolarized NMR signal was allowed to decay, and a thermal equilibrium spectrum was acquired with a $\pi/2$ pulse. The results are shown in Figure 3. A typical ^{13}C spectrum of $[1-^{13}\text{C}]$ fumarate shows a triplet peak structure from J -coupling to the strongly coupled proton pair. However, the S2hM sequence enhances only the central peak of the expected triplet [23], and this observation is supported by a SpinDynamica [31] simulation of the spectrum. The out-of-phase outer peaks are given by weakly allowed proton singlet-triplet state mixing, and are not discussed further here. There is no evidence of formation of the *cis*-product maleate in the ^{13}C NMR spectra, which would appear at 167.3 ppm.

To quantify the catalyst selectivity for *trans*-hydrogenation over *cis*-hydrogenation, we acquired ^1H NMR spectra of the reaction mixture after sample hydrogenation (see supplementary information). There is no detectable maleate signal in a 32 transient ^1H spectrum, and the ratio of *trans*:*cis* product is therefore over 500:1. If the sodium sulphite is omitted, this the ratio is only 3:1. This modification to the catalyst activity by sodium sulphite is not currently understood, and is subject to ongoing research. Additionally, succinate is produced as a side-product in 9% yield compared to fumarate. Succinate is a metabolite safe for injection *in vivo*, so this is less concerning than the formation of maleate, which is toxic. It has previously been shown that succinate forms through a Ru-carbene intermediate in a separate catalytic pathway, and not form the over-reduction of fumarate [19,20]. Importantly, this means that once fumarate is formed, it is not over-hydrogenated to succinate or isomerised to maleate.

The signal enhancement after the S2hM sequence was measured to be over 1000 at a field of 11.7 T, corresponding to a polarization level of over 1%. We plan to improve this enhancement factor and reaction yield by optimizing the reaction conditions, i.e. parahydrogen pressure, temperature, increasing the catalyst concentration, reducing the reaction time to limit losses due to relaxation, and reducing the precursor concentration to achieve complete hydrogenation. The proton singlet order generated from parahydrogen can also be increased by a factor of 3 by equilibrating the hydrogen gas over a catalyst at 25 K, instead of the 77 K used here.

Spin relaxation times were measured on a sample of 20 mM $[1-^{13}\text{C}]$ fumarate in a 40 mM phosphate buffer (pH 7) in water. The data are shown in Table 1 for a sample with and without

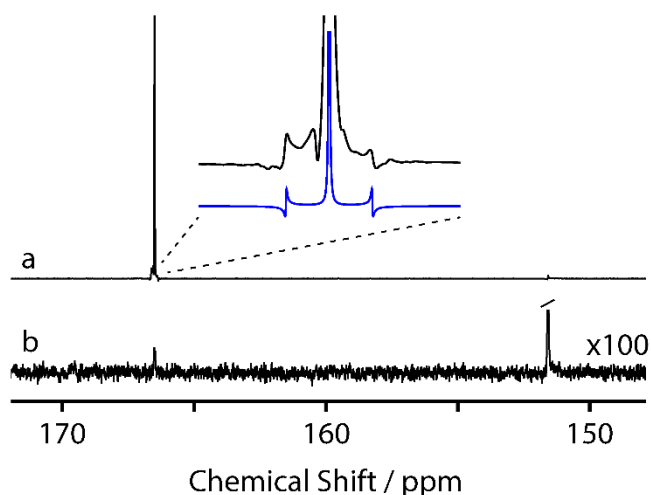


Fig 3. ^{13}C NMR spectra shown with 1 Hz line broadening, acquired without proton decoupling. a) Hyperpolarized signal acquired after applying the S2hM pulse sequence. An expansion of the signal is shown and compared to a SpinDynamica simulation in blue. b) Thermal equilibrium signal acquired in one scan, expanded by a factor of 100 for clarity. The peak assignments are as follows: acetylene $[1\text{-}^{13}\text{C}]$ dicarboxylate precursor (151.6 ppm), $[1\text{-}^{13}\text{C}]$ fumarate (166.5 ppm). The $[1\text{-}^{13}\text{C}]$ fumarate hyperpolarization is revealed when an S2hM sequence is used to convert the ^1H singlet order into ^{13}C magnetization, and the signal enhancement achieved here is over 1000.

removal of dissolved O_2 by 15 minutes of N_2 degassing, and measurement techniques are described in the supplementary information.

	$^1\text{H } T_1 / \text{s}$	$^1\text{H } T_2 / \text{s}$	$^{13}\text{C } T_1 / \text{s}$
N_2 degassed	22.6 ± 0.3	46 ± 7	28 ± 2
Not degassed	15.3 ± 0.2	24 ± 5	26 ± 2

Table 1. Spin relaxation data.

