

1 **Title: Evolutionary history of the European free-tailed-bat, a tropical affinity species**  
2 **spanning across the Mediterranean Basin**

3 **Short running title: Evolutionary history of tropical affinity taxa**

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5 Francisco Amorim<sup>a,b,\*</sup>, Orly Razgour<sup>c</sup>, Vanessa Mata<sup>a,b</sup>, Susana Lopes<sup>a</sup>, Raquel Godinho<sup>a,b,d</sup>,  
6 Carlos Ibáñez<sup>e</sup>, Javier Juste<sup>e</sup>, Stephen J. Rossiter<sup>f</sup>, Pedro Beja<sup>a,g</sup>, Hugo Rebelo<sup>a,g</sup>

7 <sup>a</sup> CIBIO-InBIO, Research Center in Biodiversity and Genetic Resources, University of Porto,  
8 Vairão, Portugal

9 <sup>b</sup> Departamento de Biologia, Faculdade de Ciências, Universidade do Porto, Porto, Portugal

10 <sup>c</sup> Biological Sciences, University of Southampton, Southampton, UK

11 <sup>d</sup> Department of Zoology, University of Johannesburg, South Africa

12 <sup>e</sup> Estación Biológica de Doñana (CSIC), Seville, Spain

13 <sup>f</sup> School of Biological & Chemical Sciences, Queen Mary University of London, London, UK

14 <sup>g</sup> CIBIO-InBIO, Research Center in Biodiversity and Genetic Resources, Institute of  
15 Agronomy, University of Lisbon, Lisbon, Portugal

16 \* Corresponding author: F. Amorim, CIBIO-InBIO, Centro de Investigação em Biodiversidade  
17 e Recursos Genéticos, Universidade do Porto, Campus de Vairão, Rua Padre Armando  
18 Quintas, nº 7, 4485-661 Vairão, Portugal. Email: famorim@cibio.up.pt

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21

22 **Abstract**

23 The Mediterranean Basin is a global biodiversity hotspot, hosting a number of native species  
24 belonging to families that are found almost exclusively in tropical climates. Yet, whether or  
25 not these taxa were able to survive in the Mediterranean region during the Quaternary  
26 climatic oscillations remains unknown. Focusing on the European-free-tailed bat (*Tadarida*  
27 *teniotis*) we aimed to i) identify potential ancient populations and glacial refugia; ii) determine  
28 the post-glacial colonization routes across the Mediterranean; and iii) evaluate current  
29 population structure and demography. Mitochondrial and nuclear markers were used to  
30 understand *T. teniotis* evolutionary and demographic history. We show that *T. teniotis* is  
31 likely restricted to the Western Palearctic, with mitochondrial phylogeny suggesting a split  
32 between an Anatolian/Middle East clade and a European clade. Nuclear data pointed to  
33 three genetic populations, one of which is an isolated and highly differentiated group in the  
34 Canary Islands, another distributed across Iberia, Morocco and France, and a third  
35 stretching from Italy to the east, with admixture following a pattern of isolation by distance.  
36 Evolutionary and demographic reconstruction supports a pre Last Glacial Maximum (LGM)  
37 colonization of Italy and the Anatolian/Middle East, while the remaining populations were  
38 colonized from Italy after the Younger Dryas. We also found support for demographic  
39 expansion following the Iberian colonization. The results show that during the LGM *T.*  
40 *teniotis* persisted in Mediterranean refugia and has subsequently expanded to its current  
41 circum-Mediterranean range. Our findings raise questions regarding the physiological and  
42 ecological traits that enabled species with tropical affinities to survive in colder climates.

43

## 44 1. Introduction

45 The Mediterranean Basin is a global biodiversity hotspot (Blondel, Aronson, Bodiou, &  
46 Boeuf, 2010; Myers, Mittermeier, Mittermeier, da Fonseca, & Kent, 2000). Despite being  
47 presently located in temperate latitudes, this region was mainly covered by tropical climates  
48 during the Tertiary (Blondel & Mourer-Chauviré, 1998). Nowadays, Europe still hosts a  
49 number of members belonging to several vertebrate groups that are almost exclusively  
50 associated with the tropics (defined here as tropical affinities), including reptiles such as  
51 geckos and chameleons, and birds such as rollers and bee-eaters (Ammerman, Lee, &  
52 Tipps, 2012; Blondel & Mourer-Chauviré, 1998; Carranza & Arnold, 2006; Townsend &  
53 Larson, 2002). However, the diversity of tropical species present in Europe is lower than that  
54 of other Holarctic areas like North America or eastern Asia (Blondel & Mourer-Chauviré,  
55 1998). The reason for such pattern is that both North America and eastern Asia remained  
56 connected to the tropics over the whole Tertiary–Quaternary. In contrast, large geographical  
57 barriers (mountain ranges, seas and desert-belts) prevented the Palearctic tropical biota  
58 from expanding their range to tropical regions further south during glacial periods, and  
59 tropical species from colonizing northern regions during inter-glacial periods (Blondel &  
60 Mourer-Chauviré, 1998). Altogether, these led to a progressive decline of the tropical  
61 species during the Pleistocene (Blondel et al., 2010). Under such circumstances, it is  
62 remarkable that some of these species were able to persist in the western Palaeartic,  
63 although mostly restricted to the circum-Mediterranean area. The population history of such  
64 lineages during periods of glaciation is poorly understood and it is not known whether these  
65 taxa were able to survive in the Mediterranean region during the climatic oscillations of the  
66 Quaternary

67 Among non-flying mammals, only a small number of species in the western Palaeartic have  
68 tropical affinities (Dobson, 1998). Although in some cases this was the result of a  
69 longstanding human-mediated introductions across the Strait of Gibraltar, in others, such as  
70 the Egyptian mongoose (*Herpestes ichneumon*), this was the result of natural dispersal into  
71 the Iberian Peninsula during the Late Pleistocene (Gaubert et al., 2011). In bats, which are  
72 likely to be able to disperse over greater distances, there is a higher number of species  
73 shared between north-west Africa and Iberia (Dobson, 1998; García-Mudarra, Ibáñez, &  
74 Juste, 2009), but even for these mammals the number of species with tropical affinities  
75 occurring in temperate regions is relatively low. The European-free-tailed bat (*Tadarida*  
76 *teniotis* Rafinesque, 1814) is the only European representative of the Molossidae family that  
77 comprises more than 110 species (Ammerman et al., 2012). All the remaining molossids are  
78 restricted to tropical regions, apart from the Mexican free-tailed bat (*Tadarida brasiliensis*)  
79 and the Big free-tailed bat (*Nyctinomops macrotis*), which reach similar Northern latitudes in

80 the American continent. Molossidae is an ancient bat family that split into Old and New  
81 World molossids ca. 29 million years ago (Ammerman et al., 2012), and fossil records of the  
82 genus *Tadarida* in Europe date from the late Eocene ca. 25 million years ago (De Bonis et  
83 al., 1973).

