**Adverse cardiovascular magnetic resonance phenotypes are associated with greater likelihood of incident coronavirus disease 2019: Findings from the UK Biobank**

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**Abstract**

**Background:** Coronavirus disease 2019 (COVID-19) disproportionately affects older people. Observational studies suggest indolent cardiovascular involvement after recovery from acute COVID-19. However, these findings may reflect pre-existing cardiac phenotypes.

**Aims:** We tested the association of baseline cardiovascular magnetic resonance (CMR) phenotypes with incident COVID-19.

**Methods:** We studied UK Biobank participants with CMR imaging and COVID-19 testing. We considered left and right ventricular (LV, RV) volumes, ejection fractions, and stroke volumes, LV mass, LV strain, native T1, aortic distensibility, and arterial stiffness index. COVID-19 test results were obtained from Public Health England. Co-morbidities were ascertained from self-report and Hospital Episode Statistics (HES). Critical care admission and death were from HES and death register records. We tested the association of each cardiovascular measure with COVID-19 test result in multivariable logistic regression models adjusting for age, sex, ethnicity, deprivation, body mass index, smoking, diabetes, hypertension, high cholesterol, and prior myocardial infarction.

**Results:** We studied 310 participants (n=70 positive). Median age was 63·8 [57·5, 72·1] years; 51·0% (n=158) were male. 78·7% (n=244) were tested in hospital, 3·5% (n=11) required critical care admission, and 6·1% (n=19) died. In fully adjusted models,smaller LV/RV end-diastolic volumes, smaller LV stroke volume, and poorer global longitudinal strain were associated with significantly higher odds of COVID-19 positivity.

**Discussion:** We demonstrate association of pre-existing adverse CMR phenotypes with greater odds of COVID-19 positivity independent of classical cardiovascular risk factors.

**Conclusions:** Observational reports of cardiovascular involvement after COVID-19 may, at least partly, reflect pre-existing cardiac status rather than COVID-19 induced alterations.

**Keywords:** Cardiovascular magnetic resonance; coronavirus disease 2019 (COVID-19); severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)

**Introduction**

Coronavirus disease 2019 (COVID-19), which disproportionately affects older people, is increasingly recognised as a multi-system disease[1]. A spectrum of cardiovascular manifestations has been reported in the context of acute infection[2–8]. The high inflammatory burden of acute COVID-19 is postulated to lead to vascular inflammation, myocarditis, and cardiac arrhythmias[9–12]. The accompanying hypercoagulable state has been linked to higher rates of thromboembolic events manifesting as myocardial and cerebral infarctions[13].

Interestingly, acute myocardial injury, occurring in the context of COVID-19, has been linked to greater mortality independent of factors such as acute respiratory distress syndrome[5, 6]. Reports of primary cardiac presentations of COVID-19 in the absence of pulmonary involvement suggest potential cardio-specific actions of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)[14–16]. Indeed, in vitro studies have demonstrated that human cardiomyocytes are permissive for SARS-CoV-2 infection and that the virus induces a cytotoxic response in these cells[17]. Similarly, autopsy studies have demonstrated direct cardiotoxic effects of SARS-CoV-2[18].

In addition to cardiac involvement linked with acute COVID-19, an emerging subset of individuals report a protracted disease course with a range of potential cardiac symptoms persisting for many weeks after the acute illness[19–21]. Limited observational studies have suggested sustained cardiac involvement, based on cardiovascular magnetic resonance (CMR) assessment, after recovery from acute COVID-19[22–25]. However, the lack of baseline CMR data severely limits any causal interpretation of such findings. We therefore investigated whether pre-existing CMR phenotypes were associated with risk of subsequent positive COVID-19 status in the UK Biobank (UKB).

**Methods**

**Setting**

UK Biobank (UKB) is a population study incorporating data from over half a million participants recruited between 2006-2010, designed primarily for study of illnesses of middle and older age. Individuals aged 40-69 years old were identified through National Health Service (NHS) registers and recruited via postal invites. There was detailed baseline characterisation of participants including socio-demographics, lifestyle, health status, and a series of physical measures[26]. Individuals who were unable to consent, or complete baseline assessment due to illness or discomfort, were not recruited. The UKB protocol is publicly available[27]. Linkages with Hospital Episode Statistics (HES) and death registers allow prospective tracking of health outcomes for all participants recorded according to international classification of disease (ICD) codes. UKB has also produced algorithmically defined outcomes for key illnesses, such as acute myocardial infarction (AMI), which integrate data from several sources[28]. The UKB Imaging Study, which includes CMR imaging, aims to scan a random subset of 100,000 participants (>48,000 completed, January 2021)[29]. Linkage with Public Health England permits tracking of COVID-19 test results for UKB participants[30, 31].

**COVID-19 tests**

We included all UKB participants with CMR imaging and COVID-19 testing between 16/03/2020 and 22/08/2020. Testing was based on real-time polymerase chain reaction (RT-PCR) assay antigen testing. For most participants, the sample tested was from combined nose and throat swabs; for patients in critical care lower respiratory samples may have been used. We considered samples labelled as inpatient to be from a hospital setting. Critical care admissions and deaths were defined based on HES and death register data.

