

# Selective anion transport mediated by strap-extended calixpyrroles

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Dedicated to Professor Atsuhiko Osuka on the occasion of his 65<sup>th</sup> birthday.

**ABSTRACT:** Synthetic anion receptors that facilitate transmembrane chloride transport are of interest as potential therapeutic agents for cancer and cystic fibrosis. Transporters selective for chloride over protons are desired for therapeutic applications to avoid autophagy inhibition and cytotoxicity. Examples of such compounds are rare because the majority of anion transporters can interact with the carboxylate head groups of fatty acids leading to proton leakage. In this paper, we report the synthesis, anion binding and transmembrane anion transport properties of two novel bis-triazole-functionalized calixpyrroles with extended straps, and compare them to previously reported shorter-strap analogues known to exhibit high  $\text{Cl}^- > \text{H}^+$  selectivity. We demonstrate improved chloride transport activities of the strap-extended compounds that likely benefits from increased lipophilicity, and reduced  $\text{Cl}^- > \text{H}^+$  selectivity due to the larger anion binding cavities facilitating interaction with fatty acids. The results are instructive for future design of ideal anion transporters with potent activity and high selectivity against proton leakage.

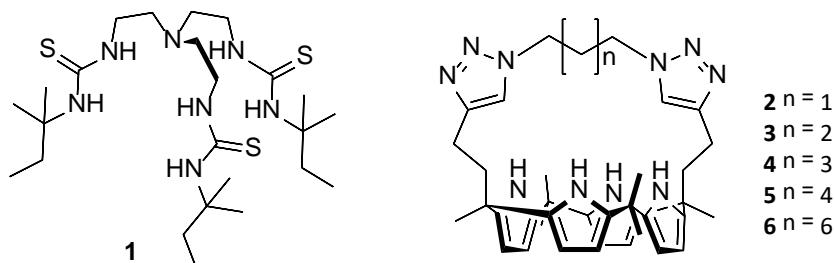
**KEYWORDS:** Anion coordination chemistry; Transmembrane anion transporters; Chloride transport; Calixpyrrole.

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## INTRODUCTION

Synthetic small molecules that bind anions and facilitate transmembrane anion transport have attracted significant research effort recently due to their potential application in the treatment of cancer and ion-channel diseases such as cystic fibrosis.[1] Several classes of synthetic anion transporters including prodigiosin analogues,[2] tambjamines,[3] tripodal tris(thioureas),[4] bisureas[5] and squaramides[6] are known to perform potent  $H^+/Cl^-$  cotransport mimicking the natural product prodigiosin[7] and induce de-acidification of lysosomal pH leading to autophagy inhibition[8] and apoptosis induction.[4,5] Although these compounds are potential anti-cancer agents, other types of compounds that solely facilitate  $Cl^-$  uniport without concomitant transport of  $H^+$  are required for channel-replacement therapy applications where pH gradient disruption and cytotoxicity are undesired side effects.[9] The design of such  $Cl^-$ -selective uniporters has proved to be highly challenging because most anion transporters can facilitate  $H^+$  transport via deprotonation (in the cases of compounds containing highly acidic hydrogen bond donors)[9] or by interacting with the carboxylate headgroup of fatty acids leading to an indirect  $H^+$  transport pathway via fatty acid flip-flop.[10] In our efforts towards making ideally  $Cl^-$ -selective uniporters, we have identified promising compounds including tripodal trithioureas with alkyl substituents (e.g. **1**)[9,11] and strapped calixpyrroles containing additional hydrogen bonding sites in the strap (e.g. compounds **2-4**) (Figure 1).[12] In particular, compound **1**[9] and an amide-stapped calixpyrrole[8a] have been subject to biological studies where the  $Cl^- > H^+$  selectivity determined in vesicles assays has been correlated with weak activity in increasing lysosomal pH and disrupting autophagy. The tripodal compounds are potent  $Cl^-$  uniporters and highly selective against  $H^+$  transport in the absence of fatty acids, but show substantially reduced selectivity when fatty acids are present in significant amounts due to their favorable interaction with carboxylates.[10,11] In contrast, the strapped calixpyrroles (compounds **2-4**) are  $Cl^- > H^+$  selective transporters that maintain the high selectivity even in the presence of fatty acids.[12] This is presumably due to the geometry of the strapped calixpyrroles allowing encapsulation of spherical halide ions by multiple hydrogen bonds from many spatial directions, whereas the Y-shape carboxylates do not allow maximum interaction with the three dimensional hydrogen bond donor arrays of these compounds.[13] The strapped calixpyrroles, however, are >200 times less active than the tripodal trithiourea **1** in  $Cl^-$  uniport.[12] To better understand the selectivity and seek further improvements for the strapped calixpyrroles, we have decided to extend the size of the strapped cavity and synthesized novel anion receptors **5** and **6**. In the current study, we report the anion binding and transport properties of the new strapped calixpyrroles **5** and **6** in comparison to the previous analogues **2-4**.



