**Natural products in drug discovery: advances and opportunities**

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**Abstract** | Natural products and their structural analogues have historically made a major contribution to pharmacotherapy, especially for cancer and infectious diseases. Nevertheless, natural products also present challenges for drug discovery, such as technical barriers for screening, isolation, characterization and optimization, which contributed to a decline in their pursuit by the pharmaceutical industry from the 1990s onwards. In recent years, several technological and scientific developments — including improved analytical tools, genome mining and engineering strategies, and microbial culturing advances — are addressing such challenges and opening up new opportunities. Consequently, interest in natural products as drug leads is being revitalized, particularly for tackling antimicrobial resistance. Here, we summarize recent technological developments that are enabling natural product-based drug discovery, highlight selected applications and discuss key opportunities.

**[H1] Introduction**

Historically, natural products (NPs) have played a key role in drug discovery, especially for cancer and infectious diseases,1,2 but also in other therapeutic areas, including cardiovascular diseases (for example, statins) and multiple sclerosis (for example, fingolimod)3–5.

NPs offer special features in comparison to conventional synthetic molecules, which confer both advantages and challenges for the drug discovery process. NPs are characterized by an enormous scaffold diversity and structural complexity. They typically have a higher molecular mass, a larger number of sp3 carbon atoms **[G]** and oxygen atoms but fewer nitrogen and halogen atoms, higher numbers of H-bond acceptors and donors, lower calculated octanol–water partition coefficients (cLogP values, indicating higher hydrophilicity) and greater molecular rigidity compared to synthetic compound libraries1,6–9. These differences can be advantageous; for example, the higher rigidity of NPs can be valuable in drug discovery tackling protein–protein interactions10. Indeed, NPs are a major source of oral drugs "beyond Lipinski's rule of five **[G]**."11 The increasing significance of drugs not conforming to this rule is illustrated by the increase in molecular masses of approved oral drugs over the last 20 years12. NPs are structurally ‘optimized’ by evolution to serve particular biological functions1, including the regulation of endogenous defence mechanisms and the interaction (often competition) with other organisms, which explains their high relevance for infectious diseases and cancer. Furthermore, their use in traditional medicine may provide insights regarding efficacy and safety. Overall, the NP pool is enriched with ‘bioactive’ compounds covering a wider area of the chemical space compared with typical synthetic small-molecule libraries13.

Despite these advantages and multiple successful drug discovery examples, several drawbacks of NPs have led pharmaceutical companies to reduce NP-based drug discovery programs. NP screens typically involve a library of extracts from natural sources (**Figure 1**), which may not be compatible with traditional target-based assays14.Identifying the bioactive compound(s) of interest can be challenging, and dereplication **[G]** tools have to be applied to avoid re-discovery of known compounds. Accessing sufficient biological material to isolate and characterize a bioactive NP may also be challenging15. Furthermore, gaining intellectual property (IP) rights for (unmodified) NPs exhibiting relevant bioactivities can be a hurdle, since naturally occurring compounds in their original form may not always be patented (legal frameworks vary among countries and are evolving16), although simple derivatives can be patent-protected (see **Box 1** for examples). An additional layer of complexity relates to the regulations defining the need for benefit sharing with countries of origin of the biological material, framed in the United Nations 1992 Convention on Biological Diversity and the Nagoya Protocol, which entered into force in 201417, as well as recent developments concerning benefit sharing linked to utilization of marine genetic resources18.

Although the complexity of NP structures can be advantageous, the generation of structural analogues to explore structure–activity relationships and to optimize NP leads can be challenging, particularly if synthetic routes are difficult. Also, NP-based drug leads are often identified by phenotypic assays **[G]**, and deconvolution of their molecular mechanisms of action can be time-consuming19. Fortunately, there have been substantial advances20 both in the development of screening assays (for example, harnessing the potential of induced pluripotent stem cells and gene editing technologies) and in strategies to identify the modes of action of active compounds (see REFS. 21–23 for reviews).

Here, we discuss recent technological and scientific advances that may help to overcome challenges in NP-based drug discovery, with an emphasis on three areas: analytical techniques, genome mining and engineering, and cultivation systems. In the concluding section, we highlight promising future directions for NP drug discovery.

**[H1] Application of analytical techniques**

Classical NP-based drug research starts with biological screening of ‘crude’ extracts to identify a bioactive ‘hit’ extract, which is further fractionated to isolate the active NP(s). Bioactivity-guided isolation is a laborious process with a number of limitations, but various strategies and technologies can be used to address some of them (**Figure 2**). For example, to create libraries that are compatible with high-throughput screening, crude extracts can be pre-fractionated into sub-fractions that are more suitable for automated liquid handling systems. In addition, fractionation methods can be adjusted such that sub-fractions preferentially contain compounds with drug-like properties (typically moderate hydrophilicity). Such approaches can increase the number of hits compared to using crude extracts, as well as enabling more efficient follow-up of promising hits24.

Metabolomics was developed as an approach to simultaneously analyse multiple metabolites in biological samples. Enabled by technological developments in chromatography and spectrometry, metabolomics was historically applied first in other research fields, such as biomedical and agricultural sciences2. Advances in the analytical instrumentation used in NP research25,26, coupled with computational approaches that can generate plausible NP analogue structures and their respective simulated spectra27, has also enabled application of ‘omics’ approaches such as metabolomics in NP-based drug discovery. Metabolomics can provide accurate information on the metabolite composition in NP extracts, thus helping to prioritize NPs for isolation, to accelerate dereplication28,29, and to annotate unknown analogues and new NP scaffolds. Moreover, metabolomics can detect differences between metabolite compositions in various physiological states of producing organisms and enable the generation of hypotheses to explain them, and can also provide extensive metabolite profiles to underpin phenotypic characterization at the molecular level30. Both options are very useful in understanding the molecular mechanisms of action of NPs.