**CMR image acquisition and analysis**

CMR imaging was performed in dedicated UKB imaging centres using uniform staff training, equipment, and according to a previously published protocol[32]. Image analysis was performed blind to all participant details using Circle Cardiovascular Imaging 42 post-processing software (Version 5.11, Circle Cardiovascular Imaging Inc., Calgary, Canada). The automated contour detection tool was used for analysis of ventricular volumes, LV mass, and LV strain (tissue tracking). There was visual quality control (QC) of all studies. Those with suboptimal contouring were manually corrected by a single reader. Native T1 was calculated from a manually drawn septal contour on a mid-ventricular short axis slice, excluding cases with poor quality maps or excess septal motion. Aortic distensibility (AoD) was calculated by dividing aortic strain (change in aortic cross-sectional area in diastole to systole) by central pulse pressure (CPP, mmHg)[33]. Aortic areas were derived from transverse cine images of the aorta using an automated tool previously developed and validated in the UKB[34]. The pipeline has inbuilt QC, which assigns a probability to correct detection of aortic areas. We limited analysis to measurements with detection probability >0·75. Aortic contours were used to calculate aortic strain at both the ascending and descending aorta, which was divided by CPP taken from Vicorder® readings at time of imaging to derive AoD at both sites. Thus, the following CMR metrics were considered: LV/RV volumes in end-diastole and end-systole, LV/RV stroke volume, LV/RV ejection fraction, LV mass, mid-ventricular radial strain, mid-ventricular circumferential strain, global longitudinal strain (GLS), torsion, septal native T1, AoD at the ascending and descending aorta.

**Arterial stiffness index**

As an additional measure of arterial health in a larger sample, we considered arterial stiffness index (ASI) derived from finger plethysmography[35]. ASI was measured at the baseline UKB visit using the PulseTrace PCA2 (CareFusion, USA) device according to a pre-defined protocol[36]. Outliers were removed from the ASI variable using a 1·5 interquartile range (IQR) rule, as previously published using this dataset[37].

**Statistical analysis**

Statistical analysis was performed using R Version 3·6·2 and RStudio Version 1·2·5019[38, 39]. Summary descriptive statistics are presented as mean (standard deviation) or median [IQR] depending on distribution. We estimated the association of each cardiovascular phenotype measure with COVID-19 test result (positive vs negative) in logistic regression models with adjustment for age, sex, ethnicity, deprivation, body mass index (BMI), smoking, diabetes, hypertension, high cholesterol, and prior AMI. We present odds ratios with corresponding 95% confidence intervals (CIs) and p-values per unit increase for each cardiac measure. We performed sensitivity analysis limiting to tests performed in a hospital setting (positive n=50). In addition, owing to the relatively small number of positive cases, we re-ran the models using Firth’s penalised logistic regression[40], which produced near-identical results. Within the sample with COVID-19 testing and ASI measurement, we tested association of ASI with death and critical care admission separately in the test positive and negative cohorts with multivariable adjustment as before. The results were near identical in sensitivity analysis with positive cases defined based on RT-PCR and clinical diagnosis of COVID-19 as per HES records (ICD10 code H07.2).

***Ascertainment of covariates***

We took sex as recorded at the baseline UKB visit and age at time of COVID-19 testing. We classified ethnicity into a binary variable of White and BAME (Black, Asian and minority ethnic) ethnicities. UKB records the Townsend score as a measure of material deprivation, with negative scores indicating less deprivation than national averages[41]. BMI was calculated from height and weight recorded at baseline. Smoking status was based on self-report. Hypertension, diabetes, and hypercholesterolaemia were defined through cross-checking across self-report and HES data. A list of International Classification of Disease (ICD) codes used is presented in Supplementary Table 1. Prior AMI was obtained from UKB algorithmically defined health outcomes.

**Ethics**

This study was covered by the ethical approval for UKB studies from the NHS National Research Ethics Service on 17th June 2011 (Ref 11/NW/0382) and extended 10th May 2016 (Ref 16/NW/0274). Access to UKB was granted through access application 2964. All procedures were performed in concordance with the Declaration of Helsinki.

**Results**

**Characteristics of study participants**

There were 315 UKB participants with CMR imaging and COVID-19 testing during the defined study period. The analysis sample comprised 310 participants (n=70 positive) with at least one analysable CMR measure (Figure 1). Of these, 78·7% (n=244) were tested in hospital, 3·5% (n=11) required critical care admission, and 6·1% (n=19) died. Median age was 63.8 [57·5, 72·1] years; 51.0% (n=158) were male (Table 1). Average interval between CMR and COVID-19 testing was 3·0 years. The rates of smoking, diabetes, hypertension, high cholesterol, and previous AMI were 45·2%, 9·4%, 40·6%, 29·4%, and 3·5% respectively (Table 1).

Those with a positive test included greater proportions of men and BAME individuals than those testing negative (Table 1). There were greater number of critical care admissions (5·7% vs 2·9%) and deaths (11·4% vs 4·6%) in the COVID-19 positive group, but this was not statistically significant. Within the COVID-19 positives, those who required critical care admission had higher average rates of all cardiometabolic morbidities, higher BMI, more deprivation, and comprised a greater proportion of men (Supplementary Table 2); those who died also had poorer cardiometabolic profiles and were older than those who survived. Similar, but less pronounced, differences were observed in the COVID-19 negative group (Supplementary Table 2).

ASI was available for 167,423 participants at baseline, of those 6,160 had COVID-19 testing within the study period. After outlier removal (n=94), 6,066 participants had analysable ASI, of whom 667 tested positive and 5,399 tested negative. The baseline characteristics for this sample are summarised in Supplementary Table 3.

**Baseline cardiovascular phenotypes**

Compared to those testing negative, the COVID-19 positive group had, on average, smaller LV end-diastolic volumes, smaller stroke volume, lower ejection fraction, and lower LV mass (Table 2, Figure 2). There was a similar pattern in the RV measures with smaller volumes in end-diastole and end-systole and lower RV stroke volume. COVID-19 positives had, on average, poorer myocardial deformation by all strain measures (circumferential, global longitudinal, radial, torsion). They also had, compared to the COVID-19 negatives, slightly higher average native T1 and higher arterial compliance (higher AoD, lower ASI), however there was near complete overlap of distributions for these variables (Table 2, Figure 2, Supplementary Table 3).

COVID-19 positive individuals who died had significantly lower LV stroke volume, poorer GLS, and lower arterial compliance (lower AoD, higher ASI) compared to those who survived (Supplementary Table 4). Those who died in the COVID-19 negative group did not have statistically different LV measures compared to those who survived, but they did have higher ASI. It is plausible that the associations with adverse CMR phenotypes may be modified with severity of COVID-19. However, we were underpowered to definitively test associations with more granular outcomes, such as death or critical care admission.