**Fig. 1.** Structures of tripodal trithiourea **1** and strapped calixpyrroles **2-6**.

## EXPERIMENTAL,

### Synthesis and vesicle preparation

See Supplemental Materials for full details of compound synthesis and vesicle preparation.

## ISE assay for Cl<sup>-</sup> uniport and H<sup>+</sup>/Cl<sup>-</sup> symport

POPC vesicles (mean diameter 200 nm) were prepared containing an internal solution of KCl (300 mM) buffered to pH 7.2 with 10 mM HEPES, suspended in an external solution of potassium D-gluconate (300 mM) buffered to pH 7.2 with 10 mM HEPES. Test solutions were made in 5 mL vials at a lipid concentration of 1 mM. In general, a 5  $\mu$ L DMSO solution of valinomycin (for testing Cl<sup>-</sup> uniport, 1  $\mu$ M or 0.1 mol%) or monensin (for testing H<sup>+</sup>/Cl<sup>-</sup> symport, 1  $\mu$ M or 0.1 mol%) was added at t = 0 s and at t = 30 s, a 10  $\mu$ L DMSO solution of the compound was added to initiate the transport and the anion efflux was monitored using a chloride or fluoride selective electrode. After t = 300 s the vesicles were lysed using Triton X-100 detergent and after t = 420 s a final anion efflux reading was taken as 100 % for calibration purposes.

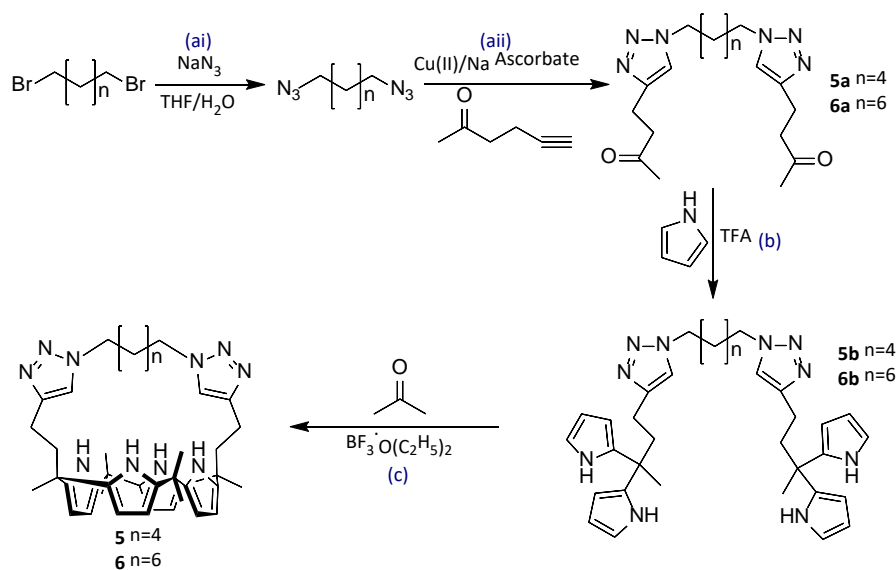
## HPTS assay

POPC vesicles (mean diameter 200 nm) were prepared containing an internal solution of HPTS (1 mM) and NMDG-Cl (100 mM) buffered to pH 7 with 10 mM HEPES, suspended in an external solution of NMDG-Cl (100 mM) buffered to pH 7 with 10 mM HEPES. Test solutions were made to 2.5 mL in cuvettes at a lipid concentration of 0.1 mM. In general, a 5  $\mu$ L DMSO solution of the compound was added followed by a base pulse (NMDG 5 mM) at t = 0 s to initiate the experiment. The fluorescence ratio acidic and basic forms of HPTS ( $\lambda_{\text{ex}}$  = 403 nm and 460 nm respectively) were measured using a fluorimeter. After t = 200 s the vesicles were lysed with Triton X-100 detergent to fully dissipate the pH gradient for calibration purposes. If the experiment involves the addition of gramicidin D (0.1  $\mu$ M or mol%) or oleic acid (2  $\mu$ M or mol%) these are added prior to the addition of the compound.