For metabolite profiling, NP extracts are analysed by nuclear magnetic resonance (NMR) spectroscopy or high-resolution mass spectrometry (HRMS), or respective hyphenated methods involving upstream liquid chromatography (LC)31,32, such as LC-HRMS, which can separate numerous isomers present in NP extracts33. Moreover, such hyphenated methods might integrate both HRMS and NMR, allowing the simultaneous use of the advantages of both techiques34,35. NMR analysis of NP extracts is simple and reproducible, and provides direct quantitative information and detailed structural information, although it has relatively low sensitivity, meaning that it generally only enables profiling of major constituents32. The applications of NMR in NP research are versatile37 and the technique is used both directly for metabolomics of unfractionated NP extracts as well as for structural characterization of compounds and fractions obtained with appropriate separation methods, most often LC. HRMS is the gold standard for qualitative and quantitative metabolite profiling33, and is most commonly applied in combination with LC. HRMS can also be used in the direct infusion mode (DIMS)38, where samples are directly profiled by MS without a chromatography step, or in MS imaging (MSI)36, which enables determination of spatial distribution of NPs within living organisms. HRMS enables routine acquisition of accurate molecular mass information, which together with appropriate heuristic filtering, can provide unambiguous molecular formulae assignment for hundreds to thousands of metabolites within a single extract over a dynamic range that may exceed 5 orders of magnitude31,39. However, challenges remain in data mining and in the unambiguous identification of the metabolites using various workflows relying on open web-based tools40.

Dereplication of secondary metabolites in bioactive extracts includes the determination of molecular mass and formula and cross-searching in the literature or structural NP databases with taxonomic information, which greatly assists the identification process. Such metadata, which are difficult to query in the literature, are often compiled in proprietary databases, such as the Dictionary of Natural Products (DNP), which encompasses all NP structures reported with links to their biological sources (see **Dictionary of Natural Products** in Related links). However, a comprehensive experimental MS/MS database of all NPs reported to date does not exist and a search for experimental spectra across various platforms is hindered by the lack of standardised collision energy conditions for fragmentation in LC-MS/MS25.

In this respect, the Global Natural Products Social (GNPS) molecular networking platform developed in the Dorrestein laboratory is an important addition to the toolbox41. The molecular networking organizes thousands of sets of MS/MS data recorded from a given set of extracts and visualizes the relationship of the analytes as clusters of structurally related molecules. This improves the efficiency of dereplication by enabling annotation of isomers and analogues of a given metabolite in a cluster42. The recorded experimental spectra can be searched against putative structures and their corresponding predicted MS/MS spectra generated by tools such as the competitive fragmentation modeling (CFM-ID)43. Based on such approaches, vast databases of theoretical NP spectra have been created and applied in dereplication44. The GNPS molecular networking approach has limitations, however, such as better applicability to some classes of NPs compared to others and the uncertainty of structural assignment among possible predicted candidates. Efforts to address such issues are ongoing45,46,47, including overlaying molecular networks of large NP extract libraries with taxonomic information to improve the confidence of annotation48. Overall, molecular networking mainly allows better prioritization of the isolation of unknown compounds by strengthening the dereplication process and elucidating relationships between NP analogues, and rigorous structure elucidation for NPs of interest should not be neglected.

Another useful platform for metabolite identification is METLIN49, which includes a high-resolution tandem mass spectrometry (MS/MS) database with a fragment similarity search function that is useful for identification of unknown compounds. Other databases and *in silico* tools such as Compound Structure Identification (CSI): FingerID and Input Output Kernel Regression (IOKR) can be used to search available fragment ion spectra, as well as generate predicted spectra of fragment ions not present in current databases50. A novel computational platform for predicting the structural identity of metabolites derived from any identified compound has also been recently reported51, which should increase the searchable chemical space of NPs.

To accelerate the identification of bioactive NPs in extracts, metabolomics data can be matched to the biological activities of these extracts52. Various chemometric methods such as multivariate data analysis can correlate the measured activity with signals in the NMR and MS spectra, enabling the active compound(s) to be traced in complex mixtures with no need for further bioassays53,54,55. Furthermore, several analytical modules involving different bioassays and detection technologies can be linked to allow simultaneous bioactivity evaluation and identification of compounds present in small amounts (analytical scale) in complex compound mixtures34,35.

Metabolomics data can be integrated with data obtained by other ‘omics’ techniques such as transcriptomics and proteomics and/or with imaging-based screens. For example, Acharya *et al.* used this approach to characterize NP-mediated interactions between a *Micromonospora* species and a *Rhodococcus* species56. In another interesting example, Kurita *et al.* developed a compound activity mapping (CAM) platform for the prediction of identities and mechanisms of action of constituents from complex NP extract libraries by integrating cytological profiling57 with untargeted metabolomics data from a library of extracts58, and identified quinocinnolinomycins as a new family of NPs causing endoplasmic reticulum stress58 (FIG. 2a).

Analytical advances that enable the profiling of responses to bioactive molecules at the single-cell level can also accelerate NP-based drug discovery. Irish, Bachmann, and colleagues developed a high-throughput platform for metabolomic profiling of bioactivity by integrating phospho-specific flow cytometry, single-cell chemical biology and cellular barcoding with metabolomic arrays (characterized chromatographic microtiter arrays originating from biological extracts)59. Using this platform, the authors studied the single-cell responses of bone marrow biopsy samples from patients with acute myeloid leukemia following exposure to microbial metabolomic arrays obtained from extracts of biosynthetically prolific bacteria, which enabled the identification of new bioactive polyketides59.

Finally, advances in analytical technologies continue to support the rigorous structure determination of NPs of interest. The progressive development of higher-field NMR instruments and probe technology60,61 has enabled NP structure determination from very small quantities (below 10 µg)62,63, which is important as the available quantities of NPs are often limited. In addition, microcrystal electron diffraction (MicroED) has recently emerged as a cryo-electron microscopy-based technique for unambiguous structure determination of small molecules64, and is already finding important applications in NP research65. The increased resolution and sensitivity of analytical equipment can also help address problems associated with “residual complexity” of isolated NPs; that is when biologically potent but unidentified impurities in an isolated NP sample (which could include structurally related metabolites or conformers) lead to an incorrect assignment of structure and/or activity66,67. To avoid futile downstream development efforts, Pauli et al. recommended that lead NPs should undergo advanced purity analysis at an early stage using quantitative NMR and LC–MS67.

**[H1] Genome mining and engineering**

Advances in knowledge on biosynthetic pathways for NPs and in developing tools for analysing and manipulating genomes are further key drivers for modern NP-based drug discovery. Two key characteristics enable the identification of biosynthetic genes in the genomes of the producing organisms. First, these genes are clustered in the genomes of bacteria and filamentous fungi. Second, many NPs are based on polyketide or peptide cores, and their biosynthetic pathways involve enzymes — polyketide synthases and non-ribosomal peptide synthetases, respectively — that are encoded by large genes with highly conserved modules68.

