Invited original article

Title:

Associations of hydroxysteroid 17-beta dehydrogenase 13 variants with liver histology in

Chinese patients with MAFLD

Short title:

HSD17B13 variants and MAFLD

Authors' name:

Wen-Yue Liu, MD^{1#}, Mohammed Eslam, MD, PhD^{2#}, Kenneth I. Zheng, MD³, Hong-Lei Ma,

MD³, Rafael S. Rios, MD³, Min-Zhi Lv, MD⁴, Gang Li, MD³, Liang-Jie Tang, MD³, Pei-Wu

Zhu, MD⁵, Xiao-Dong Wang, MD^{3,6}, Christopher D. Byrne, MB, BCh, PhD⁷, Giovanni

Targher, MD⁸, Jacob George, MD, PhD^{2*}, and Ming-Hua Zheng, MD, PhD^{3,6,9*}

Affiliations

¹Department of Endocrinology, the First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China;

²Storr Liver Centre, Westmead Institute for Medical Research, Westmead Hospital and University of Sydney, Sydney, Australia;

³MAFLD Research Center, Department of Hepatology, the First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China;

⁴Department of Biostatistics, Zhongshan Hospital, Fudan University, Shanghai, China; ⁵Department of Laboratory Medicine, the First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China; ⁶Institute of Hepatology, Wenzhou Medical University, Wenzhou, China;

⁷Southampton National Institute for Health Research Biomedical Research Centre, University

Hospital Southampton, Southampton General Hospital, Southampton, UK;

⁸Section of Endocrinology, Diabetes and Metabolism, Department of Medicine, University and Azienda Ospedaliera Universitaria Integrata of Verona, Verona, Italy;

⁹Key Laboratory of Diagnosis and Treatment for The Development of Chronic Liver Disease

in Zhejiang Province, Wenzhou, China

[#]Co-first Authors: Wen-Yue Liu and Mohammed Eslam

*Corresponding Authors:

Ming-Hua Zheng, MD, PhD

MAFLD Research Center, Department of Hepatology, the First Affiliated Hospital of Wenzhou Medical University; No. 2 Fuxue Lane, Wenzhou 325000, China. E-mail: zhengmh@wmu.edu.cn; fax: (86) 577-55578522; tel: (86) 577-55579622.

Jacob George, MD, PhD

Storr Liver Centre, Westmead Institute for Medical Research, Westmead Hospital and University of Sydney; Westmead 2145, NSW, Australia.

Email: jacob.george@sydney.edu.au; tel: 61-2-88907705; fax: 61-2-96357582.

Electronic word count: 2853

Number of figures and tables: 2 figures and 4 tables

Abbreviations:

MAFLD = metabolic associated fatty liver disease; MAFL = metabolic associated fatty liver; HCC = hepatocellular carcinoma ; BMI = body mass index; HOMA = homoeostasis model assessment; IR = Insulin resistance; GWAS = genome-wide association studies; SNP = single nucleotide polymorphism; MAF = minor allele frequency; *HSD17B13* = hydroxysteroid 17beta dehydrogenase 13; *PNPLA3* = patatin-like phospholipase domain containing protein 3; TLR3 = toll-like receptor 3; IFNL3 = interferon lambda-3 ; TLL1 = tolloid like 1; MICA = MHC class I polypeptide-related chain A

Authorship Statement:

Guarantor of the article: Ming-Hua Zheng

Author's contributions:

Study concept and design: Wen-Yue Liu, Mohammed Eslam, Jacob George and Ming-Hua Zheng

Acquisition of data: Hong-Lei Ma, Liang-Jie Tang, Gang Li and Pei-Wu Zhu

Pathology analysis: Xiao-Dong Wang

Drafting of the manuscript: Wen-Yue Liu, Mohammed Eslam, Jacob George, Kenneth I.

Zheng and Rafael S. Rios

Critical revision: Mohammed Eslam, Jacob George, Giovanni Targher and Christopher D.

Byrne

Statistical analysis: Wen-Yue Liu, Mohammed Eslam and Min-Zhi Lv

Study supervision: Jacob George and Ming-Hua Zheng

Ethical approval: Ethical approval for the study was obtained from the ethics committee of the First Affiliated Hospital of Wenzhou Medical University (2016-246, 1 December 2016). All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional research committee and with the 1964 Helsinki declaration and its later amendments.

Informed consent: Written informed consent was obtained from all participants included in the study.

Abstract

Background and objective:

In Europeans, variants in the hydroxysteroid 17-beta dehydrogenase 13 (*HSD17B13*) gene impact the liver histology of metabolic associated fatty liver disease (MAFLD). The impact of these variants in ethnic Chinese is unknown. The aim of this study was to investigate the potential associations in Chinese patients.

Methods:

427 Han Chinese with biopsy-confirmed MAFLD were enrolled. Two single nucleotide polymorphisms in *HSD17B13* were genotyped - rs72613567 and rs6531975. Logistic regression was used to test the association between the SNPs and liver histology.

