## 1 Dependence of poly-β-hydroxybutyrate accumulation in sludge on biomass concentration in SBRs

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# 5 ABSTRACT

The combination of wastewater treatment with polyhydroxyalkanoate production has attracted increasing 6 interest in the context of the circular economy. Recent studies have thus attempted to optimize the 7 8 conditions for polyhydroxyalkanoate accumulation in sludge when treating wastewater. The effects of biomass concentration and sludge morphologies in reactors on PHB storage, however, were neglected in 9 the literature. Therefore, in this study settling time and organic loading rate were manipulated to adjust 10 sludge morphology and biomass concentration in sequential batch reactors (SBRs) to investigate their 11 influence on PHB storage in the feast phase. Our study shows that reducing settling times in SBRs from 12 10 to 0 minutes under organic loading rate of 3 g L<sup>-1</sup> d<sup>-1</sup> resulted in the decrease in biomass concentration 13 at steady states from 4.2 to 1.0 g L<sup>-1</sup> and the change of sludge morphology from well-settled granules to 14 poorly settled pinpoint flocs, but PHB content in sludge at the end of feast phase increased from 7.7 to 15 16 26.7%. The well-fitted regression lines between PHB content, SRT, feast/famine and food/microorganisms ratios and biomass concentration under different settling times suggest that PHB 17 was highly dependent on biomass concentration but independent on sludge morphology. Under settling 18 time of 0 minutes, the increase in OLR from 3 to 7.5 g L<sup>-1</sup> d<sup>-1</sup> resulted in an increased biomass 19 concentration from 1.0 to 2.1 g L<sup>-1</sup> and an increase in PHB content from 26.7 to 33.8%. The batch and 20 fed-batch experiments with different biomass concentrations also showed the influence of biomass 21 22 concentration on PHB accumulation in sludge. The conclusion of the dependence of PHB content on biomass concentration under a fixed OLR and varied OLRs drawn from this study enables sludge PHB 23 content as high as possible by adjusting biomass concentration in SBRs apart from the selective enriching 24 25 strategies for PHB accumulating organisms when treating VFA-rich wastewater.

26 Keywords: poly-β-hydroxybutyrate (PHB), biomass concentration, settling time, SBR, biomass loading

27 shock, OLR

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- 30
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- 32 Introduction

Using bio-based and biodegradable plastics such as PHB to replace petroleum-based plastics is 33 believed as one of the most important strategies to solve plastic pollution problems. However, the price of 34 PHB produced from fermentation with pure culture and well-defined media at present is at least three 35 times higher than that of petroleum-based plastics (Gholami et al., 2016) although decades of research has 36 been done. With the popularity of the circular economy concept and its implementation, more studies on 37 PHB have being directed from pure culture to open mixed culture with waste streams to reduce costs of 38 both feedstock and operation by eliminating sterilisation demand (Sabapathy et al., 2020). Even more, it 39 has been gradually realised that PHB production could be integrated with the treatment of conventional 40 wastewater and waste activated sludge (WAS) and thus transform wastewater treatment plants into 41 resource recovery plants (or biorefinery plants) (Morgan-Sagastume et al., 2015). In this case, PHB is 42 produced as one of the recovered products from wastewater when achieving the main function of 43 wastewater treatment plants, i.e. pollutant removal from wastewater. This concept was demonstrated in 44 45 several pilot-scale wastewater treatment plants in the last few years (Sabapathy et al., 2020). However, the integration of PHB production with conventional wastewater treatment is still very new. The 46 fundamental study is desirable to move PHB production from wastewater forwards for real application in 47 48 the context of a circular economy.

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Although PHB storage was originally found in biological phosphate removal sludge (Fuhs and
Chen, 1975), PHB accumulation in non-phosphate removal sludge was later reported as well (Liu et al.,

52 1996). Gradually, people realised that intracellular polymer PHB storage in activated sludge is a common phenomenon. Sludge from conventional wastewater treatment plants without any particular enrichment 53 could accumulate PHB up to 7.9-24% in acetate-fed 24-h batch experiments under aerobic and nitrogen-54 and phosphorus-limited conditions (Sakai et al., 2015). PHB is more easily stored in sludge under 55 transient conditions such as sequential batch reactors (SBR) operation (Fang and Liu, 2000). SBR 56 57 operation which creates feast and famine phases (i.e. transient conditions with substrate gradient over time) is favourable for the selection and enrichment of PHB accumulating organisms because organisms 58 with PHB storage outcompete those without intracellular polymer storage during the famine phase for 59 60 stored PHB to be used as a carbon source for growth. Thus, SBR with aerobic dynamic feeding is widely adopted for the enrichment of PHB accumulating organisms. For wastewater treatment, SBR is also 61 extensively used with either activated sludge or aerobic granular sludge. PHB storage with variable 62 contents from 9% to 68% in well-settled aerobic granules in SBRs fed with palm oil mill effluent (Gobi 63 and Vadivelu, 2014), domestic wastewater (Karakas et al., 2020), and synthetic wastewater (Wang et al., 64 2014) was reported while bulking sludge with poor settleability was found to have high PHB storage 65 response (Beccari et al., 1998). From these reports, we can see that high PHB storage could be obtained 66 from either well-settled granular sludge or poor-settled bulking sludge, implying that PHB storage might 67 not be dependent on sludge settleability. The relationship between PHB storage in cells and filamentous 68 69 or non-filamentous bulking of sludge under different conditions was studied to shed light on the mechanism of ecological selection for the control of sludge bulking in conventional wastewater treatment 70 71 plants because sludge bulking can cause biomass loss from reactors and costly liquid-solid separation after wastewater treatment in downstream. However, when the enrichment of PHB accumulating 72 organisms or PHB production is one of the main research purposes, sludge settleability or sludge 73 characteristics are mostly neglected and not well studied and reported. It seems more focuses were put on 74 the achievement of PHB content in cells as high as possible or the effects of operating conditions such as 75 sludge retention time (SRT), pH, acetate concentration (Chua et al., 2003), temperature 76 (Chinwetkitvanich et al., 2004), feast/famine (F/F) ratio (Hao et al., 2018), organic loading rate (Lorini et 77

al., 2020) on PHB accumulation. PHB production efficiency from sludge depends on both PHB content
in cells and harvested sludge amount during a certain period (Johnson et al., 2009; Valentino et al., 2015).
Recently, this problem has attracted some attention from researchers and the effects of sludge settleability
and biomass concentration on PHB accumulation were investigated.

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83 It was reported that bulking sludge could accumulate PHB to a comparable content with wellsettled activated sludge and thus saved energy required for aeration in the culture enrichment stage due to 84 the lower dissolved oxygen demand of filamentous bacteria (Wen et al., 2012). However, poor sludge 85 settleability can lead to less biomass concentration in reactors, higher biomass loading rate (i.e. high F/M 86 ratio), longer feast phase in a single cycle, which in turn might affect PHB yield and productivity. In 87 addition, harvesting bulking sludge for PHB extraction could be more expensive. On the contrary, 88 aerobic granular sludge with good settleability could overcome the disadvantages of bulking sludge. 89 Meanwhile, PHB content in granular sludge could reach comparable contents to bulking and flocs-90 formers under appropriate conditions (Wen et al., 2012; Gobi and Vadivelu, 2014). It seems that sludge 91 morphology and settleability is not an important factor affecting PHB content in sludge, but the studies on 92 bulking and granular sludge were conducted under different conditions, making the fair comparison hard. 93 Furthermore, there is a requirement of hydraulic selective pressure to maintain granular sludge structure, 94 95 which makes the operation of reactors for the enrichment of PHB accumulating organisms a bit more complicated because both maintaining granule structure and enriching PHB should be achieved under the 96 97 same conditions. Thirdly, sludge settleability affects biomass retention in SBRs, and further influences biomass loading rate, feast/famine ratio, SRT, substrate gradient, which are believed to be important for 98 PHB storage. To understand the relationship between sludge characteristics and PHB accumulation, (Li et 99 al., 2019) correlated PHB storing capacity with sludge characteristics such as sludge flocs size, biomass 100 concentration and extracellular polysaccharide concentration, but this correlation was only limited to PHB 101 accumulation stage rather than enrichment stage. The correlation of biomass with PHB content in the 102 enrichment stage is more critical when PHB enrichment is combined with wastewater treatment. 103

