

1 Review: The nutritional management of multiple sclerosis with

2 propionate

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14 1 Abstract

- Over the last 15 years there has been an accumulation of data supporting the concept of a gut-brain
- axis whereby dysbiosis of the gut microbiota can impact neurological function. Such dysbiosis has
- been suggested as a possible environmental exposure triggering multiple sclerosis (MS). Dysbiosis
- has been consistently shown to result in a reduction in short-chain fatty acid (SCFA) producing
- bacteria and a reduction in stool and plasma levels of propionate has been shown for MS patients
- 20 independent of disease stage and in different geographies. A wealth of evidence supports the action
- of propionate on T-cell activity, resulting in decreased T-helper cell 1 (Th1) and T-helper cell 17
- 22 (Th17) numbers/activity and increased regulatory T cell (Treg cell) numbers/activity and an overall
- 23 anti-inflammatory profile. These different T-cell populations play various roles in the
- 24 pathophysiology of MS. A recent clinical study in MS patients demonstrated that supplementation of
- 25 propionate reduces the annual relapse rate and slows disease progression. This review discusses this
- 26 data and the relevant mechanistic background and discusses whether taming of the overactive
- immune system in MS is likely to allow easier bacterial and viral infection.

28 2 Introduction

- 29 Multiple sclerosis (MS) is a chronic, progressive autoimmune disease for which there is no current
- 30 cure. Worldwide, approximately 2.8 million people have MS, making it the most common
- 31 neurological auto-immune disease (1). The development of MS is considered to result from a
- 32 combination of genetic and environmental factors including childhood obesity, smoking, low
- Vitamin D levels and geographical latitude distant from the equator (2-4). More recently, the
- constitution and activity of the intestinal flora (microbiota) has been suggested as one of the
- environmental triggers for the development of MS (3). Here we review the data supporting the role
- 36 the microbiota and short-chain fatty acid (SCFA) metabolites, in particular propionate, play in the

- pathophysiology of MS. Additionally, we discuss the conflicting goals of immunosuppression and
- 38 the need to maintain an appropriate immune response to pathogenic bacteria and viruses.

3 MS and the gut-brain axis

- 40 Evidence of the gross pathophysiological effect of the gut content that may apply to MS has been
- 41 demonstrated through fecal transplant studies. Using a murine model of MS (experimental
- 42 autoimmune encephalomyelitis (EAE)), it was shown that MS score (based on motor deficits)
- worsened when mice received fecal material from patients with MS compared to mice receiving fecal
- 44 material from healthy individuals (5). This study was repeated using twin donors where one was
- 45 healthy and the other with MS. Fecal transplants resulted in a higher frequency of spontaneous EAE
- with the MS-human donor compared to that from the healthy human donor (6).
- 47 A major aspect of microbe–host communication receiving increased attention is the two-way
- 48 communication between the gut microbiota and the central nervous system (CNS), the so-called gut—
- brain axis (7, 8). Gut-to-brain communication can occur via metabolite effects on the blood-brain-
- barrier (BBB), entero-endocrine factors, and systemic immune effects of microbe-derived
- 51 metabolites such as SCFAs, products of tryptophan metabolism, phytoestrogens and bile acid
- metabolites (9).

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- A host of publications have described dysbiosis of gut microbiota in patients with MS and other
- autoimmune diseases compared to healthy controls (5, 6, 10-14). Interestingly, a common finding in
- 55 these reports is that the alpha diversity (the variance within an individual) and beta diversity
- 56 (variance between individuals and cohorts) are unchanged with MS. Rather, the dysbiosis is instead
- 57 manifested as changes in the number of bacteria within the particular family or taxa. Table 1
- summarizes dysbiosis associated with development of MS.

59 Table 1. Comparison of the nature of dysbiosis reported in different studies in patients with

60 MS.

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- One unifying finding throughout studies reporting dysbiosis in patients with MS is the reduction in
- the number of SCFA-producing bacteria (5, 6, 10-12), as reviewed elsewhere (15). This is important
- because it shows that there are common findings on a functional level across studies, despite different
- 64 microbiota profiles presumably due to background genetics, environmental differences and different
- 65 nutritional habits. The reduction in SCFA producing bacteria in patients with MS, as discussed later,
- 66 can have an important physiological impact.

4 Propionate deficiency in MS patients

- Five studies show that patients with MS have low levels of propionate in both feces and plasma (15-
- 69 19). All 5 studies show that propionate levels in both feces and plasma are lower than those of
- healthy controls but there is less certainty for the other SCFAs, acetate and butyrate. Zeng et al.
- 71 reported reduced fecal levels of all SCFAs in Chinese patients with MS compared to healthy controls
- 72 (15). Interestingly the microbiota profile and level of SCFAs in feces were not affected by dietary
- and health habits (e.g., vegetarianism, physical activity, smoking, and alcohol intake), indicating that
- 74 this pattern of dysbiosis may be a result of MS itself.
- 75 Park et al. assessed plasma SCFA levels in US patients with chronic MS (secondary progressive
- disease) and found significant reductions in acetate, propionate and butyrate (16). Duscha et al.
- 77 measured SCFA levels in German patients with relapsing remitting MS (RRMS) and secondary

- 78 progressive MS (SPMS) and found decreased propionate in plasma and feces for both MS subtypes,
- but no differences in butyrate and acetate (17). The findings between the two studies may be
- discordant with regards to acetate and butyrate levels due to the differences in the MS subtypes
- studied. However, both studies support a deficiency in propionate and a sub-analysis by Duscha et al.
- 82 confirms that a propionate deficiency exists in both RRMS and SPMS.
- Takewaki et al. studied 12 patients with RRMS and 9 patients with SPMS and showed reduced
- acetate, propionate and butyrate in the feces of RRMS patients, and a non-statistically significant
- reduction in SPMS patients (20). Table 2 presents the outcomes of SCFA measurements in MS
- 86 studies.

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- 87 Recently, Trend et al. have demonstrated a small but statistically significant reduction in propionate
- amongst patients with MS, without reductions in butyrate and acetate (19).
- 89 Contradictory data has come from the recent study of 58 patients with MS (a mix of RRMS and
- 90 clinically isolated syndrome) and 50 healthy controls. Here, the serum level of acetate was
- 91 significantly lower in MS patients but propionate and butyrate levels were similar in patients with
- 92 MS and healthy controls (21).

94 Table 2. SCFAs in patients with MS

- Across the studies, there is a clear reduction in propionate levels in feces and plasma in patients with
- MS, independent of the subtype of MS and across different populations. These studies therefore
- 97 provide complementary and consistent evidence that patients with MS have a dysbiosis leading to
- 98 reduced numbers of SCFA producing bacteria which results in reduced levels of propionate across
- 99 different geographies and disease forms.