84 Understanding phylogeographic patterns shaping the distributions and expansion of species  
85 is a powerful tool for predicting how future climatic changes will shape regional biodiversity  
86 (Hickerson et al., 2010). During the Quaternary ice ages, Europe experienced dramatic  
87 climatic fluctuations between glacial and interglacial cycles contributing to the contemporary  
88 distribution and genetic composition of biodiversity (G. Hewitt, 2000). The distributions of  
89 many animal species have been severely restricted to refugia to escape the harsh conditions  
90 of the glacial periods. The Last Glacial Maximum (LGM 18–20 ka BP), and the Younger  
91 Dryas (11.7–12.9 ka BP), correspond to the latest episodes where the ice sheets and cold  
92 temperatures reached their extremes. The Mediterranean region encompasses a high  
93 habitat diversity combined with topographic and geographic variability. Together with a  
94 dynamic palaeogeographic and climatic history these features contributed to marked  
95 environmental gradients (Blondel et al., 2010), strongly shaping current species and  
96 biodiversity spatial patterns, population structure and demography (G. M. Hewitt, 1999).  
97 Despite the increasing number of studies focusing on the phylogeography of species native  
98 to temperate environments, to the best of our knowledge, representatives from tropical  
99 families living in such environments have been seldom studied (but see Paulo, Pinto,  
100 Bruford, Jordan, & Nichols, 2002; Rato, Carranza, & Harris, 2011).

101 The European-free-tailed-bat is widespread throughout the Mediterranean and occurs in a  
102 variety of environments and habitats from the colder Alps to the border of the Sahara desert  
103 (Amorim, Jorge, Beja, & Rebelo, 2018; Arlettaz et al., 2000; Bendjeddou, Bakhouché, &  
104 Bouslama, 2014). However, during the Late Glacial Maximum (LGM), large parts of Europe  
105 had colder and drier habitats (Frenzel, Pécsi, & Velichko, 1992) with warmest month  
106 temperature being 10 °C cooler than present, and coldest month temperature 20 °C colder  
107 (Kageyama et al., 2006). These harsh conditions were likely unsuitable for most bat species  
108 (e.g., Bilgin et al., 2016; Kerth et al., 2008; Razgour et al., 2013; Rossiter, Benda, Dietz,  
109 Zhang, & Jones, 2007), thus raising the question of how species with tropical affinities were  
110 able to survive. Here we focus on the evolutionary history of *T. teniotis*, which belongs to a  
111 taxonomical family almost exclusively associated with the tropics and shows shorter duration  
112 of torpor bouts, and higher minimal body temperature in torpor than other temperate bats  
113 (Arlettaz et al., 2000). The high mobility and fast flight of these bats (Mata et al., 2016;  
114 McCracken et al., 2008) allows them to respond fast to environmental changes by shifting to  
115 more suitable areas. These features render *T. teniotis* a suitable model species to

116 understand how species with topical affinity reacted to the climatic oscillations of the  
117 Quaternary in temperate and subtropical regions. Therefore, our main aims were to: i)  
118 identify the location of potential ancient populations and glacial refugia; ii) determine the  
119 post-glacial colonization routes across the Mediterranean; and iii) evaluate current  
120 population structure and demography in light of the post-glacial colonisation history.

121

## 122 **2. Methods**

### 123 *2.1. Sample collection*

124 A total of 154 genetic samples collected across the Western and Central Palearctic were  
125 obtained from researchers and museum collections. Samples spanned the entire range  
126 although coverage was uneven with few samples available from some regions, particularly  
127 from Asia, Eastern Mediterranean and North Africa. For a complete list of samples, origin  
128 and providers see Appendix 1 (GenBank accession numbers MK817165 to MK817272).

### 129 *2.2. DNA extraction*

130 Due to the different nature of the samples obtained (old museum specimens and recently  
131 collected wing tissue) we used different DNA extraction methods. For older museum  
132 specimens we followed the ancient DNA extraction protocol described in Rohland &  
133 Hofreiter, (2007) with modifications described in Dabney et al. (2013). For recent tissue  
134 samples, we used DNA Micro Kits (QIAGEN) following the manufacturer's instructions.

### 135 *2.3. Validation of species identity and mitochondrial genotyping*

136 Given the poorly resolved taxonomic status of *Tadarida teniotis* (Mata, Amorim, Guillén-  
137 Servent, Beja, & Rebelo, 2017), the identity of all samples were verified using mitochondrial  
138 markers prior to microsatellite genotyping. Due to taxonomic uncertainties (Mata et al.,  
139 2017), verification was considered to be especially important for putative *T. teniotis* samples  
140 obtained from the eastern part of the distribution (Kyrgyzstan and China). Additionally,  
141 samples from Laos previously identified as *T. latouchei* were also checked.

142 Four mitochondrial primer pairs were specifically designed using Geneious v9.1.7  
143 (<http://www.geneious.com>, Kearse et al. 2012) based on an alignment of 37 mitogenomes  
144 covering the species range. The primers were designed to amplify the most variable regions  
145 of the mitogenomes (Supporting Information Table S1) and corresponded to three coding  
146 regions (*COI* - *cytochrome c oxidase subunit I*, *ATP6* - *ATP synthase subunit 6*, and *CytB* -  
147 *cytochrome b*) and one noncoding region (*D-loop*). While designing the primers took extra  
148 precautions and carefully examined the mitogenomic data to avoid the amplification of

149 nuclear copies covering almost the entire mitogenome. We did this by comparing the  
150 sequences containing nuclear copies (identified by the high prevalence of stop codons) to  
151 those without nuclear copies and selecting the regions that did not amplify nuclear copies.  
152 This way the primers designed assure that only the mitochondrial haplotype were amplified,  
153 allowing the genotyping of samples through Sanger sequencing. For highly degraded  
154 museum samples that did not amplify using the regular primers, we further developed  
155 internal primers for the *COI* (*COI-mini*) and *D-loop* (*D-loop-mini*) regions targeting key SNPs  
156 that enable to differentiate *T. teniotis* and its different haplogroups from *T. latouchei*  
157 (Supporting Information Table S1).