“Genome mining” is based on searches for genes that are likely to govern biosynthesis of scaffold structures, and can be used to identify NP biosynthetic gene clusters (for examples, see refs 69–71). Prioritizing gene clusters for further work is facilitated by advances in biosynthetic knowledge and predictive bioinformatics tools, which can provide hints about whether the metabolic products of the clusters have chemical scaffolds that are new or known, thereby supporting dereplication72,73. Such predictive tools for gene cluster analysis can be applied in combination with spectroscopic techniques to accelerate the identification of NPs65 and determine the stereochemistry of metabolic products66. Furthermore, to extend genome mining from a single genome to entire genera, microbiomes or strain collections, computational tools have been developed, such as BiG-SCAPE, which enables sequence similarity analysis of biosynthetic gene clusters, and CORASON, which uses a phylogenomic approach **[G]** to elucidate evolutionary relationships between gene clusters76.

Phylogenetic studies of known groups of talented secondary metabolite producers can also empower discovery of novel NPs. Recently, a study comparing secondary metabolite profiles and phylogenetic data in myxobacteria demonstrated a correlation between the taxonomic distance **[G]** and the production of distinct secondary metabolite families77. In filamentous fungi, it was likewise shown that secondary metabolite profiles are closely correlated to their phylogeny78. These organisms are rich in secondary metabolites, as demonstrated by LC-MS studies of their extracts under laboratory conditions79. Concurrent genomic and phylogenomic analyses implied that even the genomes of well-studied organism groups harbor many gene clusters for secondary metabolite biosynthesis with as yet unknown functions80. The phylogeny of biosynthetic gene clusters, together with analysis of the absence of known resistance determinants, was recently used to prioritize members of the glycopeptide antibiotic family that could have novel activities. This led to the identification of the known antibiotic complestatin and the newly discovered corbomycin as compounds that act through a previously uncharacterized mechanism involving inhibition of peptidoglycan remodelling81.

Many microorganisms cannot be cultured or tools for their genetic manipulation are not sufficiently developed, which makes it more challenging to access their NP-producing potential. However, biosynthetic gene clusters for NPs can be cloned and heterologously expressed in organisms that are well-characterized and easier to culture and to genetically manipulate (such as *Streptomyces coelicolor*, *Escherichia coli* and *Saccharomyces cerevisiae*)82. The aim is to achieve higher production titers in the heterologous hosts as compared to wild-type strains, improving the availability of lead compounds82–84. Vectors that can carry large DNA inserts are needed for the cloning of complete NP biosynthetic gene clusters. Cosmids (which can have inserts of 30–40 kb), fosmids (which can harbor 40–50 kb) and bacterial artificial chromosomes (BACs; which can have inserts of 100 kb to >300 kb) have been developed85. For fungal gene clusters, self-replicating fungal artificial chromosomes (FACs) have been developed, which can have inserts of >100 kb86. FACs in combination with metabolomic scoring were used to develop a scalable platform, FAC-MS, allowing the characterization of fungal biosynthetic gene clusters and their respective NPs at unprecedented scale87. The application of FAC-MS for the screening of 56 biosynthetic gene clusters from different fungal species yielded the discovery of 15 new metabolites, including a new macrolactone, valactamide A87 (FIG. 2b).

Even in culturable microorganisms, many biosynthetic gene clusters may not be expressed under conventional culture conditions, and these silent clusters could represent a large untapped source of NPs with drug-like properties88. Several approaches can be pursued to identify such NPs. One approach is sequencing, bioinformatic analysis and heterologous expression of silent biosynthetic gene clusters, which has already led to the discovery of several new NP scaffolds from cultivable strains89. Direct cloning and heterologous expression was also used to discover the new antibiotic taromycin A, which was identified upon the transfer of a silent 67 kb nonribosomal peptide synthetase biosynthetic gene cluster from *Saccharomonospora sp.* CNQ-490 into *Streptomyces coelicolor*90. To transfer a biosynthetic gene cluster of such size, a platform based on transformation-associated recombination (TAR) cloning was developed. This platform enables direct cloning and manipulation of large biosynthetic gene clusters in *Saccharomyces cerevisiae*, maintenance and manipulation of the vector in *E. coli*, and heterologous expression of the cloned gene clusters in actinobacteria (such as *Streptomyces coelicolor*) following chromosomal integration90, and is an alternative to BACs for heterologous expression of large biosynthetic gene clusters.

Heterologous expression has limitations, such as the need to clone and manipulate very large genome regions occupied by biosynthetic gene clusters and the difficulty of identifying a suitable host that provides all conditions necessary for the production of the corresponding NPs. These limitations can be circumvented by activating biosynthetic gene clusters directly in the native microorganism through targeted genetic manipulations, generally involving the insertion of activating regulatory elements or deletion of inhibitory elements such as repressors or their binding sites. For example, a de-repression strategy of deleting *gbnR*, a gene for transcriptional repressor in *Streptomyces venezuelae* ATCC 10712 was used by Sidda *et al.* in the discovery of gaburedins, a family of γ-aminobutyrate-derived ureas91. An example of the activator-based strategy is the constitutive expression of the *samR0484* gene in *Streptomyces ambofaciens* ATCC 23877, which led to the discovery of stambomycins A-D, 51-membered cytotoxic glycosylated macrolides72. Alternatively, silent biosynthetic gene clusters can be activated using repressor decoys92, which have the same DNA nucleotide sequence as the binding sites for the repressors that prevent the expression of the clusters. When these decoys are introduced into the bacteria, they sequester the respective repressors and the “endogenous” binding sites in the genome remain unoccupied, leading to de-repression of the previously silent biosynthetic genes and production of the corresponding NPs. This approach has been applied to activate eight silent biosynthetic gene clusters in multiple streptomycetes, and led to the characterization of a novel NP, oxazolepoxidomycin A92. The repressor decoy strategy is simpler, easier, and faster to perform than the deletion of genes encoding regulatory factors. However, it has the same limitation as other approaches that rely on the introduction of recombinant DNA molecules in cells: it is necessary to develop protocols for efficient introduction of DNA into the targeted host strain, and the decoy must be maintained on a high-copy plasmid to ensure efficient repressor sequestration.

Another approach focused on exchange of regulatory elements is based on the CRISPR-Cas9 technology. The promise of this technique is exemplified in a recent work by Zhang *et al.*, which demonstrated that CRISPR-Cas9-mediated targeted promoter introduction can efficiently activate diverse biosynthetic gene clusters in multiple *Streptomyces* species, leading to the production of unique metabolites, including a novel polyketide in *Streptomyces viridochromogenes*93. The CRISPR-Cas9 technology was also used to knockout genes encoding two well-known and frequently rediscovered antibiotics in several actinomycete strains, which led to the production of different rare and previously unknown variants of antibiotics that were otherwise obscured, including amicetin, thiolactomycin, phenanthroviridin, and 5-chloro-3-formylindole94.