This study thus aimed to investigate the effects of sludge settleability and morphology on PHB 105 storage under the same operating conditions by using different settling times in SBRs to form granular 106 sludge or suspended sludge with different SVI. To further understand the interaction between sludge 107 settleability, biomass concentration in reactors, and PHB storage, three levels of organic loading rates 108 were applied. The ultimate purpose of this study was to deepen the understanding of the interaction 109 between different factors caused by sludge settleability and biomass concentration in reactors, and 110 corresponding PHB storage during the culture enrichment stage, guiding engineering operating conditions 111 for the integration of PHB enrichment with wastewater treatment. 112

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#### 114 2. Materials and Methods

# 115 2.1 The enrichment of PHB accumulating organisms in Sequence batch reactors

Three bubble column reactors with a working volume of 2 L as triplicates were inoculated with 116 the excess activated sludge from the Millbrook wastewater treatment plant in Southampton, United 117 Kingdom, for the enrichment of PHB accumulating microorganisms. The reactors were operated in 118 sequential batch mode as shown in Figure 1 with a cycle of 6 hours including 8-minute feeding, 337-327-119 minute aeration, 0-10-minute settling, and 5-minute discharging. With an exchange ratio of 0.32, the 120 hydraulic retention time (HRT) was maintained at 0.8 days. Due to the variation of biomass 121 122 concentrations in the reactors under different settling times, SRT ranged between 1 to 10 days. Air was continuously supplied with a flow rate of 3.0 L min<sup>-1</sup> to provide sufficient oxygen. The temperature was 123 kept at 28°C with heating tapes and pH was not controlled with a variation from 7.5 to 8.4. 124





## Figure 1. Schematic diagram of sequential batch reactor operation

The inoculated excess activated sludge was acclimatized for 38 days by increasing the 127 concentration of COD in the synthetic wastewater until 2.4 g COD L<sup>-1</sup> to allow the sludge to adapt to 128 higher influent COD concentration and SBR operation. After the acclimatization, the reactors were fed 129 with 0.65 L of synthetic wastewater, which composed of 5.0 g  $L^{-1}$  CH<sub>3</sub>COONa·3H<sub>2</sub>O as the carbon 130 source (equivalent to 2.4 g COD L<sup>-1</sup>), 560 mg L<sup>-1</sup> (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 53 mg L<sup>-1</sup> KH<sub>2</sub>PO<sub>4</sub>, and 67 mg L<sup>-1</sup> 131 K<sub>2</sub>HPO<sub>4</sub> with carbon, nitrogen, and phosphorus ratio (C: N: P ratio) at 100:5:1, and micronutrients such 132 as 40 mg L<sup>-1</sup> of CaCl<sub>2</sub>.2H<sub>2</sub>O, 12.5 mg L<sup>-1</sup> of MgSO<sub>4</sub>  $\cdot$  7H<sub>2</sub>O, 0.307 mg L<sup>-1</sup> of FeSO<sub>4</sub>  $\cdot$  7H<sub>2</sub>O, 0.045 mg L<sup>-1</sup> 133 of H<sub>3</sub>BO<sub>3</sub>, 0.009 mg L<sup>-1</sup> of CuSO<sub>4</sub>  $\cdot$  5H<sub>2</sub>O, 0.054 mg L<sup>-1</sup> of KI, 0.036 mg L<sup>-1</sup> of MnC1<sub>2</sub>  $\cdot$  4H<sub>2</sub>O, 0.018 mg 134  $L^{-1}$  of Na<sub>2</sub>MoO<sub>4</sub> · 2H<sub>2</sub>O, 0.036 mg  $L^{-1}$  of ZnSO<sub>4</sub> · 7H<sub>2</sub>O, 0.045 mg  $L^{-1}$  of CoCl<sub>2</sub> · 6H<sub>2</sub>O, 3 mg  $L^{-1}$  of 135 EDTA. 20 mg L<sup>-1</sup> Thiourea was added to inhibit nitrification (Wang et al., 2017). The reactors were 136 operated with an organic loading rate of 3.0 g COD  $L^{-1} d^{-1}$ . 137

The reactor operation was divided into two stages. In the first stage from day 0 to day 151, settling
time was set as 10 minutes, 1 minute and 0 minute, respectively, to provide different hydraulic selective

140	pressure and thus control the sludge concentration and morphology in the reactors. In the second stage
141	from day 151 to day 189, OLR was increased from 3.0 g COD $L^{-1} d^{-1}$ to 6.0 g COD $L^{-1} d^{-1}$ , and further to
142	7.5 g COD L <sup>-1</sup> d <sup>-1</sup> by increasing the concentration of CH <sub>3</sub> COONa·3H <sub>2</sub> O in the influent from 5.0, to 10,
143	and 12.5 g $L^{-1}$ CH <sub>3</sub> COONa·3H <sub>2</sub> O, respectively, while COD: N : P ratio was still maintained at 100: 5: 1.
144	Only after reactors reached steady states with stable values of MLSS, PHB content in sludge, SVI, effluent
145	ammonium and phosphate concentrations and stable sludge morphology, reactor operational conditions were
146	changed. In both stages, samples were collected from the beginning of the aerating phase when the
147	bacteria started to consume the carbon and nutrients, the end of the feast phase when the provided
148	external carbon source in the medium was depleted which could be identified by a sharp increase in
149	dissolved oxygen (DO) by an online measurement with a DO meter (YSI 5100), and the end of the
150	aeration phase. From the depletion of the external carbon source to the end of the aerating phase, it was
151	defined as the famine phase.

Table 1. Summary of operational conditions in two stages with variable settling times and OLRs

		Operation stage	e 1		Operation stage 2	
Phase	Ι	Π	III	Ι	II	III
Settling time (min)	10	1	0	0	0	0
OLR (g COD L <sup>-1</sup> d <sup>-1</sup> )	3.0	3.0	3.0	3.0	6.0	7.5
Influent COD concentration $(q \text{ COD } I^{-1})$	2.4	2.4	2.4	2.4	4.8	6.0

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# 155 2.2 PHB accumulation in the fed-batch reactors

Fed-batch experiments depending on a feed-on-demand method (Valentino et al., 2015; Zeng et 156 al., 2018) were conducted to examine the maximum PHB storage ability of the cultures enriched in SBRs. 157 158 Airflow rate and temperature were controlled at the same levels as those in SBRs. 200 mL of the enriched culture harvested at the end of the cycle of SBR was inoculated into a 1L fed-batch reactor for 159 overnight aeration to reduce PHB content in the cells from the SBRs to facilitate the maximum PHB 160 161 accumulation in the subsequent fed-batch experiment. The first fed-batch experiment used the culture from the steady-state of reactors enriched with OLR 3.0 g L<sup>-1</sup> d<sup>-1</sup> and settling time of 1 minute, the 162 second with the same OLR but 0 settling time, and the third experiment with OLR 6.0 g  $L^{-1} d^{-1}$  and 0 163

settling time. 5 g L<sup>-1</sup> NaAc·3H<sub>2</sub>O (equivalent to 2.4 g CODL-1) and 10.0 g L<sup>-1</sup> NaAc·3H<sub>2</sub>O (equivalent 164 to 4.8 g COD L<sup>-1</sup>) were prepared, respectively, for the first fed-batch experiment and the second and third 165 fed-batch experiments accordingly. The substrates had the same concentration of COD as the feeding 166 stock of their respective enrichment processes but without nutrients. There was no residual acetate at the 167 beginning of the fed-batch experiments. An online DO probe was used to monitor the change in DO 168 169 concentration in the fed-batch reactors. 10 mL of the substrate was initially added into the fed-batch reactors. Once the reduced DO concentration due to the addition of carbon source started to rise sharply, 170 an additional 10 mL of the substrate was added (Dionisi et al., 2006). According to (Dionisi et al., 2001) 171 172 (Valentino et al., 2017) and our own results, PHB content in cells could not increase any more after DO could not drop further with the addition or presence of an external carbon source. Thus, the fed-batch 173 operation was stopped based on stable DO values with the maximum PHB storage that cells could get.. It 174 should be pointed out that there was no nutrient limitation in fed-batch experiments due to high residual 175 ammonium and phosphate concentrations in effluent from SBRs. The main purpose of the fed-batch 176 experiments in this study is to investigate if high substrate concentration in SBRs could inhibit PHB 177 accumulation in cells. 178

## 179 *2.3 Analytical methods*

180 Mixed liquor suspended sludge (MLSS), sludge volume index (SVI<sub>30</sub>), COD, ammonium, and
181 phosphate were measured in accordance with standard methods (Eaton et al., 2005).

