

1 **The Effect of Systemic Levels of TNF-alpha and Complement Pathway Activity on**  
2 **Outcomes of VEGF Inhibition in Neovascular AMD**

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51 **ABSTRACT**

52

53 **Background/Objectives:** Systemic levels of pro-inflammatory cytokines and activated  
54 complement components affect the risk and/or progression of neovascular age-related  
55 macular degeneration (AMD). This study investigated the effect of serum pro-inflammatory  
56 cytokine levels and complement pathway activity on the clinical response to vascular  
57 endothelial growth factor (VEGF) inhibition in neovascular AMD.

58

59 **Methods:** Sixty-five patients with a new diagnosis of neovascular AMD were observed over  
60 a six-month period in a single-centre, longitudinal cohort study. At each visit, the visual  
61 acuity score (VAS), central macular thickness (CMT), serum levels of CRP, pro-  
62 inflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$ , IL-2, IL-6 and IL-8), and complement pathway  
63 activity were measured. Participant DNA samples were sequenced for six complement  
64 pathway single nucleotide polymorphisms (SNPs) associated with AMD.

65

66 **Results:** A statistically significant difference in VAS was observed for serum levels of  
67 TNF- $\alpha$  only: there was a gain in VAS (from baseline) of 1.37 for participants below the 1<sup>st</sup>  
68 quartile of mean concentration compared to a reduction of 2.71 for those above the 3<sup>rd</sup>  
69 quartile. Statistical significance was maintained after Bonferroni correction (*P* value set at  
70 <0.006). No significant differences in CMT were observed. Additionally, statistically  
71 significant differences, maintained after Bonferroni correction, were observed in serum  
72 complement activity for participants with the following SNPs: *CFH* region (rs1061170),  
73 *SERPING1* (rs2511989) and *CFB* (rs641153). Serum complement pathway components did  
74 not significantly affect VAS.

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76 **Conclusions:** Lower serum TNF- $\alpha$  levels were associated with an increase in visual acuity  
77 after anti-VEGF therapy. This suggests that targeting pro-inflammatory cytokines may  
78 augment treatment for neovascular AMD.

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**Keywords**

Age-Related Macular Degeneration; Cytokines; Complement; Tumour Necrosis Factor-  
alpha; Vascular Endothelial Growth Factor

102 **INTRODUCTION**

103 Age-related macular degeneration (AMD), a progressive retinal disease that results in the loss  
104 of central vision, is predicted to affect 288 million people worldwide by 2040 (1).

105 Neovascular AMD (nAMD) is a result of choroidal neovascularisation (CNV) and leads to  
106 rapid vision loss. The mainstay of current treatment is inhibition of vascular endothelial  
107 growth factor (VEGF) (2). The evidence base for a genetic component in AMD is significant,  
108 and numerous single nucleotide polymorphisms (SNPs) have been associated with a patient's  
109 risk of developing AMD (3). SNPs in genes of the complement pathway, including the  
110 complement factor B (*CFB*) gene region (4, 5), the *C2* (4, 5) and *C3* (6) genes have been  
111 reported to affect the risk of developing AMD.

112

113 Uncontrolled activation of the complement pathway is limited by a set of complement  
114 regulatory proteins: Factor H and Factor I (encoded by the *CFH* and *CFI* genes respectively)  
115 regulate the alternative complement pathway (7), whereas the C1 inhibitor is a regulator of  
116 the classical pathway (8). Genetic variants at the Regulators of Complement Activation  
117 (RCA) locus on *chromosome* 1, which contains the *CFH* gene, contributes to AMD risk (9-  
118 11), in addition to the *CFI* gene region on *chromosome* 4 (12-14), and the *SERPING1* gene  
119 that encodes the C1 inhibitor (15, 16).