Results:

In our cohort, the minor allele TA of the rs72613567 variant was related to an increased risk of fibrosis [OR = 2.93 (1.20-7.17), P = 0.019 for the additive model; OR = 3.32 (1.39-7.91), P = 0.007 for the recessive model], an inverse association as compared to the results from European cohorts. In contrast, we observed a protective effect on fibrosis for the minor A allele carriers of the *HSD17B13* rs6531975 variant [OR = 0.48 (0.24-0.98), P = 0.043 for the additive model; OR = 0.62 (0.40-0.94), P = 0.025 for the dominant model]. *HSD17B13* variants were only associated with fibrosis but not other histological features. Furthermore, *HSD17B13* rs6531975 modulated the effect of *PNPLA3* rs738409 on hepatic steatosis.

Conclusions:

The *HSD17B13* rs72613567 variant is a risk variant for fibrosis in a Han Chinese MAFLD population but with a different direction for allelic association to that seen in Europeans. This data exemplifies the need for studying diverse populations in genetic studies in order to fine map GWAS signals.

Keywords:

Metabolic associated fatty liver disease; Non-alcoholic fatty liver disease; Hydroxysteroid 17-beta dehydrogenase 13; Single nucleotide polymorphism

Introduction

Metabolic associated fatty liver disease (MAFLD) is recognized as a leading cause of liver-related morbidity and mortality.^{1,2} In China, MAFLD burden is increasing, with prevalence rising from 18% to 29% in the last decade.³ MAFLD comprises a spectrum of disease ranging from simple steatosis or MAFL to the presence of steatohepatitis with varying degrees of fibrosis and cirrhosis.⁴ MAFLD arises from "multiple hits", with genes acting as an important modifier of the clinical phenotype.⁵ Though our understanding of the underpinnings of MAFLD has been enhanced by numerous genetic association studies, all identified polymorphisms to date explain only 10–20% of disease heritability.^{6,7} It is broadly acknowledged that there is over-representation of subjects of European ancestry in human genetics research with ~79% of all GWAS participants of European descent. This overrepresentation hinders a complete understanding of the human genetic architecture. Moreover, it can also have a negative impact including prediction accuracies between 1.6-4.9 fold lower in other ethnicities compared to Europeans.⁸ Hence, increasing the representation of diverse populations and studying other ethnicities has become a research priority.

Several variants in the hydroxysteroid 17-beta dehydrogenase 13 (*HSD17B13*) gene encoding a hepatic lipid droplet protein have been identified to impact the histological features of MAFLD. However, the impact of *HSD17B13* gene variants on MAFLD histology among those of Chinese ancestry is unknown. Notably, allele frequencies, haplotype patterns and the effect size of polymorphisms vary considerably across populations and ethnicities.⁶ As *HSD17B13* has been proposed as a therapeutic target for MAFLD, it is pivotal to explore whether the effect of this variant observed in Caucasian populations extends to other populations, as also the effect size.

It is known that the genetic association of variants in *HSD17B13* with the histological features of MAFLD is complex, with different potentially causative SNPs and various SNPs associated with different phenotypic patterns. For example, alleles of rs6834314 and rs72613567 associate with decreased injury and with increased hepatic fat.⁹ However, there are other studies that show no association of rs72613567 with steatosis.^{10,11} Noncoding SNPs (rs6531975) not in linkage disequilibrium with rs72613567 have also

been associated with decreased hepatic fat.⁹ Adding to this complexity, a recent study of 487 patients suggested that those harboring the 'protective' *TA*-allele of rs72613567 have a numerically increased risk for mortality, liver-related death and hepatic decompensation.¹² Likewise, while some reports suggested that there is a potential interaction between *HSD17B13* and variants in patatin-like phospholipase domain containing protein 3 (*PNPLA3*) in MAFLD, subsequent reports have failed to discern an association.^{13,14} Given these controversies, the aims of this study were 1) to explore the role of variants in the *HSD17B13* gene in a cohort of Han Chinese with biopsy-confirmed MAFLD, 2) to clarify the role of the variants on the various morphological features of MAFLD and 3) to discern if there is any interaction between the variants and variants in *PNPLA3*.

Materials and Methods

Study population

We recruited 427 consecutive Han Chinese with biopsy-confirmed MAFLD from the PERSONS cohort (2017.01-2019.05). The definition of MAFLD was based on the criteria proposed by an international expert panel.¹⁵ The study cohort included patients from a previously published study as well as additional subjects.¹⁶ To ascertain the effects of the *HSD17B13* variant on liver disease solely due to MAFLD, patients with other causes of liver disease including alcohol use disorder or viral hepatitis were excluded. Briefly, all consecutive patients aged \geq 18 with biopsy-proven MAFLD without alternative causes of liver disease were recruited to the study. The study protocol was approved by the ethics committee of the First Affiliated Hospital of Wenzhou Medical University (2016-246, 1 December 2016) and registered in the Chinese Clinical Trial Registry (ChiCTR-EOC-17013562). Written informed consent was obtained from each subject before their participation in the study. Patients identifiers were anonymized and replaced by the health examination number.