5 Mechanism of immune regulation by propionate

- 101 As well as providing an energy source, SCFAs such as propionate and butyrate exert effects via 2
- major mechanisms: 1) G-protein coupled receptors (GPRs) of the SCFA receptor family namely Free
- Fatty Acid Receptor 2 (FFA2 (formally known as GPR43)) and FFA3 (GPR41) (22, 23), and 2)
- histone deacetylase inhibition (HDACi) (24). A variety of immune cells express FFA2 and FFA3 as
- well as GPR109a for which butyrate is one of the proposed endogenous ligands. In contrast, T-cells
- lack the respective GPRs for mediating SCFA effects and therefore any direct modulation of T-cells
- by SCFAs is likely mediated by histone deacetylase inhibition (25).
- An overview of the transporters and receptors for SCFAs and their distribution is presented in
- 109 Table 3.

110 Table 3: SCFA transporters and receptors and tissue distribution in humans

- 111 FFA2 is expressed on myeloid cells and some granulocytes. SCFAs act via FFA2 to induce the
- chemotaxis of neutrophils (26, 27) and neutrophil degranulation (28, 29).
- Lipopolysaccharide (LPS) activated neutrophils showed diminished production of nitric oxide and
- TNF-α when co-cultured with propionate and both histone deacetylase and NF-kappa B activation
- were inhibited, suggesting their role in propionate-mediated inhibition of inflammation (30).

6 Immune regulation by SCFAs

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- The gut microbiota consists of bacteria, fungi and viruses. The mass of these micro-organisms in an
- adult is typically around 2 kg and could potentially evoke a debilitating and life-threatening immune
- response if left unchecked. In order to maintain a tolerogenic immune response in the gut, in the face
- of this considerable microbial load, communication between the commensal microbial population
- and the body's immune system is essential to maintain immunological homeostasis (8, 9). T-cell
- maturation in the intestinal tract occurs in the gut associated lymphoid tissue (GALT) and from there
- cells migrate to the intraepithelial layer or lamina propria. These T-cells are highly modifiable and
- can be induced to develop into Treg, Th1, Th2, or Th17 cells. These modifications are regulated by
- metabolites such as SCFAs but also by interaction with antigen presenting cells (e.g., dendritic cells
- (DCs)) and intestinal epithelial cells ((31). As discussed later, Treg, and Th17 cells are immune cells
- that play a central role in several auto-immune diseases and normalization of their activity may
- represent an important target for controlling MS.
- Nutritional components and microbial metabolites such as acetate, propionate, butyrate, tryptophan
- and phytoestrogens, act as immune regulators. Numerous studies demonstrate the action of
- propionate in regulating T-cell activity in vitro (32-34), in animal in vivo studies (30, 32, 33, 35), and
- in human studies (17, 36-38). Supplementation with propionate enhances the activity and numbers of
- the anti-inflammatory Treg FoxP3⁺ cells and reduces the activity and numbers of Th17 and to a lesser
- extent Th1 pro-inflammatory T-cells via histone deacetylase inhibition (33, 39-42).
- Dendritic cells are important in the activation of T-cells. Human primary DCs express FFA3 and
- GPR109a but only small amounts of FFA2, thus allowing for regulation by SCFAs. In vitro analysis
- showed that both propionate and butyrate (but not acetate) reduced the DC expression of IL-6, and
- LPS-induced IL-12 and IL-23 (43). This would have a crucial role in reducing pro-inflammatory Th1
- and Th17 populations and allow a shift to anti-inflammatory Treg cells. Additionally, the authors
- demonstrate SCFA specific effects on gene and protein expression of chemokines. Incubation of
- 141 colon cultures from colitic mice with 1 mM SCFAs (representing gut levels) led to a reduction in
- pro-inflammatory chemokines with both propionate and butyrate but not acetate. Propionate reduced
- the expression of chemokine CC ligands (CCL3, CCL5 and CXCL9, CXCL10, CXCL11)
- representing an additional indirect effect of propionate towards infiltration of immune cells.
- Propionate also has a direct effect on the inflammatory activity of non-immune cells shown by in
- 146 *vitro* studies. Following LPS stimulation, NF-kappa B activity and TNF-α release were reduced when
- 147 colon cultures were incubated with propionate or butyrate (44).

7 Modification of the immune system through SCFAs can improve auto-immune disease

- There is evidence that regulation of T-cells by SCFAs and particularly butyrate has a beneficial effect
- on Parkinson's Disease (PD). Dysbiosis and reduced levels of butyrate and propionate have been
- demonstrated in patients with PD (45, 46). Supplemental butyrate reduced the alpha-synuclein
- deposition in gut nerve cells (enteroendocrine cells) (47) and clinical studies are underway to
- determine if SCFAs play a role in reversing the pathology of PD (48).
- 154 Intestinal inflammation such as in ulcerative colitis has been a target condition for microbiota and
- SCFA research. Studies to date mostly describe associations between the disease state, the immune
- inflammatory signature, dysbiosis and reduction in SCFAs (49). Propionate reduced IL-1, IL-6 and
- iNOS production in an *in vitro* model of ulcerative colitis (44) and administration of propionate
- during a 3-week period reduced intestinal inflammation in an animal model (33). In an animal model

- of colitis, colonic SCFA levels were associated positively with Treg activity and inversely with
- disease state (32), and supplemental propionate has been shown to regulate colonic motility (50), and
- intestinal inflammation in animal models of Irritable Bowel Disease (51).
- In patients with ulcerative colitis, butyrate inhibited the pro-inflammatory transcription factor NF-
- kappa B in macrophages and improved disease state as measured by the Disease Activity Index (52).
- Propionate and other SCFAs have been investigated in relation to a number of other auto-immune
- disease states. Animal and in vitro studies have reported an association of propionate with
- metabolism, diabetes and hepatic steatosis (53-63), inflammation (64), (65), (66), (67), (68), (69),
- 167 (34), and colitis inflammation (70).

8 The effect of propionate in animal models of multiple sclerosis

- The EAE animal model is often used for studying certain pathophysiological aspects relevant to MS.
- 170 With this model, orally supplemented propionate was shown to ameliorate disease progression as
- measured by clinical scoring based on muscular function (e.g., tail tonicity, partial or total limb
- paralysis, death) (35, 71, 72). The studies consistently reported an associated increase in Treg cells
- and a decrease in Th17 cells with propionate supplementation. Of particular interest are the results
- 174 from Haghikia et al. (2015) demonstrating that feeding with propionate led to increased Treg cell
- frequency associated with the small intestine; transplantation of these cells in the EAE model
- improved the clinical outcome of the mice showing that gut associated T-cell responses were able to
- have systemic effects.