The ^1H singlet relaxation time T_2 is longer than the spin-lattice relaxation time T_1 because the singlet state is immune to intra-pair ^1H - ^1H dipolar relaxation [32]. Nevertheless, it is clear that oxygen significantly impacts both T_1 and T_2 . The $^1\text{H } T_2$ in the degassed solution is relatively long, which is appealing because time is needed during the parahydrogenation step to build up significant quantities of hyperpolarized material, for catalyst removal and sample transport.

The $^{13}\text{C } T_1$ is probably limited by the large chemical shift anisotropy, and is almost independent of oxygen concentration at 11.7 T; this means the $^{13}\text{C } T_1$ is probably longer at lower field strengths. The authors hope to combine this polarization method with a known procedure to extend the $^{13}\text{C } T_1$ by precipitation of fumarate as a solid, and store the hyperpolarization for long times [33].

Given the relatively long-lived nature of the proton singlet state, this might provide a means to perform metabolic imaging using hyperpolarized ^1H NMR, which is more sensitive than ^{13}C detection. The presence of a ^{13}C label allows the hyperpolarized singlet order in fumarate to be converted into ^1H magnetization by a S2M sequence [34]. Even in the absence of a ^{13}C label, the hyperpolarized proton singlet order of fumarate may be liberated by enzymatic conversion to malate [25].

The proton singlet lifetime is expected to be much longer (many minutes) [25,26,35] in the non ^{13}C -labelled molecule, because the ^1H - ^{13}C dipolar coupling relaxation contribution is removed. This is appealing for the production of hyperpolarized fumarate, but the oxygen sensitivity should still limit the T_2 for *in vivo* application.

In summary, a method for producing hyperpolarized fumarate from parahydrogen using a ruthenium-based catalyst in water has been presented. With radiofrequency pulse sequences we converted the hyperpolarized singlet order into magnetization on the ^{13}C spin in $[1\text{-}^{13}\text{C}]$ fumarate. Although the achieved polarization level is relatively low in this preliminary work, there is much room for optimization of the catalyst, the parahydrogen preparation, and the reaction conditions. If these important issues are addressed, this might lead to a practical route for the convenient and inexpensive preparation of hyperpolarized fumarate for magnetic resonance spectroscopy and imaging experiments.

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Conflicts of interest

Conflict of Interest: Hana Kouřilová is a co-founder of Hyperspin Scientific Ltd, U.K.

Notes and references

- Münnemann, K.; Spiess, H. W. Nuclear magnetic resonance: The art of signal enhancement. *Nat. Phys.* 2011, **7**, 522-8
- Ross, B. D.; Bhattacharya, P.; Wagner, S.; Tran, T.; Sailasuta, N. Hyperpolarized MR imaging: neurologic applications of hyperpolarized metabolism. *Am. J. Neuroradiol.* 2010, **31**, 24-33.
- Park, I.; Larson, P. E.; Zierhut, M. L.; Hu, S.; Bok, R.; Ozawa, T.; Kurhanewicz, J.; Vigneron, D. B.; VandenBerg, S. R.; James, C. D.; Nelson, S. J. Hyperpolarized ^{13}C magnetic resonance metabolic imaging: application to brain tumors. *Neuro Oncol.* 2010, **12**, 133-144.
- Gallagher, F. A.; Kettunen, M. I.; Hu, D.-E.; Jensen, P. R.; Karlsson, M.; Gisselsson, A.; Nelson, S. K.; Witney, T. H.; Bohndiek, S. E.; Hansson, G.; Peitersen, T.; Lerche, M. H.; Brindle, K. M. Production of hyperpolarized $[1, 4\text{-}^{13}\text{C}]$ malate from $[1, 4\text{-}^{13}\text{C}]$ fumarate is a marker of cell necrosis and treatment response in tumors. *Proc. Natl. Acad. Sci. U.S.A.* 2009, **106**, 19801-19806.
- Miller, J. J.; Lau, A. Z.; Nielsen, P. M.; McMullen-Klein, G.; Lewis, A. J.; Jespersen, N. R.; Ball, V.; Gallagher, F. A.; Carr, C. A.; Laustsen, C.; Bøtker, H. E.; Tyler, D. J.; Schroeder, M. A. Hyperpolarized $[1, 4\text{-}^{13}\text{C}]$ Fumarate Enables Magnetic Resonance-Based Imaging of Myocardial Necrosis. *JACC Cardiovasc. Imaging* 2017, **12**, 133-144.
- Nielsen, P. M.; Eldirdiri, A.; Bertelsen, L. B.; Jørgensen, H. S.; Ardenkjaer, Larsen, J. H.; Laustsen, C. Fumarase activity: an *in vivo*