120

121 Studies have shown elevated levels of complement activation fragments to be independently  
122 associated with AMD (17-19). Furthermore, complement activation has been demonstrated to  
123 be associated with stage of AMD (20). In addition, systemic activation of the alternative  
124 complement pathway and complement components is associated with AMD genotypes (21),  
125 including the *CFH* SNP rs1061170 (Y402H) (19) and the *CFI* region SNP rs10033900 (17,  
126 21). A meta-analysis by Hong *et al.* reported that treatment-naïve patients carrying the *CFH*

127 SNP, rs1061170 (Y402H), were more likely to achieve an improved outcome to anti-VEGF  
128 treatment (22). Furthermore, visual outcome was improved after anti-VEGF treatment for  
129 patients carrying a low-risk *CFH* genotype and low *CFH* risk score (23).

130

131 Expression of acute phase proteins and pro-inflammatory cytokines can also affect the risk of  
132 AMD development and/or progression: CRP is an acute phase protein and marker of systemic  
133 inflammation that is an independent risk factor for AMD (24). IL-6 is a known cytokine  
134 stimulus of CRP release by the liver (25), and both have been associated with AMD  
135 progression (26). CRP has been demonstrated to induce IL-8 expression by human retinal  
136 pigment epithelium (RPE) cell lines (27), and both IL-6 and IL-8 are expressed by RPE cells  
137 on complement activation (28), by degenerating RPE cells (29), and are associated with  
138 drusen formation (30). Systemic levels of IL-6 have been found to be associated with the  
139 progression rate of geographic atrophy secondary to AMD (31). Additionally, patients with  
140 AMD have been shown to express higher levels of circulating IL-1 $\beta$  than age-matched  
141 controls (32). IL-2 has been implicated in the pathogenesis of AMD as activation of IL-2  
142 signalling pathways has been observed (33) and IL-2 contributes to extracellular matrix  
143 formation and the development of fibrosis in AMD (34). TNF- $\alpha$ , a pro-inflammatory  
144 cytokine that is known to mediate CNV formation in experimental models by upregulating  
145 VEGF expression by RPE cells (35), has also been demonstrated to promote the angiogenic  
146 drive of active CNV lesions (36). Patients with elevated levels of serum TNF- $\alpha$  have been  
147 shown to respond favourably to VEGF inhibition (32).

148

149 Although the studies mentioned above have investigated the role of complement pathway  
150 SNPs, complement pathway activity and systemic concentrations of pro-inflammatory  
151 cytokines on AMD pathogenesis, relatively few studies have investigated their functional

152 effect on outcomes of VEGF inhibition. The primary aim of this study was to investigate the  
153 effect of serum levels of pro-inflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$ , IL-2, IL-6 or IL-8) and  
154 complement pathway activity on the clinical response to VEGF inhibition in neovascular  
155 AMD. A secondary aim was to investigate the effect of complement pathway SNPs,  
156 associated with AMD, on serum complement activity in the same cohort of patients.

157

## 158 **MATERIALS AND METHODS**

### 159 **Study Approval, Registration and Regulation**

160 This study was conducted in accordance with the Research Governance Framework for  
161 Health and Social Care (2005) and Good Clinical Practice. Ethical approval was obtained  
162 from the National Research Ethics Committee (NRES) South Central- Southampton A. This  
163 study adhered to the tenets of the Declaration of Helsinki. The University Hospital  
164 Southampton NHS Foundation Trust was the sponsor of this study, and The University of  
165 Southampton undertook the research study. All patient samples and data were anonymised  
166 for the purpose of this study. Patient DNA and serum samples were stored for future studies.  
167 Procedures for handling, processing and storage of patient data were in compliance with the  
168 UK Data Protection Act (1998).

