β-1,3/1,6-glucans and Immunity: State of the Art and Future Directions

**Elena De Marco Castro 1, 2, Philip C Calder3, 4 and Helen M Roche\* 1, 2, 5**

1Nutrigenomics Research Group, School of Public Health Physiotherapy and Sports Science, University College Dublin, Dublin 4, Ireland;

2UCD Institute of Food and Health / Diabetes Complications Research Centre, UCD Conway Institute, University College Dublin, Dublin 4, Ireland;

3 Faculty of Medicine, University of Southampton, Southampton SO16 6YD, United Kingdom

4 NIHR Southampton Biomedical Research Centre, University Hospital Southampton NHS Foundation Trust and University of Southampton, Southampton SO16 6YD, United Kingdom;

5 Institute for Global Food Security, Queens University Belfast, Belfast BT9 5DL, Northern Ireland, United Kingdom.

∗Correspondence to: Helen M Roche, Associate Professor of Nutrigenomics. Address: Conway Institute of Biomolecular & Biomedical Research, University College Dublin, Dublin 4, Ireland. Telephone: +353-1-7166845. Fax: +353-1-7166701. E-mail: helen.roche@ucd.ie

**Abbreviations**: APC, antigen presenting cells; AR, allergic rhinitis; CARD9, caspase recruitment domain containing protein 9; CD36, cluster of differentiation 36; DHA, docosahexaenoic acid; EFSA, European Food Safety Authority; PAMPs, pathogen-associated molecular patterns; MAMPs, microbe-associated molecular patterns; PRRs, pattern recognition receptors; SG, soluble glucans; Syk, spleen tyrosine kinase; TLR, Toll-like receptors; URTI, upper respiratory tract infection; WGP, whole glucan particles.

**Keywords:** Yeast β-glucan, metabolic-inflammation, innate immunity, trained immunity, diet and inflammation

**Abstract**

The innate immune system responds in a rapid and non-specific manner against immunologic threats; inflammation is part of this response. This is followed by a slower but targeted and specific response termed the adaptive or acquired immune response. There is emerging evidence that dietary components, including yeast-derived β-glucans, can aid host defense against pathogens by modulating inflammatory and antimicrobial activity of neutrophils and macrophages. Innate Immune Training refers to a newly recognized phenomenon wherein compounds may ‘train’ innate immune cells, such that monocyte and macrophage precursor biology is altered to mount a more effective immunological response. Although various human studies have been carried out, much uncertainty still exists and further studies are required to fully elucidate the relationship between β-glucan supplementation and human immune function. This review offers an up-to-date report on yeast-derived β-glucans as immuno-modulators, including a brief overview of the current paradigm regarding the interaction of β-glucans with the immune system. The recent pre-clinical work that has partly decrypted mode of action and the newest evidence from human trials, are also reviewed. According to pre-clinical studies, β-1,3/1,6-glucan derived from Baker’s Yeast may offer increased immuno-surveillance, although the human evidence is weaker than that gained from pre-clinical studies.

**Introduction to β-glucans and Immunity**

**Structural Diversity with Varied Health Impacts**

β-glucans are naturally occurring polysaccharides of D-glucose monomers linked by β-glycosidic bonds. They serve as energy stores and structural components of plant, algal, fungal and bacterial cell walls [1]. β-glucans show a very defined structure-activity relationship: research to date demonstrates that varied source and structure lead to varied biological activity [2-4]. For example, although oat-derived β-glucans act as effective dietary fibers that improve metabolic health parameters, including dyslipidemia and insulin resistance [4]; yeast-derived β-glucans behave as immuno-modulators, specifically targeting the innate immune response [5]. The chemistry of β-glucans is well characterized; they all share a β-1,3-glucan backbone; however, among β-glucans, there is variation regarding source as well as extraction and purification methods used in their commercial preparation [6-8], which gives rise to great diversity in branching pattern, insertions and impurities. This yields a wide range of molecular structures within the β-glucan family to ultimately determine a range of health effects [1] (Figure 1). Long side chains along the β-1,3 backbone confer insolubility in the case of some yeast and fungal β-glucans [9, 10]. The formation of purely linear β-1,3-glucans in fungi and yeast is highly uncommon; however, these are present in soluble extracts of *Saccharomyces cerevisiae* [11]. Interestingly, only β-glucans with a high molecular weight and degree of branching, like those from fungi and yeast, have been reported to exert an immunomodulatory action [3, 12-15]. In fact, it was the preliminary work of Pillemer and Ecker [16] on Fleischmann’s® Yeast (*S. cerevisiae*) that emerged during the 1940s which first suggested the connection between β-glucans and immunity. Several decades later, evidence has accumulated relating to the immunomodulatory activity of both research-exclusive and commercially available forms of β-glucans derived from various sources, and by various extraction and purification methods [5, 8, 17]; however, the translation of any nutritional immunomodulatory agent from cellular and/or pre-clinical models to use in humans is a great challenge. This specifically applies to β-glucans, since aside from their wide structural variability leading to diverse biological activities, there are differences between β-glucan receptors in humans and mice [18, 19], immune health markers are ambiguous, and variation in administration route, dose, populations studied and time points between studies are likely to contribute to inconsistent results relating to the health impact of β-glucans. As such, the first aim of this review is to collate and interpret the existing preclinical research on β-1,3/1,6-glucan with regard to immunity in order to clarify its molecular mechanism of immunomodulatory action. This will be achieved by considering its binding to immune receptors and the downstream signalling events leading to trained immunity and cytotoxic activity. The second aim of this review is to collate and evaluate the literature in order to provide a comprehensive overview of the human studies assessing the effect of supplementation with high quality, well-characterised β-1,3/1,6-glucan from commercially available sources (presented in Table 1) on immunity across multiple populations. For this, inclusion criteria consist of randomized, double-blind, placebo-controlled human studies that investigated the efficacy of orally administered β-glucan with a purity of over 75% (obtained by stripping β-glucan from the external mannoprotein layer, the interlinked chitin in the cell wall and the components inside the cell membrane) [8, 20, 21]. Exclusion criteria include β-glucan formulations that lack human studies or are used in combination with other active ingredients (e.g. β-glucan and monoclonal antibodies in cancer research). Overall, no adverse events were detected, and no major safety concerns were presented in response to any of the selected intervention studies. Statistical significance was set at P < 0.05 and all results included are by default significant unless otherwise stated.All β-glucans covered herein are by nature β-1,3/1,6-glucans, and the term β-glucan will refer to β-1,3/1,6-glucan unless otherwise specified.

**The Immune Response to Pathogen Invasion**

The immune response is composed of innate and adaptive components. The initial responder to pathogens is the innate component, comprised mainly of phagocytic cells (e.g. neutrophils, monocytes, macrophages, and dendritic cells) able to engulf and kill pathogens. After phagocytosing and digesting pathogens and then processing pathogen-derived antigens, innate immune cells release inflammatory mediators and cytokines, and present the processed antigen to T helper cells and B lymphocytes, leading to the development of the more specific adaptive immune response [22, 23]. The T cells and B cells are responsible for the antigen-specific recognition and destruction of pathogens [24, 25]. Traditionally, the classical adaptive response was attributed to T and B cells able to express different receptors for maximal specificity based on the antigen being presented, while the innate response was understood to rely on receptors only able to recognize highly conserved microbial structures. More recently, it is evident that innate cells have the ability to modify responses by pattern recognition[23], leading to the concept innate immune memory or innate immune training.

The main challenge for the innate immune system is to differentiate self or benign from pathogenic antigens. Evolution’s ingenious solution has been to identify microbe- or pathogen-associated molecular patterns (MAMPs or PAMPs), which are different motifs present in invading microorganisms that are lacking in higher eukaryotes [26, 27]. During the early phase of the immune response, MAMPs are recognized by pattern recognition receptors (PRRs) and complement binding (e.g. dectin-1 and complement receptor 3 for β-glucan signaling), which prompt a rapid detection and control of pathogen invasion via initiation of an innate immune response [5, 28]. PRRs are expressed on dendritic cells, the classical antigen presenting cells (APCs), and play an essential role in recognizing endogenous (host derived) and exogenous (environmental) ligands prior to phagocytosis. APCs are specialized phagocytes like dendritic cells and macrophages distributed across the host. Following pathogen internalization and phagosome formation, a key aspect of the antimicrobial component of the innate immune response is the production of reactive oxygen species (ROS) against pathogens, a phenomenon known as oxidative burst [29]. This response has been reported against β-glucans from fungal and yeast cell walls and results in the phosphorylation of the NADPH oxidase complex and ultimately production of ROS (e.g. superoxide anion) [30] (Figure 2).

**β-glucans as Trainers of Innate Immunity**

Initial understanding suggested that the rapid onset and non-specific innate immune response which lacks memory was followed by an adaptive response that is antigen-specific and has immunological memory. However, it was recognized that some organisms that lack adaptive immune components show immunological memory [31]. The paradox whereby a first encounter between classical innate immune cells and a pathogen triggers intracellular functional changes that expedite future defense towards the same or an unrelated pathogen has been labeled ‘trained immunity or innate immune memory’ [32]. It is now well established that PRRs expressed in innate immune cells show discrete pathogen recognition [33]. Trained immunity has been traced to epigenetic changes that result from reprogramming of chromatin marks within the progenitors of innate immune cells. This offers a mechanism whereby β-glucan is able to prime innate immune cells. This idea was strengthened by the evidence of a more robust innate response from monocytes and macrophages to various pathogens after β-glucan exposure, with enhanced antimicrobial and inflammatory properties derived from dectin-1/toll-like receptor (TLR) activation [34]. However, compared to the classical acquired immunity offered by antigen receptors as part of the adaptive immune response, PRRs do show less specificity and duration of memory.

**β-glucans as Immunomodulators: Mechanism of Action**

β-glucans are highly conserved structural components that make up the majority of the cell walls of yeast and fungi. Since β-glucans are not produced by mammalian cells, they act as biological response modifiers and are recognized as MAMPs by PRRs present on the surface of innate immune cells [11, 35, 36]. β-glucans are phagocytosed and processed by monocytes, macrophages and dendritic cells found in the upper intestinal lymphatic tissue to later be shuttled towards different immune organs like the spleen, where fragmented soluble β-1,3-glucan particles are released and prime immune cells for a more efficient antimicrobial and inflammatory response to pathogenic challenges.