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178 9 Human intervention studies

- In 2020, Duscha et al. confirmed that dysbiosis occurred in patients with MS and that levels of
- propionate, but not butyrate, were lower in MS patients compared to healthy controls (17). Previous
- 181 work by the same group had shown propionate to be unique amongst SCFAs in its ability to improve
- disease score in the EAE animal model (35).
- The study went on to investigate daily oral propionate supplementation in patients with MS. Patients
- showed significantly lower levels of Treg cells and significantly higher levels of Th17 cells at
- baseline compared to healthy controls representing a pro-inflammatory state for patients with MS.
- After 14 days of daily 1 g propionate supplementation, a significant reduction in Th17 cells and a
- significant increase in number and activity of Treg cells was seen.
- Long term supplementation with 1 g propionate daily was performed in 97 patients with MS (in
- which patients had at least 1-year supplementation). Propionate intake was associated with a
- significant benefit to MS patients as measured by annual relapse rate and expanded disability status
- 191 scale (EDSS) score.

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- The study had some design weaknesses including a non-treated control group rather than use of a
- blinded randomized placebo-controlled design. It is also unclear what medications were used for the
- treatment group vs the control group and whether medications changed during the 3 years of
- propionate supplementation. Despite these weaknesses, the data point to a beneficial effect of
- propionate supplementation in patients with MS.

10 Safety of propionate from an immunological perspective

- 198 Inflammation is a key mechanism in the defense of the body against invading pathogens. Many diets
- and nutrients are claimed to have anti-inflammatory properties with the presumption that this is
- always good. However, there could be concern that an anti-inflammatory agent could suppress
- inflammation needed during infection. At the same time, an uncontrolled inflammatory response is
- associated with chronic diseases such as diabetes, rheumatoid arthritis, allergies and auto-immune
- diseases. The question then is whether immune suppression leads to an open door to bacteria and
- 204 viruses (73).
- Interestingly, Kim et al. (74) showed propionate elicits complex immune regulation dependent on the
- 206 immunological setting. The group demonstrated that *in vitro*, propionate mediated an increase in IL-
- 207 10 production by T-cells in line with its anti-inflammatory properties, and propionate has been shown
- by others to be a central regulator of IL-10 production (75), (17). However, they also found that
- 209 propionate-induced FoxP3⁺ expression (a marker of T reg cells) was dependent on the strength of T-
- cell activation. In conditions of high T-cell activation, SCFAs could suppress FoxP3⁺ cell induction
- whereas at low T-cell activity SCFAs enhanced FoxP3⁺ cell induction (from TGFβ1). This suggests
- that T-cell modulation by propionate and other SCFAs is in accordance with the immunological
- setting the T-cell is in. The group also suggest that SCFAs could facilitate the differentiation of T-
- 214 cells to Th1 and Th17 cells given the right immunological conditions. The authors conclude that
- 215 propionate "aids to promote the right type of T-cells for specific immunological conditions".
- The concept of appropriate T-cell modulation is further supported by detailed work by Bhaskaran et
- al. (76) looking at propionate effects in the light of mucosal infection, where an immune response is
- desired to fight infection but overt inflammation can lead to tissue damage. This is a common
- 219 pathophysiological situation where initial inflammation is triggered in response to a pathogen to
- 220 combat infection, but continued inflammation results in tissue damage. In the case of the study by
- Bhaskaran et al., propionate (and other SCFAs) again increased T reg cells, but during *Candida*
- 222 albicans infection propionate also stimulated levels of Th17 cells and IL-17 and promoted clearance
- of the infection. These results in mice showed propionate improved the immune response against the
- mucosal fungal infection and at the same time promoted resolution of inflammation. Supporting in
- vitro data showed that a direct effect of propionate on Th17 cells led to reduced disease activity;
- however, co-culture with spleen and lymph node cells in a Th17 activated medium led to a switch in
- propionate activity and a promotion of IL-17 production.
- Another example is given by the equine herpesvirus which enters the horse through the upper
- respiratory tract, is spread through infection of leukocytes and T-cells and causes neurological and
- 230 reproductive disorders. Propionate (and other SCFAs) were shown to have a beneficial impact on the
- pathogenesis of the virus through several mechanisms including reduction in viral spreading through
- FFA2- and FFA3-mediated mechanisms (77). Viral spread to endothelial cells from monocytes was
- 233 inhibited via an NF-kappa-B dependent pathway and inhibition of adhesion molecule expression.
- This mechanism may also be active in suppressing the spread of measles and herpes simplex virus
- 235 (77).
- A neutral effect of propionate was demonstrated with studies of the cholera vaccine. Cholera is an
- acute diarrheal disease resulting from bacterial infection. The cholera vaccine uses inactive (killed),
- 238 whole bacteria that interact with antigen-presenting-cells and lymphocytes in the gut lymphoid tissue.
- Results from the study suggest that propionate and acetate have no detrimental effect on the response
- to the vaccine, whereas butyrate may even have beneficial effects (78).
- Some contradictory results however point towards SCFAs having a detrimental effect during an
- infection. A study designed to specifically investigate the role of SCFAs in bacteria-induced

- inflammation was performed by Correa et al. They investigated the activity of neutrophils against a
- bacterial skin infection in an animal model. They showed that SCFAs had no effect on leukocyte
- accumulation but did reduce cytokine production and neutrophil phagocytic capacity suggesting a
- 246 detrimental effect of SCFAs (79)
- 247 The relevance of the findings by Correa et al. can be seen in the light of a similar study by Ciarlo et
- 248 al. (80) where morbidity and mortality were also measured in mice. Ciarlo et al., in agreement with
- 249 Correa et al., demonstrated that propionate led to reduced activity of the innate immune system when
- 250 mouse or human cells were challenged with a range of microbes (Staph. aureus, Strep. pneumoniae,
- 251 E. coli, Klebsiella pneumoniae, Candida albicans). In this study, the production of inflammatory
- 252 cytokines such as IL-6 and IL-12 (but less so for TNF-α) was reduced by propionate in macrophages
- and monocytes and to a lesser extent in DCs. Despite these effects, 3-week supplementation of
- 254 infected mice with propionate had no effect on morbidity or mortality. Furthermore, despite the
- expected increase in Treg FoxP3⁺ cells following propionate treatment, the immunizing effect of a
- primary infection to subsequent infections from the same bacteria was not altered. The authors
- 257 conclude that this was a successful demonstration that anti-inflammatory benefits associated with
- supplemental propionate did not come at a cost of depleted immune defense to pathogens and
- 259 therefore supported the use of supplemental propionate.
- Thus, the available data support a complex interplay between SCFAs and the immune system,
- 261 whereby SCFAs in general, and propionate in particular, have a direct effect on T cell activity
- 262 mediated by histone deacetylase inhibition which can be switched according to immunological
- 263 context (e.g., chronic inflammation versus an infection).
- The aim of this review is to consider the use of propionate in patients with MS in the light of the need
- 265 for a fully functioning immune system. In the case of MS, an immune system running wild needs to
- be tamed, but not to the degree that pathogens cannot be controlled. The data collected to date
- suggest that supplemental propionate can promote a non-inflammatory T-cell profile leading to
- 268 improved clinical outcomes for MS patients and that this occurs without compromising the immune
- response to pathogens.