Clinical and biochemical data

Clinical and biochemical data were collected from all patients within 24 hours of liver biopsy. Body mass index (BMI) was calculated as weight (kg) divided by the square of height (m). Insulin resistance (IR) was estimated according by the homoeostasis model assessment (HOMA-IR).¹⁷ Diagnosis of diabetes was based on criteria of the American Diabetes Association.¹⁸

Assessment of liver histology

Liver biopsies were performed using a 16-gauge needle under ultrasound guidance. The histology was reviewed by a single liver pathologist (X.D. Wang) who was blinded to the clinical and biochemical data. Histologic scoring was based on the Activity Score.¹⁹ Steatohepatitis was diagnosed as a score \geq 4 and a score of at least one for each of steatosis, ballooning and lobular inflammation. Severe steatosis, severe ballooning and severe lobular inflammation were defined if their scores were more than or equal to 2.

Genetic analysis

Genotyping for the HSD17B13 (rs72613567 and rs6531975) and PNPLA3 (rs738409) variants were performed using the MassARRAY or TaqMan assays platform according to the manufacturer's protocol. For the purpose of genotyping, each sample used approximately 20 ng of genomic DNA. Locus-specific PCR and detection primers were designed using Assay Design Suite v3.1.

Statistical analysis

Statistical analyses were performed using R software (version 3.5.2, R Foundation for Statistical Computing, Vienna, Austria) and SPSS 19.0 (SPSS Inc. Chicago, IL, USA). Continuous variables are expressed as mean \pm SD and compared using the One-way ANOVA analysis. Categorical variables are expressed as frequency (%) and compared using the chi-square test. Multivariable logistic regression models were undertaken to test the association between the aforementioned SNPs and liver histology features. P < 0.05 was considered to be statistically significant.

Results

Patient characteristics

The study comprised 427 consecutive biopsy-confirmed MAFLD patients; their clinical, biochemical, and histological features are depicted in **Supplementary Table 1**. The average age was 41 years with 73.8% being male. About 287 (67.2%) had fibrosis (\geq F1), 226 (52.9%) had severe steatosis (S2-S3), 157 (36.8%) had severe ballooning (B2) and 84 (19.7%) had severe inflammation (A2-A3).

Genotype distribution, HWE calculations

Two single nucleotide polymorphisms in *HSD17B13* were genotyped - rs72613567 and rs6531975. Genotype distribution of rs72613567 and rs6531975 in *HSD17B13* was in Hardy–Weinberg equilibrium (All P > 0.05). The minor allele frequency (MAF) for rs72613567 and rs6531975 were 0.32 and 0.30 in our cohort, respectively. It is close to the MAF in general East Asian population in the 1000 genomes project.²⁰ The overall genotype distribution of rs72613567 T/T, T/TA and TA/TA was 47.3%, 42.0%, and 10.7%, while the distribution of rs6531975 G/G, G/A and A/A was 49.8%, 40.5%, and 9.8%, respectively.

Clinical and laboratory characteristics stratified by HSD17B13 variants

The baseline characteristics of study participants according to rs72613567 genotypes is presented in **Table 1**. There were significant differences in levels of fasting glucose, triglycerides and HDL-cholesterol among rs72613567 genotypes (All P < 0.05). **Table 2** shows the baseline characteristics of study participants according to rs6531975 genotypes. No significant differences were observed among rs6531975 genotypes.

HSD17B13 variants and hepatic steatosis

The proportion of severe steatosis in rs72613567 T/T, T/TA and TA/TA was 103 (52.0%), 91 (51.7%) and 27 (60.0%), while the proportion of severe steatosis in rs6531975 G/G, G/A and A/A was 113 (54.1%), 84 (49.4%) and 24 (58.5%) (Table 3). No association between *HSD17B13* variants and severe steatosis was observed in multivariable analysis (Table 4).

HSD17B13 variants and hepatocyte ballooning and lobular inflammation

The proportion of severe ballooning in rs72613567 T/T, T/TA and TA/TA was 73 (36.9%), 58 (33.0%) and 21(46.7%), while the proportion of severe ballooning in rs6531975 G/G, G/A and A/A was 79 (37.8%), 63 (37.1%) and 11 (26.8%). The proportion of severe inflammation in rs72613567 T/T, T/TA and TA/TA was 35 (17.7%), 35 (19.9%) and 12 (26.7%), while the proportion of severe inflammation in rs6531975 G/G, G/A and A/A was 40 (19.1%), 35 (20.6%) and 8 (19.5%) (Table 3). Both severe ballooning and inflammation were unrelated to *HSD17B13* variants in multivariable analysis (Table 4).