**Association of cardiovascular phenotypes with COVID-19 status**

In fully adjusted models, smaller LV and RV volumes in end-diastole, lower LV stroke volume, and poorer myocardial deformation by GLS (higher values) were associated with significantly higher odds of a positive COVID-19 test (Table 3, Figure 3). Associations with other strain measures, torsion, and native T1 were not statistically significant. There were no significant associations between measures of arterial stiffness (AoD, ASI) and COVID-19 status.

These relationships were unchanged when limiting analysis to the subset of participants tested in hospital (n=244, positive n=50) and, additionally, there was a significant association between smaller RV end-systolic volume and higher odds of COVID-19 positivity (Supplementary Table 5). In the sample with ASI measurement and COVID-19 testing (n=6,066), there was no association between ASI and death or critical care admission within the COVID-19 positive or negative cohorts in fully adjusted models (Supplementary Table 6).

**Discussion**

**Summary of findings**

In this sample of 310 UKB participants tested for COVID-19, we demonstrate association of pre-existing adverse morpho-functional CMR phenotypes with greater likelihood of COVID-19 positivity independent of classical cardiovascular risk factors. Specifically, in fully adjusted models, smaller LV and RV end-diastolic volume, lower LV stroke volume, and poorer GLS were associated with significantly higher odds of COVID-19 positive test. There were no significant associations between arterial stiffness (AoD, ASI) or native T1 and COVID-19 status.

**Comparison with existing research**

Our study is the first to assess association of pre-existing cardiovascular phenotypes with subsequent COVID-19 outcomes.

In a cohort of 100 patients recovered from COVID-19, Puntmann et al.[23] report significantly larger LV end-diastolic volumes, lower LV/RV ejection fraction, smaller LV mass, higher native T1, higher native T2, and greater proportion of late gadolinium enhancement (LGE) abnormalities compared to risk factor matched controls. We observed similar significant associations between measures of poorer LV function (lower LV stroke volume, higher GLS) and COVID-19 positivity. However, we found no statistically significant association between native T1 and COVID-19 status in fully adjusted models. It is possible, that associations observed in our study indicate risk factors for symptomatic COVID-19, whilst the findings of Puntmann et al. [23] reflect alterations occurring after COVID-19. These different observations may also reflect different approaches to confounder adjustment. In our study, we consider a wider range of potential confounders, including, ethnicity, BMI, deprivation, and high cholesterol (in addition to all factors matched by Puntmann et al.[23]) . Furthermore, our COVID-19 negative cohort comprise individuals tested for COVID-19, mostly in hospital (80.8%), therefore, these participants are likely to have been admitted with an acute illness, possibly with respiratory symptoms, during the same time period. The control subjects in Puntmann et al.[23] were not hospitalised or tested for COVID-19. Overall, our comparator cohort was more appropriately matched to the COVID-19 positive cases with consideration of a wider range of confounding variables. Therefore, some of the changes observed by Puntmann et al.[23], may be compromised by residual confounding. It is also possible that our study is underpowered to detect small associations with native T1.

Knight et al.[42] present a descriptive report of 29 individuals hospitalised with COVID-19, referred for CMR post-recovery with unexplained myocardial injury during the acute illness. Their participants have more severe presentations of COVID-19 with over a third requiring intensive care ventilatory support. There were non-ischaemic and ischaemic LGE patterns of myocardial injury in 11 and 5 participants respectively; 9 patients had non-specific abnormalities and four patients had dual pathology. There was no evidence of myocardial oedema based on parametric mapping sequences, suggesting that either the observed myocardial alterations are fixed post-COVID-19 changes or that they pre-existed the infection. In a similar study, Huang et al.[25] report CMR findings from patients recovered from COVID-19 with ongoing cardiac symptoms. They present three-way and cross-group comparisons between patients with COVID-19 and positive CMR findings (n=15), COVID-19 and negative CMR findings (n=11), and healthy controls (n=20). They do not report any significant differences in volumetric parameters. They report higher global native T1 in three way comparison and when comparing CMR positives to healthy and CMR negative cases, but not when comparing CMR negatives with healthy controls. The CMR positive group had higher global T2 and extracellular volume (ECV) compared to healthy cohorts. As the researchers effectively pre-select known abnormal cases (CMR positives) to compare with known normal cases (healthy comparators or CMR negatives), it is difficult to reliably compare these findings with those from our study.

Rajpal et al.[24] and Clark et al.[22] report CMR findings of young athletes (mean age 19.5 and 20.0 years respectively) recovered from mild/asymptomatic COVID-19. Rajpal et al.[24] present a descriptive report of 26 college athletes, reporting non-ischaemic patterns of LGE in 12 participants, of whom, four also had elevated T2. Clark et al.[22] compare CMR metrics of 22 college athletes with 22 healthy controls and 22 tactical athletes. The COVID-19 positive group had significantly larger RV volumes and lower RV ejection fraction compared to both control groups. The results of parametric mapping indices showed no significant difference in native T1 between the groups, higher T2 in cases vs healthy controls, and higher ECV in cases vs tactical athletes, but not healthy controls. LGE was seen in two cases, the comparator groups did not have LGE imaging for comparison. It is possible that the observed changes related to athletic cardiac adaptation. Indeed, many of the volumetric and parametric mapping difference reported by Clark et al.[22] were less pronounced when comparing COVID-19 positive athletes to tactical athletic controls than healthy controls. Overall, interpretation of these non-specific CMR findings in these rather atypical populations is challenging, particularly as the participants had no symptoms or biochemical evidence of myocardial injury.