11 Dietary Management of MS with propionate

- 271 Although pharmacological treatment of MS has progressed significantly over the last decade or so,
- the is a continuing need for improved or novel ways of managing the disease. Most patients with MS
- 273 still experience significant disease progression over time. The emerging importance of gut health and
- 274 the microbiota in MS etiology offers an opportunity for new adjunctive tools in MS management.
- 275 Propionate is classified as a food product in the European Union (81) and the United States (21 CFR
- 276 184.1784) and is therefore considered safe for the general public. In the clinical study by Duscha et
- al. no serious adverse events were reported, and mild gastrointestinal adverse events were reported in
- less than 5% of participants.
- 279 Propionate is included in some food stuffs such as some breads and dairy products for enhancing
- shelf-life, but quantities are not included in product labels and a consumer would be unable to make
- informed dietary changes in order to control propionate intake. Therefore, the management of
- patients can only be through supplementation for which some examples now exist.

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- Gold et al. (82) reported dietary supplementation with 1g propionate daily as adequate for
- 285 restoring plasma propionate concentration and immunological parameters in patients with MS to

- those of healthy individuals. Potential nutritional management of MS patients is described by Duscha
- 287 et al. (17): 2 x 0.5 g capsules were given daily as an adjunct to disease modifying therapy.
- 288 Participants under all MS drug regimens studied showed an increase in Treg cell numbers and
- function (except for where the drug glatiramer acetate was being used). Improved annual relapse
- 290 rates were noted for all treatment groups (including the non-medicated), although participant
- 291 numbers were low and results should be considered with caution.

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How MS disease modifying treatments might affect the level of propionate or its actions has, to the best of our knowledge, not been investigated. The best indication of treatment effects on propionate levels comes from information on effects on microbiota.

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Overall, few studies have been performed assessing the effect of MS-drugs on the microbiota, with inadequate data to draw conclusions. However, results to date suggest some medications may work to aid the levels of propionate. Dimethyl fumarate is an immune modifying treatment in MS with known side effects on gut health. A recent pilot study demonstrated dysbiosis in patients with MS compared with healthy controls and no significant differences were seen from dimethyl fumarate treatment apart from a trend towards increasing propionate (and butyrate) producing Bifidobacteria (83).

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- 305 Microbiota profiling was performed in glatiramer acetate treated MS, dimethyl fumarate treated MS
- and healthy subjects (84) showing a tendency of dimethyl fumarate to enhance numbers of some
- 307 SCFA producing bacteria. In a pilot study, glatiramer acetate treatment reduced the number of
- 308 propionate-forming bacteria in MS patients whilst addition of Vitamin D somewhat restored these
- 309 bacteria (10).
- 310 Given the safety profile of propionate, the potential clinical benefit in MS and its relatively
- inexpensive production, it can be considered that propionate is a good candidate agent for the dietary
- 312 management of MS.
- Thus, the current data suggests that patients with MS, either RRMS or SPMS, may have benefits
- from taking 1g propionate daily as an adjunct to their normal therapy. Studies of MS patients with
- propionate have been performed with patients on Interferon Beta, teriflunomide, glatiramer acetate,
- 316 fingolimod, rituximab, and dimethyl fumarate. The numbers of participants studied by treatment
- regimen are low and the findings should be considered with caution. Adverse events are infrequent
- 318 but may include gastrointestinal events.

12 Conclusion

- The role of propionate in MS is described as a story of dysbiosis, reduction of SCFA producing
- bacteria, reduced levels of plasma propionate coupled with the impact of propionate on T-cells
- important in the pathophysiology of autoimmunity. This broad basis supporting the mechanistic
- action of propionate, has been built up over the last 2 decades and supported by studies in the EAE
- animal model for MS, where disease score is reduced on propionate supplementation. The recent
- publication of a prospective study showing the benefit of propionate supplementation on MS disease
- 326 progression suggests this microbial metabolite may have clinical importance in the management of
- 327 MS and supplementation may be a useful adjunctive tool to current medications.
- 328 The potential use of propionate in MS management is grounded in its activity in regulating T-cell
- profiles and activity. Studies suggest that T-cell modulation is sensitive to the immunological
- challenge in the body, and this is supported by animal studies showing propionate supplementation is

- either neutral or beneficial for host immune activity when tackling bacteria and viruses. Studies
- supporting this claim show outcome data for infections or related inflammatory processes. Whilst the
- data may show a general beneficial or neutral effect of propionate supplementation on immune
- activity, the data is of insufficient volume to give a definitive picture of how propionate
- 335 supplementation can affect the immune system's response to particular pathogens.
- However, the nascent data suggest propionate may be useful in the nutritional management of MS
- 337 (85) and at the same time be neutral or contribute to a normal physiological immune response
- essential for tackling the pathogenic fungi, bacteria and viruses the body is exposed to.
- In conclusion, there is broad mechanistic support for the role of propionate in regulating the immune
- 340 system via modification of T-cell profiles and activity. In the context of auto-immune disease and gut
- regulation of immunity, propionate and other SCFAs are considered as important mediators of the
- 342 gut microbiota. In accordance with this, distal outcomes of auto-immune disease such as seen with
- MS are linked to low levels of propionate due to gut dysbiosis. The use of propionate as a
- 344 supplemental adjunct to current medical treatment has been strengthened by consistent evidence from
- animal models (EAE) and a recently published human intervention trial demonstrating long term
- improvement in disease progression across MS subtypes. Evidence that propionate may also promote
- 347 T-cell activity in the face of infection further supports that propionate may be a safe nutritional
- adjunct to MS treatments.