## RESULTS AND DISCUSSION

### Synthesis

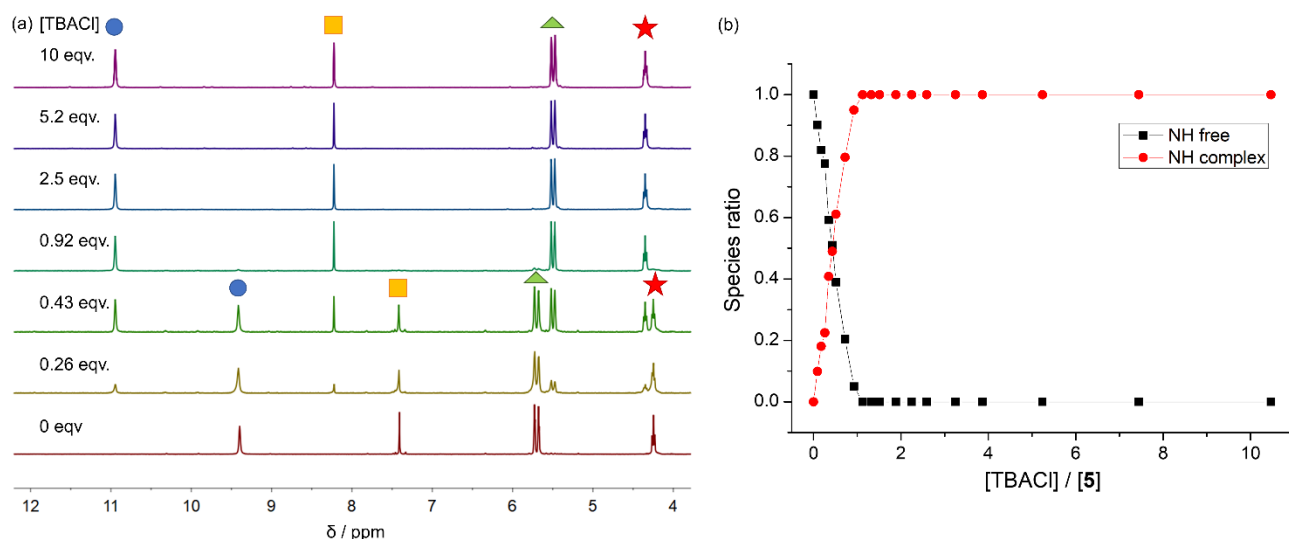
Compounds **5** and **6** were synthesised following the literature procedure outlined by Yano *et al.*[12b] as schematically shown in Fig. 2.



**Fig. 2.** Synthesis of the extended strapped calix[4]pyrroles **5** and **6**.

### Anion Binding Studies

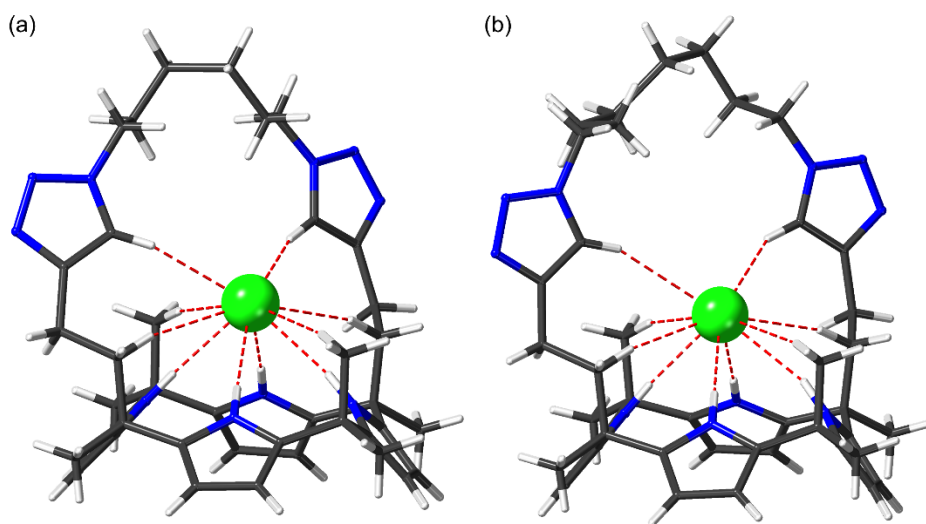
Proton NMR titration studies were conducted in DMSO-*d*<sub>6</sub>:H<sub>2</sub>O (99.5:0.5, v/v) with tetrabutylammonium acetate, fluoride, chloride, bromide, iodide (TBAOAc, TBAF, TBACl, TBABr and TBAI) for **2–6** if not previously reported (Fig. S5-S22, Supplemental Materials). As seen previously[12] slow exchange was observed on the NMR timescale for most compounds **2–6** with TBAOAc, TBAF, TBACl and TBABr. For illustration, the NMR spectra stack plot and binding curve of compound **5** with TBACl are shown in Fig. 3. The pyrrolic NH and triazole CH underwent downfield shifts by 1.5 and 0.8 ppm, respectively, upon binding of Cl<sup>-</sup>, indicating the formation of strong NH···Cl<sup>-</sup> and CH···Cl<sup>-</sup> hydrogen bonds consistent with crystallographic analysis of Cl<sup>-</sup> complexes (vide infra). The pyrrolic CH signals shifted upfield by 0.2 ppm because of increased electron density of the pyrrole ring upon Cl<sup>-</sup> binding. The species ratio determined by integration of pyrrolic NH signals from the free and bound species demonstrate a strong binding profile with ~100% saturation at 1 equivalent of TBACl, which has prevented calculation of the binding constant. Similar strong binding isotherms were observed for titrations of **2–6** with TBAF, TBACl, TBABr and TBAOAc, and in all these cases the binding constants are estimated to be >10<sup>4</sup> M<sup>-1</sup>. In contrast, weak interactions were found for titrations with TBAI and the binding constants were calculated for **5** and **6** using global fitting analysis with bindfit,[14] giving a *K<sub>a</sub>* value of 11 M<sup>-1</sup> for both compounds (Figs S16 and S21). As expected, the larger more charge diffuse iodide anion interacts weakly with the hydrogen-bond donors.