In a large multi-centre survey-based study, Dweck et al.[7] report echocardiographic findings from 1,216 patients with acute COVID-19 and clinical indication for echocardiography performed in hospital. In patients without pre-existing cardiac disease, LV abnormalities were noted in 25% and RV abnormalities in 33%. Those with an abnormal scan were older and had higher prevalence of pre-existing disease. RV abnormalities were more common in patients with more severe COVID-19 and likely reflect elevations in RV afterload due to pulmonary embolism or pneumonia. LV abnormalities were predominantly non-specific, a few cases showed patterns consistent with myocardial infarction (3%), myocarditis (3%), or takotsubo cardiomyopathy (2%). Mahmoud-Elsayed et al.[3] report similar findings in a single centre study of hospitalised patients with COVID-19 pneumonia. In this sample, as per Dweck et al.[7], RV abnormalities dominated, likely secondary to pulmonary pathology rather than cardiac involvement. Indeed, RV systolic dysfunction was significantly associated with elevated D-dimer and C Reactive Protein levels but was not with high sensitivity Troponin I. In this cohort, LV systolic function was hyperdynamic or normal in most cases (89%); 11% had impaired LV systolic function and 4% had a dilated LV. These findings likely reflect changes in cardiac phenotypes occurring secondary to the acute COVID-19 illness, but also, perhaps cardiac features that predispose to symptomatic COVID-19; acute cardiac abnormalities that may have occurred due to COVID-19 appear to be unusual.

**Clinical implications**

Our findings demonstrate association of smaller LV and RV end-diastolic volume, lower LV stroke volume, and poorer GLS were with significantly higher odds of COVID-19 positive test. Overall, this pattern of associations presents the picture of a cardiac phenotype with poorer myocardial contractility and small stiff ventricles. This suggests that adverse cardiac phenotypes, perhaps resembling a heart failure preserved ejection fraction (HFPEF) phenotype, are associated with greater odds of COVID-19. The magnitude of differences observed is small and, at the individual level, their clinical significance is highly uncertain. However, given the massive population burden of COVID-19, small effects in large number of people are likely to be important.

**Strengths and limitations**

In the present study, we used a well-defined cohort, with CMR imaging performed according to uniform technical protocols, linked to national COVID-19 test results. Uniquely, imaging in this study preceded COVID-19 testing by 1-6 years. As a result, our findings add a new and important perspective to existing work, which exclusively relies on imaging performed after COVID-19 testing. The availability of data from a hospitalised COVID-19 negative cohort provided an appropriate comparator cohort. In the present study, case definition was based on SARS-CoV-2 RT-PCR. In practice patients may be assumed to have COVID-19 based on clinical presentation and investigations in the absence of a positive PCR test. However, misclassification of cases in our analysis would lead to a conservative bias if anything, and indeed in a sensitivity analysis including RT-PCR and clinically defined cases, results were unchanged. Within the COVID-19 positive cohort, there were 8 deaths and 4 critical care admissions, as such, we had very limited ability to discern whether baseline cardiac phenotypes were related to disease severity. Future analyses with greater numbers of events will be required to clarify this point. Whilst is it plausible that baseline cardiac phenotype might be associated with risk of symptoms that might occasion COVID-19 testing, it is unlikely that this would influence our findings since UK testing is limited to symptomatic SARS-Cov2 infection (indeed at the time of our analysis predominantly disease severe enough to require hospital admission), and we compared populations according to test result within the tested population. However, our results should be interpreted in the context of the more severe spectrum of COVID-19 manifestations and may not apply across milder or asymptomatic disease manifestations*.* Furthermore, due to the observational nature of the study, we cannot exclude residual confounding or infer causality.

**Conclusions**

Our results, in a predominantly hospitalised cohort, demonstrate that several pre-existing adverse cardiac phenotypes are associated with greater risk of incident COVID-19, suggesting that these phenotypes may be risk factors for, rather than outcomes of, SARS-CoV-2 infection. Thus, observational reports of cardiovascular involvement after COVID-19 may, at least partly, reflect residual confounding and/or reverse causation from pre-existing cardiac status rather than COVID-19 induced alterations. However, whilst volumetric and ventricular function measures appeared dominant in our analysis, differences in tissue characteristics were more pronounced in studies reporting CMR phenotypes after COVID-19 infection. Thus, it is possible that whilst some adverse cardiac phenotypes pre-dispose to more severe COVID-19 and need for hospitalisation, SARS-CoV-2 infection itself might also lead to distinct phenotypic alterations. Further research in larger populations, with appropriate control groups and ideally imaging before and after COVID-19 disease, together with prospective follow-up, are required for definitive conclusions.

**Declarations**

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**Conflicts of interest**

SEP acts as a consultant for and is shareholder of Circle Cardiovascular Imaging, Inc., Calgary, Alberta, Canada. The remaining authors have nothing to disclose

**Availability of data and material**

Data used in this study is available to all bone fide researchers from UK Biobank through a formal access application: <https://www.ukbiobank.ac.uk>.

**Code availability**

Derived data will be returned to UK Biobank and made available to future researchers as per standard UK Biobank data return policy.

**Authors’ contributions**

ZRE, SEP, and NCH conceptualised the study. Image analysis was performed by KF, MYP, JMP, and ZRE. ER contributed to data collation. CM performed and JC verified and advised on the statistical analysis. ZRE wrote the manuscript with critical input from all co-authors and supervision from NCH and SEP.