349 13 References

- 350
- 1. MSIF. Atlas of MS The Multiple Sclerosis International Federation; 2020 September.
- Weng M, Walker WA. The role of gut microbiota in programming the immune phenotype. J Dev Orig
- 353 Health Dis. 2013;4(3):203-14.
- 354 3. Wekerle H. Nature plus Nurture*: The triggering of multiple sclerosis. Swiss medical weekly.
- 355 2015;145:w14189.
- 356 4. Simpson S, Jr., Blizzard L, Otahal P, Van der Mei I, Taylor B. Latitude is significantly associated with
- 357 the prevalence of multiple sclerosis: a meta-analysis. J Neurol Neurosurg Psychiatry. 2011;82(10):1132-41.
- 5. Cekanaviciute E, Yoo BB, Runia TF, Debelius JW, Singh S, Nelson CA, et al. Gut bacteria from multiple
- 359 sclerosis patients modulate human T cells and exacerbate symptoms in mouse models. Proc Natl Acad Sci U S
- 360 A. 2017;114(40):10713-8.
- Berer K, Gerdes LA, Cekanaviciute E, Jia X, Xiao L, Xia Z, et al. Gut microbiota from multiple sclerosis
- patients enables spontaneous autoimmune encephalomyelitis in mice. Proceedings of the National Academy of Sciences. 2017;114(40):10719.
- Tilocca B, Pieroni L, Soggiu A, Britti D, Bonizzi L, Roncada P, et al. Gut-Brain Axis and
- Neurodegeneration: State-of-the-Art of Meta-Omics Sciences for Microbiota Characterization. Int J Mol Sci.
- 366 2020;21(11).
- 367 8. Maslowski KM, Mackay CR. Diet, gut microbiota and immune responses. Nat Immunol. 2011;12(1):5-
- 368 9.
- Wang Y, Kasper LH. The role of microbiome in central nervous system disorders. Brain Behav Immun.
- 370 2014;38:1-12.
- 371 10. Cantarel BL, Waubant E, Chehoud C, Kuczynski J, DeSantis TZ, Warrington J, et al. Gut microbiota in
- 372 multiple sclerosis: possible influence of immunomodulators. J Investig Med. 2015;63(5):729-34.
- 373 11. Chen J, Chia N, Kalari KR, Yao JZ, Novotna M, Paz Soldan MM, et al. Multiple sclerosis patients have a
- distinct gut microbiota compared to healthy controls. Scientific Reports. 2016;6(1):28484.
- Jangi S, Gandhi R, Cox LM, Li N, von Glehn F, Yan R, et al. Alterations of the human gut microbiome in
- 376 multiple sclerosis. Nature Communications. 2016;7(1):12015.

- 377 13. Castillo-Álvarez F, Marzo-Sola ME. Role of intestinal microbiota in the development of multiple
- 378 sclerosis. Neurologia. 2017;32(3):175-84.
- Miyake S, Kim S, Suda W, Oshima K, Nakamura M, Matsuoka T, et al. Dysbiosis in the Gut Microbiota
- of Patients with Multiple Sclerosis, with a Striking Depletion of Species Belonging to Clostridia XIVa and IV
- 381 Clusters. PLOS ONE. 2015;10(9):e0137429.
- 382 15. Zeng Q, Junli G, Liu X, Chen C, Sun X, Li H, et al. Gut dysbiosis and lack of short chain fatty acids in a
- 383 Chinese cohort of patients with multiple sclerosis. Neurochem Int. 2019;129:104468.
- Park J, Wang Q, Wu Q, Mao-Draayer Y, Kim CH. Bidirectional regulatory potentials of short-chain
- fatty acids and their G-protein-coupled receptors in autoimmune neuroinflammation. Scientific Reports.
- 386 2019;9(1):8837.
- Duscha A, Gisevius B, Hirschberg S, Yissachar N, Stangl GI, Eilers E, et al. Propionic Acid Shapes the
- 388 Multiple Sclerosis Disease Course by an Immunomodulatory Mechanism. Cell. 2020;180(6):1067-80.e16.
- Takewaki D, Suda W, Sato W, Takayasu L, Kumar N, Kimura K, et al. Alterations of the gut ecological
- and functional microenvironment in different stages of multiple sclerosis. Proc Natl Acad Sci U S A.
- 391 2020;117(36):22402-12.
- 392 19. Trend S, Leffler J, Jones AP, Cha L, Gorman S, Brown DA, et al. Associations of serum short-chain fatty
- acids with circulating immune cells and serum biomarkers in patients with multiple sclerosis. Sci Rep.
- 394 2021;11(1):5244.
- 395 20. Takewaki D, Suda W, Sato W, Takayasu L, Kumar N, Kimura K, et al. Alterations of the gut ecological
- 396 and functional microenvironment in different stages of multiple sclerosis. Proceedings of the National
- 397 Academy of Sciences. 2020;117(36):22402.
- Olsson A, Gustavsen S, Nguyen TD, Nyman M, Langkilde AR, Hansen TH, et al. Serum Short-Chain
- 399 Fatty Acids and Associations With Inflammation in Newly Diagnosed Patients With Multiple Sclerosis and
- 400 Healthy Controls. Front Immunol. 2021;12:661493.
- 401 22. Nilsson NE, Kotarsky K, Owman C, Olde B. Identification of a free fatty acid receptor, FFA2R,
- 402 expressed on leukocytes and activated by short-chain fatty acids. Biochem Biophys Res Commun.
- 403 2003;303(4):1047-52.
- 404 23. Brown AJ, Goldsworthy SM, Barnes AA, Eilert MM, Tcheang L, Daniels D, et al. The Orphan G protein-
- 405 coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. J
- 406 Biol Chem. 2003;278(13):11312-9.
- 407 24. Sealy L, Chalkley R. The effect of sodium butyrate on histone modification. Cell. 1978;14(1):115-21.
- 408 25. Park J, Kim M, Kang SG, Jannasch AH, Cooper B, Patterson J, et al. Short-chain fatty acids induce both
- 409 effector and regulatory T cells by suppression of histone deacetylases and regulation of the mTOR-S6K
- 410 pathway. Mucosal Immunol. 2015;8(1):80-93.
- 411 26. Le Poul E, Loison C, Struyf S, Springael JY, Lannoy V, Decobecq ME, et al. Functional characterization
- of human receptors for short chain fatty acids and their role in polymorphonuclear cell activation. J Biol
- 413 Chem. 2003;278(28):25481-9.
- 414 27. Vinolo MA, Ferguson GJ, Kulkarni S, Damoulakis G, Anderson K, Bohlooly YM, et al. SCFAs induce
- 415 mouse neutrophil chemotaxis through the GPR43 receptor. PLoS One. 2011;6(6):e21205.
- 416 28. Eftimiadi C, Buzzi E, Tonetti M, Buffa P, Buffa D, van Steenbergen MTJ, et al. Short-chain fatty acids
- 417 produced by anaerobic bacteria alter the physiological responses of human neutrophils to chemotactic
- 418 peptide. Journal of Infection. 1987;14(1):43-53.
- 419 29. Carretta MD, Conejeros I, Hidalgo MA, Burgos RA. Propionate induces the release of granules from
- 420 bovine neutrophils. Journal of Dairy Science. 2013;96(4):2507-20.
- 421 30. Vinolo MA, Rodrigues HG, Hatanaka E, Sato FT, Sampaio SC, Curi R. Suppressive effect of short-chain
- fatty acids on production of proinflammatory mediators by neutrophils. J Nutr Biochem. 2011;22(9):849-55.
- 423 31. Ma H, Tao W, Zhu S. T lymphocytes in the intestinal mucosa: defense and tolerance. Cell Mol
- 424 Immunol. 2019;16(3):216-24.
- 425 32. Furusawa Y, Obata Y, Fukuda S, Endo TA, Nakato G, Takahashi D, et al. Commensal microbe-derived
- butyrate induces the differentiation of colonic regulatory T cells. Nature. 2013;504(7480):446-50.