**β-glucan Internalization**

Following oral β-glucan administration, PRRs expressed on the cell surface of macrophages from the microfolds of Peyer’s Patches (M cells) bind and internalize β-glucan upon arrival in the small intestinal lumen, as seen by fluorescence microscopy in murine spleen and bone marrow tissues [22, 37, 38]. Once internalized, β-glucan-containing innate immune cells travel to the different organs of the immune system where smaller and more soluble β-1,3-glucan fragments are released over several days to interact with and modulate the functional capacity of the innate immune response [3, 37, 39]. Whether β-glucan-containing macrophages are transported to the lymphoid organs via lymphatic or systemic circulation in humans is still unclear [3, 40], although rat and mouse models show intact β-glucan being shuttled from the gastrointestinal tract to the different immune organs through the systematic circulation [38, 41]. Subsequent fragmentation into smaller biologically active β-1,3-glucan particles occurs on day 3 to 5 after ingestion, β-1,3-glucan fragments are released extracellularly from days 5 to 10 and diminish after 14 to 21 days [37]. β-glucan receptors include dectin-1 [23, 26, 42-46], CR3 [26, 47-49], lactosylceramide [23, 50], TLR 2, 4 and 6 [28, 51-53], cluster of differentiation 36 (CD36) [54] and scavenger receptors like CD5 [55]. Dectin-1 and CR3 have been extensively characterized for their direct interaction with β-glucans and their ability to subsequently alter the immune response, whereas the other receptors mainly become involved later on during the intracellular signaling cascade [17]. While CR3 is highly expressed on neutrophils, monocytes, natural killer (NK) cells, and to a lesser extent on macrophages, dectin-1 is mainly expressed on macrophages, followed by granulocytes, and is absent from NK cells. Consequently, the main receptor mediating yeast phagocytosis in macrophages is dectin-1, but CR3 is the predominant receptor in granulocytes [40].

**Dectin-1 Structure and Signaling**

Murine dectin-1 and its human equivalent (also known as β-glucan receptor) are type II C-lectin-like membrane receptors from the PRR family. Although some differences in expression pattern and regulation exist between dectin-1 and its human homologue (e.g. N-linked glycosylation is present in the C-type lectin domain of murine dectin-1 receptors and only in the full-length stalks of β-glucan receptor), they both seem to serve the same function as reviewed by Willment *et al*. [56]. They share 60% sequence identity, and 71% sequence homology [57], and tend to preferentially express the stalkless isoform of the receptor [19].

Dectin-1 is expressed on the surface of non-specific myeloid immune cells (monocytes, macrophages, dendritic cells, and neutrophils), for example in M cells (in gut associated lymphoid tissue). Extensive work done by Goodridge *et al.* [17] on RAW264.7 macrophages, primary mouse bone marrow-derived macrophages and dendritic cells as well as on human monocytes, shows that β-glucans successfully bind to the C-type lectin-like carbohydrate recognition domain of dectin-1; however, insoluble whole glucan particles (WGPs) but not soluble glucans (SGs) are able to successfully elicit dectin-1 signaling [17, 19, 42]. Even when both WGP and SG are derived from the same (micro)organism (most studied is *S. cerevisiae*) they differ based on their structure, molecular weight, spatial conformation, solubility, etc. WGPs are insoluble β-glucan particles that maintain the three-dimensional yeast cellular structure with a hollow cytoplasm (i.e. only the cell wall remains) and are recognized by dectin-1 as an immobilized form of β-glucan, i.e. on the surface of a phagocytosed particle such as yeast cells. SGs like PGG-glucan, a β-1,3/1,6-glucan triple helix molecule of pharmaceutical grade, or smaller fragments of soluble β-glucan released by macrophages after WGP internalization are recognized as ‘free floating or unattached’ particles. PGG-glucan is commonly used to test this mechanism in human peripheral blood mononuclear cells (PBMCs) and neutrophils [58]. One explanation for dectin-1 only activating upon interaction with WGPs and not SGs is the ‘phagocytic-synapse’ mechanistic model, whereby dectin-1 exclusively initiates its signaling cascade to immobilized ligands as opposed to soluble ‘floating’ substances shed from their parent particle [17]. ‘Phagocytic synapse’ is required for dectin-1 hemITAM signaling regulated by CD45, and its related membrane tyrosine phosphatase CD148 [19]. CD45 and CD148 are membrane proteins with a large extracellular domain and intrinsic tyrosine phosphatase activity. Both are required for surface molecule rearrangement following macrophage-WGP binding, and to assist with the removal of inhibitory phosphatases to allow Src activation; however, to permit productive signaling, CD45 and CD148 are then isolated from the contact site of dectin-1 with WGP in the forming phagosome (Figure 3). SGs seem to fail to sequester CD45 and CD148, and therefore they are incapable of continuing the signaling cascade. Consequently, dectin-1, unlike other PRRs, plays a key role within the innate immune response by distinguishing between direct binding to microbes and binding to substances shed from microbes.

*In vitro* studies of murine and human dectin-1 clones highlights a new dectin-1 signaling model, which proposes the existence of two parallel similar structures of the entire dectin-1 unit, where the bridging of the twin dectin-1 monomers is necessary to allow dual Src and spleen tyrosine kinase (Syk) activation and recruitment, and consequent NFAT and NF-κB induced transcription for inflammatory regulation to drive the production of cytokines and chemokines [19, 59, 60]. Although the oligomerization of dectin-1 has been questioned by Brown [23], agreement exists regarding the concept of the ability of dectin-1 to recognize carbohydrates, a function unusual in the classical C-type lectin family of receptors. For the antimicrobial aspect of the response however, the phosphorylation of a unique Src domain appears sufficient for stimulation of phagocytosis and subsequent oxidative burst, as shown in murine bone marrow-derived macrophages and the RAW macrophage lines [30]. In macrophages, phagocytosis is Syk-independent, whilst in dendritic cells, it is Syk-dependent [43]. This may present another cue for ‘innate immune memory’, as it is only activated in a subspecialized population of macrophages primed for ROS production [30]. Another sign that suggests a resemblance between innate and adaptive response via regulation of transcription is the dectin-1 synergy with TLR pathways, in particular TLR2, TLR4, and TLR6, to enhance cytokine production from monocytes and macrophages against fungal pathogens [23]. As reviewed by Goodridge *et al*. [19], this synergy is mediated via caspase recruitment domain containing protein 9 (CARD9) found in the sub-population of murine bone marrow derived dendritic cells and macrophages destined for cytokine production upon β-glucan binding. TLR2-dectin-1 synergy leads to TNF-α and IL-12 secretion and promotes IL-2 and IL-10 production via Syk activation [61, 62]. Multiple other components like CD36 are involved in the complex interaction between dectin-1 and TLRs [23]. Moreover, both IL-10 and IL-2 are involved in the development of regulatory T cells during fungal infection, pointing to the innate-adaptive immune interaction [63, 64]. Lastly, the most novel role for dectin-1 involving trained immunity has been described for the regulation of cell death in murine macrophages and dendritic cells, where CARD9 has the dual role of enhancing TLR signaling through MAP kinase activation and directly promoting NF-κB activation [19, 65-67].

**Complement Receptor 3 Structure and Signaling**

The second pertinent receptor for β-glucan interaction with the innate immune system is complement receptor (CR3), a β2 integrin protein that serves as an adhesion and recognition receptor with an exceptionally broad pathogen-recognition capability [68]. CR3, while under the PRR umbrella, belongs to the complement system. The complement system is composed of proinflammatory serum proteins that ‘complement’ inflammation and circulate as sedentary precursors until activated to cause cell lysis or opsonization [23, 69]. Complement receptors are expressed on several leukocytes and are key mediators for yeast, fungal and bacterial infection; hence β-glucan recognition is essential, as it is a main component of the cell wall of these (micro)organisms [70]. CR3 triggers phagocytosis of complement opsonized particles by binding to complement activation molecule iC3 as shown in murine models [71-74], and human PBMCs and neutrophils [48, 75, 76], but it also works by direct pathogen binding [77, 78]. The CR3-β-glucan signaling cascade is yet to be fully decoded [79, 80]; however, it is thought that phosphorylation of tyrosine kinase Syk, PI3K and p38MAPK are involved in the response, as reflected in murine [37] and human immune cells [81, 82]. Fluoresce microscopy of mouse macrophages reveals how ingested β-glucans are rapidly taken up in a CR3-independent manner (dectin-1 supported) and are shuttled to the bone marrow, spleen and lymph nodes to slowly release soluble β-1,3-glucan active moiety fragments [3, 37]. As evident in human neutrophils, it is these smaller fragments, and not the parent polysaccharide, that are responsible for CR3-dependent signaling. CR3 is unique among integrins as it consists of a β-subunit (CD18) and an α-chain (CD11b) [83, 84]. As shown in Figure 4, CD11b hosts two binding sites, one at the I-domain site near the N-terminus for ligands such as iC3b, and another one at the lectin-site near the C-terminus for short carbohydrates like β-1,3-glucan. The soluble smaller β-glucan fragments prime phagocytes by binding to the lectin site of CD11b in mouse and human cells [37, 48, 73]. Signaling of CR3 requires dual ligation of CD11b, since the activation of the lectin site triggers configurational changes that expose the antigen binding site to ultimately mediate chemotaxis and phagocytosis towards the iC3b opsonized pathogenic cell. Subsequently, β-glucan primed circulating phagocytes more avidly cause cytotoxicity of iC3b opsonized particles as shown in human neutrophils [81]. Upon iC3b opsonization, the chemotactic factors C3a and C5a are produced. C3a mainly recruits eosinophils (also basophils and mast cells), whereas C5a predominantly attracts neutrophils. In murine tumor models, the major cytotoxic activity of the β-glucan C3-dependent immune response has been attributed to the C5a chemokine cascade [85].

**Enhanced Oxidative Burst of Primed Innate Immune Cells**

Following dectin-1 dependent WGP phagocytosis and CR3-dependent priming, an oxidative burst response within the phagolysosome is activated in PBMCs [18]. Research with human PBMCs revealed that the WGP-induced oxidative burst is dectin-1 mediated and activates Src and PI3K/AKT signaling pathways. On the other hand, SG-induced oxidative burst is CR3 mediated and activates actin polymerization and focal adhesion, followed by Src, Syk, PI3K/AKT, p38 MAPK, and PLC/PKC pathways [18]. Both signaling cascades (WGP via dectin-1 and SG via CR3) ultimately activate the NADPH oxidase system to generate O2- from O2 molecules. In the absence of phagocytosis, ROS are released into the environment via the signaling pathway that activates NADPH oxidase in response to focal adhered β-glucan; this involves recognition by CR3 and signaling through p38 MAPK pathway in a process called ‘frustrated phagocytosis’ , observed in mouse macrophages [86] and human neutrophils [82] (Figure 5). Furthermore, the priming of PBMCs with WGP significantly upregulates chemotaxis towards C5a and suppresses that towards IL-8, which reduces the handicap of traveling through endogenous chemotactic gradients to reach final chemoattractant C5a [87].