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**References**

1. Gupta A, Madhavan M V., Sehgal K, et al (2020) Extrapulmonary manifestations of COVID-19. Nat Med 26:1017–1032. https://doi.org/10.1038/s41591-020-0968-3

2. Xu H, Hou K, Xu R, et al (2020) Clinical Characteristics and Risk Factors of Cardiac Involvement in COVID-19. J Am Heart Assoc 9:e016807. https://doi.org/10.1161/JAHA.120.016807

3. Mahmoud-Elsayed HM, Moody WE, Bradlow WM, et al (2020) Echocardiographic Findings in Patients With COVID-19 Pneumonia. Can J Cardiol 36:1203–1207. https://doi.org/10.1016/j.cjca.2020.05.030

4. Lala A, Johnson KW, Januzzi JL, et al (2020) Prevalence and Impact of Myocardial Injury in Patients Hospitalized With COVID-19 Infection. J Am Coll Cardiol 76:533–546. https://doi.org/10.1016/j.jacc.2020.06.007

5. Shi S, Qin M, Shen B, et al (2020) Association of Cardiac Injury with Mortality in Hospitalized Patients with COVID-19 in Wuhan, China. JAMA Cardiol 5:802–810. https://doi.org/10.1001/jamacardio.2020.0950

6. Guo T, Fan Y, Chen M, et al (2020) Cardiovascular Implications of Fatal Outcomes of Patients with Coronavirus Disease 2019 (COVID-19). JAMA Cardiol 5:811–818. https://doi.org/10.1001/jamacardio.2020.1017

7. Dweck MR, Bularga A, Hahn RT, et al (2020) Global evaluation of echocardiography in patients with COVID-19. Eur Heart J Cardiovasc Imaging 21:949–958. https://doi.org/10.1093/ehjci/jeaa178

8. Kim IC, Kim JY, Kim HA, Han S (2020) COVID-19-related myocarditis in a 21-year-old female patient. Eur Heart J 41:1859. https://doi.org/10.1093/eurheartj/ehaa288

9. Madjid M, Safavi-Naeini P, Solomon SD, Vardeny O (2020) Potential Effects of Coronaviruses on the Cardiovascular System: A Review. JAMA Cardiol 10:1–10. https://doi.org/10.1001/jamacardio.2020.1286

10. Libby P, Lüscher T (2020) COVID-19 is, in the end, an endothelial disease. Eur Heart J 41:3038–3044. https://doi.org/10.1093/eurheartj/ehaa623

11. Böhm M, Frey N, Giannitsis · Evangelos, et al (2020) Coronavirus Disease 2019 (COVID-19) and its implications for cardiovascular care: expert document from the German Cardiac Society and the World Heart Federation. Clin Res Cardiol 27:1–14. https://doi.org/10.1007/s00392-020-01656-3

12. Siripanthong B, Nazarian S, Muser D, et al (2020) Recognizing COVID-19-related myocarditis: The possible pathophysiology and proposed guideline for diagnosis and management. Hear Rhythm 17:1463–1470. https://doi.org/10.1016/j.hrthm.2020.05.001

13. Lodigiani C, Iapichino G, Carenzo L, et al (2020) Venous and arterial thromboembolic complications in COVID-19 patients admitted to an academic hospital in Milan, Italy. Thromb Res 191:9–14. https://doi.org/10.1016/j.thromres.2020.04.024

14. Caballeros Lam M, de la Fuente Villena A, Hernández Hernández A, et al (2020) Cardiac magnetic resonance characterization of COVID-19 myocarditis. Rev Española Cardiol (English Ed 73:863–864. https://doi.org/10.1016/j.rec.2020.06.018

15. Hu H, Ma F, Wei X, Fang Y (2020) Coronavirus fulminant myocarditis saved with glucocorticoid and human immunoglobulin. Eur Heart J 1307800. https://doi.org/10.1093/eurheartj/ehaa190

16. Inciardi RM, Lupi L, Zaccone G, et al (2020) Cardiac Involvement in a Patient with Coronavirus Disease 2019 (COVID-19). JAMA Cardiol 5:819–824. https://doi.org/10.1001/jamacardio.2020.1096

17. Bojkova D, Wagner JUG, Shumliakivska M, et al (2020) SARS-CoV-2 infects and induces cytotoxic effects in human cardiomyocytes. bioRxiv. https://doi.org/10.1101/2020.06.01.127605

18. Buja LM, Wolf D, Zhao B, et al (2020) The emerging spectrum of cardiopulmonary pathology of the coronavirus disease 2019 (COVID-19): Report of 3 autopsies from Houston, Texas, and review of autopsy findings from other United States cities. Cardiovasc Pathol 48:107233. https://doi.org/10.1016/j.carpath.2020.107233

19. Greenhalgh T, Knight M, A’Court C, et al (2020) Management of post-acute covid-19 in primary care. BMJ 370:m3026. https://doi.org/10.1136/bmj.m3026

20. Carfì A, Bernabei R, Landi F (2020) Persistent Symptoms in Patients after Acute COVID-19. JAMA - J Am Med Assoc 324:603–605. https://doi.org/10.1001/jama.2020.12603

21. Tenforde MW, Kim SS, Lindsell CJ, et al (2020) Symptom Duration and Risk Factors for Delayed Return to Usual Health Among Outpatients with COVID-19 in a Multistate Health Care Systems Network — United States, March–June 2020. MMWR Morb Mortal Wkly Rep 69:993–998. https://doi.org/10.15585/mmwr.mm6930e1

22. Clark DE, Diamond AB, Fish FA (2020) COVID-19 Myocardial Pathology Evaluated Through scrEening Cardiac Magnetic Resonance (COMPETE CMR). medRxiv Prepr 1–25. https://doi.org/10.1101/2020.08.31.20185140

23. Puntmann VO, Carerj ML, Wieters I, et al (2020) Outcomes of Cardiovascular Magnetic Resonance Imaging in Patients Recently Recovered from Coronavirus Disease 2019 (COVID-19). JAMA Cardiol 2019:1–9. https://doi.org/10.1001/jamacardio.2020.3557

24. Rajpal S, Tong MS, Borchers J, et al (2020) Cardiovascular Magnetic Resonance Findings in Competitive Athletes Recovering from COVID-19 Infection. JAMA Cardiol E1–E3. https://doi.org/10.1001/jamacardio.2020.4916

25. Huang L, Zhao P, Tang D, et al (2020) Cardiac Involvement in Patients Recovered From COVID-2019 Identified Using Magnetic Resonance Imaging. JACC Cardiovasc Imaging. https://doi.org/10.1016/j.jcmg.2020.05.004