158 The PCR reactions were carried in volumes of 10  $\mu$ L, comprising of 5  $\mu$ L of Multiplex PCR  
159 Master Mix (QIAGEN), with 0.4  $\mu$ L of each 10  $\mu$ M primer, and 1  $\mu$ L of DNA extract. Cycling  
160 conditions for *COI*, *ATP6*, *CytB*, and *D-loop* used initial denaturing at 95  $^{\circ}$ C for 15 min,  
161 followed by 40 cycles of denaturing at 94  $^{\circ}$ C for 30 s, annealing at 59  $^{\circ}$ C for 45 s and  
162 extension at 72  $^{\circ}$ C for 45 s, with a final extension at 72  $^{\circ}$ C for 10 min. For *COI-mini* and *D-*  
163 *loop-mini* the cycling conditions were the same except the annealing temperature that was  
164 52  $^{\circ}$ C and the number of cycles was increased to 45. Successful amplifications were  
165 enzymatically purified, sequenced following the BigDye Terminator v3.1 Cycle sequencing  
166 protocol (Applied Biosystems), and sequencing products were separated using an  
167 automated Sequencer ABI3130xl Genetic Analyzer. Sequences were aligned and compared  
168 in the software SEQSCAPE 3.0 (Applied Biosystems).

#### 169 2.4. Microsatellite genotyping

170 A custom microsatellite library was developed through 454 GS-FLX Titanium  
171 pyrosequencing of enriched DNA libraries based on 12 individuals along the distribution  
172 range of *T. teniotis* (Malausa et al., 2011). This process was developed by GenoScreen  
173 ([http://www.pasteur-lille.fr/fr/recherche/plateformes/tordeux\\_plat.html](http://www.pasteur-lille.fr/fr/recherche/plateformes/tordeux_plat.html)) and included  
174 sequence data quality control, assembly and analyses, and primer design.

175 From the 159 candidate microsatellite loci, we selected 26 microsatellites with different  
176 numbers of repeat units, compatible allelic ranges and melting temperatures for multiplexing.  
177 We first tested the genotyping performance on four *T. teniotis* samples and discarded  
178 microsatellites that: i) showed no amplification, ii) had multiple bands and iii) had excessive  
179 slippage (many stutter bands). Those remaining were combined into two multiplex panels  
180 according to their allele size range and compatibility among primers, which was checked  
181 using Auto-Dimer (Vallone & Butler, 2004).

182 The optimisation of PCR conditions for multiplex loci and polymorphism detection was  
183 performed using 16 samples. From the 26 loci initially checked a total of 12 di and 2 tetra-

184 nucleotides polymorphic markers (with more than 2 alleles) were selected and genotyped for  
185 129 individuals in two multiplex panels with seven markers each. PCR fragments were  
186 fluorescent labelled following Schuelke (2000) but with FAM, VIC, NED, and PET dyes. A pig  
187 tail (GTTT) was added to the 5' end of the primer reverse in order to reduce stutter and drive  
188 the reaction to the "plusA" band (Brownstein, Carpten, & Smith, 1996). For additional details  
189 on microsatellite primers, see Supporting Information Table S2.

190 PCR amplifications were conducted as for mitochondrial fragments except that 1  $\mu$ L of primer  
191 mix was used per reaction. The PCR cycling profile was divided in four main steps:  
192 denaturation at 95 °C for 15 min; 13 cycles with denaturation at 95 °C for 30 s, annealing at  
193 58 °C for 90 s with a touchdown of 0.5 °C per cycle and extension at 72 °C for 45 s; 27  
194 cycles with denaturation at 95 °C for 30 s, annealing at 52 °C for 60 s and extension at 72 °C  
195 for 45 s; and a final extension at 60 °C for 30 min. PCR products were later separated by  
196 capillary electrophoresis on the same automatic sequencer ABI3130xl Genetic Analyzer (AB  
197 Applied Biosystems). Fragments were scored using GENEMAPPER V4.0 (Applied Biosystems)  
198 and checked independently by two people.

## 199 2.5. Genetic data analysis

### 200 2.5.1. Mitochondrial data

201 Sequences from the four mitochondrial markers were concatenated and standard molecular  
202 diversity statistics calculated in ARLEQUIN 3.5 (Excoffier & Lischer, 2010). To test for  
203 geographical genetic structure, analyses of molecular variance (AMOVA) were carried out  
204 with 10,000 permutations and diversity measures were reported for geographic groups and  
205 assessed according to the degree of differentiation between regions ( $\Phi$ CT), between  
206 populations within regions ( $\Phi$ SC) and between all populations ( $\Phi$ ST). A median-joining (MJ)  
207 haplotype network was build using POPART (Leigh & Bryant, 2015) for each marker and for  
208 the concatenated sequences. Mitochondrial diversity was assessed considering seven  
209 geographic populations based on the common population structure of European bats (e.g.  
210 Bilgin et al., 2016; Razgour et al., 2013): 1) Canary Islands; 2) Iberian Peninsula (Portugal  
211 and Spain, excluding Canary Islands); 3) Morocco; 4) France; 5) Italy; 6) Greece; 7) Anatolia  
212 and 8) Middle East (Lebanon, Israel and Palestine).

213 Phylogenetic reconstruction was performed on the CIPRES Science Gateway V. 3.3 (Miller,  
214 Pfeiffer, & Schwartz, 2010) using Bayesian inference implemented in BEAST v1.8.4  
215 (Drummond, Suchard, Xie, & Rambaut, 2012) considering unique haplotypes only (n = 65)  
216 from the concatenated sequences and with inclusion of *T. latouchei* as outgroup (Mata et al.,  
217 2017, GenBank Accession numbers: NC\_036331 and KY581662). The best substitution

218 model scheme was determined using PARTITIONFINDER v2.1.1 (Lanfear, Frandsen, Wright,  
219 Senfeld, & Calcott, 2016). We used a coalescent tree prior under constant population. Three  
220 independent runs of  $10^8$  generations sampled every 1000 were combined in TRACER V1.7  
221 (Rambaut, Drummond, Xie, Baele, & Suchard, 2018) to confirm convergence on the same  
222 posterior distribution in the MCMC runs. The first  $10^7$  runs (10%) were discarded as burn-in.