169

### 170 **Patient Recruitment, Consent, and Investigation**

171 Patients were recruited to the study after informed consent by the ophthalmology department  
172 of University Hospital Southampton NHS Foundation Trust. Patients were invited to take part  
173 if they met the principle inclusion criteria for the study: 1) over the age of 50; 2) a new  
174 diagnosis of neovascular AMD in one eye, treated with an initial loading dose of three,  
175 monthly Ranibizumab intravitreal injections; 3) White ethnicity (to limit any effects of ethnic  
176 variation on outcomes of VEGF inhibition in neovascular AMD). The exclusion criteria

177 were: 1) bilateral diagnosis of neovascular AMD (one of the exploratory endpoints of the  
178 study was the development of nAMD in the second eye); 2) a macular co-pathology; 3) poor  
179 venous access that prevents a peripheral blood samples being taken.

180

181 All patients recruited to this study had a diagnosis of neovascular AMD, confirmed on fundus  
182 fluorescein angiography, that was made by a consultant ophthalmologist specialising in  
183 medical retina diseases. Indocyanine green angiography was carried out for patients to rule  
184 out polypoidal choroidal vasculopathy (PCV)- patients with PCV were not invited to take  
185 part in the study. Patients were eligible to enrol for the study after their third intravitreal  
186 Ranibizumab injection and subsequently invited to a baseline visit (**Figure 1**). Informed  
187 consent was taken from participants at this visit, and their demographic details, medical  
188 history and baseline LogMAR visual acuity score (VAS) was recorded (number of letters on  
189 an ETDRS chart). A baseline central macular thickness (CMT) was also measured using  
190 optical coherence tomography (OCT) (Topcon, Berkshire, UK). A blood sample was taken at  
191 the baseline visit for serum cytokine and genetic analysis. Participants were reviewed by a  
192 study investigator and received treatment with an intravitreal ranibizumab injection if they  
193 had active neovascular AMD. Following the baseline visit, participants attended for six,  
194 monthly follow-up visits. At each visit, the VAS and CMT was recorded, a blood sample was  
195 taken, and the patient reviewed by a study investigator before any treatment for active  
196 disease.

197

#### 198 **Detection of Serum Cytokine levels and Activated End Components of Complement** 199 **Pathways**

200 Serum was isolated from participant blood samples using standard density-gradient  
201 ultracentrifugation at 1355 x g for 10 minutes at 21°C (Eppendorf, Stevenage, UK). Patient

202 serum cytokine levels were measured using semi-quantitative assays by Meso Scale  
203 Discovery (Rockville, Maryland, USA) as per the manufacturer's instructions. All cytokine  
204 measurements were undertaken in triplicate using the assay, and cytokine measurements were  
205 within the reading range of the kit. Functional assessment of classical and alternative pathway  
206 complement activity in patient serum samples was undertaken using Wieslab semi-  
207 quantitative ELISA Assays (SVAR Life Sciences, Malmo, Sweden) as per the manufacturer's  
208 instructions. Measurement of activated end components of classical and alternative  
209 complement pathways was expressed as a percentage relative to the fluorescence intensity of  
210 the positive control, derived from human serum components, supplied with the testing kit.

211

#### 212 **Genetic Analysis**

213 DNA was extracted from peripheral blood mononuclear cells of patient blood samples using  
214 erythrocyte lysis buffer (Fisher Scientific, Loughborough, UK) as previously described (37).  
215 DNA concentrations were measured using the Nanodrop ND1000 spectrophotometer  
216 (Thermo Scientific, Wilmington, DE, USA). Sequence analysis of participant DNA samples  
217 was undertaken by LGC Genomics (Hoddesdon, UK) on the following six SNPs associated  
218 with the complement pathway and AMD risk: *CFH* region: rs1061170; *CFI* region:  
219 rs10033900; *SERPING1 / CI-INH*: rs2511989; *CFB*: rs641153; *C2*: rs9332739; *C3*:  
220 rs2230199.