- and in vitro biomarker for acute kidney injury. *Sci Rep.* 2017, **7**, 40812.
- 7 Ardenkjær-Larsen, J. H.; Fridlund, B.; Gram, A.; Hansson, G.; Hansson, L.; Lerche, M. H.; Servin, R.; Thaning, M. & Golman, K. Increase in signal-to-noise ratio of > 10,000 times in liquid-state NMR. *Proc. Natl. Acad. Sci. U.S.A.* 2003, **100**, 10158-10163.
 - 8 Ardenkjær-Larsen, J. H. On the present and future of dissolution-DNP. *J. Magn. Reson.* 2016, **264**, 3-12.
 - 9 Bowers, C. R.; Weitekamp, D. P. Parahydrogen and synthesis allow dramatically enhanced nuclear alignment. *J. Am. Chem. Soc.* 1987, **109**, 5541-5542.
 - 10 Natterer, J.; Bargon, J. Parahydrogen induced polarization. *Prog. Nucl. Magn. Reson. Spectrosc.* 1997, **31**, 293-315.
 - 11 Chekmenev, E. Y.; Hövener, J.; Norton, V. A.; Harris, K.; Batchelder, L. S.; Bhattacharya, P.; Ross, B. D.; Weitekamp, D. P. PASADENA Hyperpolarization of Succinic Acid for MRI and NMR Spectroscopy. *J. Am. Chem. Soc.* 2008, **130**, 4212-4213.
 - 12 Iali, W.; Rayner, P. J.; Duckett, S. B. Using parahydrogen to hyperpolarize amines, amides, carboxylic acids, alcohols, phosphates, and carbonates. *Sci Adv.* 2018, **4**, No.1, eaao6250.
 - 13 Reineri, F.; Boi, T.; Aime, S. ParaHydrogen Induced Polarization of ¹³C carboxylate resonance in acetate and pyruvate. *Nat. Commun.* 2015, **6**, 5858.
 - 14 Truong, M. L.; Theis, T.; Coffey, A. M.; Shchepin, R. V.; Waddell, K. W.; Shi, F.; Goodson, B. M.; Warren, W. S.; Chekmenev, E. Y. 15N hyperpolarization by reversible exchange using SABRE-SHEATH. *J. Phys. Chem. C* 2015, **119**, 8786-8797.
 - 15 Roth, M.; Kindervater, P.; Raich, H.-P.; Bargon, J.; Spiess, H. W.; Münnemann, K. Continuous ¹H and ¹³C Signal Enhancement in NMR Spectroscopy and MRI Using Parahydrogen and Hollow-Fiber Membranes. *Angew. Chem., Int. Ed.* 2010, **49**, 8358-8362.
 - 16 Cavallari, E.; Carrera, C.; Sorge, M.; Bonne, G.; Muchir, A.; Aime, S.; Reineri, F. The ¹³C hyperpolarized pyruvate generated by ParaHydrogen detects the response of the heart to altered metabolism in real time. *Sci. Rep.* 2018, **8**, 8366.
 - 17 Schleyer, D.; Niessen, H. G.; Bargon, J. In situ ¹H-³¹P-NMR studies of the stereoselective hydrogenation of alkynes to (E)-alkenes catalyzed by a homogeneous [Cp* Ru]⁺ catalyst. *New J. Chem.* 2001, **25**, 423-426.
 - 18 Radkowski, K.; Sundararaju, B.; Fürstner, A. A Functional-Group-Tolerant Catalytic trans Hydrogenation of Alkynes. *Angew. Chem., Int. Ed.* 2013, **52**, 355-360.
 - 19 Leutzsch, M.; Wolf, L. M.; Gupta, P.; Fuchs, M.; Thiel, W.; Farès, C.; Fürstner, A. Formation of Ruthenium Carbenes by gem-Hydrogen Transfer to Internal Alkynes: Implications for Alkyne trans-Hydrogenation. *Angew. Chem., Int. Ed.* 2015, **54**, 12431-12436.
 - 20 Guthertz, A.; Leutzsch, M.; Wolf, L. M.; Gupta, P.; Rummelt, S. M.; Goddard, R.; Farès, C.; Thiel, W.; Fürstner, A. Half-Sandwich Ruthenium Carbene Complexes Link trans-Hydrogenation and gem-Hydrogenation of Internal Alkynes. *J. Am. Chem. Soc.* 2018, **140**, 3156-3169.
 - 21 Roşca, D.-A.; Radkowski, K.; Wolf, L. M.; Wagh, M.; Goddard, R.; Thiel, W.; Fürstner, A. Ruthenium-Catalyzed Alkyne trans-Hydrometalation: Mechanistic Insights and Preparative Implications. *J. Am. Chem. Soc.* 2017, **139**, 2443-2455.