Approaches relying on sequencing, bioinformatics, and heterologous expression can also enable the identification of novel NPs from bacterial strains that have not yet been cultivated (**FIG. 3a**). For example, Hover *et al.* searched the metagenomes of 2,000 soil samples for biosynthetic gene clusters for lipopeptides with calcium-binding motifs. This led to the discovery of malacidins, members of the calcium-dependent antibiotic family, via heterologous expression of a 72-kb biosynthetic gene cluster from a desert soil sample in a *Streptomyces albus* host strain95 (**FIG. 3b**). However, in comparison to some of the other above-discussed strategies72,91,92, this metagenomic-based discovery approach is more suited to finding new members of known NP classes rather than discovery of entirely new classes. In another study, Chu *et al.* developed a human microbiome-based approach that identified nonribosomal linear heptapeptides called humimycins as novel antibiotics active against methicillin-resistant *Staphylococcus aureus* (MRSA)96 (**FIG. 3c**). The structure of the NPs was predicted via bioinformatic analysis of gene clusters found in human commensal bacteria, followed by their chemical synthesis. A major strength of this innovative approach is that it is entirely independent of microbial cultivation and heterologous gene expression. Nevertheless, there are limitations related to the accuracy of computational chemical structure predictions and the feasibility of total chemical synthesis if structures are complex.

The genomes of plants or animals can also be mined for novel NPs. For example, mining of 116 plant genomes enabled by identification of a precursor gene for the biosynthesis of lyciumins, a class of branched cyclic ribosomal peptides with hypotensive action produced by *Lycium barbarum* (popularly known as goji), identified diverse novel lyciumin chemotypes in 7 other plants, including crops such as soybean, beet, quinoa, and eggplant97. Genome mining in the animal kingdom is exemplified by the work of Dutertre *et al.*, which used an integrated transcriptomics and proteomics approach to discover thousands of novel venom peptides from *Conus marmoreus* snails98. Proteomics analysis revealed that the vast majority of the conopeptide diversity was derived from a set of ~100 genes through variable peptide processing98.

Some bioactive compounds initially isolated from marine organisms might be products of symbionts, and genome mining can facilitate the characterization of such NPs. For example, it has been shown that bioactive compounds from the sponge *Theonella swinhoei* are produced by bacterial symbionts99, and characterization of the symbiont “*Candidatus* Entotheonella serta” using single-cell genomics led to the discovery of gene clusters for misakinolide and theonellamide biosynthesis100. Another example of a marine NP produced by a bacterial symbiont is ET-743 (trabectedin), originally isolated from the tunicate *Ecteinascidia turbinate. A* meta-omics approach developed by Rath *et al.* revealed that the producer of this clinically used anticancer agent is the bacterial symbiont *Candidatus Endoecteinascidia frumentensis*101.

Similarly, plant microbiomes also represent a large reservoir for the identification of novel bioactive NPs (such as the antitumor agents maytansine, taxol, and camptothecin, which were initially isolated from plants and later shown to be produced by microbial endophytes)102 that can be tapped by genome mining approaches. An illustrative example is a recent work by Helfrich *et al.*, which identified hundreds of novel biosynthetic gene clusters by genome mining of 224 bacterial strains isolated from *Arabidopsis thaliana* leaves103. A combination of bioactivity screening and imaging mass spectrometry was used to select a single species for further genomic analysis and led to the isolation of a NP with an unprecedented structure, the *trans*-acyltransferase polyketide synthase-derived antibiotic macrobrevin103.

Targeted genetic engineering of NP biosynthetic gene clusters can be of a high value, if the producing organism is difficult to cultivate or the yield of a NP is too low to allow comprehensive NP characterization. Rational genetic engineering and heterologous expression contributed to increase the production of vioprolides, a depsipeptide class of anticancer and antifungal NPs in the myxobacterium *Cystobacter violaceus* Cb vi35, by several orders of magnitude. In addition, non-natural vioprolide analogues were generated by this approach104. Similarly, promoter engineering and heterologous expression of biosynthetic gene clusters was reported to result in a 7-fold increase in the production of the cytotoxic NP disorazol105, and a 328-fold increase in the production of spinosad, an insecticidal macrolide produced by the bacterium *Saccharopolyspora spinosa*106.

Besides increasing NP yields, targeted gene manipulation can also be used to alter biosynthetic pathways in a predictable manner to produce new NP analogues with improved pharmacological properties, such as higher specific activity, lower toxicity, and better pharmacokinetics. Such biosynthetic engineering approaches depend on a solid understanding of the biosynthetic pathway leading to a specific NP, access to the genes specifying this pathway, and the ability to manipulate them either in the original or a heterologous host. Recent advances in biosynthetic engineering have enabled faster and more efficient production of NP analogues, including the development of methods for accelerated engineering and recombination of modules of polyketide synthase (PKS) gene clusters107, non-ribosomal peptide synthetases (NRPSs)108,109, and NRPS-PKS assembly lines110, as well as elucidation of mechanisms for polyketide chain release that are contributing to NP structural diversification111,112. Examples of biosynthetic engineering applied to several important NPs include the generation of analogues of the immunosuppressant rapamycin113, the antitumor agents mithramycin114 and bleomycin115, and the antifungal nystatin116.

It should be noted that biosynthetic engineering has limitations regarding the parts of the NP molecule that can be targeted for modifications, and the chemical groups that can be introduced or removed. Considering the complexity of many NPs, however, total synthesis may be prohibitively costly, and a combined approach of biosynthetic engineering and chemical modification can provide a viable alternative for identifying improved drug candidates. For example, biosynthetic engineering may create a “handle” for addition of a beneficial chemical group by synthetic chemistry, as demonstrated for the biosynthetically engineered analogues of nystatin mentioned above; further synthetic chemistry modifications resulted in compounds with improved *in vivo* pharmacotherapeutic characteristics compared to amphotericin B117,118.

**[H1] Advances in microbial culturing systems**

The complex regulation of NP biosynthesis in response to the environment means that the conditions under which producing organisms are cultivated can have a major impact on the chance to identify novel NPs89. Several strategies have been developed to improve the likelihood of identifying novel NPs compared with monoculture under standard laboratory conditions and to make ”uncultured” microorganisms grow in a simulated natural environment (**Figure 4**)119.