- 427 33. Smith PM, Howitt MR, Panikov N, Michaud M, Gallini CA, Bohlooly YM, et al. The microbial
- 428 metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. Science. 2013;341(6145):569-73.
- 429 34. Tayyeb JZ, Popeijus HE, Mensink RP, Konings M, Mokhtar FBA, Plat J. Short-Chain Fatty Acids (Except
- 430 Hexanoic Acid) Lower NF-kB Transactivation, Which Rescues Inflammation-Induced Decreased
- 431 Apolipoprotein A-I Transcription in HepG2 Cells. Int J Mol Sci. 2020;21(14).
- 432 35. Haghikia A, Jörg S, Duscha A, Berg J, Manzel A, Waschbisch A, et al. Dietary Fatty Acids Directly
- 433 Impact Central Nervous System Autoimmunity via the Small Intestine. Immunity. 2015;43(4):817-29.
- 434 36. Marzocco S, Fazeli G, Di Micco L, Autore G, Adesso S, Dal Piaz F, et al. Supplementation of Short-
- Chain Fatty Acid, Sodium Propionate, in Patients on Maintenance Hemodialysis: Beneficial Effects on
- 436 Inflammatory Parameters and Gut-Derived Uremic Toxins, A Pilot Study (PLAN Study). J Clin Med. 2018;7(10).
- 437 37. Meyer F, Seibert FS, Nienen M, Welzel M, Beisser D, Bauer F, et al. Propionate supplementation
- promotes the expansion of peripheral regulatory T-Cells in patients with end-stage renal disease. J Nephrol.
- 439 2020;33(4):817-27.
- 440 38. Su X, Yin X, Liu Y, Yan X, Zhang S, Wang X, et al. Gut Dysbiosis Contributes to the Imbalance of Treg
- and Th17 Cells in Graves' Disease Patients by Propionic Acid. J Clin Endocrinol Metab. 2020;105(11).
- Tao R, de Zoeten EF, Ozkaynak E, Chen C, Wang L, Porrett PM, et al. Deacetylase inhibition promotes
- the generation and function of regulatory T cells. Nat Med. 2007;13(11):1299-307.
- 444 40. Arpaia N, Campbell C, Fan X, Dikiy S, van der Veeken J, deRoos P, et al. Metabolites produced by
- commensal bacteria promote peripheral regulatory T-cell generation. Nature. 2013;504(7480):451-5.
- 446 41. Tan J, McKenzie C, Potamitis M, Thorburn AN, Mackay CR, Macia L. The role of short-chain fatty acids
- 447 in health and disease. Adv Immunol. 2014;121:91-119.
- 448 42. Park J, Kim M, Kang SG, Jannasch AH, Cooper B, Patterson J, et al. Short-chain fatty acids induce both
- effector and regulatory T cells by suppression of histone deacetylases and regulation of the mTOR–S6K
- 450 pathway. Mucosal Immunology. 2015;8(1):80-93.
- 451 43. Nastasi C, Candela M, Bonefeld CM, Geisler C, Hansen M, Krejsgaard T, et al. The effect of short-chain
- 452 fatty acids on human monocyte-derived dendritic cells. Scientific Reports. 2015;5(1):16148.
- 453 44. Tedelind S, Westberg F, Kjerrulf M, Vidal A. Anti-inflammatory properties of the short-chain fatty
- acids acetate and propionate: a study with relevance to inflammatory bowel disease. World J Gastroenterol.
- 455 2007;13(20):2826-32.
- 456 45. Unger MM, Spiegel J, Dillmann KU, Grundmann D, Philippeit H, Bürmann J, et al. Short chain fatty
- acids and gut microbiota differ between patients with Parkinson's disease and age-matched controls.
- 458 Parkinsonism Relat Disord. 2016:32:66-72.
- 459 46. Vascellari S, Palmas V, Melis M, Pisanu S, Cusano R, Uva P, et al. Gut Microbiota and Metabolome
- 460 Alterations Associated with Parkinson's Disease. mSystems. 2020;5(5).
- 461 47. Qiao CM, Sun MF, Jia XB, Shi Y, Zhang BP, Zhou ZL, et al. Sodium butyrate causes α-synuclein
- degradation by an Atg5-dependent and PI3K/Akt/mTOR-related autophagy pathway. Exp Cell Res.
- 463 2020;387(1):111772.
- 464 48. Effects of Resistant Starch on Bowel Habits, Fecal Short Chain Fatty Acids and Gut Microbiota in
- Parkinson Disease (RESISTA-PD) [Internet]. US National Library of Medicine. 2020 [cited November 25, 2020].
- 466 Available from: https://clinicaltrials.gov/ct2/show/NCT02784145.
- 467 49. Russo E, Giudici F, Fiorindi C, Ficari F, Scaringi S, Amedei A. Immunomodulating Activity and
- Therapeutic Effects of Short Chain Fatty Acids and Tryptophan Post-biotics in Inflammatory Bowel Disease.
- 469 Frontiers in immunology. 2019;10:2754-.
- 470 50. Tazoe H, Otomo Y, Kaji I, Tanaka R, Karaki SI, Kuwahara A. Roles of short-chain fatty acids receptors,
- 471 GPR41 and GPR43 on colonic functions. J Physiol Pharmacol. 2008;59 Suppl 2:251-62.
- 472 51. Sun M, Wu W, Liu Z, Cong Y. Microbiota metabolite short chain fatty acids, GPCR, and inflammatory
- bowel diseases. Journal of gastroenterology. 2017;52(1):1-8.
- Lührs H, Gerke T, Müller JG, Melcher R, Schauber J, Boxberge F, et al. Butyrate inhibits NF-kappaB
- activation in lamina propria macrophages of patients with ulcerative colitis. Scand J Gastroenterol.
- 476 2002;37(4):458-66.