26. Raisi-Estabragh Z, Petersen SE (2020) Cardiovascular research highlights from the UK Biobank: opportunities and challenges. Cardiovasc Res 116:e12–e15. https://doi.org/10.1093/cvr/cvz294

27. (2007) UK Biobank: Protocol for a large-scale prospective epidemiological resource. https://www.ukbiobank.ac.uk/wp-content/uploads/2011/11/UK-Biobank-Protocol.pdf. Accessed 13 Dec 2019

28. Schnier C, Bush K, Nolan J, Sudlow C (2017) Definitions of Acute Myocardial Infarction and Main Myocardial Infarction Pathological Types UK Biobank Phase 1 Outcomes Adjudication Documentation on behalf of UK Biobank Outcome Adjudication Group Definitions of Acute Myocardial Infarction. http://biobank.ndph.ox.ac.uk/showcase/showcase/docs/alg\_outcome\_mi.pdf

29. Raisi-estabragh Z, Harvey NC, Neubauer S, Petersen SE (2020) Cardiovascular magnetic resonance imaging in the UK Biobank: a major international health research resource. Eur Hear J - Cardiovasc Imaging 1–8. https://doi.org/10.1093/ehjci/jeaa297

30. Armstrong J, Rudkin JK, Allen N, et al (2020) Dynamic linkage of COVID-19 test results between Public Health England’s Second Generation Surveillance System and UK Biobank. Microb Genomics 6:mgen000397. https://doi.org/10.1099/mgen.0.000397

31. Khanji MY, Aung N, Chahal CAA, Petersen SE (2020) COVID-19 and the UK Biobank—Opportunities and Challenges for Research and Collaboration With Other Large Population Studies. Front Cardiovasc Med 7:156. https://doi.org/10.3389/fcvm.2020.00156

32. Petersen SE, Matthews PM, Francis JM, et al (2015) UK Biobank’s cardiovascular magnetic resonance protocol. J Cardiovasc Magn Reson 18:8. https://doi.org/10.1186/s12968-016-0227-4

33. Pannier BM, Avolio AP, Hoeks A, et al (2000) Methods and Devices for Measuring Arterial Compliance in Humans. Am J Hypertens 15:743–753

34. Biasiolli L, Hann E, Lukaschuk E, et al (2019) Automated localization and quality control of the aorta in cine CMR can significantly accelerate processing of the UK Biobank population data. PLoS One 14:e0212272. https://doi.org/10.1371/journal.pone.0212272

35. Laurent S, Cockcroft J, Van Bortel L, et al (2006) Expert consensus document on arterial stiffness: Methodological issues and clinical applications. Eur Heart J 27:2588–2605. https://doi.org/10.1093/eurheartj/ehl254

36. (2011) UK Biobank Arterial Pulse-Wave Velocity

37. Abdullah Said M, Eppinga RN, Lipsic E, et al (2018) Relationship of arterial stiffness index and pulse pressure with cardiovascular disease and mortality. J Am Heart Assoc 7:e007621. https://doi.org/10.1161/JAHA.117.007621

38. R Core Team (2019). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.

39. RStudio: Integrated Development for R. RStudio, Inc., Boston, MA. https://rstudio.com/. Accessed 18 Oct 2020

40. Firth D (1993) Bias reduction of maximum likelihood estimates. Biometrika 80:27–38. https://doi.org/10.1093/biomet/80.1.27

41. Townsend P, Phillimore P, Beattie A (1988) Health and Deprivation: Inequality and the North. Nurs Stand 2:34–34. https://doi.org/10.7748/ns.2.17.34.s66

42. Knight DS, Kotecha T, Razvi Y, et al (2020) COVID-19 Myocardial Injury in Survivors. Circulation 142:1120–1122. https://doi.org/10.1161/CIRCULATIONAHA.120.049252

**Table 1. Characteristics of study participants**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | Whole sample  n=310 | COVID-19 negatives  n=240 | COVID-19 positives  n=70 | p-value  [test] |
| Age | 63.8 [57.5, 72.1] | 65.1 [58.2, 72.1] | 61.4 [55.5, 72.4] | 0.154 [2] |
| Sex (Male) | 158 (51.0%) | 117 (48.8%) | 41 (58.6%) | 0.190[3] |
| White | 295 (95.2%) | 232 (96.7%) | 63 (90.0%) | 0.049 [3] |
| BAME | 15 (4.8%) | 8 (3.3%) | 7 (10.0%) |  |
| Asian | 8 (2.6%) | 5 (2.1%) | 3 (4.3%) | 0.046 [4] |
| Black | 2 (0.6%) |  | 2 (2.9%) |  |
| Mixed | 3 (1.0%) | 2 (0.8%) | 1 (1.4%) |  |
| Other | 2 (0.6%) | 1 (0.4%) | 1 (1.4%) |  |
| Townsend deprivation score | -2.5 [-3.8, 0.2] | -2.5 [-3.8, 0.0] | -2.4 [-4.1, 0.4] | 0.960 [2] |
| Smoking (current/previous) | 140 (45.2%) | 108 (45.0%) | 32 (45.7%) | 1.000 [3] |
| BMI | 27.4 (± 4.9) | 27.3 (± 4.7) | 27.6 (± 5.6) | 0.629 [1] |
| Diabetes | 29 (9.4%) | 24 (10.0%) | 5 (7.1%) | 0.641 [4] |
| Hypertension | 126 (40.6%) | 101 (42.1%) | 25 (35.7%) | 0.414 [3] |
| High cholesterol | 91 (29.4%) | 70 (29.2%) | 21 (30.0%) | 1.000 [3] |
| Prior myocardial infarction | 11 (3.5%) | 10 (4.2%) | 1 (1.4%) | 0.466 [4] |
| Tested in hospital | 244 (78.7%) | 194 (80.8%) | 50 (71.4%) | 0.127 [3] |
| Critical care admission | 11 (3.5%) | 7 (2.9%) | 4 (5.7%) | 0.276 [4] |
| Death | 19 (6.1%) | 11 (4.6%) | 8 (11.4%) | 0.069 [3] |