**Fig. 3.** (a) Stack plots for compound **5** <sup>1</sup>H NMR titration with TBACl in DMSO-*d*<sub>6</sub>:H<sub>2</sub>O (99.5:0.5, v/v). Blue circle follows the pyrrole NH signal, orange square follows the triazole CH signal, green triangle follows the pyrrole CHs signal and the red star follows an alkyl CH<sub>2</sub>. (b) Species ratio for the free **5** and **5**-Cl<sup>-</sup> complex, determined by integration of pyrrolic NH signals.

### Solid state analysis

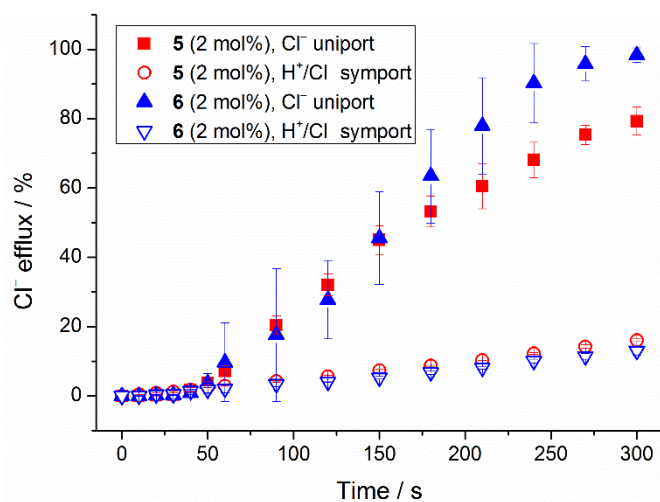
Compounds **5** and **6** were crystallized with TBACl by slow diffusion of petroleum spirits into a DCM solution of **5** with TBACl (5 equiv) and slow evaporation from a DMSO solution of **6** and excess TBACl, respectively. The solid state structures of these complexes (Fig. 4) show the anion bound within the strapped cavity of the calix[4]pyrrole compound stabilized by ten hydrogen bonds that involve four pyrrolic NH, two triazolium CH, and four methylene CH hydrogen bond donors. Compound **5** and **6** shows N···Cl<sup>-</sup> distances that range from 3.31–3.33 Å and 3.27–3.38 Å respectively. Compound **5** displays triazole C···Cl<sup>-</sup> distances of 3.72 and 3.77 Å, while longer C···Cl<sup>-</sup> distances (3.97 and 4.00 Å) were found for the triazoles of **6**. Similarly to previously reported crystal structures of shorter strapped analogues, these structures demonstrate favorable interactions with Cl<sup>-</sup> that features multiple hydrogen bonds from many spatial directions to maximize the contact with the bound Cl<sup>-</sup> ion. Such a binding mode is assumed to confer preference for spherical halide ions over non-spherical anions such as carboxylates.



**Fig. 4.** Single crystal X-ray structures of **5**·TBACl (a) and **6**·TBACl (b) complexes. Minor orientation of the disordered atoms, TBA<sup>+</sup> and DMSO are excluded for clarity. NH···Cl<sup>-</sup> and CH···Cl<sup>-</sup> hydrogen bonds are displayed as red dashed lines. Full crystallographic data can be found in the Supplementary Materials.

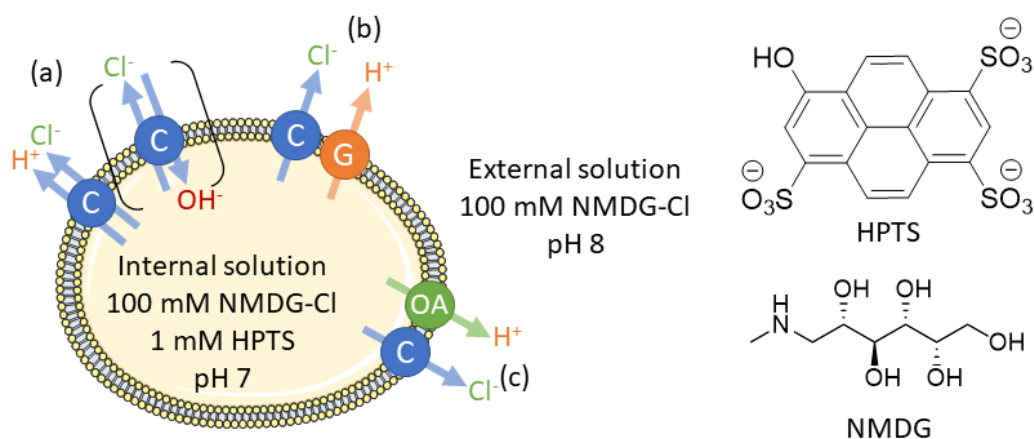
### Transmembrane anion transport studies