- 53. Deng M, Qu F, Chen L, Liu C, Zhang M, Ren F, et al. SCFAs alleviated steatosis and inflammation in
- 478 mice with NASH induced by MCD. J Endocrinol. 2020;245(3):425-37.
- Jiao A, Yu B, He J, Yu J, Zheng P, Luo Y, et al. Short chain fatty acids could prevent fat deposition in
- pigs via regulating related hormones and genes. Food Funct. 2020;11(2):1845-55.
- 481 55. Yu K, Zhang Y, Chen H, Zhu W. Hepatic Metabolomic and Transcriptomic Responses Induced by Cecal
- 482 Infusion of Sodium Propionate in a Fistula Pig Model. J Agric Food Chem. 2019;67(47):13073-81.
- 483 56. Song B, Zhong YZ, Zheng CB, Li FN, Duan YH, Deng JP. Propionate alleviates high-fat diet-induced lipid
- dysmetabolism by modulating gut microbiota in mice. J Appl Microbiol. 2019;127(5):1546-55.
- Wu Y, Ma N, Song P, He T, Levesque C, Bai Y, et al. Grape Seed Proanthocyanidin Affects Lipid
- 486 Metabolism via Changing Gut Microflora and Enhancing Propionate Production in Weaned Pigs. J Nutr.
- 487 2019;149(9):1523-32.
- 58. Duan Y, Zhong Y, Xiao H, Zheng C, Song B, Wang W, et al. Gut microbiota mediates the protective
- effects of dietary β -hydroxy- β -methylbutyrate (HMB) against obesity induced by high-fat diets. Faseb j.
- 490 2019;33(9):10019-33.
- 491 59. Pingitore A, Chambers ES, Hill T, Maldonado IR, Liu B, Bewick G, et al. The diet-derived short chain
- fatty acid propionate improves beta-cell function in humans and stimulates insulin secretion from human
- 493 islets in vitro. Diabetes Obes Metab. 2017;19(2):257-65.
- 60. Chambers ES, Byrne CS, Aspey K, Chen Y, Khan S, Morrison DJ, et al. Acute oral sodium propionate
- supplementation raises resting energy expenditure and lipid oxidation in fasted humans. Diabetes Obes
- 496 Metab. 2018;20(4):1034-9.
- 497 61. Danesi F, Larsen BD, Di Nunzio M, Nielsen R, de Biase D, Valli V, et al. Co-Administration of
- 498 Propionate or Protocatechuic Acid Does Not Affect DHA-Specific Transcriptional Effects on Lipid Metabolism
- in Cultured Hepatic Cells. Nutrients. 2020;12(10).
- 500 62. Maldonado-Contreras A, Noel SE, Ward DV, Velez M, Mangano KM. Associations between Diet, the
- 501 Gut Microbiome, and Short-Chain Fatty Acid Production among Older Caribbean Latino Adults. J Acad Nutr
- 502 Diet. 2020;120(12):2047-60.e6.
- 503 63. Frampton J, Murphy KG, Frost G, Chambers ES. Short-chain fatty acids as potential regulators of
- skeletal muscle metabolism and function. Nat Metab. 2020;2(9):840-8.
- 505 64. Wang Z, Zhang X, Zhu L, Yang X, He F, Wang T, et al. Inulin alleviates inflammation of alcoholic liver
- disease via SCFAs-inducing suppression of M1 and facilitation of M2 macrophages in mice. Int
- 507 Immunopharmacol. 2020;78:106062.
- 508 65. Filippone A, Lanza M, Campolo M, Casili G, Paterniti I, Cuzzocrea S, et al. Protective effect of sodium
- 509 propionate in Aβ(1-42) -induced neurotoxicity and spinal cord trauma. Neuropharmacology.
- 510 2020;166:107977.
- 511 66. Jeong S, Kim HY, Kim AR, Yun CH, Han SH. Propionate Ameliorates Staphylococcus aureus Skin
- 512 Infection by Attenuating Bacterial Growth. Front Microbiol. 2019;10:1363.
- 513 67. Chen D, Qiu YB, Gao ZQ, Wu YX, Wan BB, Liu G, et al. Sodium Propionate Attenuates the
- 514 Lipopolysaccharide-Induced Epithelial-Mesenchymal Transition via the PI3K/Akt/mTOR Signaling Pathway. J
- 515 Agric Food Chem. 2020;68(24):6554-63.
- 516 68. Silva LG, Ferguson BS, Avila AS, Faciola AP. Sodium propionate and sodium butyrate effects on
- 517 histone deacetylase (HDAC) activity, histone acetylation, and inflammatory gene expression in bovine
- 518 mammary epithelial cells. J Anim Sci. 2018;96(12):5244-52.
- 519 69. Zhang Y, Yu K, Chen H, Su Y, Zhu W. Caecal infusion of the short-chain fatty acid propionate affects
- 520 the microbiota and expression of inflammatory cytokines in the colon in a fistula pig model. Microb
- 521 Biotechnol. 2018;11(5):859-68.
- 522 70. Tong LC, Wang Y, Wang ZB, Liu WY, Sun S, Li L, et al. Propionate Ameliorates Dextran Sodium Sulfate-
- 523 Induced Colitis by Improving Intestinal Barrier Function and Reducing Inflammation and Oxidative Stress.
- 524 Front Pharmacol. 2016;7:253.