HSD17B13 variants and fibrosis

The prevalence of having fibrosis in rs72613567 T/T, T/TA and TA/TA was 135 (68.2%), 111 (63.1%) and 38 (84.4%). A higher prevalence of fibrosis was observed in patients with the TA/TA genotype in rs72613567 (P < 0.05) (**Table 3**). In rs6531975 genotypes, the prevalence of having fibrosis in G/G, G/A and A/A was 150 (71.8%), 109 (64.1%) and 23 (56.1%). The A allele carriers of rs6531975 showed a non-significant trend for a reduced prevalence of having fibrosis (P = 0.082) (**Table 3**).

To further understand the association between *HSD17B13* variants and liver histology in Chinese patients with MAFLD, multivariable logistic regression models were undertaken. As shown in **Table 4**, rs72613567 TA/TA increased the risk of fibrosis with an odds ratio (OR) of 2.93 (TA/TA vs. T/T, 95% CI: 1.20-7.17, P = 0.019) for the additive model and an OR of 3.32 (TA/TA vs. T/T+T/TA, 95% CI: 1.39-7.91, P = 0.007) for the recessive model after adjusting for age, sex, BMI, presence of diabetes,

fasting glucose, triglycerides and HDL-cholesterol. In contrast, the rs6531975 A allele appeared to have a protective impact on fibrosis with an OR of 0.48 (A/A vs. G/G, 95% CI: 0.24-0.98, P = 0.043) for the additive model and an OR of 0.62 (G/A + A/A vs. G/G, 95% CI: 0.40-0.94, P = 0.025) for the dominant model after adjusting for age, sex, BMI and presence of diabetes.

Interaction of PNPLA3 and HSD17B13 variants

Next, we conducted interaction analysis for *HSD17B13* (rs72613567 and rs6531975) and *PNPLA3* (rs738409) variants for their impact on liver histology. For fibrosis, no interaction effects were observed between the two genes. In contrast, there was an interaction between rs6531975 and rs738409 with regard to hepatic steatosis (**Figure 1**). For the rs738409 risk allele carriers (CG + GG), the proportion of severe steatosis was lower in patients with the rs6531975 A allele (G/A + A/A), compared to that with rs6531975 G/G (**Figure 1A**). Using the latter as reference, patients with the rs6531975 A allele (G/A + A/A) attenuated the risk effect of the rs738409 G allele (C/G + G/G) on steatosis with an OR of 0.57 (95% CI: 0.34-0.96, P = 0.034) after adjusting for age, sex, BMI and presence of diabetes (**Figure 1B**). The interaction between rs72613567 and rs738409 on liver steatosis was also performed (**Figure 2**), however no effect was observed.

Discussion

We characterized the impact of *HSD17B13* gene variants on histological features in a cohort of Han Chinese with MAFLD. This study has three key findings. First, we confirmed the *HSD17B13* region as a susceptibility locus for MAFLD related fibrosis but extend these findings to identify an inverse allelic direction of association as compared to that reported in Europeans. Second, the *HSD17B13* variants are only associated with fibrosis and not any other histological feature. Third, the *HSD17B13* variants modulates the effect of *PNPLA3* rs738409 on hepatic steatosis, but not other histological features. The association between *HSD17B13* variants and liver histological features seems to be complex, with multiple suggested functional variants. Notably, in our cohort the minor allele TA of the rs72613567 variant was related to an increased risk of fibrosis, an inverse association as compared to the results in European cohorts. Hence, if there is a shared causal variant across European and Chinese populations, it is unlikely to be rs72613567. In this regard, we observed a protective effect for the minor A allele carriers of the *HSD17B13* rs6531975 variant, but this is not in strong linkage disequilibrium with rs72613567. Thus, further fine-mapping studies in Han Chinese populations and comparison to other populations would be helpful to identify shared causal variants across different ethnicities.

The differential effect size and allele direction of variants discovered by GWAS between ethnicities is not uncommon. In one Chinese MAFLD cohort, researchers found that the neurocan (*NCAN*) rs2228603 T variant associated with a higher level of HDL,²¹ while it is positively related to liver steatosis in the US population.²² Similarly, toll-like receptor 3 (*TLR3*) rs3775290^{23,24} and interferon lambda-3 (*IFNL3*) rs12979860^{25,26} variants in Chinese hepatocellular carcinoma (HCC) populations show opposite effects to those in non-Asian populations. Inconsistent results have also been observed in other Asian populations such as among Japanese. For example, tolloid like 1 (*TLL1*) rs17047200²⁷ and MHC class I polypeptide-related chain A (*MICA*) rs2596542²⁸ variants were suggested to have protective impacts on fibrosis and HCC in Caucasians. The association was inverse to those of a Japanese cohort^{29,30}. Besides, there are several MAFLD-related SNPs in Europeans for which there has been no association in Chinese populations³¹⁻³³. Along the same line, lower genetic prediction accuracies between 1.6-4.9 fold lower was observed in other ethnicities compared to Europeans.⁸ Hence, increasing the representation of diverse populations and studying other ethnicities has recently become a research priority to enhance understanding of the human genetic architecture and its translational implications.