### 223 2.5.2. Microsatellite data

224 To test for departures from Hardy–Weinberg and linkage equilibrium, both across the whole  
225 samples and within populations, we used the ‘pegas’ R package (Paradis, 2010). Loci that  
226 violated Hardy–Weinberg equilibrium in more than two populations were excluded from  
227 further analysis (Supporting Information Table S2). Allele frequencies and number of private  
228 alleles were estimated in GENETIX v4.05 (Belkhir, Borsa, Chikhi, Raufaste, & Bonhomme,  
229 2004) and mean allele frequency across all loci was calculated for each population.  
230 Estimates of expected heterozygosity ( $H_e$ ), observed heterozygosity ( $H_{obs}$ ) and allelic  
231 richness within populations, and differentiation ( $F_{st}$ ) among populations, were all calculated  
232 using the ‘PopGenReport’ R package (Adamack & Gruber, 2014). Relatedness among  
233 individuals was measured using the triadic maximum likelihood estimator (TrioML; Wang,  
234 2007) implemented in ‘related’ R package (Pew, Muir, Wang, & Frasier, 2015). This  
235 estimator was chosen because it allows for inbreeding and accounts for genotyping errors in  
236 the data.

237 Population genetic structure was first examined using the principal component analysis in  
238 ‘PopGenReport’ R package (Adamack & Gruber, 2014) followed by the Bayesian clustering  
239 analysis implemented in STRUCTURE 2.3.4 (Pritchard, Stephens, & Donnelly, 2000) with all  
240 genotyped samples. We performed 10 replicate runs of structure for each value of K, from K  
241 = 1 to 10, and we applied the admixture model with a burn-in of  $5 \times 10^5$  and a run length of  
242  $10^6$  with and without the prior population information (LOCPRIOR). The latter can often  
243 provide accurate inference of population structure and individual ancestry in datasets where  
244 the signal of structure is too weak to be found using the standard models (Hubisz, Falush,  
245 Stephens, & Pritchard, 2009). We used STRUCTURE HARVASTER v0.6.94 to visualize  
246 likelihood and detect the number of genetic groups that best fit the data (Earl & VonHoldt,  
247 2012). The Greedy algorithm of CLUMPP (Jakobsson & Rosenberg, 2007) was used to derive  
248 symmetric similarity coefficients (SSC) among replicate runs within each value of K. Groups  
249 of runs with an  $SSC \geq 0.8$  were then combined and their outputs for each value of K were  
250 graphically displayed.

251 Spatial structuring was further analysed using multivariate analyses of spatial genetic  
252 patterns in ‘adegenet’ (Jombart, 2008). Spatial Analysis of Principal Components (sPCA)



253 allows to find the individual scores that maximize the product of variance and spatial  
254 autocorrelation (Jombart, Devillard, Dufour, & Pontier, 2008). Isolation by Distance (IBD)  
255 across all individuals within the species range was tested for in the R using the package  
256 'ade4' (Bougeard & Dray, 2018) and using a Mantel test.

## 257 2.6. ABC inference of evolutionary and demographic history

### 258 2.6.1. General overview

259 The evolutionary and demographic history of *T. teniotis* was reconstructed using  
260 Approximate Bayesian Computation (ABC) approach implemented in DIYABC v2.1 (Cornuet  
261 et al., 2014). We carried out two sets of analyses, aimed to: 1) infer the source population  
262 and patterns of range colonisation from putative refugia in the Western Palearctic; and 2)  
263 infer demographic history in the western range (Iberia, Morocco and France). In the first  
264 step, we modelled the probability of different scenarios considering 122 individuals from six  
265 populations (Iberia, Morocco, France, Italy, Greece and Anatolia/Middle East) and combining  
266 information from 12 microsatellites loci and two mitochondrial sequences (*COI* and *D-loop*).  
267 Multiple scenarios were compared representing a comprehensive range of alternative  
268 phylogeographic hypothesis and permuting the six geographic groups at the tips (Supporting  
269 Information Fig. S1 and Table S3).

270 Using the scenario topology identified in the first step, we carried out a demographic history  
271 analysis of the western range to determine changes in population size during colonisation.  
272 We compared a null model of no change in population size (Scenario 1) to a model of  
273 colonization and expansion in all populations (Scenario 2), and two models of recent change  
274 with increase or decrease in Iberian population size (Scenario 3 and Scenario 4  
275 respectively). For a schematic representation of the different scenarios, see Supporting  
276 Information Fig. S2.

277 Each scenario was tested using the combined microsatellite and mtDNA datasets and  
278 running  $10^6$  simulations. The posterior probability of scenarios was then estimated using a  
279 weighted polychotomous logistic regression. Due to the criticism of ABC model choice  
280 outlined in Robert, Cornuet, Marin, & Pillai (2011) we empirically evaluated the power of the  
281 model to discriminate among scenarios by simulating pseudo-observed datasets and  
282 calculating false allocation rates (type1 and 2 errors, Cornuet, Ravigné, & Estoup, 2010).  
283 Further details on the methods, model specifications and run parameters are presented in  
284 the following sections and in Supporting Information Table S3.

### 285 2.6.2. Specific model parameters

286 Microsatellite loci were assumed to follow a Generalized Stepwise Mutation model (GSM)  
287 and mean mutation rate was bounded between  $10^{-3}$  and  $10^{-4}$  (Balloux & Lugon-Moulin, 2002;  
288 Storz & Beaumont, 2002). For mtDNA we only considered *COI* and *D-loop* due to  
289 computational requirements and sequence completeness. We used the best substitution  
290 model scheme determined using PARTITIONFINDER v2.1.1 (Lanfear et al., 2016) as follow:  
291 HKY for the coding region (*COI*) and K80 for non-coding region (*D-loop*). Generation time  
292 was set at three years, a value in between the age of first breeding for different bat families  
293 that can go from one to five years (Crichton & Krutzsch, 2000) which meets our expectations  
294 for *T. teniotis*. We considered a mean mutation rate (per site per generation) between  $5.25E^{-8}$   
295 and  $7.2E^{-8}$  for *COI* (Juste et al., 2004; Ruedi & Mayer, 2001) and between  $9.45E^{-8}$  and  
296  $3.75E^{-7}$  for *D-loop* (Petit, Excoffier, & Mayer, 1999).