One well-established approach to promote the identification of novel NPs is the modulation of culture conditions such as temperature, pH and nutrient sources. This strategy may lead to activation of silent gene clusters, thereby promoting production of different NPs. The term “One Strain Many Compounds" (OSMAC) was coined for this approach about 20 years ago120, but the concept has a longer history121, with its use being routine in industrial microbiology since the 1960s122.

While OSMAC is still widely used for the identification of new bioactive compounds (see REFS 123,124 for recent examples), this approach has limited capacity to mimic the complexities of the natural habitats. It is difficult to predict the combination of cues (which might also involve metabolites secreted by other members of the microbial community) to which the microorganism has evolved to respond by switching metabolic programs. To account for such kind of interactions, co-culturing using "helper" strains can be applied125. This can enable the production and identification of new NPs, as illustrated by recent studies in which particular fungi were co-cultured with *Streptomcyes* species126,127.

Study of the molecular mechanisms underlying the ability of "helper" strains to increase the cultivability of previously uncultured microbes can lead to the identification of specific growth factors, allowing expansion of the number of species that can be successfully cultured. This strategy was used by D'Onofrio *et al.* for the identification of new acyl-desferrioxamine siderophores (iron-chelating compounds) as growth factors produced by "helper" strains promoting the growth of previously uncultured isolates from marine sediment biofilm119,128. The siderophore-assisted growth is based on the property of these compounds to provide iron for microbes unable to autonomously produce siderophores themselves, and the application of this approach led to the isolation of previously uncultivated microorganisms128. The development of strategies to cultivate microbial symbionts that produce NPs only upon interaction with their hosts can promote access to new NPs. Microbial symbionts interacting with insects or other organisms are a highly promising reservoir for the discovery of novel bioactive NPs produced in a unique ecological context 129–132. To stimulate NP production, culturing strategies can be developed that better mimic the native environment of microbial symbionts of insects, including the use of media either containing lyophilized dead insects133 or L-proline, a major constituent of insect hemolymph134.

Strategies to mimic the natural environment even more closely by harnessing *in situ* incubation in the environment from which the microorganism is sampled have been developed, dating back to more than 20 years ago with the biotech companies OneCell and Diversa. They developed platforms that allowed the growth of some previously uncultivated microbes from different environments based on diluting out and suspension in a single drop of medium122,135. More recently, such strategies have been highlighted by the development and application of a platform dubbed the iChip, in which diluted soil samples are seeded in multiple small chambers separated from the environment with a semipermeable membrane136. After seeding, the iChip is placed back into the soil from which the sample was taken for an *in situ* incubation period, allowing the cultured microorganisms to be exposed to influences from their native environment. The power of this culturing approach was demonstrated by the discovery of a new antibiotic teixobactin produced by a previously uncultured soil bacterium137,138 **(FIG. 4a)**. This platform may be of a great significance for NP drug discovery, given that it has been estimated that only 1% of soil organisms have so far been successfully cultured by using traditional culturing techniques139.

The “omics” strategies discussed in previous sections can complement efforts to explore NPs produced upon microbial interactions. The application of such strategy is illustrated in the work of Derewacz *et al.*, who analyzed the metabolome of a genome-sequenced *Nocardiopsis* bacterium upon co-culture with bacteria of the genera *Escherichia*, *Bacillus*, *Tsukamurella*, and *Rhodococcus*140. Around 14% of the metabolomic features found in co-cultures were undetectable in monocultures, with many of those being unique to specific co-culture genera, and the previously unreported polyketides ciromicin A and B, which possess an unusual pyrrolidinol substructure and displayed moderate and selective cytotoxicity, were identified140. Other examples include a “culturomics” approach combining multiple culture conditions with MS profiling and 16S rRNA-based taxonomy to identify prokaryotic species from the human gut141, and an ultrahigh-throughput screening platform based on microfluidic droplet single-cell encapsulation and cultivation followed by next-generation sequencing and LC-MS, which allows investigation of pairwise interactions between target microorganisms142. The latter approach enabled identification of a slow-growing oral microbiota species inhibiting the growth of Staphylococcus aureus142.

Historically early-adopted microbial culturing approaches led to a bias reflected in the predominant discovery of NPs from microorganisms that are easy to cultivate (such as Streptomycetes and some common filamentous fungi). As a result, a vast number of NPs from such “easy to culture” microbes are already characterised, and conventional screening efforts tend to yield disappointing returns associated with frequent re-discovery of known NPs and their closely related congeners. Therefore, culturing strategies aimed at previously unexplored (or under-investigated) microbial groups, with the potential to produce NPs with entirely new scaffolds and bioactivities (such as *Burkholderia*, *Clostridium* and *Xenorhabdus*) are of high interest143,144. Closthioamide, the first secondary metabolite from a strictly anaerobic bacterium, was discovered from *Clostridium cellulolyticum* by this approach145. Targeted isolation of such species is important, and a genome-guided approach to achieve this goals has recently been demonstrated for *Burkholderia* strains in environmental samples146. Another highly innovative approach to the isolation and cultivation of previously uncultured bacteria was recently reported by Cross *et al.*147, who used genomic information to engineer antibodies predicted to target selected microorganisms and to specifically capture these microorganisms from complex communities and to isolate them in pure cultures. This approach was validated by isolation and cultivation of previously uncultured bacteria from the human oral cavity147 (**FIG. 4b**), and it could be applicable to a wide range of target organisms if suitable cultivation conditions can be identified for the isolated cells.

Despite these advances in culturing strategies, artificial conditions still do not fully represent the complex environment of natural habitats. To circumvent this problem, microbial and NP diversity can also be accessed via extraction of organisms and/or their NPs *in situ*. To directly gain compounds produced in the natural marine environment (which may be missed otherwise), resin capture technology can be used to capture compounds on inert sorbent supports ready to be desorbed, analysed and tested for biological activity148. Sustainable approaches for *in situ* extraction with green solvents, such as glycerol or natural deep eutectic and ionic solvents (NADES), could be used directly during field work149,150. To improve dereplication, analytical equipment miniaturization is also facilitating *in situ* analysis; examples include the introduction of devices for physicochemical data analysis, such as micro MS and portable near infrared (NIR) spectroscopy151,152.