**Translating β-glucan immune training to humans**

Many studies have explored biological response modifiers of the innate immune system to increase host resistance to pathogens, with β-glucans ranking as one of the most widely researched. In conjunction with other standard immuno-modulatory treatments, this strategy may offer a non-specific protection potentially becoming a method of prevention and treatment for infections and enhancing the immune resistance of immuno-compromised populations including the very young and older, frail people, and athletes. However, it must be acknowledged that, as the majority of the human studies available use primarily questionnaire-based methodologies (addressing incidence, duration and symptoms of infection) to draw conclusions, much uncertainty still exists about the evidence-based relationship between β-glucan supplementation in humans and consequent immunologic changes. This review organizes the existing human studies according to population, highlighting demographic, intervention and outcome details for each study presented, according to the pre-specified inclusion criteria indicated above, and with a focus on upper respiratory tract infection (URTI). The identity of all commercial preparations used is detailed in Table 1.

**Upper Respiratory Tract Infection in Different Age Groups and in Athletes**

Due to the large body of work focusing on the potential impact of β-glucans to alleviate URTI, this condition and its prevalence will be described briefly. URTI, usually referred to as the common cold, is the most frequent infectious disease in humans, with an average prevalence of 2 to 4 episodes/year in adults and 6 to 10 episodes/year in children [88-90]. Symptoms include nasal congestion, sore throat, and headaches; and although usually self-limiting, they carry an increased risk for secondary bacterial infection and further comorbidities following inadequate recovery. Due to the heterogenic etiology of URTI, there is no reliable intervention to reduce the frequency of infection, making it the primary cause for antibiotic prescription in the UK with a total disease-associated cost of €40 billion/year nationwide [89, 91]. Hence, any alternative treatments showing a moderate alleviation of URTI incidence and severity would be vastly beneficial for population health and for the health system.

Respiratory infections are highly common in the first 5 years of life due to children’s narrower airways, and the immature lungs and immune system [92]. In fact, respiratory infections followed by diarrhea are the two main causes of morbidity and mortality in children younger than 5 years old worldwide [93]. The Department of Pediatrics, ChangPing Woman and Children Health Care Hospital in China reported that Wellmune® supplementation (35 mg/day for 12 weeks) kept children healthier during the cold and flu season by significantly decreasing the incidence and duration of the common cold by 66% compared to placebo [94]. No significant difference in efficacy or safety parameters between the Wellmune® groups (35 and 75 mg/day) was found, suggesting that 35 mg of Wellmune® daily is sufficient to achieve optimal benefit during the cold and flu season in children aged 1 to 4 years. On an immuno-cellular level, Glucan #300® is the most studied β-glucan in older children and teens and has been shown to improve mucosal innate immunity. Three studies evaluated mucosal innate immunity in children aged 8 to 12 years with recurrent respiratory problems treated with 100 mg of Glucan #300 daily [95-97]. One further study examined mucosal innate immunity of children aged 6 to 16 years [98] who were treated with 100 mg of Glucan#300 daily. Mucosal immunity represents the first line of defense against respiratory viruses and bacteria and is mainly composed of salivary IgA and antimicrobial proteins. A reduction in IgA secretion levels has been linked to an increased incidence of URTI; thus, IgA is used as a proxy measure for mucosal immune function [99]. The first two studies measured salivary albumin, lysozyme, C-reactive protein (CRP) and calprotectin levels and reported a significant increase in lysozyme and CRP, compared to placebo [95, 96]. The third study assessed salivary antibody levels (IgA, IgM and IgG), showing a significant increase in all tested antibodies, compared to a reduction in the placebo group [97]. Finally, the fourth study explored not only the effects of β-glucan on immune status using salivary IgA and salivary eNOS, but also on physical activity using the 6-minute walking test [98]. There were positive effects of β-glucan supplementation in the two age groups, 6 to 10 and 11 to 16 years, via a significant stabilization of salivary IgA compared to a decrease in the placebo counterpart, and higher physical activity scores. Taken together, these results suggest that β-glucan can improve mucosal innate immunity to reduce URTI incidence and symptom severity in children and teens.

Young children are especially susceptible to inadequate nutritional intake with a cascade effect on immunity and increased risk for infection and allergic reactions [100-102]. Mead Johnson Nutrition conducted two studies with children in day-care settings in China [103] and Brazil [104], in which the incidence of acute respiratory infection (ARI) was evaluated. Children in the experimental group consumed follow-up formula fortified with vitamins A (380 vs 630 IU, control and treatment respectively) and D (31 vs 119 IU, control and treatment respectively), 25 mg docosahexaenoic acid (DHA), 1.2 g of polydextrose/galacto-oligosaccharides and 8.7 mg of Wellmune® three times a day for 28 weeks. In the first study, children who consumed the fortified formula showed a significant decrease in the incidence and duration of ARI, antibiotic use and missed day-care hours, together with higher serum concentrations of the anti-inflammatory cytokine IL-10 and higher blood leukocyte levels, compared with children who were given an iso-caloric unfortified, cow’s milk-based beverage [103]. The second study reported a significant reduction in allergic manifestations in the skin and respiratory tract between groups, although no significant difference was reported for ARI nor diarrheal disease [104]. This may be due to an increase in serum/plasma/cellular IL-10 and its ability to inhibit Th2 mediators, which are usually elevated during allergic inflammation. These results cannot be attributed solely to β-glucan, since vitamins, prebiotics and polyunsaturated fatty acids were present in the experimental group’s beverage but not in the control drink (Table 2).

Immuno-senescence refers to the age-related decline of the immune system, which is characterized by a reduced ability to protect against and combat infections [105-107]. A study evaluating whether 250 mg/day of Wellmune® enhanced immune function during the winter months in healthy older adults, reported a reduction in symptomatic days, which approached but was not significant, compared to the placebo group [108]. Wellmune® supplementation significantly increased LPS-stimulated IFN-γ production, suggesting priming of innate immunity. However, Wellmune® supplementation did not change symptom severity, plasma cytokine or chemokine levels or salivary IgA concentration. A cross-over study investigated the humoral immune and inflammatory response in participants aged 65 years or older after consuming 2.5 mg of Lentinex® each day for 6 weeks [109], and showed a significant increase in circulating B-cell number. Other immunological and inflammatory markers, such as immunoglobulins, complement proteins and cytokines, were not significantly altered. Taken together, these study results suggest that there may be an association between β-glucan supplementation and defense against infection in the elderly. Nonetheless, it is important to note that the mechanisms behind putative health effects summarized in Table 3 require greater clarity and definition.

Athletes often experience a transient state of immuno-suppression after highly demanding physical effort leading to an ‘open window’ for opportunistic infections. This is reflected in the greater incidence of URTI among ultra-marathoners and marathoners [110-112]. A ‘J’ shaped model has been suggested to reflect the relationship between risk of URTI and amount and intensity of exercise; where sedentarism shows average infection risk, moderate-regular exercise confers maximal infection resistance and extremely heavy acute or chronic exercise decreases resistance to infection [113]. It should be highlighted that fitness level highly impacts how the body reacts to exercise-induced stress [114]. Physical stress is similar to other stressors, such as psychological stress, which can lead to a weakened immune system [115]. Psychological stress may also arise from prolonged training and competition, which is also posited to play a role in the ‘open window’. Likewise, elite athletes show deterioration in their mood during intense exercise periods, such as marathon training [116, 117]. The mechanisms explaining this immuno-suppression have not been firmly established; however it may be caused by impaired cell-mediated immunological responses and psychological factors including stress and anxiety [118]. An impaired cell-mediated immunological response during the ‘open window’ is characterized by an overall reduction in the concentration of circulating immune cells, blunted TLR expression [119], together with a concurrent state of inflammation [120], all leading to a net suppression of cellular immunity and consequent increased risk of infection. In particular, monocyte and T-cell numbers increase during and right after a bout of intense exercise to later decrease below pre-exercise levels over 2 to 6 hours, slowly recovering to baseline status within 24 hours. This, together with a suppression of LPS-stimulated cytokines [121], explains why mucosal immunity is subjected to a temporary disruption during the 24 hour-window after strenuous exercise [99].

The effect of Wellmune® supplementation (250 mg/day for 10 days) was measured after a high heat and humidity bout of aerobic exercise in two clinical trials [122, 123]. Wellmune® significantly prevented the drop of T cells and monocytes commonly seen after an intense exercise session and improved the LPS-challenged response in monocytes from recreational athletes [122] and individuals of average fitness level [123]. In the third study, Wellmune® prevented post exercise suppression of mucosal immunity as measured by a 32% increase in salivary IgA 2 hours post-exercise compared to the placebo group [99]. The plasma cytokine profile seen in the β-glucan supplemented group is linked to lymphocyte proliferation (IL-7), neutrophil-related increased chemotaxis (IL-8), anti-inflammatory action (IL-10) [122], reduced drop of circulating monocyte levels and favorable alteration in their cell-surface receptors. The last refers to receptors CD38, CD86 and TLR4, which confer improved T cell stimulation capacity, increased T-cell concentration and activity (IL-2), leukocyte priming (IFN-γ) and other markers that may suggest increased immuno-surveillance (CD8+ TCM cells) [123]. The LPS-stimulated cytokine profile showed a significant increase in IL-2, IL-4, IL-7, and IFN-γ compared to the placebo counterpart, which also suggests enhanced immuno-surveillance by the priming of monocytes, reflected in the higher Th1 cytokine production and increased TLR4 expression [123, 124].

Aside from artificially-taxing physical conditions like heat and humidity, Wellmune® supplementation was evaluated in athletes during a naturally challenging scenario, a pre- and post-marathon period. Participants were randomized to incorporate dispersible Wellmune® (250 mg Wellmune®/day) or placebo in a dairy-based beverage 45 days prior to, on the day of and 45 days post marathon [125]. In a second study, participants were randomized to receive a soluble Wellmune® formulation, a dispersible Wellmune® formulation or placebo for 28 days post-marathon [99]. Results showed a significant decrease in post-marathon URTI symptomatic days [99, 125], severity (especially nasal discharge and sore throat) and average missed workout days [125] in comparison to the placebo groups, while no significant difference was observed for average duration or incidence of URTI in either study. Another study supplementing Wellmune® for 4 weeks of post marathon training (250 or 500 mg Wellmune®/day) also evaluated URTI symptoms as well as overall health and mood using a questionnaire style health log and the profile of mood state (POMS) questionnaire, respectively [126]. The authors reported significantly lower URTI symptoms and better overall health and mood profiles in both supplement-dose groups compared to placebo.