221

#### 222 **Statistical Analyses**

223 The GraphPad Prism software version 8.2 (GraphPad Software, Lo Jolla, Ca, USA) was used  
224 for statistical analyses and graphical representation of the data obtained in this study.  
225 Assessment of normality of continuous variables was determined by quantile-quantile plots  
226 of the residuals using GraphPad Prism. The unpaired *t* test with Welch's correction was used

227 to determine statistically significant differences in changes of visual acuity scores, central  
228 macular thickness and percentage activity of activated end components of complement  
229 pathways compared to positive controls. Statistical significance was set at the  $P<0.05$  value.  
230 As this is a preliminary / pilot study, the patient sample size was determined using a rationale  
231 laid out by S.A. Julious where a sample size of at least 12 is recommended (38). Our patient  
232 cohort was stratified into quartiles of approximately 16 in line with this recommendation.

233

## 234 **RESULTS**

### 235 **Serum classical or alternative complement pathway activity and functional response to** 236 **anti-VEGF intravitreal injections**

237 A total of 65 patients with a new diagnosis of neovascular AMD were recruited to participate  
238 in this study (**Figure 1**). Participant demographics are summarised in **Table 1**. Study  
239 participants were stratified into quartiles according to average serum concentration of an  
240 inflammatory protein over seven study visits, in order to amplify the functional effects of  
241 small changes in serum concentration (**Table 1**). The study first investigated any significant  
242 differences in the visual acuity score (VAS) or central macular thickness (CMT) change from  
243 baseline at each visit between participants who had a mean serum concentration of classical  
244 pathway (**Supplementary Figure 1A-B**) or alternative pathway (**Figure 2A-B**) complement  
245 components below the first quartile and above the third quartile. There was a statistically  
246 significant difference in the VAS change from baseline, -2.78 (SD=7.01) vs. -0.34 (SD=8.51)  
247 for mean serum alternative pathway components ( $P=0.048$ ), using an unpaired  $t$  test with  
248 Welch's correction (**Figure 2A**), but significance was not maintained after a Bonferroni  
249 correction was applied ( $P$  value set at  $<0.006$ ).

250



251 **Serum inflammatory protein concentration and functional response to anti-VEGF**  
252 **intravitreal injections**

253 Study participants were also stratified into quartiles according to mean serum concentration  
254 of CRP or a pro-inflammatory cytokine (TNF- $\alpha$ , IL-1 $\beta$ , IL-2, IL-6 or IL-8) over the seven  
255 study visits. Statistically significant differences initially observed for VAS change (**Figure**  
256 **2C**) for mean CRP concentration were not maintained after Bonferroni correction ( $P$  value  
257 was set at  $<0.006$ ), and there were no significant differences in CMT change (**Figure 2D**)  
258 using the unpaired  $t$  test with Welch's correction.

259

260 Of the pro-inflammatory cytokines assessed, there was a statistically significant difference  
261 observed in the VAS change from baseline, 1.37 (SD=9.40) vs. -2.71 (SD=7.79), between  
262 participants for mean serum TNF- $\alpha$  concentration below the first quartile and above the third  
263 quartile respectively ( $P=0.0024$ ), **Figure 2E**. Significance was maintained after a Bonferroni  
264 correction was applied ( $P$  value was set at  $<0.006$ ). No significant difference was observed in  
265 CMT change in these participants (**Figure 2F**). Additionally, no significant differences were  
266 observed in the VAS or CMT change from baseline between participants for mean serum  
267 concentration of IL-1 $\beta$  (**Supplementary Figure 1C-D**), IL-2 (**Supplementary Figure 1E-F**),  
268 IL-6 (**Supplementary Figure 1G-H**), or IL-8 (**Supplementary Figure 1I-J**).