 - 22 Franzoni, M. B.; Buljubasich, L.; Spiess, H. W.; Münnemann, K. Long-lived ¹H singlet spin states originating from para-hydrogen in Cs-symmetric molecules stored for minutes in high magnetic fields. *J. Am. Chem. Soc.* 2012, **134**, 10393-10396.
 - 23 Eills, J.; Stevanato, G.; Bengs, C.; Glöggler, S.; Elliott, S. J.; Alonso-Valdesueiro, J.; Pileio, G.; Levitt, M. H. Singlet order conversion and parahydrogen-induced hyperpolarization of ¹³C nuclei in near-equivalent spin systems. *J. Magn. Reson.* 2017, **274**, 163-172.
 - 24 Bhattacharya, P.; Chekmenev, E. Y.; Perman, W. H.; Harris, K. C.; Lin, A. P.; Norton, V. A.; Tan, C. T.; Ross, B. D.; Weitekamp, D. P. Towards hyperpolarized ¹³C-succinate imaging of brain cancer. *J. Magn. Reson.* 2007, **186**, 150-155.
 - 25 Bornet, A.; Ji, X.; Mammoli, D.; Vuichoud, B.; Milani, J.; Bodenhausen, G.; Jannin, S. Long-Lived States of Magnetically Equivalent Spins Populated by Dissolution-DNP and Revealed by Enzymatic Reactions. *Chem. Eur. J.* 2014, **20**, 17113-17118.
 - 26 Zhang, Y.; Duan, X.; Soon, P. C.; Sychrovsky, V.; Canary, J. W.; Jerschow, A. Limits in Proton Nuclear Singlet-State Lifetimes Measured with para-Hydrogen-Induced Polarization. *ChemPhysChem* 2016, **17**, 2967-2971.
 - 27 Pravdivtsev, A. N.; Ivanov, K. L.; Yurkovskaya, A. V.; Vieth, H. -M.; Sagdeev, R. Z. Spontaneous transfer of para-hydrogen induced polarization to ¹³C spins in symmetric molecules. *Dokl. Phys. Chem.*, 2015, **464**, 247-250
 - 28 Stevanato, G.; Eills, J.; Bengs, C.; Pileio, G. A pulse sequence for singlet to heteronuclear magnetization transfer: S2hM. *J. Magn. Reson.* 2017, **277**, 169-178.
 - 29 Goldman, M.; Johannesson, H.; Axelsson, O.; Karlsson, M. Design and implementation of C-13 hyperpolarization from para-hydrogen, for new MRI contrast agents. *C. R. Chim.* 2006, **9**, 357-363.
 - 30 Kadlecsek, S.; Emami, K.; Ishii, M.; Rizi, R. Optimal transfer of spin-order between a singlet nuclear pair and a heteronucleus. *J. Magn. Reson.* 2010, **205**, 9-13
 - 31 Bengs, C.; Levitt, M. H. SpinDynamica: Symbolic and numerical magnetic resonance in a Mathematica environment. *Magn. Reson. Chem.* 2018, **56** (6), 374-414.
 - 32 Carravetta, M.; Johannessen, O. G.; Levitt, M. H. Beyond the T₁ Limit: Singlet Nuclear Spin States in Low Magnetic Fields. *Phys. Rev. Lett.* 2004, **92**, 153003.
 - 33 Eills, J.; Alonso-Valdesueiro, J.; Salazar Marcano, D. E.; Ferreira da Silva, J.; Alom, S.; Rees, G. J.; Hanna, J. V.; Carravetta, M.; Levitt, M. H. Preservation of Nuclear Spin Order by Precipitation. *ChemPhysChem* 2018, **19**, 40-44.
 - 34 Pileio, G.; Carravetta, M.; Levitt, M. H. Storage of nuclear magnetization as long-lived singlet order in low magnetic field. *Proc. Natl. Acad. Sci. U.S.A.* 2010, **107**, 17135-17139.
 - 35 Franzoni, M. B.; Buljubasich, L.; Speiss, H. W.; Münnemann, K. Long-Lived ¹H Singlet Spin States Originating from Para-Hydrogen in Cs-Symmetric Molecules Stored for Minutes in High Magnetic Fields. *J. Am. Chem. Soc.* 2012, **134**, 10393-10396.