Overall, Wellmune® has been shown to reduce the duration of the ‘open window’ by priming granulocytes for faster activation and cytokine production upon pathogenic challenges, increasing salivary IgA, improving circulating monocyte and T cell count and altering the balance of T helper cytokines (Th1/Th2). This helped to reduce URTI severity and improve overall health and mood in physically-induced stress independent of fitness level, as it was evident in trained [99, 125, 126] and recreational athletes [122] as well as individuals of average fitness level [123] (Table 4). This set of studies provides a good immuno-cellular insight into the β-glucan interactions with the immune system in regards to physically-induced stress. Previous research has outlined that monocyte adhesion molecule expression, such as that determined by CD38 gene expression, affects leukocyte migration capacity into damaged tissue. CD38 plays a role in the transport and adhesion mechanisms of white blood cells [127], and its downregulation on classical monocytes upon Wellmune® supplementation is in part regulated by changes in IFN-γ and may explain the observed increased levels of circulating monocytes. Additionally, Wellmune® supplementation caused CD86 upregulation in non-classic monocytes, in turn resulting in T-cell co-stimulation for rapid response upon antigen invasion [123]. Another sign of amplified immuno-surveillance is the upregulation of TLR4 and TLR2 on non-classic monocytes upon supplementation, which has been previously linked to the regulation of resistance exercise-induced inflammation [123, 128]. T-cells follow a similar pattern as monocytes during exercise and the recovery period, a slight increase during exercise followed by a drop below base-line post-exercise [129]; however, Wellmune® supplementation triggered an overall increase in circulating T cells prior to and after exercise [123]. One mechanism of action that may underlie these events is the priming of neutrophils, which increases MAMP recognition, and the priming of monocytes, which increases Th1 production capacity. The enhanced monocyte response from TLR4 upregulation supports this hypothesis [40].

**Studies Regarding Overall Health**

Allergic rhinitis (AR, commonly termed hay fever), is a leading cause of seasonal allergy symptoms and affects more than one-fifth of the adult population in Western Europe [130]. Typical symptoms include nasal congestion, sneezing, itchy eyes and difficulty breathing. AR is an IgE-mediated inflammatory allergic reaction of the nasal mucosa that is in part driven by the overproduction of Th2 cytokines (IL-4, IL-5, IL-13) and subsequent stimulation of eosinophil generation and chemotaxis. These can be suppressed by Th1 cytokines like IL-12 and IFN-γ [131, 132], where β-glucans are thought to play a role. The available standard allergen immuno-therapy for hay fever includes high doses of allergen administration with the view of desensitization [133]; however, results are variable and there is still a need for safe, effective and convenient therapy [134]. Thus, Kirmaz *et al*. [135] evaluated the effect of Imuneks® on AR with the aim of assessing whether 12-weeks of β-glucan (10 mg Imuneks® twice daily) could assist Th1-mediated immune response in *Olea europa* sensitive patients with AR. For this, they challenged the participants with a nasal provocation test of *Olea europa* followed by a nasal lavage and measured IL-4, IL-5, IL-12, IFN-γ and eosinophil count in the nasal fluid as well as peripheral blood eosinophil levels. Results from the nasal lavage showed a significant decrease in Th2 cytokines (IL-4 and IL-5) and an increase in Th1 cytokines (IL-12) together with a reduction in eosinophils compared to placebo; however, IFN-γ and peripheral blood eosinophil levels did not show a significant difference between groups [135]. Altogether, β-glucan could be considered as an adjunct to standard AR treatment due to its robust alleviation of the exaggerated Th2 activity in AR patients.

With a focus on symptom severity, Wellmune® has been studied for improving ragweed allergy symptoms in adults supplemented with 250 mg Wellmune®/day for 4 weeks [136]. The results showed significant reduction in total allergy symptoms and symptom severity, overall increased POMS score and an improvement in global mood state, physical function and all rhinoconjunctivitis. Quality of Life Questionnaire ratings including sleep problems, nasal and non-nasal symptoms, eye symptoms, activities, emotion and quality of life improved compared to placebo; although non-nasal symptoms and emotion were also significantly improved in the placebo group. Nonetheless, even though allergic rhinitis is considered to result from an IgE-mediated allergic response, no significant difference was found for serum IgE levels between groups. In addition, Yestimun®, another proprietary highly purified Baker’s Yeast β-1,3/1,6-glucan was evaluated in three studies regarding effects on incidence and symptoms of the common cold in adults with recurring infections [137-139]. In the first study, participants took 450 mg of Yestimun® twice a day for 26 weeks, whereas the second and third studies supplemented with 900 mg of Yestimun® once a day for 16 weeks. All studies showed an overall prophylactic effect, with a significant reduction of common cold incidence during the infection season; however, whilst the first and third study showed significant symptom reduction, the second study did not reach significance for this parameter, but showed significant reduction in sleep disturbances caused by infections and better efficacy rating by participants and physicians. Furthermore, a randomized phase II clinical trial was conducted to evaluate immune support during the cold and flu season by assessing symptoms and overall well-being, where participants took 500 mg of Wellmune® or placebo per day for 12 weeks [140]. While the treatment group experienced a significant increase in quality of life as rated by the SF-36V2-QOL physical component summary score and a decrease in missed days of work/school, there was no significant difference in the incidence of symptomatic respiratory infections between the groups. Lastly, with a molecular emphasis, Glucan #300® was used to evaluate β-glucan’s effects on innate immune response in adults [141]. For this, serum β-glucan levels, *ex vivo* stimulation of leukocytes and microbicidal activity in blood were analyzed in a randomized open-label intervention pilot study with 15 healthy male adults receiving 1 g of Glucan #300® daily for 7 days. At all points, β-glucan was barely detectable in serum, and leukocyte cytokine production together with microbicidal activity were unaltered compared to the placebo group. Taken together, there seems to be some evidence to indicate that individuals suffering from allergies or at increased risk of respiratory infection may benefit from β-glucan supplementation to reduce symptom severity and overall quality of life during infection season. However, aside from AR, where β-glucan’s effect on suppressing Th2 cytokine response was evident, how β-glucan affects immunity at a cellular level to achieve such benefits is not clear (Table 5).

**Dietary Regulations and Emerging Areas of Research**

There is great interest in the application of β-glucans in the food and beverage section worldwide. Food Marketing Industry data showed that the global β-glucan market was worth $307.8 million in 2016 with a prediction by Markets and Markets to reach $476.5 million in 2022 [142]. Yestimun® is the only β-glucan that has attempted to hold a European Food Safety Authority (EFSA) health claim in regards to immunity; however, it received a negative response when applying for the health claim “initiation of appropriate innate and adaptive immune responses in adults” in 2009 at a dose of 0.45 g twice daily. The EFSA Panel on Dietetic Products, Nutrition and Allergies concluded that cause and effect relationship could not be established between the consumption of Yestimun® and the initiation of appropriate innate and adaptive immune responses due to the existence of contradicting results in the literature [143]. On the other hand, Wellmune® successfully holds a claim as a novel food ingredient granted in 2011 by the EFSA Panel on Dietetic Products, Nutrition, and Allergies. This application refers to both the insoluble and soluble formulations of Wellmune® at dose levels up to 375 mg/day in food supplements and up to 600 mg/day in foods for particular nutritional uses (PARNUTS, also referred to as functional or ‘smart’ food), but not for infant or follow up formulas [144].

**Gaps in Knowledge and Perspectives for the Future, Uncertain Immunity Biomarkers**

Hard primary endpoints of innate immune functionality that are sensitive to nutritional interventions in man are lacking. In the pharma space, inflammatory biomarkers, such as CRP and other acute-phase proteins, are sometimes used as endpoints. However, it is recognized that there are significant challenges regarding biomarkers of immunity and inflammation as highlighted by an International Life Sciences Institute Europe (ILSI Europe) workshop focused on ‘Low-grade inflammation, a high-grade challenge: biomarkers and modulation by dietary strategies’ [145]. Thus, to more effectively capture innate immune changes in response to nutritional contexts in man, it was proposed that an integrated high-dimensional data approach fusing epigenetic, transcriptomic and proteomic signatures might be appropriate [146, 147]. Furthermore, given the close inter-relationship between metabolism and the immune response, there is no doubt that in time metabolic signatures may act as surrogate biomarkers of innate immune functionality. Such approaches will allow us to better dissect the dynamic inter-relationship between nutritional/metabolic status, metabolism and innate immunity, in order to more robustly define potential food-derived nutrient and/or non-nutrient sensitive immune health versus disease associations. It is only with the hard-mechanistic evidence that the field can truly advance in terms of understanding the real ability of dietary components, including β-glucans, to positively modulate metabolism and innate immune cell functionality. Additionally, there is no doubt that the impact of nutritional interventions is highly variable between individuals. This common phenomenon of high variability between subjects is particularly pertinent within the context of nutritional modulation of the immune system. Simple determinants of variability include age, sex, body mass index, medications, etc. In addition, the magnitude of response between individuals to a similar dose of a nutrient can be highly variable [148]. These additional challenges associated with personalized responses need to be factored into the research evidence underpinning any new product, including β-glucans in association with putative health effects.

Overall the balance of evidence suggests that β-1,3/1,6-glucan supplementation from Baker’s Yeast (*S. cerevisiae*) shows immune-enhancing and immune-modulatory effects across different populations. Preclinical studies have revealed major insights in relation to β-1,3/1,6-glucan signaling pathways and toxicological parameters; however, further research involving large randomized controlled trials with a clear immunological target need to be undertaken before the association of β-glucan and immuno-stimulation in humans can be more clearly understood. Studies into the cellular and molecular mechanisms of action have been established via fluorescence microscopy showing that orally administered β-1,3/1,6-glucans are engulfed and processed by macrophages and dendritic cells that later travel to the different immune organs and release fragmented soluble β-1,3-glucan particles. This results in the priming of leukocytes via dectin-1, CR3 and other collaborating receptors to enhance immuno-surveillance and counter pathogen attack by increasing the function of the complement system and innate immune cells. Altogether this leads to improved antimicrobial and inflammatory responses. β-1,3/1,6-glucans have been examined in several randomized, double-blind, placebo-controlled human studies using different proprietary β-1,3/1,6-glucan formulations. Overall, these indicate that oral β-glucan supplementation may improve symptom severity and duration in populations prone to respiratory infections and allergies. The most studied supplement is Wellmune®, which has been evaluated across the widest population range to reveal significant alleviation of URTI, allergic episodes and exercise- and age-related immuno-suppression. Other proprietary β-1,3/1,6-glucans have also shown promise; Glucan #300® improved mucosal innate immunity in older children and teens, Lentinex® was shown to alleviate immuno-senescence through an increase in immune cell count, Yestimun® had a prophylactic effect in adults with recurring infections and Imuneks® reduced the exaggerated Th2 response in allergic rhinitis. However, for many of these studies, non-significant differences between placebo and supplement groups indicate no reduction in infection incidence or cytokine production or microbial activity. Thus, to better understand β-glucan’s immuno-cellular mechanisms in humans, further well-designed human intervention trials are required to deeper evaluate immunity biomarkers that could potentially indicate the mechanisms of action behind the effect of β-glucans on immune function in humans.