The ethnic differences in the characteristics of patients with MAFLD might also contribute to the observed differences in the genetic findings. There is growing evidence for example that MAFLD disease course in Asian populations is different to that in Caucasians. As an example, for the same BMI, there is a higher prevalence of MAFLD in Asians. Published reports also indicate that lean MAFLD accounts for

36.9% of cases in China,³ but only 17.3 % of the total disease burden in the United States.³⁴ Differences in metabolic adaptation has been reported between lean and non-lean MAFLD patients suggesting that lean fatty liver disease likely has a distinct pathophysiology.³⁵

Another intriguing aspect of this study is the lack of association between HSD17B13 variants and other histological features. To date the nature of the association between the rs72613567 allelic variant and histological features of MAFLD, particularly steatosis is unclear. Abul-Husn and colleagues suggested a lack of association between the rs72613567 TA variant and steatosis in human liver,¹⁰ consistent with the study of Pirola et al.¹¹ However, a study by Ma et al. found a significant association with hepatic steatosis.9 Similarly, in animal and in-vitro studies, inconsistent results have been reported for an effect of HSD17B13 on hepatic lipid accumulation. Abul-Husn et al showed no differences in lipid accumulation according HSD17B13 isoforms.¹⁰ Similarly Ma et al. reported that HSD17B13 over-expression or knockout in HepG2 cells did not affect lipid content.⁹ On the other hand, Marion et al. noted hepatic steatosis in HSD17B13 knockout mice,³⁶ whilst Su et al. observed steatosis in mice that over-expressed HSD17B13.37 Collectively, these results imply that HSD17B13 variants could have a direct impact on fibrosis rather than effects on steatosis. These findings may be associated with retinol metabolism since retinol plays a crucial role in the activation and transformation of hepatic stellate cells to matrix secreting myofibroblasts and the development of hepatic fibrosis.³⁸ Since HSD17B13 participates in the rate limiting step of retinol metabolism,⁹ the mutant in HSD17B13 might conceivably influence the process of fibrosis.

The interaction between *HSD17B13* and *PNPLA3* variants in MAFLD is also a subject of controversy.^{14,39} In this work, we noted an interaction between these variants with regard to steatosis, but not with other histological features. Furthermore, rs72613567 TA variant in *HSD17B13* was found to be associated with obesity in Europeans.¹¹ Potential interaction effects may exist between fat distribution and *HSD17B13* variants. We performed a logistic regression model to explore the interaction effects, however, nothing was found in adiposity measures and SNP genotypes on liver phenotypes (data was not shown in Result section). As *HSD17B13* has been suggested as a potential therapeutic target for MAFLD and the growing concerns about the failure of phase 2 and 3 clinical trials in this disease^{40,41} that is at least partially attributed to clinical heterogeneity, our study highlights the importance of first understanding the functional basis of the various proposed genomic and other targets before therapeutic development.^{40,42} Collectively, our data suggest supports such an approach. The data from *HSD17B13*-knockout mice in fact suggests that HSD17B13 triggers steatosis and inflammation³⁶ which is opposite to what has been reported in humans.

The present study has limitations. Firstly, the sample size is modest. In case the opposite finding is due to the sample size, we have performed a post-hoc power analysis. The power calculated for the model was 72%. It is close to but less than 0.80. Considering the low proportion of the rs72613567 TA variant in the general population, we think it is acceptable. In addition, lack of a validation cohort from populations in other parts of China or those of Chinese ancestry living outside mainland China is another limitation. In conclusion, the *HSD17B13* rs72613567 variant is a risk variant for hepatic fibrosis in a Han Chinese MAFLD population with a different direction for allelic association to that seen in Europeans.

Funding sources

This work was supported by grants from the National Natural Science Foundation of China (81500665), High Level Creative Talents from Department of Public Health in Zhejiang Province, Project of New Century 551 Talent Nurturing in Wenzhou. GT is supported in part by grants from the University School of Medicine of Verona, Verona, Italy. CDB is supported in part by the Southampton NIHR Biomedical Research Centre (IS-BRC-20004), UK. ME and JG are supported by the Robert W. Storr Bequest to the Sydney Medical Foundation, University of Sydney; National Health and Medical Research Council of Australia (NHMRC) Program (APP1053206, APP1149976) and Project (APP1107178 and APP1108422) grants.

Competing interests

The authors declare that there are no conflicts of interest associated with this manuscript.

