

Efficacy of Docosahexaenoic Acid for the Prevention of Necrotizing Enterocolitis in Preterm Infants: A Randomized Clinical Trial

Mariela Bernabe-García¹, Philip C. Calder^{2,3,4*}, Raúl Villegas-Silva⁵, Maricela Rodríguez-Cruz¹, Luis Chávez-Sánchez⁶, Leonardo Cruz-Reynoso⁷, Leovigildo Mateos-Sánchez⁸, Gabriel Lara-Flores⁸, Augusto R. Aguilera-Joaquín⁷ and Luisa Sánchez-García⁷

- ¹ Unidad de Investigación Médica en Nutrición, UMAE Hospital de Pediatría, CMN Siglo XXI, Instituto Mexicano del Seguro Social, México City 06720, México; marielabernabe1@gmail.com (MBG); maricela.rodri-guez.cruz@gmail.com (MRC)
- ² School of Human Development and Health, Faculty of Medicine, University of Southampton, Southampton SO16 6YD, UK; pcc@soton.ac.uk (PCC)
- ³ Institute for Life Sciences, University of Southampton, Southampton SO17 1BJ, UK
- ⁴ NIHR Southampton Biomedical Research Centre, University Hospital Southampton NHS Foundation Trust and University of Southampton, Southampton SO16 6YD, UK
- ⁵ Neonatología, Hospital Infantil de México Federico Gómez, México City 06720, México; raul.villegas-silva@gmail.com (RVS)
- ⁶ Unidad de Investigación Médica en Inmunología, UMAE Hospital de Pediatría, CMN Siglo XXI, Instituto Mexicano del Seguro Social, México City 06720, México; luischz@yahoo.com (LCS)
- ⁷ Unidad de Cuidados Intensivos Neonatales, UMAE Hospital de Gineco-Obstetricia No.3, CMN La Raza, Instituto Mexicano del Seguro Social, México City 02990, México; drleonardocruz@yahoo.com.mx (LCR); draguilera357@gmail.com (ARA); lsanchezg60@hotmail.com (LSG)
- ⁸ Unidad de Cuidados Intensivos Neonatales, UMAE Hospital de Gineco-Obstetricia N° 4 “Luis Castelazo Ayala”, Instituto Mexicano del Seguro Social, México City 01090, México; lmateos95@yahoo.com.mx (LMS); drdebebes@gmail.com (GLF)
- * Correspondence: pcc@soton.ac.uk

Abstract: Necrotizing enterocolitis (NEC) is an inflammatory bowel disease and a leading cause of morbidity and mortality in preterm infants. A randomized double-blind parallel-group (1:1) trial was carried out in two neonatal intensive care units from two tertiary hospitals. Two hundred and twenty-five preterm newborns with expected functional gastrointestinal tract were recruited to receive an enteral dose of 75 mg of DHA/kg body weight or high-oleic sunflower oil daily for 14 days from the first enteral feed after birth. Confirmed NEC was evaluated with Bell’s scale from stage \geq IIa. Two hundred and fourteen randomized infants were analyzed in the intent-to-treat (DHA-group: $n = 105$; control-group: $n = 109$); data for two hundred infants were analysed per-protocol. Confirmed NEC was lower in infants from the DHA-group compared with the control-group (0/100 vs 7/100; $p = 0.007$), RR = 0.93 (95% CI 0.881 to 0.981), risk difference = -7%, (95% CI -12.00 to -1.99), number-needed-to-treat = 15 (95% CI 8.3 to 50). Intent-to-treat analysis showed a lower treatment failure in the DHA-group compared with the control-group (6/105 (6%) vs 16/109 (15%); $p = 0.03$, RR = 0.905, (95% CI 0.826 to 0.991)). Results after multivariate-regression analysis remained significant. Adverse events (apart from incidence of NEC) were not different between groups. A daily dose of DHA for 14 days starting with the first enteral feed may prevent NEC in preterm infants.

Keywords: very low birth weight; infant; prematurity; necrotizing enterocolitis; inflammation; DHA; omega-3; n-3 fatty acids; neonatal intensive care unit; hospital stay

1. Introduction

Necrotizing enterocolitis (NEC) is a multifactorial inflammatory bowel disease. This condition initiates with an unbalanced pro-inflammatory response that rapidly evolves without warning to necrotic bowel and/or death. Most cases occur in preterm infants (birth weight < 1500 g or < 32 weeks of gestational age) [1, 2], and NEC remains a leading cause of morbidity and mortality in neonatal intensive care units (NICUs) worldwide [3]. Moreover, survivors of NEC have long-term complications and high medical costs [4].

The pathophysiology of NEC involves bowel and immune immaturity including scarce mucus, low secretory IgA, poor closure between enterocytes along with an altered intestinal bacterial diversity (dysbiosis) related to antibiotic administration and the type of feeding, among other factors especially in formula-fed infants [5,6]. Once developed, NEC management is through support measures [1]. Thus, strategies to prevent NEC are needed.

Docosahexaenoic acid (DHA), a n-3 long-chain polyunsaturated fatty acid (LC-PUFA), is accreted during the last trimester of gestation [7]. This accretion is cut-off by preterm birth and so preterm infants can become deficient in DHA after birth, even if they receive LC-PUFA supplemented formula [8]. DHA is found in human breast milk, which reduces risk of NEC [7]. A global survey of human breast milk fatty acids using data from 65 studies involving 2474 women reported a mean (SD) DHA content of 0.32 (0.22) percent of total fatty acids with a range of 0.06 to 1.4% of total fatty acids [9]. The DHA content of breast milk changes with duration of lactation and is highest in colostrum [10]. Some, but not all, infant formulas contain DHA, typically at about 0.3% of total fatty acids [11]. DHA has plausible actions to modulate exacerbated inflammatory responses in several neonatal morbidities including NEC [12].

Therefore, the aim of this study was to evaluate the efficacy of the enteral administration of DHA to prevent NEC in preterm infants; considering that preterm formula may be an inflammatory stimulus, the intervention started with the first enteral feed after birth.

2. Materials and Methods

2.1. Study design and participants

A randomized double-blind parallel-group clinical trial was conducted in preterm new-born infants with birthweight ≤ 1500 g but ≥ 1000 g, with an expected functional gastrointestinal tract; infants were recruited between October 2012 to October 2017 from two hospitals affiliated with the Instituto Mexicano del Seguro Social (IMSS) in México City. Infants with congenital malformations, need for major surgery, or periventricular/intraventricular hemorrhage grade \geq II were excluded. At recruitment, none of the mothers were taking n-3 fatty acid supplements or had taken them during pregnancy.

This research was carried out in accordance with The Code of Ethics of the World Medical Association [13] and was approved by the Ethics Committee, National Committee of Scientific Research from IMSS (institutional code CNIC-2012-785-007). The trial was registered at clinicaltrials.gov (NCT01745510) prior to the enrolment of the first participant. Written informed consent was obtained from both parents/guardians (overseen by two witnesses) prior to infant randomization, conforming to the Regulation of the General Law of Health, in matters of Research for Health in Mexico [14].

2.2. Randomization and intervention

Randomization was performed using the Random Allocation Software version 1 for parallel groups, with a 1:1 intervention ratio, block sizes of 10 patients per birthweight (1000-1250 g and from 1251-1500 g) [15]; each hospital had its independent randomization assignment. The intervention groups received a code (A or B) and were packed into opaque envelopes by a researcher who did not participate in the fieldwork; all other clinical and research staff were blind to allocation.

Preterm infants received a daily dose of 75 mg of DHA/kg of body weight (DHA-group) from a DHA-rich algal oil diluted in high-oleic sunflower oil as vehicle (Neuromins® for Kids life's DHA®; DSM Nutritional Products Inc., Parsippany, NJ, USA) or sham oil (control group; high-oleic sunflower oil; PROGELA SA, México City, México) for 14 days, similar in colour and consistency. The fatty acid composition of the oils is shown in **Table 1**.

Table 1. Fatty acid composition of the intervention (DHA-rich) and sham oils (%wt of total fatty acids).

Fatty acid	GROUP	
	DHA Intervention (algal) oil	CONTROL High-oleic sunflower oil
Capric	1.0	0.0
Lauric	4.3	0.0
Myristic	14.0	0.1
Palmitic	12.2	5.2
Palmitoleic	1.9	0.1
Stearic	0.7	3.8
Oleic	18.9	58.9
Linoleic	1.3	29.2
Gamma-Linolenic	0.4	0.3
Alpha-Linolenic	< 0.1	1.2
Eicosenoic	< 0.1	0.3
Arachidonic	< 0.1	0.0
Eicosapentaenoic	< 0.1	0.0
Behenic	0.2	0.7
Docosahexaenoic	44.3	0.0
Nervonic	0.1	0.0

These fatty acids represent more than 99% of the total of fatty acid profile.