**Authors contribution**

EDMC drafted the manuscript with advice from HMR; HMR and PCC provided critical revisions. All authors approved the final version of the manuscript.

**Acknowledgment**

EDMC is supported by the Irish Research Council Employment Based PhD Scholarship Scheme. Work pertaining to this review and HMR are supported by the Irish Department of Agriculture, Food and Marine ‘ImmunoMet’ Programme (Grant number:14/F/828).

**Conflicts of interest**

PCC has received funding as part of trials investigating Wellmune®. EDMC is in receipt of a PhD studentship co-funded by the Irish Research Council and Kerry.

**References**

[1] Estrada, A., Yun, C.-H., Kessel, A. V., Li, B., Hauta, S., Laarveld, B., Immunomodulatory activities of oat β-glucan in vitro and in vivo. *Microbiology and immunology* **1997**, *41*, 991-998.

[2] Rondanelli, M., Opizzi, A., Monteferrario, F., [The biological activity of beta-glucans]. *Minerva Med* **2009**, *100*, 237-245.

[3] Volman, J. J., Ramakers, J. D., Plat, J., Dietary modulation of immune function by β-glucans. *Physiology & behavior* **2008**, *94*, 276-284.

[4] El Khoury, D., Cuda, C., Luhovyy, B., Anderson, G., Beta glucan: health benefits in obesity and metabolic syndrome. *Journal of nutrition and metabolism* **2011**, *2012*.

[5] Novak, M., Vetvicka, V., β-glucans, history, and the present: immunomodulatory aspects and mechanisms of action. *Journal of immunotoxicology* **2008**, *5*, 47-57.

[6] Talbott, S. M., Talbott, J. A., Talbott, T. L., Dingler, E., beta-Glucan supplementation, allergy symptoms, and quality of life in self-described ragweed allergy sufferers. *Food Sci Nutr* **2013**, *1*, 90-101.

[7] Kim, H. S., Hong, J. T., Kim, Y., Han, S.-B., Stimulatory Effect of β-glucans on Immune Cells. *Immune Netw* **2011**, *11*, 191-195.

[8] Stier, H., Ebbeskotte, V., Gruenwald, J., Immune-modulatory effects of dietary Yeast Beta-1, 3/1, 6-D-glucan. *Nutrition journal* **2014**, *13*, 38.

[9] Zeković, D. B., Kwiatkowski, S., Vrvić, M. M., Jakovljević, D., Moran, C. A., Natural and modified (1→ 3)-β-D-glucans in health promotion and disease alleviation. *Critical reviews in biotechnology* **2005**, *25*, 205-230.

[10] Hewawasam, R., Udawatte, C., Weliwegamage, S., Sotheeswaran, S., Rajapakse, S., Immobilization of selected microbes at some selected solid supports for enhanced fermentation process. *Ferment Technol* **2015**, *4*, 2.

[11] Ruiz-Herrera, J., Ortiz-Castellanos, L., Cell wall glucans of fungi. a review. *The Cell Surface* **2019**, 100022.

[12] Demleitner, S., Kraus, J., Franz, G., Synthesis and antitumour activity of sulfoalkyl derivatives of curdlan and lichenan. *Carbohydrate research* **1992**, *226*, 247-252.

[13] Driscoll, M., Hansen, R., Ding, C., Cramer, D. E., Yan, J., Therapeutic potential of various beta-glucan sources in conjunction with anti-tumor monoclonal antibody in cancer therapy. *Cancer biology & therapy* **2009**, *8*, 218-225.

[14] Bohn, J. A., BeMiller, J. N., (1→ 3)-β-D-Glucans as biological response modifiers: a review of structure-functional activity relationships. *Carbohydrate polymers* **1995**, *28*, 3-14.

[15] Wang, Q., Sheng, X., Shi, A., Hu, H., Yang, Y., Liu, L., Fei, L., Liu, H., β-Glucans: Relationships between modification, conformation and functional activities. *Molecules* **2017**, *22*, 257.

[16] Pillemer, L., Ecker, E. E., Anti-complementary factor in fresh yeast. *Journal of Biological Chemistry* **1941**, *137*, 139-142.

[17] Goodridge, H. S., Reyes, C. N., Becker, C. A., Katsumoto, T. R., Ma, J., Wolf, A. J., Bose, N., Chan, A. S., Magee, A. S., Danielson, M. E., Activation of the innate immune receptor Dectin-1 upon formation of a ‘phagocytic synapse’. *Nature* **2011**, *472*, 471.

[18] Bose, N., Wurst, L.R., Chan, A.S., Dudney, C.M., LeRoux, M.L., Danielson, M.E., Will, P.M., Nodland, S.E., Patchen, M.L., Dalle Lucca, J.J. and Lebeda, F.J., Differential regulation of oxidative burst by distinct β-glucan-binding receptors and signaling pathways in human peripheral blood mononuclear cells. *Glycobiology,* **2014**, *24*, 379-391.

[19] Goodridge, H. S., Wolf, A. J., Underhill, D. M., β‐glucan recognition by the innate immune system. *Immunological reviews* **2009**, *230*, 38-50.

[20] EFSA Panel on Dietetic Products, N. a. A. N., in: Authority, E. F. S. (Ed.), *9(5):2137*, EFSA Journal 2011.

[21] Babíček, K., Čechová, I., Simon, R., Harwood, M., Cox, D., Toxicological assessment of a particulate yeast (1, 3/1, 6)-β-D-glucan in rats. *Food and chemical toxicology* **2007**, *45*, 1719-1730.

[22] Hong, F., Hansen, R. D., Yan, J., Allendorf, D. J., Baran, J. T., Ostroff, G. R., Ross, G. D., β-Glucan functions as an adjuvant for monoclonal antibody immunotherapy by recruiting tumoricidal granulocytes as killer cells. *Cancer research* **2003**, *63*, 9023-9031.

[23] Brown, G. D., Dectin-1: a signalling non-TLR pattern-recognition receptor. *Nature Reviews Immunology* **2006**, *6*, 33.

[24] Hoffmann, J. A., Kafatos, F. C., Janeway, C. A., Ezekowitz, R., Phylogenetic perspectives in innate immunity. *Science* **1999**, *284*, 1313-1318.

[25] Aderem, A., Underhill, D. M., Mechanisms of phagocytosis in macrophages. *Annual review of immunology* **1999**, *17*, 593-623.

[26] Brown, G. D., Taylor, P. R., Reid, D. M., Willment, J. A., Williams, D. L., Martinez-Pomares, L., Wong, S. Y., Gordon, S., Dectin-1 is a major β-glucan receptor on macrophages. *Journal of Experimental medicine* **2002**, *196*, 407-412.

[27] Fesel, P. H., Zuccaro, A., β-glucan: Crucial component of the fungal cell wall and elusive MAMP in plants. *Fungal Genetics and Biology* **2016**, *90*, 53-60.

[28] Aderem, A., Ulevitch, R. J., Toll-like receptors in the induction of the innate immune response. *Nature* **2000**, *406*, 782.

[29] Nathan, C., Neutrophils and immunity: challenges and opportunities. *Nature reviews immunology* **2006**, *6*, 173.

[30] Underhill, D. M., Rossnagle, E., Lowell, C. A., Simmons, R. M., Dectin-1 activates Syk tyrosine kinase in a dynamic subset of macrophages for reactive oxygen production. *Blood* **2005**, *106*, 2543-2550.

[31] Bowdish, D. M. E., Loffredo, M. S., Mukhopadhyay, S., Mantovani, A., Gordon, S., Macrophage receptors implicated in the “adaptive” form of innate immunity. *Microbes and Infection* **2007**, *9*, 1680-1687.

[32] Netea, M. G., Joosten, L. A., Latz, E., Mills, K. H., Natoli, G., Stunnenberg, H. G., O’Neill, L. A., Xavier, R. J., Trained immunity: a program of innate immune memory in health and disease. *Science* **2016**, *352*, aaf1098.

[33] Netea, M. G., Quintin, J., van der Meer, J. W., Trained immunity: a memory for innate host defense. *Cell host & microbe* **2011**, *9*, 355-361.

[34] Quintin, J., Saeed, S., Martens, J. H., Giamarellos-Bourboulis, E. J., Ifrim, D. C., Logie, C., Jacobs, L., Jansen, T., Kullberg, B.-J., Wijmenga, C., Candida albicans infection affords protection against reinfection via functional reprogramming of monocytes. *Cell host & microbe* **2012**, *12*, 223-232.

[35] McCann, F., Carmona, E., Puri, V., Pagano, R. E., Limper, A. H., Macrophage internalization of fungal beta-glucans is not necessary for initiation of related inflammatory responses. *Infect Immun* **2005**, *73*, 6340-6349.

[36] Bowman, S. M., Free, S. J., The structure and synthesis of the fungal cell wall. *Bioessays* **2006**, *28*, 799-808.

[37] Li, B., Allendorf, D. J., Hansen, R., Marroquin, J., Ding, C., Cramer, D. E., Yan, J., Yeast β-glucan amplifies phagocyte killing of iC3b-opsonized tumor cells via complement receptor 3-Syk-phosphatidylinositol 3-kinase pathway. *The Journal of Immunology* **2006**, *177*, 1661-1669.

[38] Rice, P. J., Adams, E. L., Ozment-Skelton, T., Gonzalez, A. J., Goldman, M. P., Lockhart, B. E., Barker, L. A., Breuel, K. F., DePonti, W. K., Kalbfleisch, J. H., Oral delivery and gastrointestinal absorption of soluble glucans stimulate increased resistance to infectious challenge. *Journal of Pharmacology and Experimental Therapeutics* **2005**, *314*, 1079-1086.