The ability of the compounds to facilitate Cl<sup>-</sup> uniport and H<sup>+</sup>/Cl<sup>-</sup> symport was tested in a KCl cotransport assay with the addition of valinomycin (for Cl<sup>-</sup> uniport) and monensin (for H<sup>+</sup>/Cl<sup>-</sup> symport) as previously described,[9,15] with Cl<sup>-</sup> efflux monitored by a Cl<sup>-</sup> ion selective electrode (ISE). KCl (300 mM) was used the internal medium and K-gluconate (300 mM) as the external medium. Here gluconate was used as an untransportable anion to prevent anion exchange. However, it should be noted that gluconate could potentially competitively bind to anion transporters without being transported and slow down anion transport as suggested by Moran *et al.*[16] In the valinomycin-coupled assay, the anion transporter facilitates Cl<sup>-</sup> uniport that coupled to valinomycin-facilitated K<sup>+</sup> uniport to give overall KCl transport (Figure 5). In the monensin-coupled assay, by contrast, the process studied is the H<sup>+</sup>/Cl<sup>-</sup> symport facilitated by anion transporter with the resultant pH gradient dissipated by monensin-facilitated K<sup>+</sup>/H<sup>+</sup> antiport, leading to overall KCl transport. Both **5** and **6** show efficient Cl<sup>-</sup> uniport whereas only a minor amount of H<sup>+</sup>/Cl<sup>-</sup> symport at 2 μM (or 2 mol%, compound to lipid molar ratio), indicating that these compounds are selective for Cl<sup>-</sup> over H<sup>+</sup>/OH<sup>-</sup>. Control experimentals with **5** or **6** alone show no Cl<sup>-</sup> efflux, confirming that these compounds do not facilitate metal ion transport or non-specific membrane leakage (Fig. S25). Because of the poor NO<sub>3</sub><sup>-</sup> transport activity of the strapped calixpyrroles,<sup>12b</sup> we did not employ the Cl<sup>-</sup>/NO<sub>3</sub><sup>-</sup> exchange assay to characterize these transporters.



**Fig. 5.** Cl<sup>-</sup> uniport and H<sup>+</sup>/Cl<sup>-</sup> symport facilitated by compounds **5** and **6** (both tested at 2 μM or 2 mol% compound to lipid molar ratio), determined using an ISE assay using POPC vesicles in the presence of either valinomycin (for Cl<sup>-</sup> uniport) or monensin (for H<sup>+</sup>/Cl<sup>-</sup> symport). Error bars represent standard deviations from two or three repeats. Note the errors for compound are high presumably because of the tendency of this compound to precipitate from the aqueous solution when added to the lipid suspensions as DMSO solutions.

To quantify the Cl<sup>-</sup> uniport activity and Cl<sup>-</sup> over H<sup>+</sup>/OH<sup>-</sup> selectivity of the compounds **5** and **6**, we performed a different assay, i.e. a modified HPTS assay[17] with *N*-methyl-D-glucamine chloride (NMDG-Cl) as the medium, which involved the use of gramicidin D (Gra or G) a naturally occurring proton channel, and oleic acid (OA)[9,15] as schematically illustrated in Fig. 6. The large hydrophilic cation NMDG<sup>+</sup> was used to prevent metal ion transport. The assay measures the rate of pH gradient dissipation induced by anion transporters or a combination of anion transporters and Gra facilitating H<sup>+</sup>/Cl<sup>-</sup> symport or OH<sup>-</sup>/Cl<sup>-</sup> antiport. Note that the thermodynamic driving forces and the amount of ion fluxes are different in the HPTS assay compared with the ISE assay. The transport process in the HPTS assay was driven by a pH gradient and required a H<sup>+</sup> and Cl<sup>-</sup> efflux (or an OH<sup>-</sup> influx and a Cl<sup>-</sup> efflux) of 5 mM to reach equilibrium, whereas in the ISE assay, the transport was driven by a Cl<sup>-</sup> concentration gradient and required a K<sup>+</sup> and Cl<sup>-</sup> efflux of 300 mM to reach equilibrium. In the absence of gramicidin D, the assay determines the activity of the anion transporter in facilitating H<sup>+</sup>/Cl<sup>-</sup> or OH<sup>-</sup>/Cl<sup>-</sup> antiport, where the transport of Cl<sup>-</sup> or H<sup>+</sup>/OH<sup>-</sup> may be rate-limiting (Fig. 6a). Neutral hydrogen bond-based anion transporters are known to facilitate H<sup>+</sup> transport either by deprotonation of the hydrogen bond donors[9] or by a fatty acid shuttling mechanism.[10] For the latter mechanism, the process is fatty acid-dependent (Fig. 6c) and therefore the assay was also conducted with oleic acid added to the vesicles. In the presence of Gra that facilitates rapid H<sup>+</sup> uniport, the anion transporter only needs to facilitate Cl<sup>-</sup> uniport for dissipation of pH gradient (Fig. 6b). Therefore if the anion transporter is Cl<sup>-</sup> > H<sup>+</sup>/OH<sup>-</sup> selective, gramicidin would enhance the rate of pH gradient dissipation by replacing the rate-limiting step of H<sup>+</sup>/OH<sup>-</sup> transport, thus allowing the intrinsic Cl<sup>-</sup> uniport activity to be determined.