297 In the colonization analysis, uniform priors were assumed for all demographic parameters.  
298 Effective population size ( $N_e$ ) was kept as equal for all populations, ranging between  $1E^3$   
299 and  $1E^6$ . Population divergence time priors were bounded between  $1E^3$  and  $2E^5$  generations  
300 and varied depending on model analysis. Divergence times between source populations  
301 were set at either pre-LGM ( $1E^4$  -  $2E^5$ ) or flexible pre-post LGM ( $1E^3$  -  $2E^4$ ). Priors for  
302 admixture rates were bounded between 0.01 and 0.99. In the demographic history analysis,  
303 we used variable Effective population size ranging from 10 to  $1E^6$ . Population divergence  
304 time priors were bounded to post-LGM (10 and  $1E^4$ ) and varied depending on model  
305 analysis.

306 In each ABC analysis we used 269 summary statistics. For the microsatellite loci we used  
307 three single sample statistics (mean number of alleles, mean Nei's genetic diversity index  
308 and mean allele size variance), and five between-sample statistics ( $F_{ST}$ , mean number of  
309 alleles, mean genic diversity, mean allele size variance and shared allele distance). For the  
310 mtDNA sequence we used seven single sample statistics (number of distinct haplotypes,  
311 number of segregating sites, mean pairwise differences, variance of pairwise distance,  
312 Tajima's D statistics, private segregating sites, mean of numbers of the rarest nucleotide at  
313 segregation site) and four between-sample statistics ( $F_{ST}$ , number of haplotypes, number of  
314 segregating sites, mean within sample pairwise differences and number of segregating  
315 sites). The demographic history analysis included only 47 summary statistics due to the  
316 small number of groups compared.

317 The complete list of parameters used in the ABC analysis, respective priors and estimated  
318 results for the most supported colonization scenario (SC2) and the most supported  
319 demographic history scenario (SC2) can be found in Supporting Information Table S4.

### 320 2.6.3. Colonization analysis

321 This analysis included the potential range colonization from an ancient unsampled  
322 population with unknown origin. For a schematic representation of the different scenarios,  
323 see Supporting Information Fig. S1.

324 *Scenario 1* considered an Iberian colonization from an ancient unsampled population before  
325 the LGM, and a long-range colonization of the Eastern Mediterranean through an admixture  
326 event from Iberia and the ancient unsampled population. The Iberian population later  
327 colonized Morocco and the later colonized Italy. Admixture events between Iberia and Italy  
328 and between the Eastern Mediterranean and Italy resulted in the French and Greek  
329 populations, respectively.

330 *Scenario 2* considered an Italian colonization from an ancient unsampled population before  
331 the LGM, and a colonization of the Eastern Mediterranean through an admixture event from  
332 Italy and the ancient unsampled population. The Italian population then colonized Morocco  
333 and France, while Iberia and Greece were colonized through admixture events between  
334 France and Morocco and Italy and Eastern Mediterranean, respectively.

335 *Scenario 3* considered a colonization of the Eastern Mediterranean from an ancient  
336 unsampled population before the LGM, and a colonization of the Greek population through  
337 an admixture event between the Eastern Mediterranean population and the ancient  
338 unsampled population. Italy was later colonized from Greece, while Morocco and France  
339 were both be colonized from Italy. Finally, Iberia was colonized through an admixture event  
340 between the Moroccan and French populations.

341 *Scenario 4* considered an Italian colonization from an ancient unsampled population before  
342 the LGM, and a colonization of the Greek population from an admixture event between the  
343 Italian population and the ancient unsampled population. Eastern Mediterranean was then  
344 be colonized from Greece, while Italy colonized both Morocco and France. Finally, Iberia  
345 was colonized through an admixture event between the French and Moroccan populations.

346 *Scenario 5* considered a colonization of the Eastern Mediterranean from an ancient  
347 unsampled population before the LGM, and an Italian colonization through an admixture  
348 event between the Eastern Mediterranean and the ancient unsampled populations. Greece  
349 was also colonized through an admixture event, this time between the Italian and the  
350 Eastern Mediterranean populations. The Italian population then colonized France, while the  
351 Eastern Mediterranean population colonized Morocco. Finally, Iberia was colonized through  
352 an admixture event between the Moroccan and French populations.

353

### 354 **3. Results**

355 3.1. Mitochondrial data

356 We were able to amplify DNA from 136 samples. Samples from Kyrgyzstan were sequenced  
357 using *COI*-mini marker and showed a high mitochondrial divergence from *T. teniotis* (ca.  
358 13%) and aligned with sequences belonging to *T. latouchei* from Laos (99% similarity, Mata  
359 et al., 2017). Additionally, from the four samples from China identified as *T. teniotis* in  
360 museum collections (Appendix 1), we were able to sequence two, both aligning with  
361 *Chaerephon plicatus* (vouchers: MVZ:Mamm:192571 and MVZ:Mamm:193379). According  
362 to the available information, the four samples were collected in the same event at a bat cave  
363 in Southern China, and thus assumed to belong to the same species. According to the  
364 International Union for Conservation of Nature (IUCN) the species has a highly fragmented  
365 distribution in central and eastern Asia (Benda & Piraccini, 2016) and our results suggest  
366 that *T. teniotis* could be absent or rare in this region. Therefore samples from Kyrgyzstan  
367 eastwards were excluded from further analysis.

368 A total of 120 samples belonging to *T. teniotis* were successfully sequenced for *COI* (566 bp  
369 final alignment) and *D-loop* (307 bp final alignment), 114 for *CytB* (509 bp final alignment)  
370 and 109 for *ATP6* (639 bp final alignment). The number of unique haplotypes ranged from  
371 17 for *ATP6* to 33 for *D-loop*. After concatenation the length of the resulting sequences was  
372 between 873 and 2020 bp (average = 1937 bp, Alignment in Supporting Information) and  
373 included 56 unique haplotypes (N = 109, 2020 bp). The Bayesian phylogenetic tree showed  
374 maximum posterior probability support (> 0.9) for the split of two main lineages,  
375 Anatolian/Middle East clade (AMh) and a European clade (EUh) further splitting into two  
376 subgroups but in this case with low support (EUh-A and EUh-B) (Fig. 1).