[39] Ganda Mall, J.-P., Casado-Bedmar, M., Winberg, M. E., Brummer, R. J., Schoultz, I., Keita, Å. V., A β-glucan-based dietary fiber reduces mast cell-induced hyperpermeability in ileum from patients with Crohn’s disease and control subjects. *Inflammatory bowel diseases* **2017**, *24*, 166-178.

[40] Hong, F., Yan, J., Baran, J. T., Allendorf, D. J., Hansen, R. D., Ostroff, G. R., Xing, P. X., Cheung, N.-K. V., Ross, G. D., Mechanism by which orally administered β-1, 3-glucans enhance the tumoricidal activity of antitumor monoclonal antibodies in murine tumor models. *The Journal of Immunology* **2004**, *173*, 797-806.

[41] Sandvik, A., Wang, Y., Morton, H., Aasen, A., Wang, J., Johansen, F. E., Oral and systemic administration of β‐glucan protects against lipopolysaccharide‐induced shock and organ injury in rats. *Clinical & Experimental Immunology* **2007**, *148*, 168-177.

[42] Brown, G. D., Gordon, S., Immune recognition: a new receptor for β-glucans. *Nature* **2001**, *413*, 36.

[43] Tsoni, S. V., Brown, G. D., β‐Glucans and Dectin‐1. *Annals of the New York Academy of Sciences* **2008**, *1143*, 45-60.

[44] Taylor, P. R., Brown, G. D., Reid, D. M., Willment, J. A., Martinez-Pomares, L., Gordon, S., Wong, S. Y., The β-glucan receptor, dectin-1, is predominantly expressed on the surface of cells of the monocyte/macrophage and neutrophil lineages. *The Journal of immunology* **2002**, *169*, 3876-3882.

[45] Brown, G. D., Herre, J., Williams, D. L., Willment, J. A., Marshall, A. S., Gordon, S., Dectin-1 mediates the biological effects of β-glucans. *Journal of Experimental medicine* **2003**, *197*, 1119-1124.

[46] Steele, C., Rapaka, R. R., Metz, A., Pop, S. M., Williams, D. L., Gordon, S., Kolls, J. K., Brown, G. D., The beta-glucan receptor dectin-1 recognizes specific morphologies of Aspergillus fumigatus. *PLoS pathogens* **2005**, *1*, e42.

[47] Ross, G. D., Cain, J. A., Myones, B. L., Newman, S. L., Lachmann, P. J., Specificity of membrane complement receptor type three (CR3) for ß-glucans. *Complement* **1987**, *4*, 61-74.

[48] Thornton, B. P., Vĕtvicka, V., Pitman, M., Goldman, R. C., Ross, G. D., Analysis of the sugar specificity and molecular location of the beta-glucan-binding lectin site of complement receptor type 3 (CD11b/CD18). *The Journal of Immunology* **1996**, *156*, 1235-1246.

[49] Czop, J. K., Austen, K. F., Generation of leukotrienes by human monocytes upon stimulation of their beta-glucan receptor during phagocytosis. *Proceedings of the National Academy of Sciences* **1985**, *82*, 2751-2755.

[50] Zimmerman, J. W., Lindermuth, J., Fish, P. A., Palace, G. P., Stevenson, T. T., DeMong, D. E., A novel carbohydrate-glycosphingolipid interaction between a β-(1–3)-glucan immunomodulator, PGG-glucan, and lactosylceramide of human leukocytes. *Journal of Biological Chemistry* **1998**, *273*, 22014-22020.

[51] Luther, K., Torosantucci, A., Brakhage, A. A., Heesemann, J., Ebel, F., Phagocytosis of Aspergillus fumigatus conidia by murine macrophages involves recognition by the dectin‐1 beta‐glucan receptor and Toll‐like receptor 2. *Cellular microbiology* **2007**, *9*, 368-381.

[52] Chan, G. C.-F., Chan, W. K., Sze, D. M.-Y., The effects of β-glucan on human immune and cancer cells. *Journal of hematology & oncology* **2009**, *2*, 25.

[53] Meier, A., Kirschning, C., Nikolaus, T., Wagner, H., Heesemann, J., Ebel, F., Toll‐like receptor (TLR) 2 and TLR4 are essential for Aspergillus‐induced activation of murine macrophages. *Cellular microbiology* **2003**, *5*, 561-570.

[54] Li, X., Zhang, X., Pang, L., Yao, L., ShangGuan, Z., Pan, Y., Agaricus bisporus-derived β-glucan enter macrophages and adipocytes by CD36 receptor. *Natural product research* **2019**, 1-4.

[55] Rice, P. J., Kelley, J. L., Kogan, G., Ensley, H. E., Kalbfleisch, J. H., Browder, I. W., Williams, D. L., Human monocyte scavenger receptors are pattern recognition receptors for (1→ 3)‐β‐D‐glucans. *Journal of leukocyte biology* **2002**, *72*, 140-146.

[56] Willment, J. A., Marshall, A. S. J., Reid, D. M., Williams, D. L., Wong, S. Y. C., Gordon, S., Brown, G. D., The human β-glucan receptor is widely expressed and functionally equivalent to murine Dectin-1 on primary cells. *European journal of immunology* **2005**, *35*, 1539-1547.

[57] Hernanz-Falcon, P., Arce, I., Roda-Navarro, P., Fernandez-Ruiz, E., Cloning of human DECTIN-1, a novel C-type lectin-like receptor gene expressed on dendritic cells. *Immunogenetics* **2001**, *53*, 288-295.

[58] Bose, N., Chan, A. S., Guerrero, F., Maristany, C., Walsh, R., Ertelt, K., Jonas, A., Gorden, K., Dudney, C., Wurst, L., Danielson, M., Elmasry, N., Magee, A., Patchen, M., Qiu, X., Vasilakos, J., Binding of Soluble Yeast β-Glucan to Human Neutrophils and Monocytes is Complement-Dependent. *Frontiers in immunology* **2013**, *4*.

[59] Kerrigan, A. M., Brown, G. D., Syk-coupled C-type lectins in immunity. *Trends in immunology* **2011**, *32*, 151-156.

[60] Kerrigan, A. M., Brown, G. D., Syk-coupled C-type lectin receptors that mediate cellular activation via single tyrosine based activation motifs. *Immunological Reviews* **2010**, *234*, 335-352.

[61] Ferwerda, G., Meyer-Wentrup, F., Kullberg, B. J., Netea, M. G., Adema, G. J., Dectin-1 synergizes with TLR2 and TLR4 for cytokine production in human primary monocytes and macrophages. *Cell Microbiol* **2008**, *10*, 2058-2066.

[62] Goodridge, H. S., Underhill, D. M., Fungal Recognition by TLR2 and Dectin-1. *Handbook of experimental pharmacology* **2008**, 87-109.

[63] Rogers, N. C., Slack, E. C., Edwards, A. D., Nolte, M. A., Schulz, O., Schweighoffer, E., Williams, D. L., Gordon, S., Tybulewicz, V. L., Brown, G. D., Reis e Sousa, C., Syk-dependent cytokine induction by Dectin-1 reveals a novel pattern recognition pathway for C type lectins. *Immunity* **2005**, *22*, 507-517.

[64] Osorio, F., LeibundGut-Landmann, S., Lochner, M., Lahl, K., Sparwasser, T., Eberl, G., Reis e Sousa, C., DC activated via dectin-1 convert Treg into IL-17 producers. *European journal of immunology* **2008**, *38*, 3274-3281.

[65] Inoue, M., Okinaga, T., Usui, M., Kawano, A., Thongsiri, C., Nakashima, K., Ariyoshi, W., Nishihara, T., β-glucan suppresses cell death of ASC deficient macrophages invaded by periodontopathic bacteria through the caspase-11 pathway. *FEMS Microbiology Letters* **2019**, *366*.

[66] Li, X., Luo, H., Ye, Y., Chen, X., Zou, Y., Duan, J., Xiang, D., β‑glucan, a dectin‑1 ligand, promotes macrophage M1 polarization via NF‑κB/autophagy pathway. *International journal of oncology* **2019**, *54*, 271-282.

[67] Ding, J., Ning, Y., Bai, Y., Xu, X., Sun, X., Qi, C., β-Glucan induces autophagy in dendritic cells and influences T-cell differentiation. *Medical microbiology and immunology* **2019**, *208*, 39-48.

[68] Schittenhelm, L., Hilkens, C. M., Morrison, V. L., beta2 Integrins As Regulators of Dendritic Cell, Monocyte, and Macrophage Function. *Frontiers in immunology* **2017**, *8*, 1866.

[69] Moticka, E. J., *A historical perspective on evidence-based immunology*, Newnes 2015.

[70] Cramer, D. E., Allendorf, D. J., Baran, J. T., Hansen, R., Marroquin, J., Li, B., Ratajczak, J., Ratajczak, M. Z., Yan, J., Beta-glucan enhances complement-mediated hematopoietic recovery after bone marrow injury. *Blood* **2006**, *107*, 835-840.

[71] Lu, H., Smith, C. W., Perrard, J., Bullard, D., Tang, L., Shappell, S. B., Entman, M. L., Beaudet, A. L., Ballantyne, C. M., LFA-1 is sufficient in mediating neutrophil emigration in Mac-1-deficient mice. *The Journal of clinical investigation* **1997**, *99*, 1340-1350.

[72] Coxon, A., Rieu, P., Barkalow, F. J., Askari, S., Sharpe, A. H., von Andrian, U. H., Arnaout, M. A., Mayadas, T. N., A novel role for the β2 integrin CD11b/CD18 in neutrophil apoptosis: a homeostatic mechanism in inflammation. *Immunity* **1996**, *5*, 653-666.

[73] Vetvicka, V., Thornton, B. P., Ross, G. D., Soluble beta-glucan polysaccharide binding to the lectin site of neutrophil or natural killer cell complement receptor type 3 (CD11b/CD18) generates a primed state of the receptor capable of mediating cytotoxicity of iC3b-opsonized target cells. *The Journal of clinical investigation* **1996**, *98*, 50-61.

[74] Cain, J. A., Newman, S. L., Ross, G. D., Role of complement receptor type three and serum opsonins in the neutrophil response to yeast. *Complement* **1987**, *4*, 75-86.

[75] Ross, G., Cain, J., Lachmann, P., Membrane complement receptor type three (CR3) has lectin-like properties analogous to bovine conglutinin as functions as a receptor for zymosan and rabbit erythrocytes as well as a receptor for iC3b. *The Journal of Immunology* **1985**, *134*, 3307-3315.

[76] Krych, M., Atkinson, J. P., Holers, V. M., Complement receptors. *Current opinion in immunology* **1992**, *4*, 8-13.