**Fig. 6.** Overview of NMDG-Cl HPTS assay. Conditions- Internal: NMDG-Cl 100 mM and HPTS 1 mM buffered to pH 7 with HEPES buffer 10 mM. External: NMDG-Cl 100 mM buffered to pH 7 with HEPES buffer 10 mM, pulse: NMDG to bring external pH to 8. The transport was initiated by addition of a DMSO solution of the transporter. (a) The transporter was added alone and the electroneutral ( $H^+/Cl^-$  symport or  $Cl^-/OH^-$  antiport) transport was monitored by following the pH gradient dissipation. (b) Gramicidin (G) was added which facilitates  $H^+$  transport, this will allow electrogenic  $Cl^-$  transporters to function within the assay, the transport was monitored by following the pH gradient dissipation. (c) Oleic acid (OA) was added to evaluate the compounds ability to uphold selectivity in the presence of fatty acids.

Quantification of  $Cl^-$  uniport activity and  $Cl^- > H^+/OH^-$  selectivity and was undertaken by dose-dependent Hill analyses which afforded an effective concentration to reach 50% of maximum transport at 200 s ( $EC_{50}$ ) expressed as the transporter concentration or the transporter to lipid molar ratio, where a low  $EC_{50}$  value indicates a high transport activity. To simplify the analysis, we examined the dependence of fluorescence intensity ratio between the basic and acidic forms of HPTS ( $I_{base}/I_{acid}$ ) on the amount of base added in the pH range of 7-8 after the vesicles were lysed with a detergent. The results (Fig. S26) demonstrate that  $I_{base}/I_{acid}$  is approximately linear to the amount of base added, thus justifying the use of  $I_{base}/I_{acid}$  to indicate the progress of  $H^+$  efflux. The  $EC_{50}$  Gra value provides a measure of  $Cl^-$  uniport activity as Gra removes the need for the anion transporter to facilitate the potentially rate-limiting  $H^+$  transport. The ratio between  $EC_{50}$  values in the absence and presence of Gra gives a selectivity factor ( $S_G$ ) that quantifies  $Cl^-$  over  $H^+/OH^-$  selectivity. The selectivity factor was additionally determined in the presence of oleic acid ( $S_{OAG}$ ) to evaluate the  $Cl^-$  over  $H^+/OH^-$  selectivity dependent on the fatty acid shuttling pathway. The  $S_{OAG}$  value is more biologically relevant than  $S_G$  because biological membranes contain significant amounts of free fatty acids. The  $EC_{50}$  and selectivity factors for **5** and **6** are summarized in Table 1, along with previously reported results for **2-4**. We also performed an initial rate analysis at a fixed transporter concentration and the results are summarized in Table S1.

In the absence of fatty acids, compounds **5** and **6** show acceleration of pH gradient dissipation by Gra indicating high selectivity for  $Cl^-$  over  $H^+/OH^-$  with  $S_G$  values of 18, and 9.5, respectively. Remarkably, the  $EC_{50}$  Gra values of **5** and **6** are substantially lower than previously reported shorter strapped compounds **2-4**, demonstrating significantly improved  $Cl^-$  uniport activities attributable to increased lipophilicity of **5** and **6** due to the extended alkyl linker length. However, **5** and **6** show modestly accelerated  $H^+$  transport by oleic acid, leading to reduced  $Cl^-$  over  $H^+/OH^-$  selectivity in the presence of oleic acid ( $S_{OAG}$ ). Previously the  $Cl^-$  over  $H^+/OH^-$  selectivity of the shortest strapped compounds **2** and **3** were found to be unaffected by fatty acids (i.e.  $S_{OAG} \approx S_G$ ), while that of **4** was slightly diminished by fatty acids ( $S_{OAG} < S_G$ ).<sup>[12a]</sup> In the current work, fatty acid induced a more pronounced attenuation of  $Cl^-$  over  $H^+/OH^-$  selectivity for the longest strapped compounds **5** and **6**, thus supporting the hypothesis that expanding the size of the anion binding cavity to some extent favors the binding and transport of carboxylate ions over the smaller  $Cl^-$  ions. Despite the compromised selectivity for the longer straps, the strapped calixpyrroles still represents the class of compounds with the highest  $Cl^-$

over H<sup>+</sup>/OH<sup>-</sup> selectivity to date that can resist H<sup>+</sup> transport via the fatty acid shuttling mechanism due to three-dimensional encapsulation of chloride (Fig. 4). Note that the Hill coefficients (*n* values) are higher than 1 in many cases which can be attributed to ion transport by an aggregated (transporter)<sub>*n*</sub> ion species.