Sludge samples taken from the SBR reactors were placed in centrifuge tubes with each 5 drops of formaldehyde to stop all biological activity before centrifugation, which was conducted in a centrifuge (Thermo, Sorvall Legend XTR) at 8500 rpm (equivalent to 7916 RCF) for 15 minutes. The supernatant was then removed, and the sludge pellets were vortexed with deionized water and centrifuged at the same condition to remove sodium acetate residuals from the sludge. The washed sludge pellets were frozen at -80 °C and freeze-dried (VirTis® BenchTop<sup>TM</sup>, K) at a condenser temperature of -80 °C for 10 to 14 hours. 5 to 8 mg dried sludge sample was placed into a 15 mL vial and then mixed with 1.5 mL of

dichloromethane solution containing 300 mg L<sup>-1</sup> benzoic acid as an internal standard, and 1.5 mL 189 acidified 1-propanol (4 volumes of hydrochloric acid and 1 volume of 1-propanol). The vial was then 190 kept at 65 °C for 12 hours for the esterification of the extracted PHB and benzoic acid with 1-propanol. 191 After that, the vial was cooled down at ambient temperature and vortexed with 2 mL deionized water to 192 separate the dichloromethane laver, in which esterified PHB and benzoic acid were dissolved. 1 mL of 193 dichloromethane layer was placed into a GC vial with 50 to 80 mg of Na<sub>2</sub>CO<sub>3</sub> to dry the residual water in 194 the organic layer. 1 µL of the dried dichloromethane solution was injected into a gas chromatograph 195 (Varian CP – 3800) equipped with a flame ionization detector (FID) and an Agilent DB-FFAP 0.53mm 196 capillary column with a thickness of 0.25  $\mu$ m and a length of 30 m. Helium was used as the carrier gas 197 with a 50:1 split volume ratio between helium and sample. The temperatures of the sample injector and 198 the detector were set at 230°C and 250°C, respectively. The temperature of the oven with the column was 199 increased from 80°C by a rate of 10°C min<sup>-1</sup> until 120 °C, and then the heating rate was increased to 45°C 200 min<sup>-1</sup> until the temperature reached 240°C, at which it was held for three minutes. 201

Poly(3-hydroxybutyric acid-co-3-hydroxyvaleric acid) solutions with 5 different concentrations
 (sigma-Aldrich) were prepared as external standards against the peaks of internal standard (i.e. benzoic
 acid) for the quantification of PHB and PHV in sludge samples.

# 205 2.4 Calculation of stoichiometric parameters

The feast/famine (F/F) ratio was calculated by dividing the length of the feast Phase by the length of the famine phase in the same cycle. PHB content (wt%) in sludge was calculated by dividing PHB mass with the dry mass of sludge as shown in equation 1.

209 PHB content (wt%) = 
$$\frac{\text{PHB amount (g)}}{\text{dry mass of a sample (g)}} \times 100$$
 (eq1)

PHB yield, PHB productivity, biomass yield and the specific uptake rate of acetate were calculated as shown in equations 2-5, respectively. In the calculation, the mass of PHB was converted into Cmol based on that PHB monomer with a chemical formula of  $C_4H_6O_2$  and a molecular weight of 213 21.5 g C mol<sup>-1</sup> (Marang et al., 2014). The mass of biomass was converted into Cmol based on the 214 assumption that biomass had a general chemical formula of  $CH_{1.8}O_{0.5}N_{0.2}$  with a molecular weight of 215 24.6 g C mol<sup>-1</sup> (Beun et al., 2002).

216 PHB yield (PHB Cmol/Acetate Cmol) = 
$$\frac{\Delta PHB_{feast} (wt\%) * MLSS_r (g L^{-1}) * 68 (g Cmol^{-1})}{\text{Influent acetate } (g L^{-1}) * \text{Exchange ratio } * 21.5 (g Cmol^{-1}) * 100} (eq2)$$

217 PHB productivity(g PHB/L·h) = 
$$\frac{\Delta PHB_{feast} (wt\%) * MLSS_r (g L^{-1})}{6h}$$
 (eq3)

218 Biomass yield(xCmol/Acetate Cmol) = 
$$\frac{\Delta NH4 - N_{feast}(g L^{-1}) * 68 g C mol^{-1}}{Influent Acetate (g L^{-1}) * Exchange ratio * 0.114 * 24.6 g C mol^{-1}} (eq4)$$

219 Sodium acetate specific uptake rate = 
$$\frac{\Delta Acetate \text{ concentration } (g L^{-1}) * 24.6 \text{ g C mol}^{-1}}{MLSS_r(g L^{-1}) * 68 \text{ g C mol}^{-1} * \text{ Length of feast Phase } (h)}$$
(eq5)

Where  $\Delta PHB_{feast}$  (wt%) the different of PHB contents in cells between the initial and ends of feast 220 phase, MLSS<sub>r</sub> biomass concentration in the reactors at the end of a cycle, V<sub>r</sub> effective reactor volume, 221 reactor exchange ratio in this study is 0.32, 0.114 is nitrogen percentage of biomass, 68 g Cmol<sup>-1</sup> is the 222 molecular weight of CH<sub>3</sub>COONa·3H<sub>2</sub>O,  $\Delta$ Acetate concentration (g  $L^{-1}$ ) is the difference between 223 acetate concentrations in a cycle. The specific uptake rate of sodium acetate per unit of biomass was 224 calculated with assumptions of a zero-order sodium acetate consumption rate and a constant active 225 biomass concentration in the cycle (Beun et al., 2002). It needs to point out that only MLSS at the end of 226 a cycle as a single reference point was used for PHB productivity, so the productivity values were only 227 used for the purpose of comparison under different conditions. 228

## 229 3. Results and discussion

- 230 3.1 Sludge morphology and settleability, biomass retention in SBRs and PHB storage in sludge under
- 231 *different settling times*

After reactors reached a steady state, i.e., MLSS, PHB content, ammonium and phosphate concentrations in reactors were stable for at least three SRTs, sludge samples were taken for morphology observation as shown in Figure 1. Activated sludge in SRBs with a settling time of 10 min was gradually converted into aerobic granular sludge as shown in Figure 2a. The formed granular sludge had a compact 236 structure and large size, and the SVI of granules was less than 90 mL g<sup>-1</sup>, indicating good sludge settleability. Short settling time is usually believed to be critical for the formation or maintenance of 237 aerobic granular sludge (Liu and Tay, 2015). After a 100-day operation, the settling time was reduced 238 from 10 min to 1 min to create stronger hydraulic selective pressure. Aerobic granules, however, 239 disintegrated due to the extreme condition (i.e. settling time of 1min) applied. At the steady-state with a 240 settling time of 1 minute, SVI was 194 mL g<sup>-1</sup>, a significantly increased SVI, indicating a dramatically 241 reduced sludge settleability compared with that with a 10-minute settling time. It can be seen from Figure 242 2b that at this condition, large suspended flocs were dominant without aerobic granules observed. This 243 244 might be due to the extreme hydraulic selective pressure exerted on reactors, causing aerobic granules to be unstable. Although a short settling time is usually required as a selective pressure for the cultivation of 245 aerobic granular sludge, the settling time used for the operation of SBRs is usually longer than 5 minutes 246 to maintain the long-term stability of granular sludge (Chen et al., 2015; Franca et al., 2018). After a 29-247 day operation with a settling time of 1 min, the settling time was further reduced to 0 min (i.e. no settling 248 time applied). It was found that the settleability of sludge deteriorated further, resulting in an SVI of 273 249 mL g<sup>-1</sup>. Pinpoint flocs were dominant in SBRs at the steady state as shown in Figure 2c. Since only 250 251 settling time was changed, it can be concluded that settling time is a critical factor to control the 252 settleability of sludge in SBRs when other operating conditions are the same. A longer settling time is 253 favourable to form more compact sludge with lower SVI.