Reference

- [1] EASL-EASD-EASO Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease. Journal of hepatology 2016;64(6):1388-1402. doi: 10.1016/j.jhep.2015.11.004. PMID: 27062661
- [2] Younossi ZM, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease-Meta-analytic assessment of prevalence, incidence, and outcomes. Hepatology (Baltimore, Md) 2016;64(1):73-84. doi: 10.1002/hep.28431. PMID: 26707365
- [3] Zhou F, Zhou J, Wang W, Zhang XJ, Ji YX, Zhang P, et al. Unexpected Rapid Increase in the Burden of NAFLD in China From 2008 to 2018: A Systematic Review and Meta-Analysis. Hepatology (Baltimore, Md) 2019;70(4):1119-1133. doi: 10.1002/hep.30702. PMID: 31070259
- [4] Masuoka HC, Chalasani N. Nonalcoholic fatty liver disease: an emerging threat to obese and diabetic individuals. Ann N Y Acad Sci 2013;1281:106-122. doi: 10.1111/nyas.12016. PMID: 23363012
- [5] Buzzetti E, Pinzani M, Tsochatzis EA. The multiple-hit pathogenesis of non-alcoholic fatty liver disease (NAFLD). Metabolism: clinical and experimental 2016;65(8):1038-1048. doi: 10.1016/j.metabol.2015.12.012. PMID: 26823198
- [6] Eslam M, George J. Genetic contributions to NAFLD: leveraging shared genetics to uncover systems biology. Nature reviews Gastroenterology & hepatology 2020;17(1):40-52. doi: 10.1038/s41575-019-0212-0. PMID: 31641249
- [7] Eslam M, Valenti L, Romeo S. Genetics and epigenetics of NAFLD and NASH: clinical impact. Journal of hepatology 2018;68(2):268-279. PMID:
- [8] Martin AR, Kanai M, Kamatani Y, Okada Y, Neale BM, Daly MJ. Current clinical use of polygenic scores will risk exacerbating health disparities. Nature genetics 2019;51(4):584. PMID:
- [9] Ma Y, Belyaeva OV, Brown PM, Fujita K, Valles K, Karki S, et al. 17-Beta Hydroxysteroid Dehydrogenase 13 Is a Hepatic Retinol Dehydrogenase Associated With Histological Features of Nonalcoholic Fatty Liver Disease. Hepatology (Baltimore, Md) 2019;69(4):1504-1519. doi: 10.1002/hep.30350. PMID: 30415504
- [10] Abul-Husn NS, Cheng X, Li AH, Xin Y, Schurmann C, Stevis P, et al. A Protein-Truncating HSD17B13 Variant and Protection from Chronic Liver Disease. The New England journal of medicine 2018;378(12):1096-1106. doi: 10.1056/NEJMoa1712191. PMID: 29562163
- [11] Pirola CJ, Garaycoechea M, Flichman D, Arrese M, San Martino J, Gazzi C, et al. Splice variant rs72613567 prevents worst histologic outcomes in patients with nonalcoholic fatty liver disease. Journal of lipid research 2019;60(1):176-185. doi: 10.1194/jlr.P089953. PMID: 30323112
- [12] Scheiner B, Stättermayer AF, Schwabl P, Bucsics T, Paternostro R, Bauer D, et al. Impact of HSD17B13 rs72613567 genotype on hepatic decompensation and mortality in patients with portal hypertension. Liver International 2020;40(2):393-404. PMID:
- [13] Kallwitz E, Tayo BO, Kuniholm MH, Daviglus M, Zeng D, Isasi CR, et al. Association of HSD17B13 rs72613567:TA with non-alcoholic fatty liver disease in Hispanics/Latinos. Liver international : official journal of the International Association for the Study of the Liver 2020;40(4):889-893. doi: 10.1111/liv.14387. PMID: 31965669