Every 3 days, research staff updated infant's weight to modify the dosage. Research staff directly observed administration of the oils, which were administered from the first feed after birth, flushed through an orogastric tube before the milk and/or enteral formula. The DHA dose mimics the high physiological supply of DHA from human milk [9] and has been well tolerated in previous studies in neonates [16, 17]. DHA administration was suspended if any persisting bleeding was identified, if platelet count was < 80,000 mm³ or if the infant was fasting due to acute illness.

2.3. Main outcome measure: Confirmed necrotizing enterocolitis

Confirmed NEC was determined prospectively using Bell's scale modified by Walsh [18]. Briefly, stage Ia to Ib are suspected NEC, stage IIa to IIb are confirmed NEC and, severe/advanced NEC is from stage IIIa with intact bowel and stage IIIb with perforated bowel. The attendant neonatologist identified the concordance of the X rays with systemic and intestinal signs; then a second neonatologist confirmed or discarded the diagnosis. In case of non-concordance, a third neonatologist decided the diagnosis. Confirmed NEC was considered from stage IIa or greater [18].

2.4. Adverse events

Adverse events included platelet count < 80,000 mm³ (collected from routine biochemical analyses), bleeding events such as periventricular/intraventricular haemorrhage grade ≥II, pulmonary and gastric bleeding, and death.

2.5. Clinical course and management 128

At recruitment, infant sex, gestational age, severe asphyxia at birth, being small for gestational age, being a twin or singleton, and severity of disease measured with the Clinical Risk Index for Babies (CRIB) Score [19] were recorded. Clinical course such as presence of patent ductus arteriosus, respiratory distress syndrome, suspected sepsis, apnoea events, as well as medical management such as antibiotics, ibuprofen, omeprazole, red blood cell (RBC) transfusions, and duration of low oxygen saturation ($\text{SpO}_2 < 85\%$) were recorded. 129
130
131
132
133
134
135

Details of nutritional support were collected until hospital discharge. Patients who required total/partial parenteral nutrition (TPN) received 20% soybean oil-based lipid emulsion. Intake of mother's own milk was recorded. Data are presented during the first 2 weeks of the intervention and during total hospital stay; hospital discharge happened when the infant was able to have stable body temperature and after reaching a body weight of at least 1.8 kg. 136
137
138
139
140
141

2.6. Fatty acid profile in erythrocytes and human milk 142

The fatty acid profile of infant erythrocytes was measured. To avoid additional punctures for research purposes, an additional blood sample was collected into EDTA tubes when consultants ordered blood for clinical tests. The method to measure the erythrocyte fatty acid profile by gas chromatography was reported elsewhere [17]. Briefly, after total lipid extraction from erythrocytes by the modified Folch method, fatty acid methyl esters (FAMES) were produced and separated by gas chromatography (Agilent 7820A, Agilent Technologies, Santa Clara, CA, USA). Identification of FAMES was according to retention time from specific FAME standards (Poly Sciences, Niles, IL, USA) and heptadecanoic acid as an internal standard [20]. The fatty acid profile of human milk samples was determined using the same methods. Fatty acids are reported as weight percentage of total fatty acids (%wt of total fatty acids). The fatty acid profile of enteral formula was estimated from the composition reported by manufacturers. 143
144
145
146
147
148
149
150
151
152
153
154

2.7. Sample size and statistical analysis 155

Sample size was estimated with a 2-sided $\alpha = 0.05$ and a power = 80%, where P1 for control = 15% and P2 for DHA intervention = 4%. A sample size of 111 patients/group was obtained. However, a bivariate interim analysis performed after 5 years identified significant differences between groups and recruitment was stopped. 156
157
158
159

IBM SPSS software version 21 (IBM Corp. Armonk, NY, USA) was used for data analysis. Data distribution was inspected. Data are presented as median and interquartile range [quartile 25, quartile 75] or [minimum, maximum] for variables with scarce patients. DHA and control-groups were compared with Mann-Whitney-U test, χ square test, and relative risk (RR) with 95% confidence interval (CI). However, as both groups had similar covariates, the infants in the control-group who developed NEC were also compared as an independent group with the DHA-group and the remainder of the infants in control-group without NEC, using Fisher's Exact, Kruskal-Wallis and Mann-Whitney-U tests. Multivariate analysis with decision trees of CHAID regression was applied to identify the predictors of confirmed NEC. In all cases, a p -value < 0.05 was considered as statistically significant. Those preterm infants who received at least one dose of DHA or sham, were included in the intent-to-treat analysis using χ square test, and RR (95% CI). 160
161
162
163
164
165
166
167
168
169
170
171

3. Results 172

From the two hospitals, 225 preterm infants were recruited and randomized; 214 infants received at least 1 dose of DHA or sham and from them 100 infants per group were analysed per protocol (Figure 1). Of these 200 infants, 179 were recruited at one hospital ($n = 90$ in control group, $n = 89$ in DHA group) and 21 at the other ($n = 10$ in control group, $n = 11$ in DHA group). Baseline characteristics of the infants were comparable between DHA and control groups (Table 2). 173
174
175
176
177
178

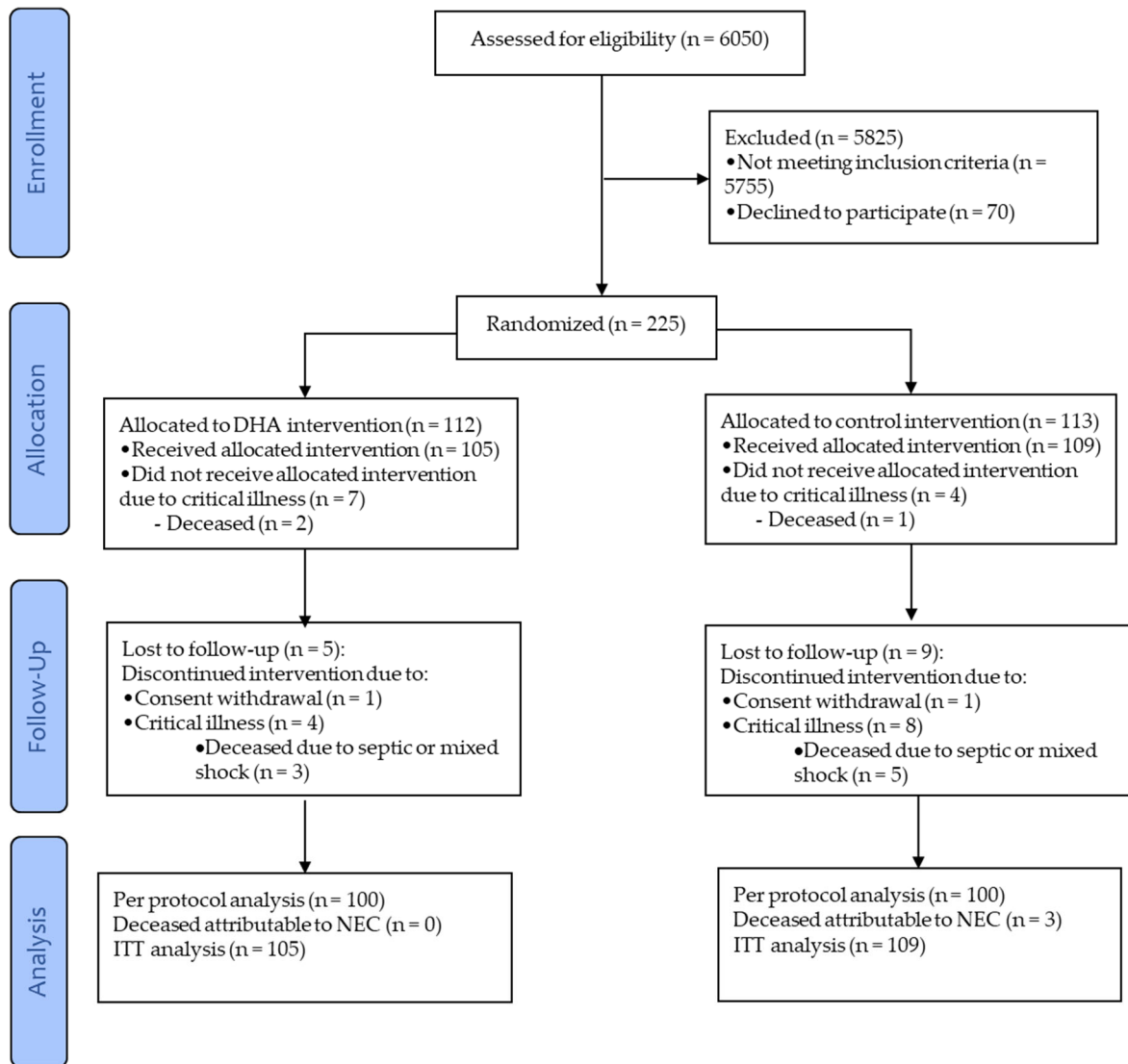


Figure 1. CONSORT diagram depicting the flow of the infants through the study. DHA, Docosahexaenoic acid; NEC, Necrotizing enterocolitis; ITT, Intent-to-treat.