Figure 2. Morphologies of sludge at steady states in SBRs with a settling time of (a) 10 minutes, (b) 1 minute, and

(c) 0 minute

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257	Due to the change in sludge settleability with different settling times, biomass retention was
258	changed accordingly with more biomass washed out at a shorter settling time, resulting in lower biomass
259	concentrations in the reactors. Table 2 shows biomass concentration in reactors, sludge PHB storage and
260	other parameters in the PHB enrichment stage from three different settling times. It can be seen that the
261	change in sludge settleability caused by settling time led to the changes in biomass concentration in the
262	reactors and the effluent, thus the changes in biomass loadings (F/M ratio), feast/famine (F/F) ratio, and
263	SRT, which further affected PHB storage in sludge, biomass yield and PHB yield and productivity. It is
264	widely believed that higher biomass concentration results in lower biomass yield (Liu and Liu, 2004),
265	which was demonstrated as well by the biomass yields shown in Table 2. This might benefit the direction
266	of carbon flux from new biomass growth to PHB accumulation. However, higher biomass concentration
267	might be unfavorable to the enrichment of PHB accumulating organisms in sludge because of the
268	retention of biomass in reactors. It has been widely reported that aerobic granules can retain slow-
269	growing bacteria and anaerobic bacteria into the reactors due to their compact structure, large size and
270	excellent retention (Tay et al., 2002; Liu et al., 2004b; Liu et al., 2007). Thus, it is reasonable to
271	speculate that non-PHB accumulating microorganisms could be retained in aerobic granules while
272	aerobic granules were retained in the reactors, leading to a very low PHB content even at low F/F. In
273	addition, more carbon is demanded for aerobic respiration at higher biomass concentrations.

Table 2. Comparison of sludge settling ability, PHB storage and other reactor parameters at steady states with
 different settling times

Settling	SVI	MLSS	MLSS	SRT	F/M	F/F	Biomass yield	PHB	PHB yield	PHB
time	(mL	in	in	(day)	(g COD g <sup>-1</sup>	(feast/	(xCmol Cmol <sup>-1</sup>	content	(PHB Cmol	productivity
(min)	g <sup>-1</sup> )	reactors	effluent		MLSS·day <sup>-1</sup> )	famine)	Acetate)	(wt%)	Cmol <sup>-1</sup>	(g PHB L <sup>-</sup>
		(g L <sup>-1</sup> )	(g L <sup>-1</sup> )		-				Acetate)	<sup>1</sup> ·h <sup>-1</sup> )
10	89	4.23	0.74	4 47	0.71	0.07	0.02	7.7	0.28	0.41
		±0.20	±0.11	4.47			0.05	±0.5	0.28	0.41
1	194	1.55	0.80	1 5 1	1.94	0.28	0.15	22.0	0.33	0.14
		±0.11	±0.08	1.51			0.15	±1.4	0.55	0.14
0	273	0.98	1.06	0.72	3.07	0.30	0.19	26.7	0 27	0 11
		$\pm 0.11$	±0.15	0.72			0.16	$\pm 1.0$	0.27	0.11

\* biomass yield, PHB content, PHB yield and PHB productivity are at the end of feast phase

278 3.2 Correlation of PHB content, F/F and F/M ratios and SRT with biomass concentration in reactors with

279 *different settling times during the long-term operation period* 

280 Variable PHB contents in well-settled aerobic granule sludge from 9% to 68% were reported (Fang et al., 2009; Gobi and Vadivelu, 2014; Wang et al., 2014; Karakas et al., 2020). In addition, 281 comparable PHB content such as 53% was found in poorly-settled bulking sludge with well-settled 282 sludge (Wen et al., 2012). Thus, it could be speculated that the low PHB content in aerobic granular 283 sludge in this study was partially caused by high biomass concentration, which reduced the ratios of F/F 284 285 and F/M, and increased SRT accordingly. To validate this, PHB content in cells at the end of feast phase, SRT, F/F and F/M during transient and steady states from three triplicate reactors were correlated with 286 biomass concentration at three different settling times from 10 min to 0 min as shown in Figure 2. It can 287 288 be seen that although sludge morphologies (Figure 2) and settleability at the settling times of 10, 1 and 0 minutes (Table 2) were different, experimental data between PHB content, F/F and F/M and biomass 289 concentration were fitted very well with regression models with  $R^2$  above 0.93, suggesting that sludge 290 morphology (e.g. granular sludge or suspended sludge) and sludge settleability (high or low SVI) are not 291 decisive factors to affect PHB storage in cells. Although higher contents of extracellular polymeric 292 293 substances (EPSs) were reported in aerobic granular sludge (Liu et al., 2004a), there is no evidence to show that the synthesis of more EPS would reduce the storage of PHB in cells although it was reported 294 that EPS removal in granules facilitates PHB recovery in downstream (Gobi and Vadivelu, 2015). Thus, 295 296 sludge morphologies and settleability are not critical for PHB accumulation. The well-fitted regression 297 lines against biomass concentration with different sludge morphologies and settleability indicate that biomass concentration plays an important role while effects of sludge morphologies and settleability 298 299 were negligible in this study.

If there was no extra control of biomass concentration in reactors by sludge discharge, biomass concentration would be determined by sludge settleability, which was influenced by settling time in this study. Other parameters such as SRT and F/F ratio have been intensively investigated for PHB storage capacity. It was found that generally shorter SRT was beneficial to PHB production capability (Chua et al., 2003; Johnson et al., 2010), but too short SRT such as 0.5 days led to reduced PHB storage and unstable reactor operation (Johnson et al., 2010). Some researchers reported that a lower F/F ratio less

306 than 0.28 in SBRs allowed the good selection of culture with high polymer storage capacities (Dionisi et al., 2006; Johnson et al., 2009; Albuquerque et al., 2010; Jiang et al., 2011; Moita et al., 2014) while 307 F/F ratios higher than 0.55 increased the biomass growth and the storage mechanisms started to be 308 negligible. F/F ratios in this study under OLR of 3.0 g  $L^{-1}$  d<sup>-1</sup> ranging between 0.02-0.2 as shown in 309 Figure 3 fell well within the range of less than 0.28. Compared with SRT and F/F ratio, the studies on the 310 311 effects of F/M on sludge PHB storage were much less even though a wide range of F/M ratios from 0.03 to 6.0 were used in the literature (Serafim et al., 2004; Ou and Liu, 2009; Chakravarty et al., 2010; 312 Khumwanich et al., 2014). As shown in Figure 3, F/F and F/M ratios decreased exponentially with the 313 314 increase in biomass concentration while SRT increased linearly with biomass concentration. Therefore, it could be interpreted that in this study biomass concentration affected PHB storage capacity by changing 315 SRT, F/F and F/M ratios. It has to be pointed out that biomass concentration, SRT, F/F and F/M ratios 316 interact with each other while they are also affected by OLR and initial substrate concentration in cycles. 317 Thus, it is very challenging to derive the decisive factor for PHB storage capacity. Even though, the 318 experiment designed in this study is helpful to understand the importance of biomass concentration, 319 which is widely ignored and much less reported in PHB studies with open mixed culture compared with 320 other parameters. Although the effects of biomass concentration on enrichment or PHB accumulation 321 were occasionally touched in literature (Chakravarty et al., 2010; Chen et al., 2017), all of them just 322 focused on the comparison of PHB storage under two or three different biomass concentrations, resulting 323 in difficulty to look into the precise relationship between biomass concentration and PHB storage or 324 325 other parameters. In addition, those very limited studies on biomass concentration did not consider the determining effects of biomass concentration on SRT, F/F and F/M ratios to elucidate the interaction of 326 different parameters. Our experimental design in this study with triplicate reactors at three different 327 settling times allowed us to obtain dozens of data under multiple biomass concentrations to establish 328 reliable correlations as shown in Figure 3. As far as we know, this is the first report about the 329 correlations between PHB storage and biomass concentration, and between other parameters such as SRT, 330 F/F and F/M and biomass concentration during the long-term operation period, which shed light on the 331

- physiological response of sludge for PHB storage on the top of the enrichment theory on PHB
- accumulating organisms.