377 The haplotype network divided the haplotypes into three separate groups, of which one was  
378 exclusive to Iberia and Morocco (EUh-A) and one was distributed elsewhere in central and  
379 western Mediterranean (EUh-B) (Fig. 1 and Supporting Information Fig. S3). The third group  
380 comprised all the haplotypes from Anatolia and Middle East and one additional haplotype  
381 from eastern Crete, broadly supporting the phylogenetic tree. The most common haplotypes  
382 from EUh-A and EUh-B were separated by only one mutational step (percent differences  
383 <0.05 %), while AMh shows a divergence of 0.70% from EUh-A and 0.59 % from EUh-B.

384 Despite the split between the eastern and western clades, the phylogenetic tree and  
385 haplotype network based on mtDNA showed low levels of geographic structuring within each  
386 haplogroup. Mitochondrial haplotype diversity was highest and equal to one in the Middle  
387 East (N = 7), France (N = 4) and Morocco (N = 6), while nucleotide diversity was highest in  
388 Anatolia (Pi = 0.0040, N = 3) and the Middle East (Pi = 0.0036, N = 7) (Table 1). The lowest  
389 values for both haplotype and nucleotide diversity were found in the Canary Islands.

390 Genetic differentiation at mitochondrial DNA was seen between all populations ( $\chi^2 = 532.49$ ,  
391  $P < 0.001$ , overall  $\theta_{ST} = 0.57$ ), with Anatolia and Middle East being genetically differentiated  
392 from all populations except for each other (Supporting Information Table S4). The general  
393 pattern showed a higher mitochondrial diversity in Anatolia/Middle East and equally low  
394 diversity in all the three peninsula.

### 395 *3.2. Microsatellite data*

396 A total of 128 individuals were successfully genotyped. Of the 14 microsatellite loci, two  
397 markers (TAD5 and TAD9) were removed due to violation of Hardy-Weinberg equilibrium  
398 (Supporting Information Table S2). After removing these markers all populations and  
399 markers were overall in Hardy-Weinberg. Our final dataset contained a total of 146 alleles,  
400 with an average number of  $12.17 \pm 2.44$  alleles per locus (range 7-15) and 24 private alleles.

401 Genetic diversity in terms of allelic richness was highest in Anatolia and the Middle East,  
402 followed by Italy and the Iberian Peninsula (Table 1 and Supporting Information Fig. S4).  
403 Expected heterozygosity was high in all populations with the exception of the Canary  
404 population, where the relatedness was particularly high (mean TrioML = 0.40). Overall  
405 population differentiation was low, suggesting a meaningful gene flow. Canaries showed the  
406 highest  $F_{ST}$  values with some degree of differentiation with Greek and Anatolian populations  
407 (Supporting Information Table S5).

408 Model-based clustering method implemented in STRUCTURE without prior population  
409 information did not identify any population structure (Supporting Information Fig. S5).  
410 However, when using this prior, models revealed three main genetic populations (Supporting  
411 Information Fig. S6 and Table S6). Individuals from the Canary Islands formed a separate  
412 population, while all individuals from the Iberian Peninsula, Morocco and France showed a  
413 higher estimated membership fraction to a second inferred cluster, and individuals from Italy  
414 eastwards consistently showed higher estimated membership fraction to a third inferred  
415 cluster (Fig. 2). The three clusters topology was further supported by the Spatial Analysis of  
416 Principal Components (sPCA), although the pattern was not significant (Monte-Carlo test,  
417  $p=0.082$ ) (Fig. 3). Both analyses showed that, except for the Canary population, most  
418 individuals had high levels of admixture, and only a west to east geographic gradient was  
419 evident. An overall observed pattern of isolation by distance was significant (Monte-Carlo  
420 test,  $p = 0.001$ ) (Supporting Information Fig. S7).

### 421 *3.3. ABC inference of evolutionary and demographic history*

422 Model-based inference showed high support (86 %) for a pre-LGM colonization of Italy from  
423 an unsampled population (Supporting Information Fig. S1), while the Anatolian/Middle East

424 population was also colonized pre-LGM from an admixture event between Italy and the  
425 unsampled population, with a similar contribution from both (proportion of admixture from  
426 unsampled population 0.46). The remaining European populations were colonized from Italy  
427 after the Younger Dryas, either directly or via a stepping stone manner with admixture (Fig.  
428 4). However, the Greek population showed some level of admixture between Italy and  
429 Anatolia/Middle East (Fig. 4 and Supporting Information Table S3). Overall, our models  
430 identified two glacial refugia, in Italy and the Anatolia/Middle East with high confidence and  
431 low error rates (type I = 0.04; type II = 0.05).

432 Within the western edge of the range, ABC inference indicated a colonization and population  
433 expansion in Iberia with a generation time similar to that of the colonization analysis  
434 (Supporting Information Table S4). This scenario received high support (99 %) (Supporting  
435 Information Fig. S2) and error rates were estimated at 0.19 and 0.17 for type I and II errors  
436 respectively.

437

#### 438 **4. Discussion**

439 We reconstructed the evolutionary history of a European bat species with tropical affinities.  
440 We show that *T. teniotis* populations were able to survive in Italy and Anatolia/Middle East  
441 during the LGM, and have subsequently colonized the current species range. The species  
442 has experienced a strong population expansion during the post-glacial colonization of its  
443 western range. Our results also point to the occurrence of another population in the  
444 Anatolian/Middle East area. Yet, the high haplotype diversity and network pattern found  
445 suggests that our samples did not cover the eastern refugium, which is likely located further  
446 east (Rossiter et al., 2007) or perhaps towards the Caucasus as suggested for the bat  
447 *Myotis bechsteinii* (Kerth et al., 2008).

##### 448 *4.1. Postglacial colonization and demographic expansion*

449 Our inferences of demographic history indicate two main refugia during the LGM, one in the  
450 Italian Peninsula and another further east in the Anatolian/Middle East region. During this  
451 period, the species may have been extinct throughout the rest of southern Europe, with  
452 subsequent recolonization from the Italian Peninsula. Although the origin of the ancestral  
453 population is unclear, ABC indicates some degree of gene flow between Europe and  
454 Anatolia/Middle East before the LGM. Central and western Mediterranean areas were  
455 subsequently colonized in a stepping-stone manner, and through gene flow between  
456 populations originating from North Africa and France leading to an admixed population in the  
457 Iberian Peninsula. Although samples obtained provided a good coverage of the species

458 range in the western Palaearctic, only a limited number of samples were available from  
459 North Africa. This is a common caveat of phylogeographic studies (Husemann, Schmitt,  
460 Zachos, Ulrich, & Habel, 2014) and we stress that our models do not negate the possibility of  
461 north African or Asian glacial refugium. While such a refugium could be the origin of the  
462 unknown ancestral population inferred in this study, our evolutionary history models show  
463 that a species with tropical affinities was able to survive in Italy during the LGM, from where  
464 it expanded across its current European range.