Table 2. Characteristics of the preterm infants.

	GROUP		<i>p</i>
	DHA n = 100	CONTROL n = 100	
<u>At birth</u>			
Born by caesarean section, n (%)	96 (96)	99 (99)	0.391
Received antenatal steroid, n (%)	48 (48)	42 (42)	0.601
Gestational age, weeks	30 [29,32]	31 [30,32]	0.513
Male sex, n (%)	52 (52)	47 (47)	0.572
APGAR score at minute 5 of ≥ 7 , n (%)	96 (96)	95 (95)	1.000
<u>At baseline</u>			
Corrected gestational age, weeks	31.0 [29.8,32.3]	31.1 [28,32.5]	0.306

Data are presented as median [Q25, Q75] unless otherwise stated.

3.1. Confirmed necrotizing enterocolitis

Seven infants developed NEC; all were in the control-group. Therefore, infants with NEC were separated from the rest of the infants in the control-group, but the characteristics remained similar (**Table 3**). All infants who developed NEC had sepsis prior to developing NEC and all were from one of the two hospitals; this was the hospital that recruited the higher number of infants.

Confirmed NEC was lower in infants from the DHA-group compared with the control-group [0/100 vs 7/100, $p = 0.007$; with a RR of 0.93 (95% CI 0.881 to 0.981, $p = 0.008$); a risk difference of -7%, 95% CI (-12.00 to -1.99), and a number needed to treat of 15 (95% CI 8.3 to 50). Among NEC cases, four were female and three were male. The stages of NEC were two cases IIa, three cases IIb, one case IIIa and one case IIIb.

The median [minimum, maximum] postnatal age at diagnosis of confirmed NEC was 26 [6, 29] days. The post-enteral feeding time to diagnosis of confirmed NEC was 4 [0, 23] days.

The analysis per intent-to-treat (ITT) showed a lower treatment failure in the DHA-group compared with the control-group [6/105 (6%) vs 16/109 (15%); $p = 0.031$, with a RR of 0.905 (95% CI 0.826 to 0.991)].

Table 3. Clinical course and medical management during hospital stay of the very low birth weight infants stratified by confirmed NEC.

	GROUP			<i>p</i>
	DHA n = 100	CONTROL n = 93	NEC [‡] n = 7	
Birthweight 1000-1250 g, n (%)	40 (40)	44 (47)	4 (57)	0.460
Birthweight 1251-1500 g, n (%)	60 (60)	49 (53)	3 (43)	
Small for gestational age, n (%)	14 (14)	12 (13)	1 (14)	0.974
Twins, n (%)	23 (23)	30 (32)	2 (29)	0.354
Severe asphyxia at birth, n (%)	9 (9)	10 (11)	0	0.627
Severity of disease (CRIB) at baseline	2.0 [1, 5.0]	1.0 [1, 4.5]	1.0 [0, 6.0]	0.760
Haemoglobin at baseline, g/dl	15.7 [14.2, 17.7]	16.5 [14.8,18.0]	14.1 [12.9, 16.8]	0.150
Respiratory distress syndrome, n (%)	90 (90)	84 (90)	7 (100)	0.681
Suspected or confirmed sepsis, n (%)	66 (66)	57 (61)	7 (100)	0.112
PDA, n (%)	18 (18)	21 (23)	3 (43)	0.259
Apnoea, n (%)	36 (36)	24 (26)	4 (57)	0.110
Apnoea events in week 1, n	1 [1, 2]	1 [1, 2]	1	0.780
Apnoea events in week 2, n	2 [1, 2]	1.5[1, 3]	1[1, 1.5]	0.669
Apnoea events during hospitalization, n	3.0 [2, 5]	3.0 [1, 7]	4.5 [1.3, 8]	0.881
Requirement for phase III ventilator support, n (%)	65 (66)	62 (67)	6 (86)	0.552
Duration with SpO ₂ < 85% in week 1, h	5.5 [2.0, 12.0]	4 [2.0, 6.0]	0	0.219
Duration with SpO ₂ < 85% in week 2, h	7.0 [5.0, 13.5]	4 [3.5, 8.0]	0	0.146
Retinopathy of the prematurity, n (%)	23 (23)	23 (25)	0*	0.325
Antibiotics used, n (%)	97 (97)	92 (99)	7 (100)	0.589
Postnatal age at antibiotic start, h	8 [4, 24]	8 [4, 24]	8 [4, 48]	0.937
Antibiotic duration, days	26 [17, 42]	27 [19, 42]	28 [25, 38]	0.950
Postnatal steroid use, n (%)	21 (21)	21 (23)	3 (43)	0.408
Ibuprofen use, n (%)	9 (9)	6 (7)	1 (14)	0.665
Omeprazole during intervention period, n (%)	43 (43)	41 (46)	2 (29)	0.671
Duration of omeprazole use during intervention period, days	4 [1, 6]	4 [2, 7]	4.5 [3, 6]	0.759
Infants with RBC transfusion during intervention period, n (%)	38 (38)	32 (34)	3 (43)	0.821
Number of RBC transfusions during intervention period, n	1 [1, 1]	1 [1, 1]	1.5 [1, 2]	0.467
Length of NICU stay, days	9.5 [2, 18]	12.0 [3, 21]	18.0 [9, 31]	0.166
Length of hospital stay, days	45 [35, 55]	46 [38, 53]	47 [26, 66]*	0.888

Data are presented median [Q25, Q75] unless otherwise stated. [‡]All infants were from the control group; CRIB: Clinical risk index for babies with weight at born < 1500g; PDA: Persistence of ductus arteriosus; SpO₂: Oxygen saturation measured with pulse oximeter; RBC: Red blood cell; *Not determined in 3 infants due to death.

Multivariate analysis identified that being in the control-group was the strongest predictor for developing NEC (adjusted-*p* = 0.002, **Figure 2**). Among infants in the control-group, the best predictor to develop NEC was gestational age (adjusted-*p* = 0.043, **Figure 2**). The probability to develop NEC was higher in infants older than 29 weeks of gestational age at birth who presented with apnoea (adjusted-*p* = 0.014, **Figure 2**). The intake of any volume of the own mother’s milk was a non-statistically significant, but clinically protective predictor of NEC in the control-group (**Figure 3**).