Figure 3. Correlations between (a) PHB content, (b) SRT, and (c) F/F ratio d) F/M ratio and biomass concentration at OLR of 3 g  $L^{-1}d^{-1}$  in three reactors with different settling times at transient and steady states: •, experimental data; — fitted regression lines

341

3.3 The immediate response of reactor sludge regarding PHB storage to biomass loading shock caused
by the change in the settling time

Figure 4 shows the changes in biomass concentration and PHB content in sludge with a reduced settling time from 10 minutes, to 1 and 0 minutes during the operating period from day 39 to 150. Two peaks regarding PHB content were observed just after the reduction in the settling time from 10 minutes

347 to 1 minute and from 1 minute to 0 minute, i.e. around days 100 and 129. Once the settling time was reduced from 10 minutes to 1 minute on day 100, biomass was immediately washed out due to a very 348 short settling time, which resulted in a drastic decrease in biomass concentration and thus a significant 349 increase in biomass loading shock (i.e. a surge of F/M ratio) immediately. Meanwhile, sludge PHB 350 content at the end of the feast phase surged from 7% to 28.5% within one day and further increased to 351 352 32.4% within 3 days. Sludge PHB content after three days, however, reduced and stabilised at around 15%. On day 129, the setting time was reduced from 1 minute to 0 minutes, which again resulted in a 353 quick biomass wash-out and a biomass loading shock. Within 3 days, sludge PHB content surged from 354 355 15% to 32.5% again, but decreased later and stabilized at around 25%. The surge of sludge PHB content and the subsequent decrease to respond to the change in the settling time is hard to be explained by the 356 enrichment theory of PHB accumulating organisms. To a large extent, this surge of sludge PHB content 357 should come from the quick physiological response of cells to a sudden increase in biomass loading 358 shock. Once biomass adapted to the new biomass loading rate, PHB storage was reduced. 359

360 As far as we know, this is the first study of settling time effects on PHB content and the first report about the surge of PHB content responding to biomass loading shock. According to (Mannina et 361 al., 2020), under appropriate conditions, once PHB accumulating organisms were enriched, stable PHB 362 363 content could be achieved while it was also reported that PHB accumulation was unstable at some conditions such as high OLRs (Valentino et al., 2014). However, there was no report in the literature 364 about the surge of PHB content during a very short period, i.e. within 1-3 days by changing settling times 365 and biomass loading rates. It could be speculated that some microorganisms in sludge that do not 366 accumulate PHB at a steady state could synthesize PHB or those PHB accumulating organisms could 367 368 synthesize more by a rapid stress response mechanism when suddenly being exposed to excessive loading pressure. When excessive biomass loading pressure becomes routine, these microorganisms 369 could adapt to the new biomass loading pressure without synthesizing PHB at all or only synthesizing 370 371 reduced PHB amount. This speculation needs to be validated by further study. But the phenomenon of PHB content surge under biomass loading shock could be tapped into by future studies for the increased 372

- 373 sludge PHB content before the purge of sludge particularly for the simultaneous enrichment and
- accumulation of PHB in a single reactor.



Figure 4. The profiles of biomass concentration, sludge PHB content and biomass loading rate with a
reduced settling time from 10 minutes to 1 and 0 minutes during the operating period from days 39 to
150 under OLR of 3 g L<sup>-1</sup>d<sup>-1</sup>

- 381 *3.4 Effects of organic loading rate on sludge settleability, biomass retention and PHB storage*
- 382

To further study the effects of biomass concentration on sludge PHB content, OLR was increased 383 from 3.0 g  $L^{-1}d^{-1}$  to 6.0 g  $L^{-1}d^{-1}$  and then to 7.5 g  $L^{-1}d^{-1}$  from operating days 151 to 189 with a settling 384 time of 0 minutes. The profiles of biomass concentration and sludge PHB content under different OLRs 385 are shown in Figure 5. It can be seen that biomass concentration in the reactor increased with the rise in 386 OLR almost linearly. For instance, the average biomass concentration increased by 95% from 0.93 to 387 1.81 g L<sup>-1</sup> and by 24% from 1.81 to 2.45 g L<sup>-1</sup>, respectively, corresponding to the increase of OLR by 100% 388 from 3.0 to 6.0 g  $L^{-1}d^{-1}$  and by 25% from 6.0 to 7.5 g  $L^{-1}d^{-1}$ , respectively. A similar result was found by 389 (Dionisi et al., 2006), who reported that an increase in an OLR from 8 to 12.8 g L<sup>-1</sup>d<sup>-1</sup> (by 50%) and 12.8 390 to 20.0 g  $L^{-1}d^{-1}$  (by 56%) resulted in an increase in biomass concentration by 49% and 52%, respectively. 391 (Kanimozhi and Vasudevan, 2014) also found the corresponding increase in biomass concentration when 392

393 OLRs were increased although the increase was not linear. Meanwhile, it was observed that sludge PHB content increased accordingly with the increase in OLRs, but sludge PHB content increased only by 16.8% 394 and 8.3%, respectively. This is consistent with those found by (Reddy and Mohan, 2012) that an increase 395 in OLR from 1.51 to 3.03 (100%) and from 3.03 to 4.54 kg  $L^{-1}$ day<sup>-1</sup> (50%) resulted in an increase in PHB 396 content from 32 to 37.4 wt% (16.87%) and 40.3 wt% (7.75%), respectively, although only one batch 397 398 instead of steady state was investigated in their study. This suggests that an increase in OLR could not lead to the same level of increase in sludge PHB content. In addition, it was found that the increase in 399 sludge PHB content did not necessarily lead to an increase in PHB yield. As shown in Table 3, both 400 401 biomass and PHB yields were more or less the same under three different OLRs, indicating that the carbon flux was neither directed to biomass nor sludge PHB with the increased OLRs. This result might 402 have practical significance as reactors could be operated with different influent COD concentrations and 403 OLRs within certain ranges to get similar biomass and sludge PHB yields with regard to carbon source 404 utilization. But too-high influent COD concentrations and OLRs above 20 g L<sup>-1</sup> d<sup>-1</sup> would inhibit the 405 activity of microorganisms and thus reduce PHB accumulation or stable PHB accumulation (Dionisi et 406 al., 2006). Comparable results regarding biomass and sludge PHB yields when biomass loading rate 407 (F/M) reduced from 3.07 to 1.43 g COD g<sup>-1</sup> MLSS d<sup>-1</sup> and F/F ratio increased from 0.32 to 0.62 are 408 contradictory with that was reported by (Albuquerque et al., 2010), in which that PHB yield decreased 409 410 linearly with the increase of F/F ratio in the range of 0.2-0.6 by increasing OLRs. Also, (Qu and Liu, 2009) found that an increased F/M ratio from 1.0 to 4.5 enhanced PHB storage in sludge in batch tests 411 412 while PHB accumulation decreased when F/M was increased further to 6.0, but it has been pointed out the results from Qu and Liu (2009) were only from batch experiments instead of from steady-states of 413 long-term SBR operation. Instead, in this study, PHB contents in sludge increased when F/M decreased 414 from 3.07to 1.43 during the steady states of long-term operation with OLRs increased from 3.0 to 7.5 g 415  $L^{-1} d^{-1}$  (Table 3) while increased when F/M increased from 0.71 to 3.07 with OLR of 3.0 g  $L^{-1} d^{-1}$  under 416 three different settling times (Table 2). Thus, the correlations in Figure 3 are only applicable to the 417 constant OLR. Once OLR increased as shown in Table 3, the relationships between biomass 418

concentration and PHB content was inverse. This partially suggests that the relationships between 419 different parameters obtained in literature are only limited to the conditions where the results were 420 obtained. Furthermore, these types of discrepancies indicate the intricated interaction between different 421 operating conditions and the difficulty to identify critical factors or universal rules affecting PHB storage. 422 Meanwhile, the discrepancies also suggest that F/F and F/M ratios could not be decisive factors to 423 determine PHB contents or yields. 424 In addition, it was noticed that both SVI and sludge PHB productivity increased by 40-45%, 425 which was much less than the OLR increasing percentage, i.e. 150%, in this study. An increase in SVI 426 with OLR in SBRs was observed by some researchers as well (Kanimozhi and Vasudevan, 2014) 427 although intermittently fed operations such as SBRs could be used to control filamentous growth and 428 sludge bulking (Chiesa and Irvine, 1985). The increase in SVI would lead to an increased cost for liquid-429 solid separation for the sludge PHB extraction downstream while the increase in productivity would 430 result in decreased capital and operating costs. Meanwhile, a higher sludge PHB content at higher OLRs 431 is beneficial to PHB extraction. Thus, the optimal OLR should be determined based on the cost 432 evaluation from the whole process with other specific operating conditions. 433