**Table 1 caption:** [1] Welch Two Sample t-test (numeric data with unequal variances); [2] Wilcoxon rank sum test with continuity correction (numeric skewed); [3] Two-sample test for equality of proportions with continuity correction (where minimum count > 5); [4] Fisher's Exact Test for Count Data (where minimum count 5). Abbreviations: BAME: Black, Asian, and minority ethnic; BMI: body mass index; COVID-19: coronavirus disease 2019

**Table 2. Summary of Cardiovascular magnetic resonance measures**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | Whole sample  *n*=310 | COVID-19 negatives  *n*=240 | COVID-19 positives  *n*=70 | p-value [test] |
| LVEDVi (ml/m2) | 80.0 (± 14.2) | 80.8 (± 14.3) | 77.1 (± 13.2) | 0.046 [1] |
| LVESVi (ml/m2) | 31.5 (± 8.3) | 31.6 (± 8.4) | 31.4 (± 8.1) | 0.863 [1] |
| LVSVi (ml/m2) | 48.5 (± 9.3) | 49.2 (± 9.4) | 45.7 (± 8.4) | 0.004 [1] |
| LVEF (%) | 60.8 (± 6.5) | 61.1 (± 6.4) | 59.6 (± 6.5) | 0.083 [1] |
| LVMi (g/m2) | 46.7 (± 8.8) | 46.9 (± 8.8) | 45.7 (± 8.4) | 0.318 [1] |
| RVEDVi (ml/m2) | 79.2 (± 15.5) | 79.9 (± 15.6) | 76.9 (± 15.0) | 0.148 [1] |
| RVESVi (ml/m2) | 30.8 (± 8.4) | 31.0 (± 8.5) | 30.0 (± 8.1) | 0.393 [1] |
| RVSVi (ml/m2) | 48.4 (± 10.4) | 48.9 (± 10.5) | 46.8 (± 9.8) | 0.133 [1] |
| RVEF (%) | 61.3 (± 6.6) | 61.3 (± 6.8) | 61.1 (± 6.2) | 0.808 [1] |
| Native T1 (ms) | 923.2 (± 39.1) | 922.9 (± 40.1) | 924.2 (± 35.9) | 0.808 [1] |
| MRS (%) | 35.5 (± 9.1) | 35.6 (± 9.3) | 34.9 (± 8.5) | 0.558 [1] |
| MCS (%) | -20.0 (± 3.2) | -20.1 (± 3.2) | -19.8 (± 3.1) | 0.562 [1] |
| GLS (%) | -15.4 (± 2.6) | -15.5 (± 2.5) | -15.0 (± 2.6) | 0.206 [1] |
| Torsion (degrees) | 0.9 (± 0.8) | 0.9 (± 0.7) | 0.8 (± 1.0) | 0.437 [1] |
| AA AoD (× 10-3 mmHg-1) | 1.4 [0.8, 2.3] | 1.3 [0.8, 2.3] | 1.4 [0.9, 2.3] | 0.665 [2] |
| PDA AoD (× 10-3 mmHg-1) | 2.2 [1.7, 3.1] | 2.2 [1.7, 3.1] | 2.4 [1.6, 3.2] | 0.511 [2] |

**Table 2 caption:** [1] Welch Two Sample t-test (numeric data with unequal variances); [2] Wilcoxon rank sum test with continuity correction (numeric skewed). Abbreviations: AA: ascending aorta; AoD: aortic distensibility; COVID-19: coronavirus disease 2019; LVEDV: left ventricular endo-diastolic volume; LVEF: left ventricular ejection fraction; LVESV: left ventricular endo-systolic volume; LVSV: left ventricular stroke volume; GLS: global longitudinal strain; MCS: circumferential strain at the mid short axis level; MRS: radial strain at the mid short axis level; PDA: proximal descending aorta; RVEDV: right ventricular endo-diastolic volume; RVEF: right ventricular ejection fraction; RVESV: right ventricular end-systolic volume; RVSV: right ventricular stroke volume

**Table 3. Odds ratios from logistic regression models demonstrating association of cardiovascular phenotype measures with COVID-19 status**