Table 1 Summary of Cl<sup>-</sup> transport activity and selectivity over H<sup>+</sup>/OH<sup>-</sup> for **2-6**.

Compound	Compound alone		with Gra <sup>a</sup>		S <sub>G</sub> <sup>b</sup>	with OA <sup>c</sup>		S <sub>OAG</sub> <sup>d</sup>
	EC <sub>50</sub> / μM (mol%)	<i>n</i> <sup>e</sup>	EC <sub>50</sub> / μM (mol%)	<i>n</i> <sup>e</sup>		EC <sub>50</sub> OA / μM (mol%)	<i>n</i>	
<b>2</b> <sup>f</sup>	11 ± 0	2.9 ± 0.2	0.60 ± 0.03	1.7 ± 0.2	18	11 ± 0	2.3 ± 0.2	18
<b>3</b> <sup>f</sup>	4.1 ± 0.1	3.8 ± 0.5	0.38 ± 0.03	1.3 ± 0.1	11	3.6 ± 0.0	4.1 ± 0.2	9.5
<b>4</b> <sup>f</sup>	2.7 ± 0.0	4.5 ± 0.3	0.18 ± 0.01	1.0 ± 0.0	15	1.6 ± 0.1	1.8 ± 0.2	8.9
<b>5</b>	1.5 ± 0.1	2.0 ± 0.3	0.092 ± 0.004	1.4 ± 0.1	16	0.43 ± 0.03	1.6 ± 0.1	4.7
<b>6</b>	1.1 ± 0.0	2.3 ± 0.2	0.15 ± 0.01	1.2 ± 0.1	7.0	0.49 ± 0.03	1.7 ± 0.2	3.3

Notes: <sup>a</sup> Hill analysis in the presence of gramicidin D; this value shows the total H<sup>+</sup>/Cl<sup>-</sup> symport (Cl<sup>-</sup>/OH<sup>-</sup> antiport) activity possible, with no rate-limiting H<sup>+</sup>/OH<sup>-</sup> transport. The Gra concentration has been optimised at 0.1 μM to prevent this having a limiting effect. <sup>b</sup> Cl<sup>-</sup> over H<sup>+</sup>/OH<sup>-</sup> selectivity factor S<sub>G</sub> is calculated by dividing EC<sub>50</sub> in the absence of Gra by the EC<sub>50</sub> Gra. S<sub>G</sub> > 1 indicates Cl<sup>-</sup> selectivity. <sup>c</sup> Hill analysis in the presence of oleic acid (2 μM); this value shows the effect of fatty acids on the selectivity. <sup>d</sup> Cl<sup>-</sup> over H<sup>+</sup>/OH<sup>-</sup> selectivity value in the presence of oleic acid, calculated by dividing the EC<sub>50</sub> OA by the EC<sub>50</sub> Gra. S<sub>OAG</sub> > 1 indicates Cl<sup>-</sup> selectivity. <sup>e</sup> Hill coefficients determined by the Hill analysis. The Hill coefficient indicates the stoichiometry of the receptor-anion complex formed to facilitate anion transport. <sup>f</sup> Values for previous reported compounds are taken from [12a]. <sup>g</sup> Errors in this table are statistical errors of fitting.

## CONCLUSION

Based on our previous work on selective halide transport facilitated by strapped calixpyrroles, we have synthesized strap-extended calixpyrroles **5** and **6** featuring three-dimensional encapsulation of chloride ions via multiple NH and CH hydrogen bond interactions as confirmed by solid-state analysis. Extension of the alkyl linker led to improved Cl<sup>-</sup> uniport activity but compromised Cl<sup>-</sup> over H<sup>+</sup>/OH<sup>-</sup> selectivity due to the larger anion binding cavity facilitating binding and transport of deprotonated fatty acids. The structure-selectivity relationship demonstrated in this work is informative for future development of highly active and selective Cl<sup>-</sup> uniporters for channel replacement therapies where H<sup>+</sup> transport leading to pH disruption and autophagy inhibition should be avoided.

General experimental details, synthetic procedures, NMR spectra and titration, data, further details of the crystallography and anion transport assays (FIGS S1-S32 and Table S1) are given in the supplementary material. This material is available free of charge *via* the Internet at <http://www.worldscinet.com/jpp/jpp.shtml>.