436 Figure 5. The profiles of biomass concentration and sludge PHB content under different OLRs from



## operating days from 135 to 189

Table 3. Comparison of sludge settleability, PHB storage and other reactor parameters at steady stateswith different OLRs and a settling time of 0 minutes

						0 1111100000					
	OLR	SVI	MLSS	MLSS	SRT	F/M	F/F	Biomass yield*	PHB	PHB yield *	PHB
	(g L <sup>-1</sup>	(mL g⁻	in	in	(day)	(g COD g <sup>-1</sup>	(feast	(xCmol Cmol <sup>-1</sup>	content*	(PHB Cmol	productivity
	day-1)	1)	reactors	effluent		MLSS.day <sup>-</sup>	/famine)	Acetate)	(wt%)	Cmol <sup>-1</sup>	* (g PHB
	-		(g L <sup>-1</sup> )	((g L <sup>-1</sup> )		1)				Acetate)	$L^{-1} h^{-1}$ )
	2.0	296	0.98	1.06	0.72	3.07	0.32	0.18	26.7	0.27	0.11
	5.0		±0.12	±0.10	0.72			0.18	$\pm 1.0$	0.27	0.11
	6.0	366	1.70	1.74	0.76	1.77	0.42	0.22	31.2	0.24	0.14
0.0	0.0		±0.09	±0.06	0.76			0.25	±0.9	0.24	0.14
	75	415	2.10	2.26	0 72	1.43	0.62	0.10	33.8	0.28	0 16
	1.5		$\pm 0.05$	±0.05	0.75			0.19	$\pm 1.0$	0.28	0.10

440 \* PHB content, PHB yield and PHB productivity represents the situation at the end of feast phase

## 442 3.5 Sludge PHB storage in SBRs and fed-batch reactors

To understand the effects of substrate and nutrient concentrations on sludge PHB accumulation in SBR cycles, the profiles of acetate, nutrients, DO and sludge PHB contents in batch cycles with different settling times at steady states were analysed as shown in Figure 5. Ammonium was consumed mainly for biomass synthesis, which could be used to represent new biomass growth. It can be seen from Figure 5 that ammonium consumption was much faster in the feast phase than in the famine phase when the settling time was 10 minutes with a biomass concentration of 4.3 g L<sup>-1</sup> while equivalent ammonium

<sup>441</sup> 

449 consumption rates during feast and famine phases were found when setting time was 0 minute with biomass concertation of around 0.7 g  $L^{-1}$ . This is because with the biomass concentration of 0.7 g  $L^{-1}$ , the 450 more PHB content accumulated in the feast phase could be used for biomass growth in the famine phase 451 for a higher specific PHB degradation rate (i.e. 5% PHB g<sup>-1</sup> MLSS·h<sup>-1</sup>) while less biomass growth 452 accompanied with a lower specific PHB degradation rate (i.e. 0.3%PHB g<sup>-1</sup> MLSS·h<sup>-1</sup>h) with the biomass 453 concentration of 4.3 g L<sup>-1</sup>. In addition, according to Table 2, it can be seen that at the end of the feast 454 phase, PHB yields were equivalent while biomass yield with a biomass concentration of 0.98 g L<sup>-1</sup> was 6 455 times higher than that with a biomass concentration of 4.3 g  $L^{-1}$ . This result is interesting because the 456 amount of carbon that flowed to PHB (i.e. PHB yield) was not affected by biomass concentration and 457 biomass loading rate in the reactors although biomass yield was influenced drastically. Sludge PHB 458 content, however, was much higher at the lower biomass concertation due to higher biomass loading rate. 459 460 From this perspective, lower biomass concentration is favourable due to an equivalent PHB yield and higher sludge PHB content. 461

The specific acetate uptake rate during the feast phase in Figure 5a was 1.06 acetate Cmol  $\cdot$  Cmol<sup>-1</sup> biomass h<sup>-1</sup> while 2.51 acetate Cmol  $\cdot$  Cmol<sup>-1</sup> biomass h<sup>-1</sup> in Figure 5b. Thus, low biomass concentration or high F/M encourages the microorganism to consume carbon faster. Similar results were found with a specific ammonium uptake rate, which was 0.0048 NH<sub>4</sub><sup>+</sup>-N mol·Cmol<sup>-1</sup> biomass h<sup>-1</sup> with 4.4 g L<sup>-1</sup> biomass concentration while 0.021 NH<sub>4</sub><sup>+</sup>-N mol  $\cdot$  Cmol<sup>-1</sup> biomass h<sup>-1</sup> with 0.7g L<sup>-1</sup> biomass concentration. Thus, the high specific ammonium uptake rate indicates that biomass growth occurred in the feast phase as well although PHB was stored in this phase.



471



Figure 6. Profiles of acetate, nutrients, DO and sludge PHB contents at the end of the feast phases in
batch cycles of SBRs at steady states with (a) settling time of 10 minutes and a biomass concentration of
4.3 g L<sup>-1</sup>, (b) settling time of 0 minute and a biomass concentration of 0.7 g L<sup>-1</sup>



- the feed-on-demand method were compared as shown in Table 4. For both batch and bed-batch
- 479 experiments, no nutrient limitation was used to study the effects of biomass and substrate concentrations

on PHB accumulation. The initial acetate concentration was reduced to 3.7 Cmmol acetate L<sup>-1</sup> in the fed-480 batch experiment from 24 Cmmol acetate L<sup>-1</sup> in the batch experiment to study the possible inhibition of 481 acetate on PHB accumulation with two different biomass concentrations at a settling time of 1 min and 0 482 min, respectively. It was found that when biomass concentrations were the same at around 1.3 g  $L^{-1}$  in 483 the batch and fed-batch experiments. PHB contents in batch and fed-batch experiments were almost the 484 same, suggesting there was no inhibition from the substrate at the concentration of 24 Cmmol acetate L<sup>-1</sup>. 485 When the biomass concentration in the fed-batch experiment was reduced to 0.8 g L<sup>-1</sup>, PHB content 486 reached 25.7%, an increase by 70% compared with the biomass concentration at 2.4 g  $L^{-1}$  in the batch 487 experiment although the same sludge was used. This again indicates that lower biomass concentration 488 has a positive impact on PHB accumulation in cells as the effects from substrate inhibition or different 489 microbial population were ruled out. (Chua et al., 2003) also found out that PHB contents of 31% and 21% 490 with biomass concentrations of 0.7 and 2.5 g L<sup>-1</sup>, respectively, in the batch experiments with the same 491 sludge. This finding is beneficial to the downstream PHB extraction as adjusting biomass concentration 492 could be conducted to maximise sludge PHB content apart from the enrichment of PHB accumulating 493 organisms. 494