207
208

209
210
211
212

213
214
215
216
217
218
219

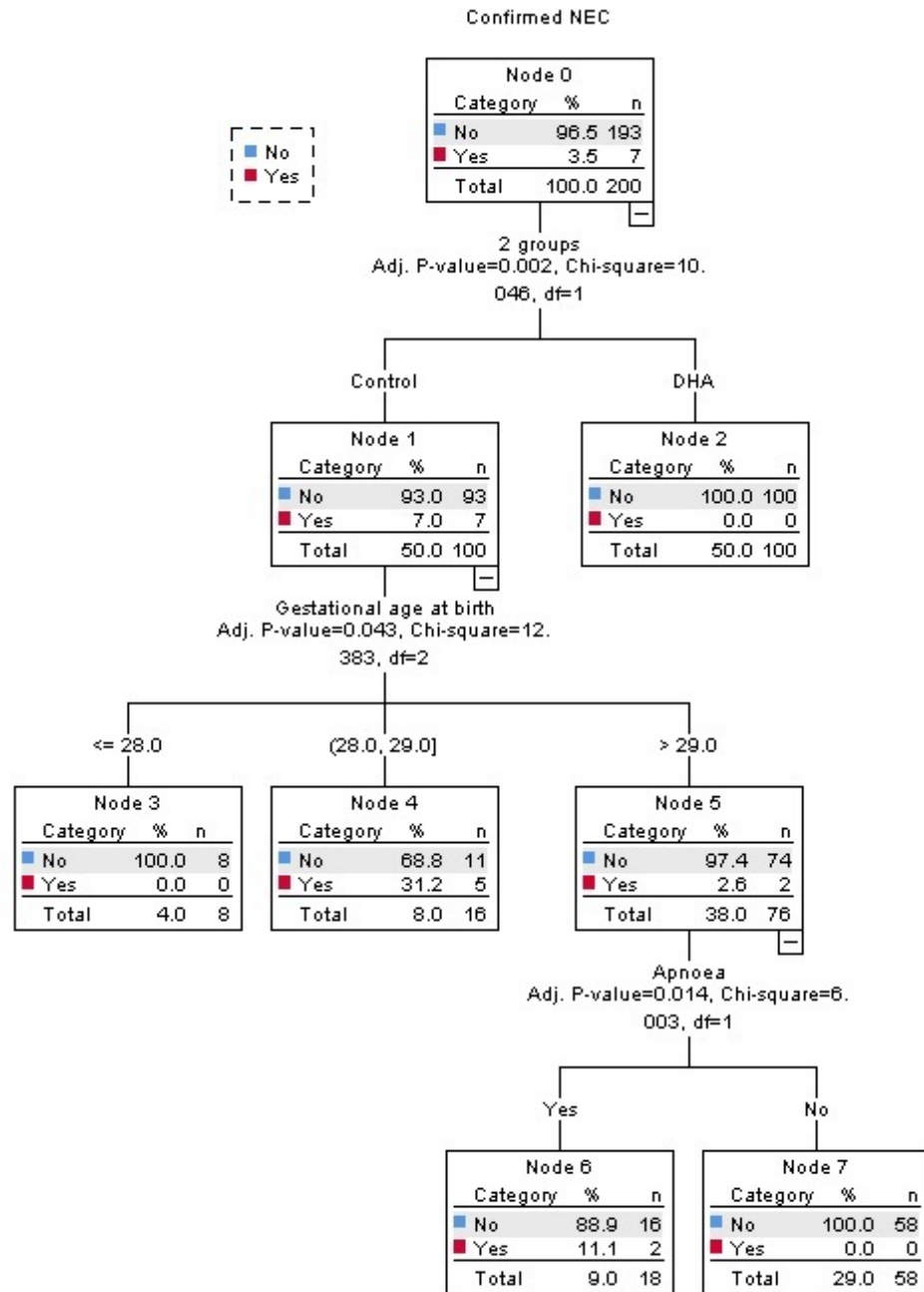


Figure 2. Decision tree from multiple regression for prediction of confirmed necrotizing enterocolitis (NEC). The order of the variables, from top to bottom, shows the ranking for prediction of confirmed NEC; the first was the intervention, the second was gestational age and the last one was presenting apnoea.

220
221
222
223
224

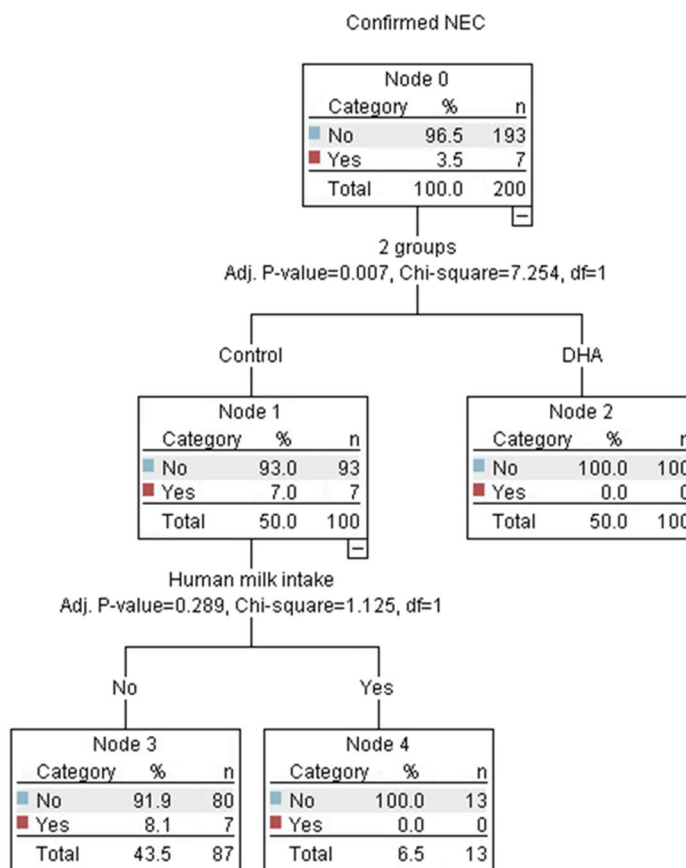


Figure 3. The decision tree from the multiple regression showed that no infants in the control group who received any volume of human milk developed necrotizing enterocolitis (NEC).

3.2. Adverse events

The median [minimum, maximum] of platelet counts was not different between the DHA and control groups at baseline and after the intervention (196,000 mm³ [107,000-464,000] vs. 182,000 [103,000 - 473,000], $p = 0.647$ and 345,000 mm³ [120,000 - 598,000] vs. 347,000 [131,000 - 776,000], $p = 0.884$, respectively). No infant showed a platelet count < 80,000 mm³.

The per protocol analysis showed no difference in death between the DHA and control groups (1/100 compared to 5/100, one-sided hypothesis $p = 0.106$), with a RR of 0.960 for DHA-group (95% CI 0.914 to 1.008). Of the five deaths in control-group, three were attributable to NEC; the death in the DHA-group was not attributable to NEC. Likewise, in the intent to treat analysis, the mortality was not different between the DHA-group and control-group [3/105 vs. 8/109, $p = 0.119$; with a RR = 0.954 for infants in the DHA-group (95% CI 0.896 to 1.015)].

During total hospital stay, there was no difference between the DHA and control groups in the presence of at least one of the following bleeding entities: periventricular/intraventricular haemorrhage grade \geq II, upper gastrointestinal tract and/or pulmonary bleeding (68/100 compared with 73/100, $p = 0.438$); the median of the bleeding events was similar (1 [1, 2] vs. 1 [1, 2], $p = 0.814$). Periventricular/intraventricular haemorrhage grade \geq II was the most common type of bleeding (57% in DHA-group vs. 54% in control-group, $p = 0.669$).

3.3. Clinical course and management.

Covariates from clinical course and management were similar and there was no collinearity among them. Therefore, data are presented stratified by the two treatments and the infants with confirmed NEC as an independent group (Table 3).

Regarding nutritional support, total parenteral nutrition did not contain eicosapentaenoic acid, arachidonic acid (AA) or DHA. The volume of fluids, osmolarity (239 mOsm/L) and the enteral formula were not different among groups. The enteral formula for preterm infants contained 19.4 mg of both DHA and AA per 100 kcal; cumulative intake of DHA and AA from the formula was not different (Table 4). The cumulative intake of DHA and AA estimated from human milk, in those infants receiving human milk during the two weeks of intervention, was a median of 0.31 [0.12 to 1.13] mg/kg/day and 0.84 [0.35 to 2.52] mg/kg/day in DHA-group, while the control-group received 0.25 [0.12 to 0.38] mg/kg/day and 0.35 [0.18 to 0.63] mg/kg/day, respectively. Infants in the DHA-group received an additional 75 mg DHA/kg body weight/day. The infants who developed NEC showed a non-statistical trend to have higher postnatal age when starting enteral feeding and to receive lower advances in enteral feeding (maximum increases do not exceed 25 ml/kg/day), resulting in a statistical trend to lower nutritional support (Table 4). No infant who developed NEC received breastmilk (Table 4), and as consequence, they did not receive fatty acids from human milk (Table 5). Other aspects of nutritional support were not different among groups.

Table 4. Nutritional support of the preterm infants during hospital stay.