|  |  |  |  |
| --- | --- | --- | --- |
|  | Univariate | Age and Sex Adjusted | Fully Adjusted |
| LVEDVi (ml/m2) | 0.98 [0.96, 1.00] | 0.97\* [0.95, 0.99] | 0.97\* [0.95, 1.00] |
|  | 0.057 | 8.50x10-3 | 0.022 |
| LVESVi (ml/m2) | 1.00 [0.96, 1.03] | 0.99 [0.95, 1.02] | 1.00 [0.96, 1.03] |
|  | 0.865 | 0.416 | 0.806 |
| LVSVi (ml/m2) | 0.96\* [0.93, 0.99] | 0.95\* [0.92, 0.98] | 0.95\* [0.92, 0.98] |
|  | 6.4x10-3 | 1.4x10-3 | 2.2x10-3 |
| LVEF (%) | 0.96 [0.93, 1.00] | 0.97 [0.93, 1.01] | 0.96 [0.91, 1.00] |
|  | 0.081 | 0.127 | 0.059 |
| LVMi (g/m2) | 0.98 [0.95, 1.02] | 0.96\* [0.92, 1.00] | 0.96 [0.92, 1.00] |
|  | 0.328 | 0.036 | 0.087 |
| RVEDVi (ml/m2) | 0.99 [0.97, 1.00] | 0.98\* [0.96, 1.00] | 0.98\* [0.96, 1.00] |
|  | 0.155 | 0.023 | 0.042 |
| RVESVi (ml/m2) | 0.99 [0.95, 1.02] | 0.96\* [0.92, 1.00] | 0.97 [0.93, 1.00] |
|  | 0.404 | 0.050 | 0.087 |
| RVSVi (ml/m2) | 0.98 [0.95, 1.01] | 0.97 [0.95, 1.00] | 0.98 [0.95, 1.00] |
|  | 0.146 | 0.065 | 0.093 |
| RVEF (%) | 1.00 [0.96, 1.04] | 1.01 [0.97, 1.06] | 1.01 [0.96, 1.06] |
|  | 0.816 | 0.682 | 0.746 |
| Native T1 (ms) | 1.00 [0.99, 1.01] | 1.00 [1.00, 1.01] | 1.00 [1.00, 1.01] |
|  | 0.818 | 0.536 | 0.483 |
| MRS (%) | 0.99 [0.96, 1.02] | 1.00 [0.97, 1.04] | 1.00 [0.96, 1.03] |
|  | 0.575 | 0.828 | 0.972 |
| MCS (%) | 1.02 [0.94, 1.11] | 0.99 [0.90, 1.09] | 1.01 [0.91, 1.11] |
|  | 0.569 | 0.914 | 0.851 |
| GLS (%) | 1.07 [0.96, 1.19] | 1.10 [0.98, 1.24] | 1.14\* [1.01, 1.29] |
|  | 0.197 | 0.113 | 0.039 |
| Torsion (degrees) | 0.86 [0.62, 1.20] | 0.88 [0.63, 1.24] | 0.92 [0.65, 1.31] |
|  | 0.366 | 0.443 | 0.628 |
| AA AoD (× 10-3 mmHg-1) | 1.06 [0.84, 1.33] | 0.93 [0.68, 1.23] | 0.92 [0.67, 1.24] |
|  | 0.602 | 0.625 | 0.606 |
| PDA AoD (× 10-3 mmHg-1) | 1.12 [0.89, 1.41] | 1.04 [0.79, 1.35] | 1.06 [0.79, 1.39] |
|  | 0.372 | 0.863 | 0.812 |
| ASI (m/s) | 0.99 [0.96, 1.02] | 0.99 [0.96, 1.02] | 0.99 [0.96, 1.02] |
|  | 0.367 | 0.549 | 0.481 |

**Table 3 caption:** Modelling is with sample tested for COVID-19 with analysable CMR data. Model outcome is set as COVID-19 test result (positive vs negative). Fully adjusted model includes adjustment for age, sex, ethnicity, deprivation, body mass index, smoking, diabetes, hypertension, high cholesterol, and prior myocardial infarction. Results are odds ratio [95% confidence interval] and p-value, each belonging to a separate logistic regression model with covariate adjustment as indicated in columns. Abbreviations: AA: ascending aorta; AoD: aortic distensibility; ASI: arterial stiffness index; COVID-19: coronavirus disease 2019; LVEDV: left ventricular endo-diastolic volume; LVEF: left ventricular ejection fraction; LVESV: left ventricular endo-systolic volume; LVSV: left ventricular stroke volume; GLS: global longitudinal strain; MCS: circumferential strain at the mid short axis level; MRS: radial strain at the mid short axis level; PDA: proximal descending aorta; RVEDV: right ventricular endo-diastolic volume; RVEF: right ventricular ejection fraction; RVESV: right ventricular end-systolic volume; RVSV: right ventricular stroke volume.

**Figure 1. Approach to selection of participants for inclusion in the analysis**

UK Biobank participants recruited at baseline (2006-2010)

n=502,506

Completed UK Biobank Imaging study (2014-2020)

n=48,000

Tested for SARS-CoV-2 between 16/3/20 and 22/8/20

n=18,162

CMR scan and SARS-CoV-2 test result data available

n=315

LV/RV volumes/ LV strain

n=309

Exclusions n=6:

missing images n=4 breathing artefact n=1 incorrect planning n=1

Native T1

n=289

Exclusions n=26:

missing images n=16 poor image quality n=5 poor planning n=1

acquisition error n=2 artefact n=2

AoD

n=264

Exclusions n=51:

Pulse wave analysis data not available n= 37

Implausible Vicorder reading n= 5

Not analysable n= 5

Outlier n= 4

Checks for QC and completeness of sequences\*

**Figure 1 caption.** \*There were 310 participants with at least one analysable CMR measure, this included one participant with analysable native T1, but not volumetric images. AoD: aortic distensibility; CMR: cardiovascular magnetic resonance; LV: left ventricle; RV: right ventricle; SARS-CoV-2: severe acute respiratory syndrome coronavirus 2.

**Figure 2. Summary of cardiovascular phenotype measures in the COVID-19 positive and negative groups**

**Graphical user interface

Description automatically generated**

**Figure 2 caption.** AA AoD: aortic distensibility at the ascending aorta; ASI: arterial stiffness index; CMR: cardiovascular magnetic resonance; COVID-19: coronavirus disease 2019; ED: end-diastole; ES: end-systole; GLS: global longitudinal strain; MCS: circumferential strain at the mid short axis level; MRS: radial strain at the mid short axis level; LV: left ventricle; PDA AoD: aortic distensibility at the proximal descending aorta. Intervals for AoD show the 95% confidence interval for the median, all others are 95% confidence interval for the mean.

**Figure 3.** **Odds ratios from fully adjusted multivariable logistic regression models demonstrating association of cardiovascular phenotype measures with COVID-19 status**

**Chart, box and whisker chart

Description automatically generated**

**Figure 3 caption.** Association of each cardiovascular measure with COVID-19 test result (positive vs negative) in multivariable logistic regression models adjusting for age, sex, ethnicity, deprivation, body mass index, smoking, diabetes, hypertension, high cholesterol, and prior myocardial infarction. Results are from individual models and expressed as odds ratio and 95% confidence interval (CI) corresponding to each cardiovascular measure. Green: 95% CI includes 1. Red: 95% CI does not include 1.