- [14] Stickel F, Lutz P, Buch S, Nischalke HD, Silva I, Rausch V, *et al.* Genetic variation in HSD17B13 reduces the risk of developing cirrhosis and hepatocellular carcinoma in alcohol misusers. Hepatology (Baltimore, Md) 2019 Published Online First: 2019/10/21. doi: 10.1002/hep.30996. PMID: 31630428
- [15] Eslam M, Newsome PN, Anstee QM, Targher G, Gomez MR, Zelber-Sagi S, et al. A new definition for metabolic associated fatty liver disease: an international expert consensus statement. Journal of hepatology 2020 Published Online First: 2020/04/12. doi: 10.1016/j.jhep.2020.03.039. PMID: 32278004
- [16] Liu WY, Zheng KI, Pan XY, Ma HL, Zhu PW, Wu XX, et al. Effect of PNPLA3 polymorphism on diagnostic performance of various noninvasive markers for diagnosing and staging nonalcoholic fatty liver disease. Journal of gastroenterology and hepatology 2019 Published Online First: 2019/11/05. doi: 10.1111/jgh.14894. PMID: 31677195
- [17] Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. Diabetologia 1985;28(7):412-419. doi: 10.1007/bf00280883. PMID: 3899825
- [18] Association AD. 1. Improving Care and Promoting Health in Populations: Standards of Medical Care in Diabetes-2020. Diabetes care 2020;43(Suppl 1):S7-s13. doi: 10.2337/dc20-S001. PMID: 31862744
- [19] Kleiner DE, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW, et al. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. Hepatology 2005;41(6):1313-1321. doi: 10.1002/hep.20701. PMID: 15915461
- [20] Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, Korbel JO, et al. A global reference for human genetic variation. Nature 2015;526(7571):68-74. doi: 10.1038/nature15393. PMID: 26432245
- [21] Wu MJ, Yuan C, Lu LL, An BQ, Xuan SY, Xin YN. Role of NCAN rs2228603 polymorphism in the incidence of nonalcoholic fatty liver disease: a case-control study. Lipids in health and disease 2016;15(1):207. doi: 10.1186/s12944-016-0367-4. PMID: 27887608
- [22] Hernaez R, McLean J, Lazo M, Brancati FL, Hirschhorn JN, Borecki IB, et al. Association between variants in or near PNPLA3, GCKR, and PPP1R3B with ultrasound-defined steatosis based on data from the third National Health and Nutrition Examination Survey. Clinical gastroenterology and hepatology : the official clinical practice journal of the American Gastroenterological Association 2013;11(9):1183-1190.e1182. doi: 10.1016/j.cgh.2013.02.011. PMID: 23416328
- [23] Huang X, Li H, Wang J, Huang C, Lu Y, Qin X, et al. Genetic polymorphisms in Toll-like receptor 3 gene are associated with the risk of hepatitis B virus-related liver diseases in a Chinese population. Gene 2015;569(2):218-224. doi: 10.1016/j.gene.2015.05.054. PMID: 26024592
- [24] Sghaier I, Zidi S, Mouelhi L, Ghazoueni E, Brochot E, Almawi WY, et al. TLR3 and TLR4 SNP variants in the liver disease resulting from hepatitis B virus and hepatitis C virus infection. British journal of biomedical science 2019;76(1):35-41. doi: 10.1080/09674845.2018.1547179. PMID: 30421643
- [25] Hou W, Qiao K, Huo Z, Du Y, Wang C, Syn WK. Association of IFNL3 rs12979860 polymorphism with HCV-related hepatocellular carcinoma susceptibility in a Chinese

population. Clinical and experimental gastroenterology 2019;12:433-439. doi: 10.2147/ceg.S206194. PMID: 31807049

- [26] Buivydiene A, Liakina V, Kashuba E, Norkuniene J, Jokubauskiene S, Gineikiene E, et al. Impact of the Uridine⁻Cytidine Kinase Like-1 Protein and IL28B rs12979860 and rs8099917 SNPs on the Development of Hepatocellular Carcinoma in Cirrhotic Chronic Hepatitis C Patients-A Pilot Study. Medicina (Kaunas, Lithuania) 2018;54(5) doi: 10.3390/medicina54050067. PMID: 30344298
- [27] John M, Metwally M, Mangia A, Romero-Gomez M, Berg T, Sheridan D, et al. TLL1 rs17047200 Increases the Risk of Fibrosis Progression in Caucasian Patients With Chronic Hepatitis C. Gastroenterology 2017;153(5):1448-1449. doi: 10.1053/j.gastro.2017.04.056. PMID: 28993163
- [28] Lange CM, Bibert S, Dufour JF, Cellerai C, Cerny A, Heim MH, et al. Comparative genetic analyses point to HCP5 as susceptibility locus for HCV-associated hepatocellular carcinoma. Journal of hepatology 2013;59(3):504-509. doi: 10.1016/j.jhep.2013.04.032. PMID: 23665287
- [29] Kumar V, Kato N, Urabe Y, Takahashi A, Muroyama R, Hosono N, et al. Genome-wide association study identifies a susceptibility locus for HCV-induced hepatocellular carcinoma. Nature genetics 2011;43(5):455-458. doi: 10.1038/ng.809. PMID: 21499248
- [30] Matsuura K, Sawai H, Ikeo K, Ogawa S, Iio E, Isogawa M, et al. Genome-Wide Association Study Identifies TLL1 Variant Associated With Development of Hepatocellular Carcinoma After Eradication of Hepatitis C Virus Infection. Gastroenterology 2017;152(6):1383-1394. doi: 10.1053/j.gastro.2017.01.041. PMID: 28163062
- [31] Yuan C, Lu L, An B, Jin W, Dong Q, Xin Y, et al. Association Between LYPLAL1 rs12137855 Polymorphism With Ultrasound-Defined Non-Alcoholic Fatty Liver Disease in a Chinese Han Population. Hepatitis monthly 2015;15(12):e33155. doi: 10.5812/hepatmon.33155. PMID: 26977168
- [32] Peng XE, Chen FL, Liu W, Hu Z, Lin X. Lack of association between SREBF-1c gene polymorphisms and risk of non-alcoholic fatty liver disease in a Chinese Han population. Scientific reports 2016;6:32110. doi: 10.1038/srep32110. PMID: 27572914
- [33] Niu TH, Jiang M, Xin YN, Jiang XJ, Lin ZH, Xuan SY. Lack of association between apolipoprotein C3 gene polymorphisms and risk of nonalcoholic fatty liver disease in a Chinese Han population. World journal of gastroenterology 2014;20(13):3655-3662. doi: 10.3748/wjg.v20.i13.3655. PMID: 24707151
- [34] Younossi ZM, Stepanova M, Negro F, Hallaji S, Younossi Y, Lam B, et al. Nonalcoholic fatty liver disease in lean individuals in the United States. Medicine 2012;91(6):319-327. doi: 10.1097/MD.0b013e3182779d49. PMID: 23117851
- [35] Chen F, Esmaili S, Rogers GB, Bugianesi E, Petta S, Marchesini G, et al. Lean NAFLD: A Distinct Entity Shaped by Differential Metabolic Adaptation. Hepatology (Baltimore, Md) 2020;71(4):1213-1227. doi: 10.1002/hep.30908. PMID: 31442319
- [36] Adam M, Heikela H, Sobolewski C, Portius D, Maki-Jouppila J, Mehmood A, *et al.* Hydroxysteroid (17beta) dehydrogenase 13 deficiency triggers hepatic steatosis and inflammation in mice. Faseb j 2018;32(6):3434-3447. doi: 10.1096/fj.201700914R. PMID: 29401633