269

270 **Complement Pathway SNPs and Activated Complement End Components**

271 All 65 study participants underwent DNA sequencing for the detection of six complement  
272 pathway SNPs reported to affect AMD risk. Measurement of classical or alternative pathway  
273 complement activity in the serum was undertaken at each study visit. Complement activity  
274 was expressed as a percentage relative to the positive control (based on human serum  
275 components) of the assay and could thus exceed 100%. For the *CFH* SNP rs1061170, a

276 statistically significant reduction was observed: 1) in mean classical pathway complement  
277 activity in homozygous participants (16.7% reduction;  $n=24$ ;  $P=0.0016$ ) (**Figure 3A**); 2) in  
278 mean alternative pathway activity in both homozygous (20.1% reduction;  $n=24$ ;  $P=0.0019$ )  
279 and heterozygous participants (19.4% reduction;  $n=35$ ;  $P=0.0025$ ), **Figure 3B**. For the *CFI*  
280 region SNP rs10033900, there was a statistically significant increase of 7.7%, after  
281 Bonferroni correction, in mean classical pathway activity, in both homozygous ( $n=16$ ;  
282  $P=0.0037$ ) and heterozygous ( $n=32$ ;  $P=0.0002$ ) participants, **Supplementary Figure 2A-B**.  
283 For the *SERPING1 / C1-INH* SNP rs2511989, there was a significant increase in only the  
284 mean alternative pathway complement activity of 11.8% in homozygous participants ( $n=11$ ;  
285  $P=0.005$ ), maintained after Bonferroni correction **Figure 3C-D**.

286

287 A significant increase in mean classical pathway complement activity of 16.3% was  
288 observed, after Bonferroni correction ( $P$  value set at  $<0.008$ ), in patients who were  
289 homozygous for the *CFB* SNP rs641153 ( $n=2$ ;  $P<0.0001$ ) and 9.7% who were heterozygous  
290 ( $n=10$ ;  $P<0.0001$ ), **Figure 3E**. Similarly, a significant increase in alternative complement  
291 pathway activity of 7.3% was observed, after Bonferroni correction, in heterozygous patients  
292 ( $n=10$ ;  $P=0.0069$ ), **Figure 3F**. No differences were observed for the *C2* SNP rs9332739 or  
293 *C3* SNP rs2230199 (**Supplementary Figure 2C-F**).

294

## 295 **DISCUSSION**

296 The primary aim of this study was to investigate the effect of serum pro-inflammatory  
297 cytokine levels and complement pathway activity on the clinical response to VEGF inhibition  
298 in neovascular AMD. After Bonferroni correction, a statistically significant difference was  
299 observed only in VAS (change from baseline) between participants stratified into quartiles by  
300 mean TNF- $\alpha$  serum concentration [a gain of 1.37 for participants below the 1<sup>st</sup> quartile

301 compared to a reduction of 2.71 above the 3<sup>rd</sup> quartile]. This was not associated with  
302 significant changes in CMT. This study supports both pre-clinical and clinic findings  
303 showing a small, but significant overall impact of systemic levels of TNF- $\alpha$  on CNV lesions  
304 and clinical responses to VEGF inhibition. In a previous study using a murine model of laser-  
305 induced CNV, inhibition of TNF- $\alpha$  with intraperitoneal injections of infliximab or etanercept  
306 led to significantly reduced CNV lesion size and pathological fluorescein leakage (39).  
307 Furthermore, in a non-controlled trial, infusions of the anti-TNF- $\alpha$  chimeric monoclonal  
308 antibody, infliximab, in neovascular AMD demonstrated non-progression of the disease in  
309 almost half of the treated patients and regression of exudative lesions without significant  
310 fibrous scarring (40). There was, however, no placebo group in this trial. Other small studies  
311 using intravitreal anti-TNF- $\alpha$  therapy combined with bevacizumab showed beneficial effects  
312 (41).