Nutritional intake variable	GROUP			p
	DHA n = 100	CONTROL n = 93	NEC [†] n = 7	
<i>Week one of postnatal age</i>				
Total fluid volume, ml/kg/day	100 [88, 108]	97 [88, 105]	89 [75, 104]	0.437
Infants receiving TPN before EF, n (%)	50 (57)	48 (59)	6 (86)	0.327
Postnatal age at starting TPN, days	2.0 [1.7, 2.4]	2.7 [1.0, 2.4]	3.0 [1.8, 4.0]	0.851
Lipid supply by TPN, g/kg/d	2.0 [0.6, 3.8]	2.1 [0.4, 2.9]	2.0 [1.4, 2.4]	0.877
Postnatal age at starting EF, days	4.0 [2.0, 6.5]	4.3 [2.5, 6.2]	7.0 [4.1, 9.1]	0.176
Lipid intake-EFF, g/kg/day	1.1 [0.5, 2.3]	1.2 [0.6, 2.0]	0.5 [0.2, 1.2]	0.151
<i>Fatty acid intake from EFF, mg/kg/day</i>				
Linoleic acid	69 [32, 231]	100 [31, 217]	38 [11, 107]	0.282
Alpha-linolenic acid	26 [10, 44]	23 [10, 39]	9 [4, 22]	0.154
Arachidonic acid	5 [2, 9]	5 [2, 8]	2 [0.8, 5]	0.171
Docosahexaenoic acid	5 [2, 9]	5 [2, 8]	2 [0.8, 5]	0.161
<i>Week two of postnatal age</i>				
Total fluid volume, ml/kg/day	136 [125, 147]	138 [123, 147]	127 [99, 131]	0.081
Lipid supply by TPN, g/kg/d	1.8 [0.8, 2.4]	1.8 [0.8, 2.3]	2.1 [1.4, 2.5]	0.717
Lipid intake-EFF, g/kg/day	4.5 [2.5, 5.8]	4.2 [2.9, 5.7]	1.2 [0.4, 3.9]	0.059
<i>Fatty acid intake from EFF, mg/kg/day</i>				
Linoleic acid	576 [209, 880]	506 [119, 833]	82 [21, 505]	0.106
Alpha-linolenic acid	86 [48, 111]	79 [35, 108]	23 [8, 74]	0.064
Arachidonic acid	17 [10, 22]	16 [7, 22]	7 [1.4, 15]	0.088
Docosahexaenoic acid	17 [10, 22]	16 [7, 22]	7 [1.4, 15]	0.088
<i>Human milk intake</i>				
Infants fed any volume of human milk, n (%)	18 (18)	13 (14)	0	0.382
Infants fed human milk during first week post-enteral feeding, n (%)	11 (11)	4 (4.2)	0	0.157
Intake during first week post-enteral feeding, ml/kg/day	3.9 [2.1, 10.3] ^a	4.9 [0.7, 9.9] ^b	0 ^c	0.003
Infants fed human milk during second week post-enteral feeding, n (%)	11 (11)	6 (6.5)	0	0.376
Intake during second week post-enteral feeding, ml/kg/day	8.3 [2.4, 17.2] ^a	5.4 [1.8, 20.3] ^b	0 ^c	0.052
Infants fed human milk during third week post-EF, n (%)	9 (9)	7 (7.5)	0	0.679
Intake during third week post-enteral feeding, ml/kg/day	6.7 [2.6, 23.1] ^a	7.9 [4.1, 13.6] ^a	0 ^b	0.029

Infants fed human milk during fourth week post-EF, ml/kg/day, n (%)	12 (12)	7 (7.5)	0	0.390
Intake during fourth week post-enteral feeding, ml/kg/day	5.6 [2.6, 12.7] ^a	4.3 [1.5, 6.2] ^b	0 ^c	0.043
Intake during hospital stay, ml/kg/day	14 [7, 37]	12 [3, 60]	0	0.448
Time to reach full enteral feeding, days	15 [12, 22]	17 [12, 22]	20 [16, 46]	0.161

Median [Q25, Q75]; [‡]All infants were from the control group; different superscripts indicate significant differences among groups ($p < 0.05$, Mann-Whitney-U test). TPN: Total Parenteral Nutrition; EF: Enteral feeding; EE: enteral feeding from formula; Note: The calculated intake of DHA is from formula in both groups and excludes the additional supplemental intake of 75 mg/kg/day in the DHA-group.

3.4. Fatty acid profile in erythrocytes and human milk

The fatty acid profile of erythrocytes collected at baseline was similar between groups, except that alpha-linolenic acid was higher in the DHA-group (Table 5). The human milk received by the infants in the DHA-group contained less eicosapentaenoic acid and DHA than the human milk received by the infants in the control-group; the remainder of the fatty acids were similar between human milk of both groups, including AA (Table 5).

Table 5. Fatty acid profile of erythrocyte membranes from preterm infants and of human milk (%weight of total fatty acids).

Fatty acid	DHA n = 100	CONTROL n = 93	NEC n = 7	<i>p</i>	DHA n = 100	CONTROL n = 93	NEC n = 7
In baseline erythrocyte membranes					In human milk during hospital stay		
Lauric	0.34 [0.15, 0.67]	0.38 [0.22, 0.79]	0.36 [0.15, 0.67]	0.534	7.6 [7.0, 9.6]	6.6 [6.6, 7.2]	—*
Myristic	0.71 [0.57, 1.06]	0.80 [0.61, 1.24]	1.02 [0.71, 1.06]	0.252	9.0 [7.7, 9.5]	8.8 [8.2, 9.4]	—*
Palmitic	32.5 [29.9, 40.7]	33.4 [32.2, 42.3]	31.7 [30.3, 48.0]	0.463	22.0 [20.1, 26.9]	24.7 [24.2, 25.2]	—*
Palmitoleic	1.12 [0.78, 1.42]	1.18 [0.80, 1.48]	1.47 [1.0, 2.50]	0.055	2.3 [2.1, 2.6]	2.3 [2.2, 2.4]	—*
Stearic	17.4 [15.9, 18.9]	16.9 [4.3, 23.6]	17.1 [15.8, 19.0]	0.485	6.6 [6.1, 6.7]	7.0 [6.6, 7.3]	—*
Oleic	16.7 [15.2, 18.8]	16.6 [14.6, 19.3]	16.7 [16.0, 18.8]	0.894	33.7 [31.6, 34.9]	32.0 [31.9, 32.9]	—*
Linoleic	5.1 [4.0, 7.4]	5.0 [4.1, 7.3]	3.6 [2.2, 7.3]	0.473	16.1 [15.9, 17.4]	15.5 [14.4, 17.0]	—*
Alpha-linolenic	0.12 ^a [0.09, 0.30]	0.10 ^b [0.07, 0.17]	0.07 ^b [0.07, 0.17]	0.042	1.2 [1.1, 1.5]	1.1 [1.0, 1.2]	—*
Arachidonic	14.67 [5.07, 20.11]	15.50 [5.51, 20.2]	13.5 [4.3, 18.9]	0.829	0.6 [0.5, 0.6]	0.5 [0.5, 0.6]	—*
Eicosapentaenoic	0.54 [0.24, 0.83]	0.53 [0.27, 0.70]	0.40 [0.31, 0.69]	0.782	0.03 ^a [0.03, 0.06]	0.10 ^b [0.09, 0.12]	—*
Nervonic	2.82 [2.0, 3.4]	2.62 [2.11, 3.4]	2.90 [2.0, 3.1]	0.896	0.03 [0.04, 0.05]	0.03 [0.03, 0.04]	—*
Docosahexaenoic	2.92 [1.2, 3.4]	2.99 [0.95, 3.7]	3.0 [2.0, 4.0]	0.812	0.22 ^a [0.16, 0.27]	0.35 ^b [0.33, 0.36]	—*

Median [Q25, Q75]; *no infant who developed NEC received human milk; different superscripts indicate significant differences between groups ($p < 0.05$; Mann-Whitney-U test).

4. Discussion

This trial demonstrates that enteral administration of 75 mg/kg/day of DHA starting at the first enteral feed prevents NEC in preterm infants. The efficacy of DHA remained significant in both intent-to-treat and multivariate analysis. To our knowledge, this is the

first trial to evaluate the efficacy of isolated enteral DHA administration on confirmed NEC.

Our findings are consistent with those of Lu et al., who found reduced NEC incidence (25%) in neonatal rats that received formula with DHA (0.5% of fatty acids) compared with incidence of 35% in those with AA (0.7%) plus DHA (0.5%), and 50% in the control-group [21]. Although that study did not specify whether neonatal rats were preterm, a separate study found similar results in preterm rats: 26% receiving DHA (0.5%) developed NEC, compared with 35% in a group with AA (0.7%) plus DHA (0.5%) and 50% in controls [22]. Those studies suggest a more protective effect of DHA in the absence of AA. Although AA is necessary for growth and for cognitive development [11], it is also considered to have some pro-inflammatory properties [12].