465 The inferred scenario of an Italian refugium and post-glacial European recolonization  
466 concurs with the widely accepted phylogeographic paradigms for the western Palearctic (G.  
467 M. Hewitt, 1999). Among bats, Italy has been identified as a glacial refugium for *Myotis*  
468 *myotis* (Ruedi et al., 2008) and a possible refugium for *Rhinolophus ferrumequinum*  
469 (Rossiter et al., 2007). In a recent paper, Bogdanowicz et al. (2015) suggested that this  
470 pattern might be widespread among bat species. Focusing on *Miniopterus schreibersii*,  
471 Bilgin et al. (2016) suggested a new paradigm of European colonization from Anatolian  
472 populations, and although we identified an ancient population in Anatolia/Middle East, our  
473 results do not support the hypothesis of a European recolonization from this region, a similar  
474 pattern to *R. ferrumequinum* (Rossiter et al., 2007). In fact, samples from Anatolia and the  
475 Middle East formed a distinct clade at the mitochondrial level (AMh), with no haplotypes  
476 shared with Europe. Interestingly, the high haplotype diversity (nine haplotypes in 10  
477 samples) and the absence of a star-like pattern in the haplotype network for this region,  
478 suggests that the eastern refugium could be located further east.

479 High levels of relatedness and reduced genetic diversity in the Canary Islands likely reflect  
480 inbreeding in an isolated population. Increased inbreeding relative to mainland populations  
481 has been described for different taxa in insular populations (Frankham, 2008), including  
482 bats. Our results suggest that Canary Islands were colonized following a model of long-  
483 distance dispersal and establishment with limited subsequent gene flow from the parent  
484 population (Crisp, Trewick, & Cook, 2011). A general pattern of continental dispersion to the  
485 Canary Islands driven by stochastic events such as storms was described by Juan,  
486 Emerson, Oromí, & Hewitt (2000).

487 The star-like topology in the European mitochondrial groups (EUh-A and EUh-B) indicates  
488 population expansion (Slatkin & Hudson, 1991). This hypothesis was further supported by  
489 the ABC inference, which shows a demographic expansion following the Iberian  
490 colonization. Such expansion could be the result of a natural process (e.g., Bilgin et al.,  
491 2016; Razgour et al., 2013) or might be mediated by human activity, such as through  
492 increased roost availability from tall buildings and other structures including bridges, many of

493 which were built during the 20th century (Amorim, Alves, & Rebelo, 2013; Russo &  
494 Ancillotto, 2014).

495 Post-glacial population growth appear to be common in taxa with that underwent the same  
496 climatic changes since the LGM (Branco, Monnerot, Ferrand, & Templeton, 2002; Korsten et  
497 al., 2009), and was also suggested for another fast flying bat species, *Nyctalus noctula* (Petit  
498 et al., 1999). Microsatellites have a fast mutation rate when compared to other molecular  
499 markers, but it has been questioned whether this rate is fast enough to detect recent  
500 population changes (Barrett & Schluter, 2008). Therefore, it is difficult to ascertain if these  
501 populations, especially the ones located in the western edge of the species' range are still  
502 expanding.

#### 503 4.2. Barriers to gene flow

504 Our results show high differentiation at mitochondrial markers between the populations from  
505 the Anatolia and Middle East region and those from central and western Mediterranean. We  
506 also found evidence of genetic differentiation within the European clade, whereby  
507 populations from Canary Islands, Morocco and Iberia seemed to form a distinct group from  
508 Central Mediterranean populations (Italy, France and Greece). Genetic structuring at the  
509 mitochondrial level suggests that, once established, females will not disperse freely,  
510 supporting some degree of philopatry, a common trait among several bat species (reviewed  
511 in Burland & Worthington Wilmer, 2001). In fact, the Iberian Peninsula seems to have been  
512 colonized following a first-come, first-served pattern, as indicated by the presence of  
513 haplotypes from both the central Mediterranean and North African haplogroups. Even though  
514 *T. teniotis* females are physically capable of crossing geographical barriers (e.g. mountain  
515 ranges and large bodies of water), philopatric behaviour may have a strong effect on female  
516 dispersal, thus explaining the absence of Iberian/north African haplotypes in central  
517 Mediterranean. Contrary to mtDNA, at the nuclear level we confirmed some degree of gene  
518 flow between Europe and the Anatolia/Middle East. We also found high levels of gene flow  
519 within the European range and North Africa, whereas the Gibraltar strait does not act as a  
520 barrier to current or even past gene flow (García-Mudarra et al., 2009). Yet, the Canaries  
521 show high levels of isolation from mainland Africa. Combined, these results reflect a typical  
522 pattern of male-mediated gene flow (Castella, Ruedi, & Excoffier, 2001).

523 Gene flow inferred from nuclear markers seemed to be solely restricted by geographic  
524 distance, showing a clear pattern of isolation by distance and the absence of strong  
525 geographic barriers to dispersal. *T. teniotis* performs fast and direct flights while foraging  
526 with median speeds of 50 km.h<sup>-1</sup> and covering linear distances of up to 70 km (Marques,  
527 Rainho, Carapuço, Oliveira, & Palmeirim, 2004). Although flight altitudes have not been



528 reported for *T. teniotis*, the species is known to prey on large moths that migrate at high  
529 altitudes (Mata et al., 2016). Indeed the smaller congeneric species, *T. brasiliensis* (approx.  
530 12 g compared to 30 g of *T. teniotis*), can fly up to 1 km above ground level (McCracken et  
531 al., 2008). Thus, the absence of geographic barriers to gene flow in our focal taxa is not  
532 surprising.