- 525 71. Chitrala KN, Guan H, Singh NP, Busbee B, Gandy A, Mehrpouya-Bahrami P, et al. CD44 deletion
- 526 leading to attenuation of experimental autoimmune encephalomyelitis results from alterations in gut
- 527 microbiome in mice. Eur J Immunol. 2017;47(7):1188-99.
- 528 72. Mizuno M, Noto D, Kaga N, Chiba A, Miyake S. The dual role of short fatty acid chains in the
- pathogenesis of autoimmune disease models. PLoS One. 2017;12(2):e0173032.
- 530 73. Baker D, Amor S, Kang AS, Schmierer K, Giovannoni G. The underpinning biology relating to multiple
- sclerosis disease modifying treatments during the COVID-19 pandemic. Mult Scler Relat Disord.
- 532 2020;43:102174.
- 533 74. Kim CH, Park J, Kim M. Gut microbiota-derived short-chain Fatty acids, T cells, and inflammation.
- 534 Immune network. 2014;14(6):277-88.
- 535 75. Cavaglieri CR, Nishiyama A, Fernandes LC, Curi R, Miles EA, Calder PC. Differential effects of short-
- 536 chain fatty acids on proliferation and production of pro- and anti-inflammatory cytokines by cultured
- 537 lymphocytes. Life Sci. 2003;73(13):1683-90.
- 538 76. Bhaskaran N, Quigley C, Paw C, Butala S, Schneider E, Pandiyan P. Role of Short Chain Fatty Acids in
- 539 Controlling T(regs) and Immunopathology During Mucosal Infection. Frontiers in microbiology. 2018;9:1995-.
- 77. Poelaert KCK, Van Cleemput J, Laval K, Descamps S, Favoreel HW, Nauwynck HJ. Beyond Gut Instinct:
- Metabolic Short-Chain Fatty Acids Moderate the Pathogenesis of Alphaherpesviruses. Frontiers in
- 542 Microbiology. 2019;10(723).
- 543 78. Sim JR, Kang SS, Lee D, Yun CH, Han SH. Killed Whole-Cell Oral Cholera Vaccine Induces CCL20
- Secretion by Human Intestinal Epithelial Cells in the Presence of the Short-Chain Fatty Acid, Butyrate. Front
- 545 Immunol. 2018;9:55.
- 546 79. Corrêa RO, Vieira A, Sernaglia EM, Lancellotti M, Vieira AT, Avila-Campos MJ, et al. Bacterial short-
- 547 chain fatty acid metabolites modulate the inflammatory response against infectious bacteria. Cell Microbiol.
- 548 2017;19(7).
- 549 80. Ciarlo E, Heinonen T, Herderschee J, Fenwick C, Mombelli M, Le Roy D, et al. Impact of the microbial
- derived short chain fatty acid propionate on host susceptibility to bacterial and fungal infections in vivo. Sci
- 551 Rep. 2016;6:37944.
- Additives EPanel of, Food NSat. Safety of the extension of use of sodium propionate (E 281) as a food
- 553 additive. EFSA Journal. 2016;14(8):e04546.
- 554 82. Gold R, Montalban X, Haghikia A. Multiple sclerosis and nutrition: back to the future? Ther Adv
- 555 Neurol Disord. 2020;13:1756286420936165.
- 556 83. Storm-Larsen C, Myhr KM, Farbu E, Midgard R, Nyguist K, Broch L, et al. Gut microbiota composition
- during a 12-week intervention with delayed-release dimethyl fumarate in multiple sclerosis a pilot trial.
- 558 Mult Scler J Exp Transl Clin. 2019;5(4):2055217319888767.
- 559 84. Katz Sand I, Zhu Y, Ntranos A, Clemente JC, Cekanaviciute E, Brandstadter R, et al. Disease-modifying
- therapies alter gut microbial composition in MS. Neurol Neuroimmunol Neuroinflamm. 2019;6(1):e517.
- 561 85. Haase S, Wilck N, Haghikia A, Gold R, Mueller DN, Linker RA. The role of the gut microbiota and
- microbial metabolites in neuroinflammation. Eur J Immunol. 2020;50(12):1863-70.
- 563 86. Sivaprakasam S, Prasad PD, Singh N. Benefits of short-chain fatty acids and their receptors in
- inflammation and carcinogenesis. Pharmacol Ther. 2016;164:144-51.

566 **13.1 Tables**

Table 1: Comparison of the nature of dysbiosis reported in different studies in patients with MS

Country	Subjects (n = those with MS)	Bacteria increased in MS	Bacteria decreased in MS	Reference
USA	RRMS n=31	Pseudomonas, Mycoplana, Haemophilus, Blautia, Dorea, Pedobacter, Flavobacterium	Prevotella, Parabacteroides, Adlercreutzia, Collinsella, Lactobacillus, Coprobacillus, Haemophilius	(11)
USA	RRMS n=60	Methanobrevibacter, Akkermansia	Butyricimonas, Prevotella	(12)
USA	RRMS n= 43	Ruminococcus	Fecalibacterium	(10)
USA	RRMS n=7	Akkermansia, Acinetobacter		(5)
Germany	MS twins n=34 (each twin pair had 1 with MS and one healthy)	Akkermansia		(6)
UK	RRMS n=39	Prevotella copri		(13)
Japan	RRMSn=20	Bifidobacteria, Streptococcus, Thermophilus, Eggerthella lenta	Bacteroides, Fecalibacterium, Prevotella, Anaerostipes, Clostridium, Sutterella	(14)
China	RRMS n=34	Streptococcus	Prevotella	(15)
Germany	RRMS and SPMS	Akkermansia, Faecalibacteria	Butyricimonas, Bacteroides, Romboutsia	(17)
Japan	RRMS n=62, SPMS n=15, Atypical MS n= 21, Controls n=55	RRMS: Bifidobacteria, Streptococcus. SPMS: Streptococcus	RRMS: Megamonas SPMS: Roseburia	(20)

Adapted from (15, 17, 20)

Table 2. SCFAs in patients with MS

	Ace	etate	Propio	onate	Buty	rate
Author	Feces	Blood	Feces	Blood	Feces	Blood
(17) All types MS	No change from healthy controls	No change from healthy controls	Reduction p=0.0045	Reduction p=0.0006	No change from healthy controls	No change from healthy controls
(15)	Reduction	NA	Reduction	NA	Reduction	NA
MS	p=0.0001		p=0.0001		p=0.05	
(16) Patients with SPMS	NA	Reduction p=0.001	NA	Reduction p=0.01	NA	Reduction p=0.0001
(20) Patients with RRMS and SPMS	RRMS: Reduction p<0.001 SPMS: Reduction in levels but without statistical significance	NA	RRMS: Reduction p<0.001 SPMS: Reduction in levels but without statistical significance	NA	RRMS: Reduction p<0.001 SPMS: Reduction in levels but without statistical significance	NA
(Trend et al., 2021) Clinically Isolated Syndrome, MS and healthy controls	NA	No difference between MS and healthy control	NA	Reduction reported for CIS/MS group. p=0.0008	NA	No difference between MS and healthy control
(21) RRMS and Clinically Isolated Syndrome	NA	Small but significant reduction in MS	NA	No difference between MS and healthy control	NA	No difference between MS and healthy control

573 Table 3: SCFA transporters and receptors and tissue distribution in humans

	Ligand	Tissue		
Transporters				
MCT-1	Butyrate, lactate, pyruvate	Colon, Blood cells (monocytes, granulocytes, lymphocytes)		
SMCT-1	Butyrate>propionate>acetate	Intestine (Ileum, proximal colon and distal colon)		
G-coupled pr	rotein receptors			
FFA3 (GPR41)	Propionate=butyrate>acetate	Adipose tissue Peripheral blood mononuclear cells (PBMCs) Pancreas Spleen Placenta Monocytes, neutrophils, monocyte-derived dendritic cells (DCs)		
FFA2 (GPR43)	Acetate=propionate=butyrate	Intestinal epithelium Monocytes, neutrophils, PBMCs, T and B cells Treg cells (colonic>spleen and mesenteric lymph node), colonic myeloid cells		
GPR109a	Butyrate	Adipose tissue Colon Monocytes and macrophages		
Ref: (41, 86)			

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14 Conflict of Interest

576 577	D Tobin and R Vige have ownership in DAHRT Biocare AS.
578 579	P Calder declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.
580	15 Author Contributions
581 582	DT drafted the manuscript; RV and PCC contributed and commented on the manuscript; all authors approved the final version of the manuscript.