Interestingly, an *in vitro* study using epithelial cells from resected small intestine from a neonate with NEC demonstrated that treatment with DHA significantly decreased interleukin (IL)-1 β induced production of pro-inflammatory cytokines IL-8 and IL-6 compared with controls; AA did not show an effect on those cells [23].

Our findings are to some extent consistent with the results of Carlson et al., who reported reduced incidence of NEC in infants receiving formula with AA (0.41%) plus DHA (0.13%) compared with a control formula; NEC incidence was 2.9% and 17.6%, respectively [24]. However, that study showed a high incidence of NEC. In another study, Innis et al. found 0, 2, and 1 cases of suspected/confirmed NEC in preterm infants receiving AA (0.60%) plus DHA (0.33%), DHA (1%), or a control formula, respectively [25]. In that study NEC was sometimes suspected rather than confirmed and the incidence was low.

A meta-analysis [26] and a single study in preterm infants [27], reported no effect of formulas supplemented with AA plus DHA to prevent NEC. Authors of the meta-analysis reported that studies had small sample sizes and high risk of bias. A large study that administered a dosage of DHA similar to *in utero* accretion (60 mg/kg/day) did not find differences in bronchopulmonary dysplasia (primary outcome) nor NEC [28]. Noteworthy, these studies estimated their power based on primary outcomes other than NEC; thus they may be under-powered to detect an effect on NEC or they may inaccurately evaluate NEC (some studies do not report how NEC was evaluated). The present study had NEC as its primary outcome, selected a population at risk of NEC and was powered accordingly. It is possible that preterm infants from our study may be better responders to DHA compared with infants in other studies. This is speculated because low maternal intake of DHA would result in poor neonatal status: a study from central Mexico showed a low maternal intake of DHA (median 0.11 mg/day) [29]. In the current study, infant erythrocyte DHA averaged ~3% of total fatty acids. This is lower than reported in many other studies. For example, in a recent US study, DHA in erythrocytes from 100 one month old infants ranged from 3.96% to 7.75% [30].

Preterm infants have enterocytes prone to inflammatory responses. This has been explained to be due to exaggerated toll like receptor (TLR) 4 expression and inflammatory signalling and can upregulate platelet-activating factor (PAF) production that plays an important role in pathogenesis of NEC. Preterm infants also exhibit under-expression of the inhibitory sub-unit (I κ B) of the nuclear transcription factor (NF) κ B, which results in easier synthesis of pro-inflammatory cytokines, chemokines, and other inflammatory mediators [2, 31]. Some studies have shown that pro-inflammatory cytokines can weaken intestinal barrier function, escalating inflammation, injury, and gut damage and enabling bacteria from the bowel to enter the circulation [32, 33]. Moreover, monocytes from preterm infants produce lower levels of IL-10 [34], an anti-inflammatory cytokine critical for intestinal homeostasis [5], increasing the risk of an exaggerated bowel inflammation.

There is much evidence that DHA reduces inflammation, preventing gut disruption due to several mechanisms such as decreased NF κ B activation, decreased pro-inflammatory cytokine and PAF synthesis, and increased production of pro-resolving mediators such as resolvins, protectins and maresins, which in turn reduce cytokine production and inflammatory cell recruitment [12, 35].

The dose of DHA used in the current study (75 mg/kg/day) was used peri-operatively in term neonates who underwent cardiovascular surgery [17]. In that study, circulating monocytes showed an early increase of anti-inflammatory cytokine expression (IL-1 receptor antagonist and IL-10) along with a non-significant increase of pro-inflammatory cytokine expression (IL-1 β , TNF- α and IL-6) [17]. Thus, this DHA dosage seems relevant to modify inflammatory processes and switch to a less inflammatory state. It is biologically plausible that administration of DHA in anticipation of an inflammatory challenge, as in the current study, represents a narrow window of opportunity for a protective anti-inflammatory strategy. The approach is feasible and inexpensive; the dose of DHA-containing oil can be adjusted and flushed before the milk or enteral formula with a syringe coupled to a feeding tube.

Regarding covariates of clinical course and medical management, in the current study the second predictor of NEC (after treatment allocation) in multivariate analysis was gestational age, consistent with the developmental window for the susceptibility of NEC onset [6, 36]. The third predictor was apnoea events, which result in intestinal hypoxia/ischemia. Nonetheless, these were significant predictors only in the control-group; there was no predictor of NEC in the DHA group since NEC did not occur in that group (Figure 2). All other potential confounders were not significant and therefore, the regression did not select them.

Human milk from mothers with infants in the control-group presented higher DHA and eicosapentaenoic acid (Table 5), but the estimated cumulative intake of DHA, although negligible, tended to be higher in the DHA-group than in the control-group. This is explained because the average volume of human milk received in DHA-group tended to be higher than in the control-group; also the human milk intake per week did not increase steadily due to critical illness in those infants but was different among groups (Table 4). Therefore, to evaluate the effect of human milk on confirmed NEC, it was intentionally added along with the treatment allocation into the regression model. Although the effect of human milk was not-statistically significant on confirmed NEC, this analysis confirmed a clinically significant effect because it separated those infants from the control-group receiving any volume of human milk and no development of NEC (Right square, Figure 3). This analysis was not possible in the DHA group because this group had no NEC events. It is well known that bioactive factors contained in human milk protect against onset of NEC [37], giving an absolute risk reduction of 4% for any stage of NEC for preterm infants [38]. Interestingly, in the present study the absolute risk reduction with DHA was 7%. Therefore, this intervention should be considered as a strategy for NEC prevention. In this study, preterm birth was often due to maternal critical illness; thus, only 31/200 infants received any volume of human milk, 15/200 received it at least for 2 weeks, and no infant was exclusively-fed with human milk.

A review identified that antenatal steroids, ibuprofen/indomethacin, for treatment of patent ductus arteriosus were protective factors, while lower oxygen saturations (85-89%) increased NEC risk [39]. In the current study, ibuprofen was also used to treat patent ductus arteriosus in 8% of the total studied infants but without differences among groups. Likewise, some causes of ischemic intestinal necrosis such as duration of oxygen saturations < 85% were similar among groups, while major congenital heart disease was an exclusion criterion in the current study [40]. Other known risk factors for NEC were also similar among groups [2, 6], and no patients had haemoglobin \leq 8 g/dl [41].

The percentage of alpha-linolenic acid was higher in erythrocytes from the DHA-group compared with those in the infants from control-group and NEC-group (Table 5). However, this difference is probably of little relevance because alpha-linolenic acid has low anti-inflammatory potential compared with DHA [42, 43].

The current study has several strengths. The trial was blinded, randomized and placebo controlled. NEC was the primary outcome and was confirmed. The trial was amply powered for this outcome. The DHA and control interventions were delivered by clinical staff and administration was supervised by research staff to ensure compliance. Loss to follow-up was limited. Both intent to treat and per protocol analysis were consistent. DHA

dosage was regularly adjusted to maintain similar daily DHA delivery per kilogram of body weight. The two groups had similar characteristics and covariates, possibly due to the randomization in blocks of birth weight and recruiting centre. Unfortunately, nearly 85% of mothers were not able to provide their milk for feeding infants but this allowed to evaluate the DHA effect without the important protective effect of human milk.

There are also some limitations. We did not assess fatty acid status at the end of intervention. We did not assess markers of immune function or inflammation or faecal microbiota. Also, these results are not generalizable to extremely low birth weight infants because the lower limit of birthweight to recruit patients permitted by the Mexican National Committee of Scientific Research was 1 kg. Therefore, whether this intervention is useful to prevent NEC development in preterm infants with lower weight and gestational age at birth needs to be elucidated.

5. Conclusions

A daily enteral dose of DHA for 14 days starting with the first enteral feeding may be a preventive strategy for NEC in preterm infants.

Author Contributions: Conceptualization, M.B.-G. and R.V.-S.; methodology, M.B.-G., P.C.C., R.V.-S., M.R.-C., and L.C.-S.; validation, R.V.-S.; formal analysis, M.B.-G.; investigation, M.B.-G., A.R.A.-J.; resources, M.B.-G., M.R.-C., L.C.-S., L.C.-R., L.M.-S., G.L.-F., L.S.-G.; data curation, including randomization and blinding, M.R.-C.; writing—original draft preparation, M.B.-G., R.V.-S., M.R.-C., P.C.C.; writing—review and editing, P.C.C. and M.R.-C.; supervision, M.B.-G., R.V.-S., L.S.-G.; project administration, M.B.-G.; funding acquisition, M.B.-G., R.V.-S., M.R.-C., L.M.-S. All authors have read and agreed to the published version of the manuscript.

Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsinki and was approved by the Institutional Ethics Committee (National Committee of Scientific Research) of the Instituto Mexicano del Seguro Social (protocol code CNIC-2012-785-007, date of approval February 28th, 2012).

Informed Consent Statement: Informed consent was obtained from parents or tutors of all subjects involved in the study.

Clinical Trial registration ID: NCT01745510. <https://clinicaltrials.gov/ct2/show/NCT01745510>

Funding: This research was funded by Consejo Nacional de Ciencia y Tecnología (CONACYT), México, grant number Salud-2011-01-161643 in support of the research work (to M.B.-G.).

Acknowledgments: We thank the parents who agreed to participate on behalf of their infants. We also thank Drs Mardia Lopez-Alarcón, José Magdaleno-Lara, José Ramón Jimenez, Marisol Millán, José M. González, Jorge I. Gutiérrez, Xóchitl Rodríguez, Elvira Palacios, neonatologists and nursery staff for their contribution and support. We also thank Julio Cesar Flores Castro from Experiencia Analítica, México City, México for statistical advice and support.

Conflicts of Interest: P.C.C. acts as a consultant to BASF AS, Smartfish, DSM, Cargill, Danone/Nutricia and Fresenius-Kabi. All other authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

1. Gupta, A.; Paria, A. Etiology and medical management of NEC. *Early Hum Dev* **2016**, *97*, 17-23, doi:10.1016/j.earlhumdev.2016.03.008.
2. Neu, J.; Walker, W.A. Necrotizing enterocolitis. *New Engl J Med* **2011**, *364*, 255-264, doi:10.1056/NEJMra1005408.
3. Jones, I.H.; Hall, N.J. Contemporary outcomes for infants with necrotizing enterocolitis—a systematic review. *J Pediatr* **2020**, *220*, 86-92 e83, doi:10.1016/j.jpeds.2019.11.011.
4. Bazacliu, C.; Neu, J. Necrotizing enterocolitis: long term complications. *Curr Pediatr Rev* **2019**, *15*, 115-124, doi:10.2174/1573396315666190312093119.
5. Denning, T.L.; Bhatia, A.M.; Kane, A.F.; Patel, R.M.; Denning, P.W. Pathogenesis of NEC: Role of the innate and adaptive immune response. *Sem Perinatol* **2017**, *41*, 15-28, doi:10.1053/j.semperi.2016.09.014.
6. Neu, J.; Pammi, M. Pathogenesis of NEC: Impact of an altered intestinal microbiome. *Sem Perinatol* **2017**, *41*, 29-35, doi:10.1053/j.semperi.2016.09.015.

7. Smith, S.L.; Rouse, C.A. Docosahexaenoic acid and the preterm infant. *Matern Health Neonatol Perinatol* **2017**, *3*, 22, doi:10.1186/s40748-017-0061-1. 450
451
8. Martin, C.R.; Dasilva, D.A.; Cluette-Brown, J.E.; Dimonda, C.; Hamill, A.; Bhutta, A.Q.; Coronel, E.; Wilschanski, M.; Stephens, A.J.; Driscoll, D.F., et al. Decreased postnatal docosahexaenoic and arachidonic acid blood levels in premature infants are associated with neonatal morbidities. *J Pediatr* **2011**, *159*, 743-749 e741-742, doi:10.1016/j.jpeds.2011.04.039. 452
453
454
9. Brenna, J.T.; Varamini, B.; Jensen, R.G.; Diersen-Schade, D.A.; Boettcher, J.A.; Arterburn, L.M. Docosahexaenoic and arachidonic acid concentrations in human breast milk worldwide. *Am J Clin Nutr* **2007**, *85*, 1457-1464. doi: 10.1093/ajcn/85.6.1457. 455
456
10. Moltó-Puigmartí, C.; Castellote, A.I.; Carbonell-Estrany, X.; López-Sabater, M.C. Differences in fat content and fatty acid proportions among colostrum, transitional, and mature milk from women delivering very preterm, preterm, and term infants. *Clin Nutr* **2011**, *30*, 116-123. doi: 10.1016/j.clnu.2010.07.013. 457
458
459
11. Koletzko, B.; Bergmann, K.; Brenna, J.T.; Calder, P.C.; Campoy, C.; Clandinin, M.T.; Colombo, J.; Daly, M.; Decsi, T.; Demmelmair, H.; Domellöf, M.; FidlerMis, N.; Gonzalez-Casanova, I.; van Goudoever, J.B.; Hadjipanayis, A.; Hernell, O.; Lapillonne, A.; Mader, S.; Martin, C.R.; Matthäus, V.; Ramakrishan, U.; Smuts, C.M.; Strain, S.J.J.; Tanjung, C.; Tounian, P.; Carlson, S.E. Should formula for infants provide arachidonic acid along with DHA? A position paper of the European Academy of Paediatrics and the Child Health Foundation. *Am J Clin Nutr* **2020**, *111*, 10-16. doi: 10.1093/ajcn/nqz252. 460
461
462
463
464
12. Frost, B.L.; Caplan, M.S. Can fish oil reduce the incidence of necrotizing enterocolitis by altering the inflammatory response? *Clin Perinatol* **2019**, *46*, 65-75, doi:10.1016/j.clp.2018.09.004. 465
466
13. World Medical Association. World Medical Association Declaration of Helsinki: ethical principles for medical research involving human subjects. *JAMA* **2013**, *310*, 2191-2194, doi:10.1001/jama.2013.281053. 467
468
14. Regulation of the General Law of Health in matters of Research for Health [REGLAMENTO de la Ley General de Salud en Materia de Investigación para la Salud. Diario Oficial de la Federación]. Published February 7, 1984. Available online: <http://www.salud.gob.mx/unidades/cdi/nom/compi/rlgsmis.html> (accessed on 13 January 2021). 469
470
471
15. Saghaei, M. Random allocation software for parallel group randomized trials. *BMC Med Res Methodol* **2004**, *4*, 26, doi:10.1186/1471-2288-4-26. 472
473
16. Bernabe-Garcia, M.; Villegas-Silva, R.; Villavicencio-Torres, A.; Calder, P.C.; Rodriguez-Cruz, M.; Maldonado-Hernandez, J.; Macias-Loaiza, D.; Lopez-Alarcon, M.; Inda-Icaza, P.; Cruz-Reynoso, L. Enteral docosahexaenoic acid and retinopathy of prematurity: a randomized clinical trial. *J Parent Ent Nutr* **2019**, *43*, 874-882, doi:10.1002/jpen.1497. 474
475
476
17. Bernabe-Garcia, M.; Lopez-Alarcon, M.; Villegas-Silva, R.; Mancilla-Ramirez, J.; Rodriguez-Cruz, M.; Maldonado-Hernandez, J.; Chavez-Rueda, K.A.; Blanco-Favela, F.; Espinoza-Garcia, L.; Lagunes-Salazar, S. Beneficial effects of enteral docosahexaenoic acid on the markers of inflammation and clinical outcomes of neonates undergoing cardiovascular surgery: an intervention study. *Ann Nutr Metab* **2016**, *69*, 15-23, doi:10.1159/000447498. 477
478
479
480
18. Walsh, M.C.; Kliegman, R.M. Necrotizing enterocolitis: treatment based on staging criteria. *Pediatr Clin North Am* **1986**, *33*, 179-201, doi:10.1016/s0031-3955(16)34975-6. 481
482
19. The Internatonal Neonatal Network. The CRIB (clinical risk index for babies) score: a tool for assessing initial neonatal risk and comparing performance of neonatal intensive care units. The International Neonatal Network. *Lancet* **1993**, *342*, 193-198. 483
484
20. Evershed, R. Gas chromatography of lipids. In *Lipid analysis. A practical approach*, Hamilton RJ, H.S., Ed. Oxford University Press: Oxford, 1992; pp. 113-151. 485
486
21. Lu, J.; Jilling, T.; Li, D.; Caplan, M.S. Polyunsaturated fatty acid supplementation alters proinflammatory gene expression and reduces the incidence of necrotizing enterocolitis in a neonatal rat model. *Pediatr Res* **2007**, *61*, 427-432, doi:10.1203/pdr.0b013e3180332ca5. 487
488
489
22. Ohtsuka, Y.; Okada, K.; Yamakawa, Y.; Ikuse, T.; Baba, Y.; Inage, E.; Fujii, T.; Izumi, H.; Oshida, K.; Nagata, S., et al. Omega-3 fatty acids attenuate mucosal inflammation in premature rat pups. *J Ped Surg* **2011**, *46*, 489-495, doi:10.1016/j.jpedsurg.2010.07.032. 490
491
492
23. Wijendran, V.; Brenna, J.T.; Wang, D.H.; Zhu, W.; Meng, D.; Ganguli, K.; Kothapalli, K.S.; Requena, P.; Innis, S.; Walker, W.A. Long-chain polyunsaturated fatty acids attenuate the IL-1beta-induced proinflammatory response in human fetal intestinal epithelial cells. *Pediatr Res* **2015**, *78*, 626-633, doi:10.1038/pr.2015.154. 493
494
495
24. Carlson, S.E.; Montalto, M.B.; Ponder, D.L.; Werkman, S.H.; Korones, S.B. Lower incidence of necrotizing enterocolitis in infants fed a preterm formula with egg phospholipids. *Pediatr Res* **1998**, *44*, 491-498, doi:10.1203/00006450-199810000-00005. 496
497
25. Innis, S.M.; Adamkin, D.H.; Hall, R.T.; Kalhan, S.C.; Lair, C.; Lim, M.; Stevens, D.C.; Twist, P.F.; Diersen-Schade, D.A.; Harris, C.L., et al. Docosahexaenoic acid and arachidonic acid enhance growth with no adverse effects in preterm infants fed formula. *J Pediatr* **2002**, *140*, 547-554, doi:10.1067/mpd.2002.123282. 498
499
500
26. Smithers, L.G.; Gibson, R.A.; McPhee, A.; Makrides, M. Effect of long-chain polyunsaturated fatty acid supplementation of preterm infants on disease risk and neurodevelopment: a systematic review of randomized controlled trials. *Am J Clin Nutr* **2008**, *87*, 912-920. 501
502
503
27. Henriksen, C.; Haugholt, K.; Lindgren, M.; Aurvag, A.K.; Ronnestad, A.; Gronn, M.; Solberg, R.; Moen, A.; Nakstad, B.; Berge, R.K., et al. Improved cognitive development among preterm infants attributable to early supplementation of human milk with docosahexaenoic acid and arachidonic acid. *Pediatrics* **2008**, *121*, 1137-1145, doi:10.1542/peds.2007-1511. 504
505
506
28. Collins, C.T.; Makrides, M.; McPhee, A.J.; Sullivan, T.R.; Davis, P.G.; Thio, M.; Simmer, K.; Rajadurai, V.S.; Travadi, J.; Berry, M.J., et al. Docosahexaenoic acid and bronchopulmonary dysplasia in preterm infants. *New Engl J Med* **2017**, *376*, 1245-1255, doi:10.1056/NEJMoa1611942. 507
508
509