[77] Ross, G. D., Cain, J. A., Myones, B. L., Newman, S. L., Lachmann, P. J., Specificity of membrane complement receptor type three (CR3) for beta-glucans. *Complement* **1987**, *4*, 61-74.

[78] Ross, G. D., Regulation of the adhesion versus cytotoxic functions of the Mac-1/CR3/alphaMbeta2-integrin glycoprotein. *Critical reviews in immunology* **2000**, *20*, 197-222.

[79] Berton, G., Lowell, C. A., Integrin signalling in neutrophils and macrophages. *Cellular signalling* **1999**, *11*, 621-635.

[80] Mocsai, A., Abram, C. L., Jakus, Z., Hu, Y., Lanier, L. L., Lowell, C. A., Integrin signaling in neutrophils and macrophages uses adaptors containing immunoreceptor tyrosine-based activation motifs. *Nature immunology* **2006**, *7*, 1326-1333.

[81] O'Brien, X. M., Heflin, K. E., Lavigne, L. M., Yu, K., Kim, M., Salomon, A. R., Reichner, J. S., Lectin site ligation of CR3 induces conformational changes and signaling. *The Journal of biological chemistry* **2012**, *287*, 3337-3348.

[82] Lavigne, L. M., Albina, J.E. and Reichner, J.S., β-Glucan is a fungal determinant for adhesion-dependent human neutrophil functions. . *The Journal of Immunology* **2006**, *177*, 8667-8675.

[83] Nathan, C., Srimal, S., Farber, C., Sanchez, E., Kabbash, L., Asch, A., Gailit, J., Wright, S. D., Cytokine-induced respiratory burst of human neutrophils: dependence on extracellular matrix proteins and CD11/CD18 integrins. *The Journal of cell biology* **1989**, *109*, 1341-1349.

[84] Ehlers, M. R., CR3: a general purpose adhesion-recognition receptor essential for innate immunity. *Microbes and infection* **2000**, *2*, 289-294.

[85] Allendorf, D. J., Yan, J., Ross, G.D., Hansen, R.D., Baran, J.T., Subbarao, K., Wang, L. and Haribabu, B., C5a-mediated leukotriene B4-amplified neutrophil chemotaxis is essential in tumor immunotherapy facilitated by anti-tumor monoclonal antibody and β-glucan. *The Journal of Immunology* **2005**, *174*, 7050-7056.

[86] Kruskal, B. A., Maxfield, F. R., Cytosolic free calcium increases before and oscillates during frustrated phagocytosis in macrophages. *The Journal of cell biology* **1987**, *105*, 2685-2693.

[87] Tsikitis, V. L., Albina, J. E., Reichner, J. S., Beta-glucan affects leukocyte navigation in a complex chemotactic gradient. *Surgery* **2004**, *136*, 384-389.

[88] Monto, A. S., Epidemiology of viral respiratory infections. *The American journal of medicine* **2002**, *112*, 4-12.

[89] Heikkinen, T., Järvinen, A., The common cold. *The Lancet* **2003**, *361*, 51-59.

[90] Burt, C. W., McCaig, L. F., Rechtsteiner, E. A., Ambulatory medical care utilization estimates for 2005. **2007**.

[91] Bramley, T. J., Lerner, D., Sarnes, M., Productivity losses related to the common cold. *Journal of Occupational and Environmental Medicine* **2002**, *44*, 822-829.

[92] Darrow, L. A., Klein, M., Flanders, W. D., Mulholland, J. A., Tolbert, P. E., Strickland, M. J., Air pollution and acute respiratory infections among children 0-4 years of age: an 18-year time-series study. *American journal of epidemiology* **2014**, *180*, 968-977.

[93] Liu, L., Oza, S., Hogan, D., Perin, J., Rudan, I., Lawn, J. E., Cousens, S., Mathers, C., Black, R. E., Global, regional, and national causes of child mortality in 2000-13, with projections to inform post-2015 priorities: an updated systematic analysis. *Lancet (London, England)* **2015**, *385*, 430-440.

[94] Meng, F., Baker’s Yeast Beta-Glucan Decreases Episodes of Common Childhood Illness In 1 to 4 Year Old Children during Cold Season in China. *J Nutr Food Sci* **2016**, *6*, 2.

[95] Vetvicka, V., Richter, J., Svozil, V., Rajnohová Dobiášová, L., Král, V., Placebo-driven clinical trials of yeast-derived β-(1-3) glucan in children with chronic respiratory problems. *Ann Transl Med* **2013**, *1*, 26-26.

[96] Richter, J., Svozil, V., Kral, V., Rajnohova Dobiasova, L., Stiborova, I., Vetvicka, V., Clinical trials of yeast-derived beta-(1,3) glucan in children: effects on innate immunity. *Ann Transl Med* **2014**, *2*, 15.

[97] Vaclav, V., Josef, R., Vladimir, S., Lucie, R. D., Vlastimil, K., Placebo-driven clinical trials of transfer point Glucan #300 in children with chronic respiratory problems: antibody production. *American Journal of Immunology* **2013**, *9*, 43.

[98] Richter, J., Svozil, V., Král, V., Rajnohová Dobiášová, L., Vetvicka, V., β-glucan affects mucosal immunity in children with chronic respiratory problems under physical stress: clinical trials. *Ann Transl Med* **2015**, *3*, 52-52.

[99] McFarlin, B. K., Carpenter, K. C., Davidson, T., McFarlin, M. A., Baker's yeast beta glucan supplementation increases salivary IgA and decreases cold/flu symptomatic days after intense exercise. *Journal of dietary supplements* **2013**, *10*, 171-183.

[100] Briefel, R., Reidy, K., Karwe, V., Devaney, B., *Feeding Infants and Toddlers Study: Improvements needed in meeting infant feeding recommendations*, 2004.

[101] Calder, P. C., Feeding the immune system. *The Proceedings of the Nutrition Society* **2013**, *72*, 299-309.

[102] Smithers, L. G., Golley, R. K., Brazionis, L., Lynch, J. W., Characterizing whole diets of young children from developed countries and the association between diet and health: a systematic review. *Nutrition reviews* **2011**, *69*, 449-467.

[103] Li, F., Jin, X., Liu, B., Zhuang, W., Scalabrin, D., Follow-up formula consumption in 3-to 4-year-olds and respiratory infections: an RCT. *Pediatrics* **2014**, *133*, e1533-1540.

[104] Pontes, M. V., Ribeiro, T. C. M., Ribeiro, H., de Mattos, A. P., Almeida, I. R., Leal, V. M., Cabral, G. N., Stolz, S., Zhuang, W., Scalabrin, D. M. F., *Nutrition journal* 2016, p. 19.

[105] Ventura, M. T., Casciaro, M., Gangemi, S., Buquicchio, R., Immunosenescence in aging: between immune cells depletion and cytokines up-regulation. *Clin Mol Allergy* **2017**, *15*, 21-21.

[106] Aw, D., Silva, A. B., Palmer, D. B., Immunosenescence: emerging challenges for an ageing population. *Immunology* **2007**, *120*, 435-446.

[107] Pawelec, G., Age and immunity: What is "immunosenescence"? *Experimental gerontology* **2018**, *105*, 4-9.

[108] Fuller, R., Moore, M. V., Lewith, G., Stuart, B. L., Ormiston, R. V., Fisk, H. L., Noakes, P. S., Calder, P. C., Yeast-derived beta-1,3/1,6 glucan, upper respiratory tract infection and innate immunity in older adults. *Nutrition* **2017**, *39-40*, 30-35.

[109] Gaullier, J.-M., Sleboda, J., Ofjord, E. S., Ulvestad, E., Nurminiemi, M., Moe, C., Albrektsen, T., Gudmundsen, O., Supplementation with a soluble beta-glucan exported from Shiitake medicinal mushroom, Lentinus edodes (Berk.) singer mycelium: a crossover, placebo-controlled study in healthy elderly. *International journal of medicinal mushrooms* **2011**, *13*.

[110] M Peters, E., D Bateman, E., *Ultramarathon running and upper respiratory tract infections. An epidemiological survey*, 1983.

[111] Nieman, D. C., Johanssen, L. M., Lee, J. W., Arabatzis, K., Infectious episodes in runners before and after the Los Angeles Marathon. *The Journal of sports medicine and physical fitness* **1990**, *30*, 316-328.

[112] Robson-Ansley, P., Howatson, G., Tallent, J., Mitcheson, K., Walshe, I., Toms, C., G, D. U. T., Smith, M., Ansley, L., Prevalence of allergy and upper respiratory tract symptoms in runners of the London marathon. *Medicine and science in sports and exercise* **2012**, *44*, 999-1004.

[113] Nieman, D. C., Exercise, infection, and immunity. *International journal of sports medicine* **1994**, *15 Suppl 3*, S131-141.

[114] Walsh, N. P., Gleeson, M., Shephard, R. J., Gleeson, M., Woods, J. A., Bishop, N. C., Fleshner, M., Green, C., Pedersen, B. K., Hoffman-Goetz, L., Rogers, C. J., Northoff, H., Abbasi, A., Simon, P., Position statement. Part one: Immune function and exercise. *Exercise immunology review* **2011**, *17*, 6-63.

[115] Mackinnon, L. T., Immunity in athletes. *International journal of sports medicine* **1997**, *18 Suppl 1*, S62-68.

[116] Achten, J., Halson, S. L., Moseley, L., Rayson, M. P., Casey, A., Jeukendrup, A. E., Higher dietary carbohydrate content during intensified running training results in better maintenance of performance and mood state. *Journal of applied physiology (Bethesda, Md. : 1985)* **2004**, *96*, 1331-1340.

[117] Hassmen, P., Blomstrand, E., Mood change and marathon running: a pilot study using a Swedish version of the POMS test. *Scandinavian journal of psychology* **1991**, *32*, 225-232.

[118] Campbell, J. P., Turner, J. E., Debunking the Myth of Exercise-Induced Immune Suppression: Redefining the Impact of Exercise on Immunological Health Across the Lifespan. *Frontiers in immunology* **2018**, *9*, 648.

[119] Gleeson, M., Bishop, N. C., URI in athletes: are mucosal immunity and cytokine responses key risk factors? *Exercise and sport sciences reviews* **2013**, *41*, 148-153.

[120] Nieman, D. C., Henson, D. A., Smith, L. L., Utter, A. C., Vinci, D. M., Davis, J. M., Kaminsky, D. E., Shute, M., Cytokine changes after a marathon race. *Journal of applied physiology (Bethesda, Md. : 1985)* **2001**, *91*, 109-114.

[121] Weinstock, C., Konig, D., Harnischmacher, R., Keul, J., Berg, A., Northoff, H., Effect of exhaustive exercise stress on the cytokine response. *Medicine and science in sports and exercise* **1997**, *29*, 345-354.