- [37] Su W, Wang Y, Jia X, Wu W, Li L, Tian X, et al. Comparative proteomic study reveals 17beta-HSD13 as a pathogenic protein in nonalcoholic fatty liver disease. Proc Natl Acad Sci U S A 2014;111(31):11437-11442. doi: 10.1073/pnas.1410741111. PMID: 25028495
- [38] Puche JE, Saiman Y, Friedman SL. Hepatic stellate cells and liver fibrosis. Comprehensive Physiology 2013;3(4):1473-1492. doi: 10.1002/cphy.c120035. PMID: 24265236
- [39] Bellan M, Colletta C, Barbaglia MN, Salmi L, Clerici R, Mallela VR, et al. Severity of Nonalcoholic Fatty Liver Disease in Type 2 Diabetes Mellitus: Relationship between Nongenetic Factors and PNPLA3/HSD17B13 Polymorphisms. Diabetes & metabolism journal 2019;43(5):700-710. doi: 10.4093/dmj.2018.0201. PMID: 31694082
- [40] Ratziu V, Friedman SL. Why do so many NASH trials fail? Gastroenterology 2020 Published Online First: 2020/05/23. doi: 10.1053/j.gastro.2020.05.046. PMID: 32439497
- [41] Eslam M, George J. Genetic Insights for Drug Development in NAFLD. Trends in pharmacological sciences 2019;40(7):506-516. doi: 10.1016/j.tips.2019.05.002. PMID: 31160124
- [42] Eslam M, Sanyal AJ, George J. MAFLD: A Consensus-Driven Proposed Nomenclature for Metabolic Associated Fatty Liver Disease. Gastroenterology 2020;158(7):1999-2014.e1991. doi: 10.1053/j.gastro.2019.11.312. PMID: 32044314

Table Legends

 Table 1. Baseline characteristics of biopsy-confirmed MAFLD patients according to rs72613567
 genotypes.

 Table 2. Baseline characteristics of biopsy-confirmed MAFLD patients according to rs6531975
 genotypes.

Table 3. Liver histology features of biopsy-confirmed MAFLD patients according to *HSD17B13*

 genotypes.

Table 4. Association between HSD17B13 variants and liver histology features in Chinese MAFLD

 patients.

Supplemental Table 1. Baseline characteristics of biopsy-confirmed Chinese MAFLD patients.

Figure Legends

Figure 1. The interaction of HSD17B13 rs6531975 and PNPLA3 rs738409 on liver steatosis.

Panel A shows the prevalence of mild steatosis and severe steatosis according to rs6531975 and rs738409 genotypes. Panel B shows the interaction effect of rs6531975 and rs738409 on steatosis after adjusting for age, sex, BMI and presence of diabetes. Patients with the rs6531975 A allele (G/A + A/A) attenuated the risk effect of the rs738409 G allele (C/G + G/G) on steatosis with an OR of 0.57 (95% CI: 0.34-0.96, P = 0.034).

Figure 2. The interaction of HSD17B13 rs72613567 and PNPLA3 rs738409 on liver steatosis. Panel A shows the prevalence of mild steatosis and severe steatosis according to rs72613567 and rs738409 genotypes. Panel B shows the interaction effect of rs72613567 and rs738409 on steatosis after adjusting for age, sex, BMI and presence of diabetes. No interaction effect was observed in rs72613567 and rs738409.