29. Parra-Cabrera, S.; Moreno-Macias, H.; Mendez-Ramirez, I.; Schnaas, L.; Romieu, I. Maternal dietary omega fatty acid intake and auditory brainstem-evoked potentials in Mexican infants born at term: cluster analysis. *Early Hum Dev* **2008**, *84*, 51-57, doi:10.1016/j.earlhumdev.2007.03.005. 510
511
30. Farahnak, Z.; Yuan, Y.; Vanstone, C.A.; Weiler, H.A. Maternal and neonatal red blood cell n-3 polyunsaturated fatty acids inversely associate with infant whole-body fat mass assessed by dual-energy X-ray absorptiometry. *Appl Physiol Nutr Metab* **2020**, *45*, 318-326, doi:10.1139/apnm-2019-0311. 513
514
515
31. Hackam, D.J.; Afrazi, A.; Good, M.; Sodhi, C.P. Innate immune signaling in the pathogenesis of necrotizing enterocolitis. *Clin Dev Immunol* **2013**, *2013*, 475415, doi:10.1155/2013/475415. 516
517
32. Bruewer, M.; Luegering, A.; Kucharzik, T.; Parkos, C.A.; Madara, J.L.; Hopkins, A.M.; Nusrat, A. Proinflammatory cytokines disrupt epithelial barrier function by apoptosis-independent mechanisms. *J Immunol* **2003**, *171*, 6164-6172, doi:10.4049/jimmunol.171.11.6164. 518
519
520
33. Halpern, M.D.; Clark, J.A.; Saunders, T.A.; Doelle, S.M.; Hosseini, D.M.; Stagner, A.M.; Dvorak, B. Reduction of experimental necrotizing enterocolitis with anti-TNF-alpha. *Am J Physiol Gastrointest Liver Physiol* **2006**, *290*, G757-764, doi:10.1152/ajpgi.00408.2005. 521
522
523
34. Chheda, S.; Palkowetz, K.H.; Garofalo, R.; Rassin, D.K.; Goldman, A.S. Decreased interleukin-10 production by neonatal monocytes and T cells: relationship to decreased production and expression of tumor necrosis factor-alpha and its receptors. *Pediatr Res* **1996**, *40*, 475-483, doi:10.1203/00006450-199609000-00018. 524
525
526
35. Calder, P.C. n-3 PUFA and inflammation: from membrane to nucleus and from bench to bedside. *Proc Nutr Soc* **2020**, *79*, 404-416. doi:10.1017/S0029665120007077. 527
528
36. Llanos, A.R.; Moss, M.E.; Pinzon, M.C.; Dye, T.; Sinkin, R.A.; Kendig, J.W. Epidemiology of neonatal necrotizing enterocolitis: a population-based study. *Paediatr Perinat Epidemiol* **2002**, *16*, 342-349, doi:10.1046/j.1365-3016.2002.00445.x. 529
530
37. Patel, A.L.; Kim, J.H. Human milk and necrotizing enterocolitis. *Semin Pediatr Surg* **2018**, *27*, 34-38, doi:10.1053/j.sempedsurg.2017.11.007. 531
532
38. Miller, J.; Tonkin, E.; Damarell, R.A.; McPhee, A.J.; Sukanuma, M.; Sukanuma, H.; Middleton, P.F.; Makrides, M.; Collins, C.T. A systematic review and meta-analysis of human milk feeding and morbidity in very low birth weight infants. *Nutrients* **2018**, *10*, 707. doi:10.3390/nu10060707. 533
534
535
39. Xiong, T.; Maheshwari, A.; Neu, J.; Ei-Saie, A.; Pammi, M. An Overview of systematic reviews of randomized-controlled trials for preventing necrotizing enterocolitis in preterm infants. *Neonatology* **2020**, *117*, 46-56, doi:10.1159/000504371. 536
537
40. Spinner, J.A.; Morris, S.A.; Nandi, D.; Costarino, A.T.; Marino, B.S.; Rossano, J.W.; Shamszad, P. Necrotizing enterocolitis and associated mortality in neonates with congenital heart disease: a multi-institutional study. *Pediatr Crit Care Med* **2020**, *21*, 228-234, doi:10.1097/PCC.0000000000002133. 538
539
540
41. Patel, R.M.; Knezevic, A.; Shenvi, N.; Hinkes, M.; Keene, S.; Roback, J.D.; Easley, K.A.; Josephson, C.D. Association of red blood cell transfusion, anemia, and necrotizing enterocolitis in very low-birth-weight infants. *JAMA* **2016**, *315*, 889-897, doi:10.1001/jama.2016.1204. 541
542
543
42. Marion-Letellier, R.; Butler, M.; Dechelotte, P.; Playford, R.J.; Ghosh, S. Comparison of cytokine modulation by natural peroxisome proliferator-activated receptor gamma ligands with synthetic ligands in intestinal-like Caco-2 cells and human dendritic cells—potential for dietary modulation of peroxisome proliferator-activated receptor gamma in intestinal inflammation. *Am J Clin Nutr* **2008**, *87*, 939-948, doi:10.1093/ajcn/87.4.939. 544
545
546
547
43. Baker, E.J.; Valenzuela, C.A.; De Souza, C.O.; Yaqoob, P.; Miles, E.A.; Calder, P.C. Comparative anti-inflammatory effects of plant- and marine-derived omega-3 fatty acids explored in an endothelial cell line. *Biochim Biophys Acta Mol Cell Biol Lipids* **2020**, *1865*, 158662, doi: 10.1016/j.bbalip.2020.158662. 548
549
550