**[H1] Outlook for NPs in drug discovery**

The technological advances discussed above have the potential to reinvigorate NP-based drug discovery in both established and emerging areas. NPs have long been the key source of new drugs against infectious diseases, especially antibiotics (see REFS 153,154 for reviews). Selected NPs with antimicrobial properties discovered by leveraging advances discussed in the sections above, including strategies to exploit the human microbiome for novel NPs96,155 are highlighted in **Figure 3** and **Figure 4**. Along with the search for new NPs with antimicrobial activities, researchers are continuing to develop and optimize already known NP classes, making use of advances in biosynthetic engineering (for example, REF. 156), total synthesis (for example, REF. 157) or semi-synthetic strategies (for example, REF. 158,159). In addition, antivirulence strategies could represent an alternative approach to fighting infections,160 for which NPs targeting bacterial quorum sensing could be of interest (for example, REF. 161).

NPs also have a successful history as cancer therapeutics, which has been well-covered in other reviews162–165. An important new opportunity in this field is the capacity of some NPs to trigger selective yet potent host immune reaction against cancer cells, particularly given the intense interest at present in strategies that could improve response rates to immune checkpoint inhibitors by turning “cold” tumors “hot”166. For example, NPs such as cardiac glycosides167 can increase the immunogenicity of stressed and dying cancer cells by triggering immunogenic cell death (ICD), characterized by the release of damaged-associated molecular patterns (DAMPs), which could open new avenues for drug discovery or repurposing168–170.

Botanical therapies containing complex mixtures of NPs have long attracted interest owing to the potential for synergistic therapeutic effects of components within the mixture171,172. However, the variability of the NP composition in the starting plant material owing to factors such as environmental variations in the location the plants were collected at is a major challenge for the development of botanical drugs1. With the advances in technology for their characterization such as metabolomics discussed above, as well as development of regulatory guidance for complex mixtures of NPs (see Related links), it is becoming more feasible to develop such mixtures as therapeutics, rather than to identify and purify a single active ingredient173.

Since gut microbiota are considered to play a major role in health and disease174–176 and NPs are known to affect the gut microbiome composition177–180, this area is an emerging opportunity for NP-based drug discovery. However, drug discovery efforts in this area are still in their infancy, with many open questions remaining181. A future direction may be the characterization of single microbiota-derived species for particular therapeutic applications, and the above-discussed advances in culturing strategies, genome mining and analytics will be of great importance in this respect.

Many advances discussed above are supported by computational tools including databases (such as genomic, chemical, or spectral analysis data; see REF.182 for a recent review on NP databases) and tools that enable the analysis of genetic information, the prediction of chemical structures and pharmacological activities183, the integration of datasets with diverse information (such as tools for multi-omics analysis184) and machine learning applications185.

Although this article focuses on technologies that are enabling the discovery of novel NPs, it is important to acknowledge that unmodified NPs may possess sub-optimal efficacy or ADMET (absorption, distribution, metabolism, excretion, and toxicity) properties. So, for development of NP hits into leads and ultimately into successful drugs, chemical modification may be required. In addition, bringing a compound into clinical development requires a sustainable and economically viable supply of sufficient quantities of the compound. Total chemical synthesis, semi-synthesis using a NP as a starting point for analogue generation and biosynthetic engineering modifying biosynthetic pathways of the producing organism will be of great importance in this context (**Figure 5**). Recent advances in chemical synthesis and biosynthetic engineering technologies are strongly empowering NP-based drug discovery and development by enabling property optimization of complex NP scaffolds that were previously regarded as inaccessible. This allows the enrichment of screening libraries with NPs, NP-hybrids, NP analogues and NP-inspired molecules, as well as superior structure functionalization approaches (including late-stage functionalization) for optimization of NP leads96,107–110,186–190.

Finally, although NP-based drug discovery offers a unique niche for diverse forms of academia–industry collaborations, a key challenge is that scientific and technological expertise is often scattered over many academic institutions and companies. Focused efforts are needed to support translational NP research in academia, which has become more difficult in recent years given the decline in the number of large companies actively engaged in NP research. A conventional solution to improve academia-industry interaction is to focus the relevant expertise “under one umbrella” and in close spatial proximity. For example, the Phytovalley Tirol, centered in Innsbruck, Austria, brings together several research institutions and companies (among others, the Austrian Drug Screening Institute (ADSI), the Michael Popp Research Institute for New Phyto-Entities, Bionorica Research, and Biocrates Life Sciences AG) with the aim of accelerating NP-based drug discovery. Another solution could be virtual consortia, such the International Natural Product Sciences Taskforce (INPST) that we have recently established (see INPST in Related links), which provides a platform for integration of expertise, technology, and materials from the participating academic and industrial entities.

In conclusion, NPs remain a promising pool for the discovery of scaffolds with high structural diversity and various bioactivities that can be directly developed or used as starting points for optimisation into novel drugs. While drug development overall continues to be challenged by high attrition rates, there are additional hurdles for NPs due to issues such as accessibility, sustainable supply, and intellectual property constraints. We, however, believe that the scientific and technological advances discussed in this review provide a strong basis for NP-based drug discovery to continue making major contributions to human health and longevity.

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**Competing interests**

A.G.A. is executive administrator of the International Natural Product Sciences Taskforce (INPST) and Digital Health and Patient Safety Platform (DHPSP). M.B. has served on the speaker’s bureau of Abbott/Mylan, Abbott Vascular, Actavis, Akcea, Amgen, Biofarm, KRKA, MSD, Novo-Nordisk, Novartis, Sanofi-Aventis, Servier and Valeant, has served as a consultant to Abbott Vascular, Akcea, Amgen, Daichii Sankyo, Esperion, Freia Pharmaceuticals, Lilly, MSD, Novartis, Polfarmex, Resverlogix, Sanofi-Aventis, and received grants from Amgen, Mylan, Sanofi and Valeant. R.B. collaborates with Bayer Consumer Health and Dr Willmar Schwabe GmbH & Co KG, and is scientific advisory committee member of PuraPharm International (HK) Limited and ISURA™. G.K.B. is a board member of Bionorica SE. M.D. has received consultancy honorarium from Pfizer Italia and Mylan for training courses for chemists, and is a member of the INPST board of directors. A.T.D.K. is a member of the Scientific and Medical Advisory Board of Evgen Pharma plc. I.E.O. is Dean of Faculty of Pharmacy, Gazi University, Ankara, Turkey, member of Traditional Chinese Medicine Experts Group in European Pharmacopeia, and principal member of Turkish Academy of Sciences (TUBA). B.L.F. is a member of the INPST Board of Directors and received research funding from Dr Willmar Schwabe GmbH & Co KG. K.M.G. has received reimbursement for speaking at conferences sponsored by companies selling nutritional products, and is part of an academic consortium that has received research funding from Abbott Nutrition, Nestec and Danone. C.W.G. is chairman of the scientific advisory board of Cyxone AB, SE. M.H.'s research group has received charitable donations from Dr Willmar Schwabe GmbH & Co KG and recently completed a research project sponsored by Pukka Herb, UK. A.L. is a member of the board of directors of Kaisa Health. M.J.S.M. is president of Kaiviti Consulting and consults for Gnosis by LeSaffre. F.N. is cofounder and shareholder of OncoNox and Aura Biopharm. G.P. is on the board of Neurotez and Neurotrope. M.R. serves as an advisor for the Nestlé Institute of Health Sciences. G.L.R. is a member of the board of directors of INPST. N.T.T. is Founder and CEO of NTZ Lab Ltd. and advisory board member of INPST. M.W. collaborates with Finzelberg GmbH and Schwabe GmbH. J.L.W. collaborates with Nestlé and Firmenich. M.A.P. is CEO and Owner of Bionorica SE. J.H. is an employee and hold shares in UCB Pharma Ltd. M.M. is Founder and Chairman of Sami–Sabinsa Group of Companies. D.S.B. is an employee of Janssen R&D. M.B. is an employee of Evotec (UK) Ltd.