When acetate concentration was doubled in SBRs and increased to 47 Cmmol L<sup>-1</sup>, it was found 495 496 that sludge PHB content in the fed-batch experiment was 38.7%, 18% higher than 32.8% in the batch experiments with the same sludge under the same biomass concentrations. (Serafim et al., 2004) also 497 reported that 67.5% of the maximum PHB content was obtained with 180 Cmmol/l of acetate supplied in 498 one pulse of a batch test while it reached 78.5% when 180 Cmmol L<sup>-1</sup> of acetate was fed in three pulses 499 of 60 Cmmol  $L^{-1}$  each. This might suggest that the higher acetate concentration used in this study, i.e. 47 500 Cmmol L<sup>-1</sup>, slightly inhibited PHB accumulation in the batch experiment. Reducing the acetate 501 concentration to 7.4 Cmmol L<sup>-1</sup> in the fed-batch was advantageous regarding the alleviation of substrate 502 inhibition for the maximum accumulation with the same sludge, but the increased percentage is not as 503 high as that by reducing biomass concentration. From the comparison results in this study, it could be 504 known that controlling both substrate and biomass concentrations could enhance PHB accumulation in 505

cells. This could provide guidance for the operation of SBRs by controlling settling time (to control 506 biomass concentration), and exchange ratio (to control substrate concentration) to achieve sludge PHB 507 content as high as possible in SBRs, which has a practical significance for harvesting sludge at the end of 508 the feast phase instead of using an extra fed-batch reactor for PHB accumulation to reduce capital and 509 operating costs when only wastewater streams containing excessive nutrients are available for PHB 510 accumulation. Furthermore, it has been reported that higher PHB content in sludge could lead to high 511 biogas yield from digestion due to higher biodegradability of PHB than cells (SATOH and MINO, 2013), 512 thus, the treatment of VFA-rich wastewater containing excessive nutrients in SBRs could be combined 513 with sludge PHB production in a single reactor, from which sludge harvested from the end of feast phase 514 could be digested for biogas production to increase energy recovery efficiency from wastewater when 515 PHB content is not high enough for extraction. 516

517

Table 4. Sludge PHB contents in SBRs and fed-batch reactors

518	Table 4. Sludge PHB contents in SBRs and fed-batch reactors								
	OLR (g	Settling	MLSS	The highest acetate	PHB content	MLSS in	The highest acetate	PHB content	
	L <sup>-1</sup> day <sup>-</sup>	time	in	concentration in the	(wt%) at the end of	fed-batch	concentration in the fed-	(wt%) at the end	
	1)	(minute)	SBRs	batch cycle (Cmmol	feast phase in	reactors (g	batch experiment	of fed-batch	
			(g L <sup>-1</sup> )	L-1)	SBRs	L-1)	(Cmmol L <sup>-1</sup> )	experiment	
	3.0	1	2.4	24	15.1	0.8	3.7	25.7	
	3.0	0	1.3	24	25.7	1.2	3.7	26.3	
	6.0	0	2.0	47	32.8	1.9	7.4	38.7	

519

#### 520 4. Conclusions

This study revealed the relationships between sludge morphologies and biomass concentration in 521

SBRs with sludge PHB storage at varied settling times and organic loading rates without nutrient 522

limitations. The conclusions are summarised as follows: 523

Under OLR of 3 g L<sup>-1</sup> d<sup>-1</sup>, sludge PHB content was highly dependent on biomass concentration, 524

which could be described by a well-established correlation equation. Sludge morphologies (i.e. 525

- granular sludge, suspended sludge, bulking sludge) have no impact on PHB accumulation in cells. 526
- Under OLR of 3 g L<sup>-1</sup> d<sup>-1</sup>, the sludge experienced biomass loading shock induced a surge of PHB 527 content in sludge within 1 day from a physiological response. 528

529	•	With the increased OLRs from 3 to 7.5 g $L^{-1} d^{-1}$ under the same settling time of 0 minutes, the
530		corresponding increase in biomass concentration led to slightly increased PHB content in sludge,
531		which is inverse dependence on biomass concentration compared with the constant OLR of 3 g $L^{-1}$
532		$d^{-1}$ .

The different dependence of sludge PHB content on biomass concentration under different operational conditions indicated the intricate nature of operating conditions with parameters interactedg each other. No universal rule could be achieved and the pattern obtained is applicable to a restricted range of operational conditions. But the dependence of sludge PHB content on biomass concentration implies a practical significance for the operation of combined PHB accumulation and wastewater

treatment in SBRs by adjusting biomass concentration to maximize PHB content in sludge.

### 539 **References**

Albuquerque, M., Torres, C., Reis, M., 2010. Polyhydroxyalkanoate (PHA) production by a mixed microbial culture

using sugar molasses: effect of the influent substrate concentration on culture selection. Water research 44,

542 3419-3433.

543 Beccari, M., Majone, M., Massanisso, P., Ramadori, R., 1998. A bulking sludge with high storage response selected 544 under intermittent feeding. Water Research 32, 3403-3413.

Beun, J., Dircks, K., Van Loosdrecht, M., Heijnen, J., 2002. Poly-β-hydroxybutyrate metabolism in dynamically fed
mixed microbial cultures. Water Research 36, 1167-1180.

547 Chakravarty, P., Mhaisalkar, V., Chakrabarti, T., 2010. Study on poly-hydroxyalkanoate (PHA) production in pilot

548 scale continuous mode wastewater treatment system. Bioresource Technology 101, 2896-2899.

549 Chen, F.-Y., Liu, Y.-Q., Tay, J.-H., Ning, P., 2015. Rapid formation of nitrifying granules treating high-strength

ammonium wastewater in a sequencing batch reactor. Applied microbiology and biotechnology 99, 4445-4452.

551 Chen, Z., Huang, L., Wen, Q., Zhang, H., Guo, Z., 2017. Effects of sludge retention time, carbon and initial biomass

552 concentrations on selection process: From activated sludge to polyhydroxyalkanoate accumulating cultures.

553 Journal of Environmental Sciences 52, 76-84.

- Chiesa, S.C., Irvine, R.L., 1985. Growth and control of filamentous microbes in activated sludge: an integrated
  hypothesis. Water Research 19, 471-479.
- 556 Chinwetkitvanich, S., Randall, C., Panswad, T., 2004. Effects of phosphorus limitation and temperature on PHA
  557 production in activated sludge. Water Science and Technology 50, 135-143.
- 558 Chua, A.S., Takabatake, H., Satoh, H., Mino, T., 2003. Production of polyhydroxyalkanoates (PHA) by activated
- sludge treating municipal wastewater: effect of pH, sludge retention time (SRT), and acetate concentration in
- 560 influent. Water Research 37, 3602-3611.
- Dionisi, D., Majone, M., Ramadori, R., Beccari, M., 2001. The storage of acetate under anoxic conditions. Water
  Research 35, 2661-2668.
- 563 Dionisi, D., Majone, M., Vallini, G., Di Gregorio, S., Beccari, M., 2006. Effect of the applied organic load rate on
- 564 biodegradable polymer production by mixed microbial cultures in a sequencing batch reactor. Biotechnology and
- 565 bioengineering 93, 76-88.
- 566 Eaton, A., Clesceri, L.S., Rice, E.W., Greenberg, A.E., Franson, M., 2005. APHA: standard methods for the
- 567 examination of water and wastewater. Centennial Edition., APHA, AWWA, WEF, Washington, DC.
- 568 Fang, F., Liu, X.-W., Xu, J., Yu, H.-Q., Li, Y.-M., 2009. Formation of aerobic granules and their PHB production at
- various substrate and ammonium concentrations. Bioresource technology 100, 59-63.
- 570 Fang, H.H., Liu, Y., 2000. Intracellular polymers in aerobic sludge of sequencing batch reactors. Journal of
- 571 Environmental Engineering 126, 732-738.
- 572 Franca, R.D., Pinheiro, H.M., van Loosdrecht, M.C., Lourenço, N.D., 2018. Stability of aerobic granules during long-
- term bioreactor operation. Biotechnology Advances 36, 228-246.
- 574 Fuhs, G.W., Chen, M., 1975. Microbiological basis of phosphate removal in the activated sludge process for the
- treatment of wastewater. Microbial Ecology 2, 119-138.
- 576 Gholami, A., Mohkam, M., Rasoul-Amini, S., Ghasemi, Y., 2016. Industrial production of polyhydroxyalkanoates by
- 577 bacteria: opportunities and challenges. Minerva Biotechnol 28, 59-74.
- 578 Gobi, K., Vadivelu, V., 2014. Aerobic dynamic feeding as a strategy for in situ accumulation of
- 579 polyhydroxyalkanoate in aerobic granules. Bioresource technology 161, 441-445.