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## REFERENCES

- (1) (a) Li H, Valkenier H, Thorne AG, Dias CM, Cooper JA, Kieffer M, Busschaert N, Gale PA, Sheppard DN and Davis AP. *Chem. Sci.* 2019; DOI:10.1039/c9sc04242c; (b) Gale PA, Davis JT and Quesada R. *Chem. Soc. Rev.* 2017; **46**: 2497-2519; (c) Haynes CJE and Gale PA. *Chem. Commun.* 2011: 8203-8209.
- (2) (a) Gale PA, Light ME, McNally B, Navakhun K, Sliwinski KE and Smith BD. *Chem. Commun.* 2005: 3773-3775; Jowett LA, Howe ENW, Soto-Cerrato V, Van Rossom W, Pérez-Tomás R and Gale PA. *Sci. Rep.* 2017; **7**: 9397.
- (3) Hernandez PI, Moreno D, Javier AA, Torroba T, Pérez-Tomás R and Quesada R. *Chem. Commun.* 2012; **48**: 1556-1558.
- (4) Busschaert N, Wenzel M, Light ME, Iglesias-Hernández P, Pérez-Tomás R and Gale PA. *J. Am. Chem. Soc.* 2011; **133**: 14136-14148.
- (5) Moore SJ, Haynes CJE, González J, Sutton JL, Brooks SJ, Light ME, Herniman J, Langley GJ, Soto-Cerrato V, Pérez-Tomás R, Marques I, Costa PJ, Felix V and Gale, PA. *Chem. Sci.* 2013; **4**: 103-117.
- (6) Busschaert N, Kirby IL, Young S, Coles SJ, Horton PN, Light ME and Gale PA. *Angew. Chem., Int. Ed.* 2012; **51**: 4426-4430.
- (7) Sato T, Konno H, Tanaka Y, Kataoka T, Nagai K, Wasserman HH and Ohkuma S *J. Biol. Chem.* 1998; **273**: 21455-21462.
- (8) (a) Park S-H, Park S-H, Howe ENW, Hyun JY, Chen LJ, Huang I, Vargas-Zuñiga G, Busschaert N, Gale PA, Sessler JL and Shin I. *Chem* 2019; **5**: 1210-1222; (b) Zhang Z, Wang Y, Xie W, Howe ENW, Busschaert N, Sauvat A, Leduc M, Gomes-da-Silva LC, Chen G, Martins I, Deng X, Maiuri L, Kepp O, Soussi T, Gale PA, Zamzami N and Kroemer G. *Cell Death & Disease* 2019; **10**:242 (c) Busschaert N, Park S-H, Baek K-H, Choi YP, Park J, Howe ENW, Hiscock JR, Karagiannidis LE, Marques I, Félix V, Namkung W, Sessler JL, Gale PA, and Shin I. *Nat. Chem.* 2017; **9**: 667-675; (d) Rodilla AM, Korrodi-Gregório L, Hernando E, Manuel-Manresa P, Quesada R, Pérez-Tomás R and Soto-Cerrato V. *Biochem. Pharmacol.* 2017; **126**: 23-33.
- (9) (a) Wu X, Judd LW, Howe ENW, Withecombe AM, Soto-Cerrato V, Li H, Busschaert N, Valkenier H, Pérez-Tomás R, Sheppard DN, Jiang Y-B, Davis AP and Gale PA. *Chem* 2016; **1**: 127-146; (b) Wu X, Small JR, Cataldo A, Withecombe AM, Turner P and Gale PA. *Angew. Chem. Int. Ed.* 2019; **58**: 15142-15147.
- (10) (a) Wu X and Gale PA. *J. Am. Chem. Soc.* 2016; **138**: 16508-16514; see also (b) Howe ENW and Gale PA. *J. Am. Chem. Soc.* 2019; **141**: 10654-10660.
- (11) Jowett LA, Ricci A, Wu X, Howe ENW and Gale PA. *Molecules* 2019; **24**: 1278.
- (12) (a) Clarke HJ, Howe ENW, Wu X Sommer F, Yano M, Light ME, Kubik S and Gale, PA. *J. Am. Chem. Soc.* 2016; **138**: 16515-16522; (b) Yano M, Tong CC, Light ME, Schmidtchen FP and Gale PA. *Org, Biomol. Chem.* 2010; **8**: 4356-4363.
- (13) Dabrowa K, Ulatowski F, Lichosyt D and Jurczak J. *Org. Biomol. Chem.* 2017; **15**: 5927-5943.
- (14) Thordarson P. in *Supramolecular Chemistry: from Molecules to Nanomaterials*; Gale PA and Steed JW Editors; John Wiley & Sons, Ltd: 2012.
- (15) Wu X, Howe ENW and Gale PA. *Acc. Chem. Res.* 2018; **51**: 1870-1879.
- (16) Fiore M, Cossu C, Capurro V, Picco C, Ludovico A, Mielczarek M, Carreira-Barral I, Caci E, Baroni D, Quesada R and Moran, O. *Brit. J. Pharmacol.* 2019; **176**: 1764-1779

(17) Matile S and Sakai N. in *Analytical Methods in Supramolecular Chemistry*; Schalley, CA, Editor; Wiley-VCH: Weinheim, 2012, pp711-742.