[122] Carpenter, K. C., Breslin, W. L., Davidson, T., Adams, A., McFarlin, B. K., Baker's yeast beta-glucan supplementation increases monocytes and cytokines post-exercise: implications for infection risk? *Br J Nutr* **2013**, *109*, 478-486.

[123] McFarlin, B. K., Venable, A. S., Carpenter, K. C., Henning, A. L., Ogenstad, S., Oral Supplementation with Baker's Yeast Beta Glucan Is Associated with Altered Monocytes, T Cells and Cytokines following a Bout of Strenuous Exercise. *Front Physiol* **2017**, *8*, 786.

[124] Iijima, N., Mattei, L. M., Iwasaki, A., Recruited inflammatory monocytes stimulate antiviral Th1 immunity in infected tissue. *Proc Natl Acad Sci U S A* **2011**, *108*, 284-289.

[125] Mah, E., Kaden, V. N., Kelley, K. M., Liska, D. J., Beverage Containing Dispersible Yeast beta-Glucan Decreases Cold/Flu Symptomatic Days After Intense Exercise: A Randomized Controlled Trial. *Journal of dietary supplements* **2018**, 1-11.

[126] Talbott, S., Talbott, J., Effect of BETA 1, 3/1, 6 GLUCAN on upper respiratory tract infection symptoms and mood state in marathon athletes. *Journal of sports science & medicine* **2009**, *8*, 509.

[127] Musso, T., Deaglio, S., Franco, L., Calosso, L., Badolato, R., Garbarino, G., Dianzani, U., Malavasi, F., CD38 expression and functional activities are up-regulated by IFN-gamma on human monocytes and monocytic cell lines. *J Leukoc Biol* **2001**, *69*, 605-612.

[128] McFarlin, B. K., Flynn, M. G., Campbell, W. W., Stewart, L. K., Timmerman, K. L., TLR4 is lower in resistance-trained older women and related to inflammatory cytokines. *Medicine and science in sports and exercise* **2004**, *36*, 1876-1883.

[129] Kruger, K., Mooren, F. C., T cell homing and exercise. *Exercise immunology review* **2007**, *13*, 37-54.

[130] Bauchau, V., Durham, S., Prevalence and rate of diagnosis of allergic rhinitis in Europe. *European Respiratory Journal* **2004**, *24*, 758-764.

[131] Bousquet, J., Khaltaev, N., Cruz, A. A., Denburg, J., Fokkens, W., Togias, A., Zuberbier, T., Baena‐Cagnani, C., Canonica, G., Van Weel, C., Allergic rhinitis and its impact on asthma (ARIA) 2008. *Allergy* **2008**, *63*, 8-160.

[132] Abbas, A. K., Murphy, K. M., Sher, A., Functional diversity of helper T lymphocytes. *Nature* **1996**, *383*, 787.

[133] Bousquet, J., Dhivert, H., Michel, F. B., Current trends in the management of allergic diseases. *Allergy* **1994**, *49*, 31-36.

[134] Kim, H. S., Hong, J. T., Kim, Y., Han, S. B., Stimulatory Effect of beta-glucans on Immune Cells. *Immune Netw* **2011**, *11*, 191-195.

[135] Kirmaz, C., Bayrak, P., Yilmaz, O., Yuksel, H., Effects of glucan treatment on the Th1/Th2 balance in patients with allergic rhinitis: a double-blind placebo-controlled study. *European cytokine network* **2005**, *16*, 128-134.

[136] Talbott, S. M., Talbott, J. A., Talbott, T. L., Dingler, E., β-Glucan supplementation, allergy symptoms, and quality of life in self-described ragweed allergy sufferers. *Food science & nutrition* **2013**, *1*, 90-101.

[137] Graubaum, H.-J., Busch, R., Stier, H., Gruenwald, J., *Food Nutr Sci* 2012, pp. 738-746.

[138] Auinger, A., Riede, L., Bothe, G., Busch, R., Gruenwald, J., Yeast (1,3)-(1,6)-beta-glucan helps to maintain the body's defence against pathogens: a double-blind, randomized, placebo-controlled, multicentric study in healthy subjects. *European journal of nutrition* **2013**, *52*, 1913-1918.

[139] Dharsono, T., Rudnicka, K., Wilhelm, M., Schoen, C., Effects of Yeast (1, 3)-(1, 6)-Beta-Glucan on Severity of Upper Respiratory Tract Infections: A Double-Blind, Randomized, Placebo-Controlled Study in Healthy Subjects. *Journal of the American College of Nutrition* **2019**, *38*, 40-50.

[140] Feldman, S., Schwartz, H. I., Kalman, D. S., Mayers, A., Kohrman, H. M., Clemens, R., Krieger, D. R., Randomized phase II clinical trials of wellmune WGP [R] for immune support during cold and flu season. *Journal of Applied Research* **2009**, *9*, 30-43.

[141] Leentjens, J., Quintin, J., Gerretsen, J., Kox, M., Pickkers, P., Netea, M. G., The effects of orally administered beta-glucan on innate immune responses in humans, a randomized open-label intervention pilot-study. *PloS one* **2014**, *9*, e108794.

[142] Bai, J., Ren, Y., Li, Y., Fan, M., Qian, H., Wang, L., Wu, G., Zhang, H., Qi, X., Xu, M., Physiological functionalities and mechanisms of β-glucans. *Trends in Food Science & Technology* **2019**.

[143] EFSA Panel on Dietetic Products, N., Allergies, Guidance on the scientific requirements for health claims related to the immune system, the gastrointestinal tract and defence against pathogenic microorganisms. *EFSA Journal* **2016**, *14*, 4369.

[144] EFSA Panel on Dietetic Products, N., Allergies, Scientific Opinion on the safety of ‘yeast beta‐glucans’ as a Novel Food ingredient. *EFSA Journal* **2011**, *9*, 2137.

[145] Minihane, A. M., Vinoy, S., Russell, W. R., Baka, A., Roche, H. M., Tuohy, K. M., Teeling, J. L., Blaak, E. E., Fenech, M., Vauzour, D., McArdle, H. J., Kremer, B. H., Sterkman, L., Vafeiadou, K., Benedetti, M. M., Williams, C. M., Calder, P. C., Low-grade inflammation, diet composition and health: current research evidence and its translation. *Br J Nutr* **2015**, *114*, 999-1012.

[146] Furman, D., Davis, M. M., New approaches to understanding the immune response to vaccination and infection. *Vaccine* **2015**, *33*, 5271-5281.

[147] Manzoni, C., Kia, D. A., Vandrovcova, J., Hardy, J., Wood, N. W., Lewis, P. A., Ferrari, R., Genome, transcriptome and proteome: the rise of omics data and their integration in biomedical sciences. *Briefings in Bioinformatics* **2016**, *19*, 286-302.

[148] Zeevi, D., Korem, T., Zmora, N., Israeli, D., Rothschild, D., Weinberger, A., Ben-Yacov, O., Lador, D., Avnit-Sagi, T., Lotan-Pompan, M., Suez, J., Mahdi, J. A., Matot, E., Malka, G., Kosower, N., Rein, M., Zilberman-Schapira, G., Dohnalova, L., Pevsner-Fischer, M., Bikovsky, R., Halpern, Z., Elinav, E., Segal, E., Personalized Nutrition by Prediction of Glycemic Responses. *Cell* **2015**, *163*, 1079-1094.

**Figure legends**

**Figure 1. β-glucan Structure-Activity Relationship.** Variability inβ-glucan structure is due to differences in source and extraction and/or purification methods, which likely explains the divergent functionalities that exist among β-glucans. Depending on the source, differences arise such as the nature of molecular linkages and the degree of branching, as well as variability in mass, charge, solubility and configuration in solution (single helix, triple helix or random coil), as well as in impurity levels and content. These variabilities will result in different interactions with the host. Bacterial β-glucans represent the most basic form of the polysaccharide with a linear β-1,3 structure; cereal β-glucans follow the same pattern with dominant β-1,4 stretches; fungal (e.g. mushroom) and yeast (i.e. single cell fungi) β-glucans have frequent β-1,3-D-glucose side chains at β-1,6 branching points that are short and spaced in fungal species (e.g. mushroom) and longer in yeast species. Further variations to these general structures are common. Only highly purified β-1,3-1,6-glucans with a high degree of branching along the β-1,3-glucan backbone and a high molecular weight are able to exert immunomodulatory properties.

**Figure 2. Immune Response Overview.** The immune system has the ability to recognize and eliminate pathogens by first activating the innate response, which, without prior antigen exposure, acts in a fast and un-targeted manner to phagocytose and kill the invader. APCs use PRRs to recognize pathogens directly (e.g. β-glucan recognition by dectin-1) or indirectly (e.g. β-glucan recognition by CR3 by binding to iC3b opsonized β-glucan). After PRR-pathogen binding, APCs proceed with an antimicrobial response followed by an inflammatory response with a complex intracellular signaling cascade that culminates in the activation of transcription factors to produce inflammatory mediators and antigen presentation aided by MHC Class I and II molecules.

**Figure 3. Dectin-1 Structure and Signalling.** Murine and human dectin-1 consist of an extracellular C-type lectin domain (which recognizes β-1,3-glucans) joined to a short stalk that continues into a single transmembrane domain followed by an intracellular ITAM-like motif signaling domain (hemITAM). Upon whole glucan particle (WGP) attachment and Src phosphorylation, CD45 and CD148 are isolated from the contact site of dectin-1-WGP to allow hemITAM signaling. The downstream responses include 1) a rapid antimicrobial response lead by ligand-uptake, phagocytosis and oxidative burst, and 2) a consequent pro-inflammatory response that is mediated by the production of cytokines and chemokines as a result of gene transcription modulation, together with antigen presentation to T and B cells to further continue with adaptive cell differentiation to the specific antigen. The binding to both dectin stalks is necessary to induce downstream Src activation and Syk recruiting.

**Figure 4. CR3 Signalling.** Dual ligation of the α-chain (CD11b) by complement iC3b and soluble glucan (SG) is needed to activate CR3. iC3b coats the antigen and attaches to the I-domain, while SG, derived from the fragmentation of its parent β-glucan by phagocytes, attaches to the lectin-1 domain.

**Figure 5. Oxidative Burst.** β-glucan primed immune cells produce reactive oxygen species within the lysosome by activating NADPH oxidase; however, the activation of NADPH oxidase is achieved via different molecular signalling according to the form of β-glucan, i.e. WGP, SG, or focal adhered β-glucan.