- 580 Gobi, K., Vadivelu, V., 2015. Polyhydroxyalkanoate recovery and effect of in situ extracellular polymeric
- substances removal from aerobic granules. Bioresource technology 189, 169-176.
- Hao, J., Wang, H., Wang, X., 2018. Selecting optimal feast-to-famine ratio for a new polyhydroxyalkanoate (PHA)
- 583 production system fed by valerate-dominant sludge hydrolysate. Applied microbiology and biotechnology 102,
- 584 3133-3143.
- Jiang, Y., Marang, L., Kleerebezem, R., Muyzer, G., van Loosdrecht, M.C., 2011. Polyhydroxybutyrate production
   from lactate using a mixed microbial culture. Biotechnology and bioengineering 108, 2022-2035.
- Johnson, K., Jiang, Y., Kleerebezem, R., Muyzer, G., Van Loosdrecht, M.C., 2009. Enrichment of a mixed bacterial
  culture with a high polyhydroxyalkanoate storage capacity. Biomacromolecules 10, 670-676.
- Johnson, K., Kleerebezem, R., van Loosdrecht, M.C., 2010. Influence of the C/N ratio on the performance of
- 590 polyhydroxybutyrate (PHB) producing sequencing batch reactors at short SRTs. Water research 44, 2141-2152.
- 591 Kanimozhi, R., Vasudevan, N., 2014. Effect of organic loading rate on the performance of aerobic SBR treating
- 592 anaerobically digested distillery wastewater. Clean Technologies and Environmental Policy 16, 467-476.
- 593 Karakas, I., Sam, S.B., Cetin, E., Dulekgurgen, E., Yilmaz, G., 2020. Resource recovery from an aerobic granular
- 594 sludge process treating domestic wastewater. Journal of Water Process Engineering 34, 101148.
- 595 Khumwanich, P., Napathorn, S.C., Suwannasilp, B.B., 2014. Polyhydroxyalkanoate production with a feast/famine
- 596 feeding regime using sludge from wastewater treatment plants of the food and beverage industry. Journal of
- 597 Biobased Materials and Bioenergy 8, 641-647.
- 598 Li, H., Zhang, J., Shen, L., Chen, Z., Zhang, Y., Zhang, C., Li, Q., Wang, Y., 2019. Production of
- 599 polyhydroxyalkanoates by activated sludge: correlation with extracellular polymeric substances and
- 600 characteristics of activated sludge. Chemical Engineering Journal 361, 219-226.
- Liu, W.-T., Mino, T., Nakamura, K., Matsuo, T., 1996. Glycogen accumulating population and its anaerobic
- 602 substrate uptake in anaerobic-aerobic activated sludge without biological phosphorus removal. Water Research
- 603 30, 75-82.
- Liu, Y.-Q., Liu, Y., Tay, J.-H., 2004a. The effects of extracellular polymeric substances on the formation and stability
   of biogranules. Applied Microbiology and Biotechnology 65, 143-148.

- 606 Liu, Y.-Q., Moy, B.Y.-P., Tay, J.-H., 2007. COD removal and nitrification of low-strength domestic wastewater in
- 607 aerobic granular sludge sequencing batch reactors. Enzyme and microbial technology 42, 23-28.
- Liu, Y.-Q., Tay, J.-H., 2015. Fast formation of aerobic granules by combining strong hydraulic selection pressure
   with overstressed organic loading rate. Water research 80, 256-266.
- 610 Liu, Y., Liu, D., 2004. Kinetic study on glycerol production by repeated batch fermentation using free Candida
- 611 krusei. Process Biochemistry 39, 1507-1510.
- 612 Liu, Y., Yang, S.-F., Tay, J.-H., 2004b. Improved stability of aerobic granules by selecting slow-growing nitrifying
- 613 bacteria. Journal of Biotechnology 108, 161-169.
- 614 Lorini, L., di Re, F., Majone, M., Valentino, F., 2020. High rate selection of PHA accumulating mixed cultures in
- 615 sequencing batch reactors with uncoupled carbon and nitrogen feeding. New biotechnology 56, 140-148.
- 616 Mannina, G., Presti, D., Montiel-Jarillo, G., Carrera, J., Suárez-Ojeda, M.E., 2020. Recovery of
- 617 polyhydroxyalkanoates (PHAs) from wastewater: A review. Bioresource technology 297, 122478.
- 618 Marang, L., Jiang, Y., van Loosdrecht, M.C., Kleerebezem, R., 2014. Impact of non-storing biomass on PHA
- production: an enrichment culture on acetate and methanol. International journal of biological macromolecules71, 74-80.
- 621 Moita, R., Freches, A., Lemos, P., 2014. Crude glycerol as feedstock for polyhydroxyalkanoates production by
- 622 mixed microbial cultures. Water research 58, 9-20.
- 623 Morgan-Sagastume, F., Hjort, M., Cirne, D., Gérardin, F., Lacroix, S., Gaval, G., Karabegovic, L., Alexandersson, T.,
- Johansson, P., Karlsson, A., 2015. Integrated production of polyhydroxyalkanoates (PHAs) with municipal
- 625 wastewater and sludge treatment at pilot scale. Bioresource technology 181, 78-89.
- 626 Qu, B., Liu, J., 2009. Determination of optimum operating conditions for production of polyhydroxybutyrate by
- 627 activated sludge submitted to dynamic feeding regime. Chinese Science Bulletin 54, 142-149.
- 628 Reddy, M.V., Mohan, S.V., 2012. Effect of substrate load and nutrients concentration on the
- 629 polyhydroxyalkanoates (PHA) production using mixed consortia through wastewater treatment. Bioresource
- 630 technology 114, 573-582.

- 631 Sabapathy, P.C., Devaraj, S., Meixner, K., Anburajan, P., Kathirvel, P., Ravikumar, Y., Zabed, H.M., Qi, X., 2020.
- 632 Recent developments in Polyhydroxyalkanoates (PHAs) production–A review. Bioresource technology 306,
- 633 123132.
- 634 Sakai, K., Miyake, S., Iwama, K., Inoue, D., Soda, S., Ike, M., 2015. Polyhydroxyalkanoate (PHA) accumulation
- 635 potential and PHA-accumulating microbial communities in various activated sludge processes of municipal
- 636 wastewater treatment plants. Journal of applied microbiology 118, 255-266.
- 637 SATOH, H., MINO, T., 2013. Anaerobic digestion of polyhydroxybutyrate accumulated in excess activated sludge.
- 538 Journal of Water and Environment Technology 11, 429-438.
- 639 Serafim, L.S., Lemos, P.C., Oliveira, R., Reis, M.A., 2004. Optimization of polyhydroxybutyrate production by mixed
- 640 cultures submitted to aerobic dynamic feeding conditions. Biotechnology and bioengineering 87, 145-160.
- Tay, J.H., Ivanov, V., Pan, S., Tay, S.L., 2002. Specific layers in aerobically grown microbial granules. Letters in
- 642 Applied Microbiology 34, 254-257.
- Valentino, F., Beccari, M., Fraraccio, S., Zanaroli, G., Majone, M., 2014. Feed frequency in a sequencing batch
- reactor strongly affects the production of polyhydroxyalkanoates (PHAs) from volatile fatty acids. New
- 645 biotechnology 31, 264-275.
- 646 Valentino, F., Karabegovic, L., Majone, M., Morgan-Sagastume, F., Werker, A., 2015. Polyhydroxyalkanoate (PHA)
- 647 storage within a mixed-culture biomass with simultaneous growth as a function of accumulation substrate
- 648 nitrogen and phosphorus levels. Water Research 77, 49-63.
- 649 Valentino, F., Morgan-Sagastume, F., Campanari, S., Villano, M., Werker, A., Majone, M., 2017. Carbon recovery
- 650 from wastewater through bioconversion into biodegradable polymers. New biotechnology 37, 9-23.
- Wang, J., Li, W.-W., Yue, Z.-B., Yu, H.-Q., 2014. Cultivation of aerobic granules for polyhydroxybutyrate production
- from wastewater. Bioresource technology 159, 442-445.
- 653 Wang, X., Oehmen, A., Freitas, E.B., Carvalho, G., Reis, M.A., 2017. The link of feast-phase dissolved oxygen (DO)
- 654 with substrate competition and microbial selection in PHA production. Water research 112, 269-278.
- 655 Wen, Q., Chen, Z., Wang, C., Ren, N., 2012. Bulking sludge for PHA production: Energy saving and comparative
- storage capacity with well-settled sludge. Journal of Environmental Sciences 24, 1744-1752.