#### 533 4.3. Implications for the phylogeography of Western Palearctic species with tropical affinity

534 The importance of refugia for conservation planning has been widely recognized because  
535 they can facilitate the persistence of biodiversity under changing climates (Keppel et al.,  
536 2012), and their relevance is even greater in the face of anthropogenic climate change.  
537 Common refugia in the Western Palearctic have been widely acknowledge for a number of  
538 species (G. M. Hewitt, 1999; Husemann et al., 2014), however of the 914 studies focusing on  
539 taxa that occur in the western Palearctic (Keppel et al., 2012) only very few focus on species  
540 with tropical affinities (but see Rato et al., 2011). The location of refugia are often similar  
541 between species sharing climatic and environmental requirements, though it has been  
542 shown that species may respond differently to changes in habitat availability resulting from  
543 climatic changes at the end of the LGM (Taberlet, Fumagalli, Wust-Saucy, & Cosson, 1998).  
544 In a recent paper, Carstens, Morales, Field, & Pelletier (2018) showed that species' traits in  
545 bats can influence the response to climatic oscillations. Most importantly, they found that  
546 heavier bat species and those with longer wings were more likely to suffer a bottleneck at  
547 the LGM, and although this was mostly driven by frugivorous species from the neotropics, it  
548 highlights the importance of phylogeographic studies on species showing different traits in  
549 similar environments.

550 In this study we show that a species with tropical affinities was able to survive in the harsh  
551 environments of glacial Europe when a large area of the Western Palearctic was covered in  
552 ice sheets and permafrost, and temperatures were 10-20 °C cooler than today (Kageyama et  
553 al., 2006). Yet, these results raise new questions regarding how these species survived in  
554 colder climates where the environment carrying capacity was lower (Frenzel et al., 1992).  
555 Moreover, free-tailed bats, such as *T. teniotis*, are thought to be poor hibernators. Although  
556 Arlettaz et al. (2000) found that in the Swiss Alps *T. teniotis* can go through torpor bouts that  
557 can last up to 8 days, average body temperature during hibernation and mean arousal  
558 frequency was much higher than in other temperate bat species.

559 This study contributes to understanding the evolutionary history of species with tropical  
560 affinities living in temperate regions, and raises questions regarding the physiological,  
561 behavioural and ecological traits that enabled them to survive in colder climates. The lack of  
562 phylogeographic studies focusing on these species highlights the importance of such studies

563 for informing their population management and conservation, in particular under future  
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581

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843

844 **Figure 1** – Map showing the study area, with colour representing geographic origins of  
845 samples. Grey lined filled area represents IUCN species range within the study area.  
846 Bayesian phylogenetic tree and Median-joining haplotype network for *T. teniotis* based on  
847 2020 bp of mtDNA (concatenated genes *COI*, *ATP6*, *CytB*, and *D-Loop*). Bayesian posterior  
848 probabilities (BPP) equal to 1 (\*) and greater than 0.9 (†) are marked above branches.  
849 Proportional geographic origin of shared haplotypes is indicated in colour at the branch tips  
850 along with total number of samples. Major supported clades (EUh and AMh) and subgroups  
851 (EUh-A and EUh-B) are indicated (EUh and AMh). Median-joining haplotype networks for  
852 each supported clade as well as the European subgroups (EUh-A and EUh-A) are shown  
853 below where branch lengths are not proportional to base-pair changes. Sampling locations  
854 and haplotype frequency scale are shown in inset. The Bayesian phylogeny used unique  
855 haplotypes only (n = 56) and is shown with out-group (*T. latouchei*). For the median-joining  
856 network all concatenated mtDNA sequences (n = 109) were used.

857

858 **Figure 2** – *Tadarida teniotis* population structure based on the microsatellite dataset. Cluster  
859 membership plots from STRUCTURE analysis using prior population information (LOCPRIOR)  
860 including all samples. Results from 3 to 5 cluster are presented (K = 3 gets the highest rank  
861 according to the Evanno method, Supporting Information Fig. S6 and Table S6).

862

863 **Figure 3** – Spatial Analysis of Principal Components (sPCA) showing the spatial genetic  
864 pattern of *Tadarida teniotis* population based on the microsatellite dataset. The Canaries  
865 form a separate cluster in the left down part, and with less support Greece, Anatolia and the  
866 Middle East also cluster together (top left). The two PCs explain 55.4% of the spatial genetic  
867 pattern. See also the sPCA Eigenvalues histogram in the inset. Dots indicate individual  
868 genotypes.

869

870 **Figure 4** – Colonization patterns across the range of *T. teniotis* according to the best  
871 supported scenario (86 %) based on Approximate Bayesian Computation model inference  
872 (presented in the inset). The geographical location of *T. teniotis* genetic samples included in  
873 the study are plotted over an elevation map, with the location of the six populations marked  
874 and colour coded following the inset. Arrows indicate patterns of pre and post-Last Glacial  
875 Maximum range colonisation. Map coordinate system: Aitoff (sphere-based).

876

877 **List of Supporting Information**

878 Table S1  
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891

892 **Table 1** – Genetic diversity of *T. teniotis* populations based on microsatellite (first five  
 893 columns) and mtDNA (last two columns) datasets. Sample sizes in brackets. Mean allelic  
 894 richness and mean allele frequency across all loci ( $\pm$  SD).  $H_e$  – Expected Heterozygosity;  
 895  $H_{obs}$  – Observed Heterozygosity.

	Mean Allele frequency	Mean Allelic richness	Number of private alleles	$H_e$	$H_{obs}$	Haplotypic diversity	Nucleotide diversity (Pi)
<b>Canary (5)</b>	0.34 $\pm$ 0.10	2.62 $\pm$ 0.45	0	0.58	0.63	0.40	0.0004
<b>Morocco (6)</b>	0.19 $\pm$ 0.04	3.74 $\pm$ 0.45	1	0.76	0.81	1.00	0.0022
<b>Iberia (60)</b>	0.1 $\pm$ 0.020	3.91 $\pm$ 0.37	14	0.80	0.78	0.92	0.0013
<b>France (7)</b>	0.19 $\pm$ 0.03	3.72 $\pm$ 0.31	1	0.76	0.78	1.00	0.0011
<b>Italy (16)</b>	0.12 $\pm$ 0.03	4.00 $\pm$ 0.40	3	0.80	0.77	0.83	0.0010
<b>Greece (5)</b>	0.22 $\pm$ 0.06	3.55 $\pm$ 0.47	3	0.73	0.73	0.90	0.0011
<b>Anatolia (3)</b>	0.25 $\pm$ 0.08	3.56 $\pm$ 0.57	0	0.71	0.83	0.67	0.0040
<b>Middle East (7)</b>	0.15 $\pm$ 0.03	4.00 $\pm$ 0.44	2	0.79	0.78	1.00	0.0036

896