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**Figure legends**

**Figure 1 | Outline of traditional bioactivity-guided isolation steps in natural product drug discovery.** Steps in the process are shown in purple boxes, with associated key limitations shown in red boxes and advances that are helping to address these limitations in modern natural product (NP)-based drug discovery shown in green boxes. The process begins with extraction of NPs from organisms such as bacteria. The choice of extraction method determines which compound classes will be present in the extract (for example, the use of more polar solvents will result in a higher abundance of polar compounds in the crude extract). To maximize the diversity of the extracted NPs, the biological material can be subjected to extraction with several solvents of different polarity. Upon the identification of a crude extract with promising pharmacological activity, the next step is its (often multiple) consecutive bioactivity-guided fractionation until the pure bioactive compound(s) are isolated. A key limitation for the potential of this approach to identify novel NPs is that many potential source organisms cannot be cultured or stop producing relevant NPs when taken out of their natural habitat. These limitations are being addressed through development of new methods for culturing, for *in situ* analysis, for NP synthesis induction and for heterologous expression of biosynthetic genes. At the crude extracts step, challenges include the presence in the extracts of NPs that are already known, NPs that do not have drug-like properties or insufficient amounts of NPs for characterization. These challenges can be addressed through the development of methods for de-replication, extraction and pre-fractionation of extracts. Finally, at the last stage when bioactive compounds are identified by phenotypic assays, significant time and effort are typically needed to identify the affected molecular targets. This challenge can be addressed by the development of methods for accelerated elucidation of molecular modes of action, such as the nematic protein organization technique (NPOT), drug affinity responsive target stability (DARTS), stable isotope labeling with amino acids in cell culture and pulse proteolysis (SILAC-PP), the cellular thermal shift assay (CETSA) and an extension known as thermal proteome profiling (TPP), stability of proteins from rates of oxidation (SPROX), the similarity ensemble approach (SEA) and bioinformatics-based analysis of connectivity (connectivity map, CMAP)23,191–194.

Figure 2 | **Applications of advanced analytical technologies empowering modern natural-product-based drug discovery**. **a** | An illustrative example of the application of LC-HRMS metabolomics in the screening of natural product (NP) extracts is the work of Kurita et al.58, in which 234 bacterial extracts were subjected to image-based phenotypic bioactivity screening and LC-HRMS metabolomics. Clustering of the resulting data allowed prioritization of promising extracts for further analysis, resulting in the discovery of the new NPs, quinocinnolinomycins A–D. **b** | Another illustrative example of LC–HRMS screening of NP extracts is the work of Clevenger et al.87, who obtained novel NP extracts through heterologous expression of FACs containing uncharacterized biosynthetic gene clusters (BGCs) from diverse fungal species in *Aspergillus nidulans.* Analysis of the LC–HRMS metabolomics data with a FAC-score algorithm directed the simultaneous discovery of 15 new NPs and the characterization of their BGCs.

**Figure 3 | Strategies for genome mining-driven discovery of natural products and natural-product-like compounds. a** | Genome mining-based approaches to explore the biosynthetic capacity of microorganisms rely on DNA extraction, sequencing, and bioinformatics analysis. The vast majority of microbes from different environments and microbiota communities have not been cultured, and their capacity to produce natural products (NPs) was largely inaccessible until recently. In the case of unculturable microorganisms, the bioinformatics analysis step can be followed by either targeted heterologous expression of biosynthetic gene clusters prioritized as being likely to yield relevant new NPs or direct chemical synthesis of ‘synthetic–bioinformatic’ NP-like compounds. **b**,**c** | These two approaches are exemplified by the recent discoveries of malacidins (panel **b**) and humimycins (panel **c**), respectively95,96. A major strength of the ‘synthetic–bioinformatic’ approach is that it is entirely independent of microbial culture and gene expression. Its limitations are the accuracy of computational chemical structure predictions and the feasibility of total chemical synthesis.

Figure 4 | **Application of advanced microbial culturing approaches to identify new natural products**. New strategies for isolating previously uncultured microorganisms can enable access to new natural products (NPs) produced by them. **a** | To recapitulate the effect of complex signals coming from the native environment, microorganisms can be cultivated directly in the environment from which they were isolated. This concept is used with the iChip platform, in which diluted environmental samples are seeded in multiple small chambers separated from the native environment with a semipermeable membrane. The potential of this approach is illustrated by the recent discovery of teixobactin, a new antibiotic with activity against Gram-positive bacteria136. **b** | Another important recent development involves obtaining information from environmental samples using ‘omics’ techniques such as metagenomics to identify and partially characterize microorganisms present in a specific environment prior to culturing. An approach relying on such preliminary information was recently used to engineer capturing antibodies based on genetic information, which resulted in the successful cultivation of previously uncultured bacteria from the human mouth147. This reverse genomics workflow was validated by the isolation and cultivation of three species of Saccharibacteria (TM7) along with their interacting Actinobacteria hosts, as well as SR1 bacteria that are members of a candidate phylum with no previously cultured representatives.

**Figure 5 | Strategies to obtain natural product analogues with superior properties.** Unmodified natural products (NPs) often possess sub-optimal properties, and superior analogues need to be obtained in order to yield valuable new drugs. **a** | NP analogues can be accessed through the development of total chemical synthesis followed by chemical derivatization, through semisynthesis using a NP as a starting point for the introduction of chemical modifications, and through biosynthetic engineering using manipulations of biosynthetic pathways of the producing organism to generate NP analogues. **b**,**c** | Tetracyclines are an example of NP-derived antibiotics that already yielded several generations of successfully marketed semisynthetic and synthetic derivatives. The first generation of tetracyclines such as chlortetracycline and tetracycline) were unmodified NPs, while the two following generations of analogues with optimized properties were semisynthetic (second-generation, doxycycline, minocycline; third-generation, tigecycline), and the most recently developed fourth generation (eravacycline) are entirely synthetic analogues accessed *via* total synthesis195,196. More recent examples of property optimization of other classes of NPs through total chemical synthesis followed by chemical derivatization or through semisynthesis are illustrated by studies focused on analogues of chrysomycin A (panel **b**)197 and arylomycins (panel **c**)159, respectively. **d** | The biosynthetic engineering approach has also shown potential; for example, in the generation of analogues of rapamycin113, bleomycin115 (illustrated in panel **d**) and nystatin116.

Box 1 | **Natural products that activate the KEAP1/NRF2 pathway**

An example of a pathway affected by diverse NPs is the KEAP1/NRF2 pathway. This pathway regulates the expression of networks of genes encoding proteins with versatile cytoprotective functions, and has essential roles in the maintenance of redox and protein homeostasis, mitochondrial biogenesis, and the resolution of inflammation198–201.

Activation of this pathway can protect against damage by most types of oxidants and pro-inflammatory agents, and it restores redox and protein homeostasis202. The pathway has therefore attracted attention for the development of drugs for the prevention and treatment of complex diseases, including neurological conditions such as relapsing-remitting multiple sclerosis203 and autism spectrum disorder204.

Dimethyl fumarate (DMF), the methyl ester of the NP fumarate (a tricarboxylic acid cycle intermediate that is found in both animals and plants), is one of the earliest discovered inducers of the KEAP1/NRF2 pathway205,206. The origins of the development of DMF as a drug date back to the use in traditional medicine of the plant *Fumaria officinalis*. Initially, fumaric acid derivatives were used for the treatment of psoriasis as it was thought that psoriasis is caused by a metabolic deficiency in the TCA cycle, which could be compensated for by repletion of fumarate207. Despite this erroneous assumption, DMF is effective in treating psoriasis, both topically and orally, and is the active principle of Fumaderm, which has been used clinically for several decades in the treatment of plaque psoriasis in Germany. More recently, a DMF formulation developed by Biogen has been tested in other immunological disorders, with successful phase III trials in multiple sclerosis208,209 leading to its approval by the FDA and EMA in 2013.

The isothiocyanate sulforaphane, isolated from broccoli (*Brassica oleracea*)210, is among the most potent naturally-occurring inducers of the KEAP1/NRF2 pathway211 and has protective effects in animal models of Parkinson’s212, Huntington’s213 and Alzheimer’s214 diseases, traumatic brain injury215, spinal cord contusion injury216, stroke217, depression218, and multiple sclerosis219. Sulforaphane-rich broccoli extract preparations are being developed as preventive intervention in areas of the world with unavoidable exposures to environmental pollutants, such as China; the initial results of a randomized clinical trial showed rapid and sustained, statistically significant increases in the levels of excretion of the glutathione-derived conjugates of benzene and acrolein220 and a follow-up trial (NCT02656420) also demonstrated dose-response dependent benzene detoxication221. In a placebo-controlled, double-blind, randomized clinical trial in young individuals (aged 13-27) with autism spectrum disorder, sulforaphane reversed many of the clinical abnormalities204; these encouraging findings led to a recently completed clinical trial in children (aged 3–12) (NCT02561481; results of the trial are not yet publicly available). An alpha-cyclodextrin complex of sulforaphane known as SFX-01 (developed by Evgen Pharma) is being clinically studied for its potential to reverse resistance to endocrine therapies in patients with ER+HER2- metastatic breast cancer (phase II trial completed222) and in patients with subarachnoid haemorrhage (phase II trial NCT02614742 recently completed; results are not yet publicly available). Currently, a clinical trial of SFX-01 in patients hospitalized with COVID-19 is in its final stages of preparation.

Finally, the pentacyclic triterpenoids bardoxolone methyl (also known as RTA 402) and omaveloxolone (RTA 408), which are semi-synthetic derivatives of the NP oleanolic acid, are the most potent (active at nanomolar concentrations) activators of the KEAP1/NRF2 pathway known to date223. These compounds have shown protective effects in numerous animal models of chronic disease224, and are currently in clinical trials for a wide range of indications, such as chronic kidney disease in type 2 diabetes, pulmonary arterial hypertension, melanoma, radiation dermatitis, ocular inflammation and Friedreich's ataxia202. Most recently, bardoxolone methyl has entered a clinical trial in patients hospitalized with confirmed COVID-19 (NCT04494646).

**Glossary**

**sp3 carbon atoms**

Tetravalent carbon atoms forming single covalent bonds with other atoms within the molecular structure. A higher fraction of sp3 carbons within molecules is a descriptor that indicates more complex 3D structures.

**Lipinski's rule of five**

This guideline for the likelihood of a compound having oral bioavailability is based on several characteristics containing the number 5. It predicts that a molecule is likely to have poor absorption or permeation if it has more than one of the following characteristics: there are > 5 H-bond donors and >10 H-bond acceptors; the molecular weight is >500; or the partition coefficient LogP is >5. Notably, natural products were identified as common exceptions at the time of publication in 1997.

**Dereplication**

Pharmacological screening of natural product extracts yields hits potentially containing multiple natural products that need to be considered for further study to identify the bioactive compounds. Dereplication is the process of recognizing and excluding from further study such hit-mixtures that contain already known bioactive compounds.

**Phenotypic assays**

Phenotypic assays rely on the ability of tested compounds to exert desired phenotypic changes in cells, isolated tissues, organs or animals. They offer a complementary strategy to target-based assays for identifying new potential drugs.

**Phylogenomic approach**

Phylogenomics is the use of genomic data to reveal evolutionary relationships. In the context of natural product drug discovery, the use of phylogenomics is based on the assumption that organisms that have closer evolutionary relationships are more likely to produce similar natural products.

**Taxonomic distance**

The distance of compared taxa on a constructed phylogenetic tree (also known as an evolutionary tree). Closer distance of compared taxa indicates a closer evolutionary relationship.

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