313

314 This study also investigated the effect of complement pathway SNPs, associated with AMD,  
315 on serum classical or alternative pathway complement activity in the same cohort of patients.  
316 A statistically significant, but modest, reduction (after Bonferroni correction) of classical  
317 pathway activity was observed in participants who were homozygous for the *CFH* region  
318 SNP rs1061170, in addition to a reduction in alternative pathway activity in participants who  
319 were either homozygous or heterozygous for this SNP. Furthermore, participants who were  
320 either homozygous or heterozygous for the *CFI* region SNP rs10033900 had a statistically  
321 significant increase in classical pathway activity, despite Factor I being better recognised as a  
322 regulator of the alternative pathway (7). Despite these differences, this study demonstrated no  
323 significant differences in VAS or CMT change from baseline between participants below the  
324 1<sup>st</sup> quartile and above the 3<sup>rd</sup> quartile of mean serum classical or alternative pathway  
325 complement components. Therefore, although statistically-significant, modest, differences in

326 serum complement activity were observed in participants with *CFI*, *CFH* and other  
327 complement pathway SNPs tested, this did not translate to real-world, significant differences  
328 in visual response to anti-VEGF treatment.

329

330 A recent study by Heesterbeek *et al.* demonstrated higher systemic levels of activated  
331 complement in patients with intermediate AMD (who demonstrated the highest serum  
332 complement activation), geographic atrophy and inactive neovascular AMD compared to  
333 patients with active nAMD (20). This raises the question of whether significant increases in  
334 complement activity were not observed in this study as all patients had active nAMD.

335 Interestingly, it was demonstrated in a study by Keir *et al.*, that anti-VEGF intravitreal  
336 injections in neovascular AMD patients resulted in increased levels of complement  
337 components (C3a, C4a and C5a) in the aqueous humour (42), and this was elevated in  
338 patients with earlier relapses of active nAMD compared to those with later relapses.

339

340 This study focused on measuring overall complement pathway activation (via the activated  
341 end products of complement activation) rather than a specific activated complement  
342 component, e.g. C3d. Commercially-available Wieslab assays (Svar Life Sciences) were used  
343 in this study, which are optimised to detect activation of the complement pathway using  
344 human serum. This assay was previously used to demonstrate a significant elevation in the  
345 activity of the alternative complement pathway in AMD patients with genetic variants in *CFB*  
346 and *C3* compared to controls (19). An alternative method to detect elevated systemic  
347 complement activation in our study could have been to calculate the C3d/C3 ratio from the  
348 plasma concentration of these complement components. This method was also used (in  
349 addition to the Wieslab assays) in recent studies to detect systemic complement activation in  
350 AMD patients with genetic variants (19, 20). It will be interesting to see whether there is any

351 difference between the Wieslab assay method and C3d/C3 ratio to measure systemic  
352 complement activation in our cohort of participants.

353

354 This study analysed individual genetic variants and their effect on complement pathway  
355 activation, which demonstrated some statistically-significant effects. Previous studies have  
356 demonstrated that the association of gene variants with complement activation in AMD  
357 patients may be stronger when undertaking haplotype analysis (43) compared to single  
358 variant analysis. An additive effect of complement pathway risk SNPs has been suggested to  
359 lead to an additive risk of disease (44). In the recent study by Heesterbeek *et al*, the  
360 association of AMD stage with complement activation was greatest in patients with  
361 haplotypes that were associated with the highest levels of complement activation (20). It will  
362 be interesting to undertake haplotype analysis to investigate the effect of overall complotype  
363 on the outcomes of VEGF inhibition in AMD in our patient cohort.

364

365 Studies have suggested that AMD pathogenesis is driven primarily by dysregulation of  
366 immune mediators locally within the eyes rather than circulating levels of these mediators. A  
367 study by Agawa *et al*. demonstrated that intravitreal anti-VEGF treatment (with  
368 bevacizumab) itself significantly raised intraocular levels of IL-6 and IL-8 (45), both  
369 implicated in AMD pathogenesis. A subsequent study, in contrast, demonstrated a reduced  
370 intraocular concentration of IL-6 after intravitreal aflibercept injection (46). Peripheral blood  
371 mononuclear cells (PBMCs), particularly monocytes, from AMD patients have been  
372 demonstrated to produce higher levels of IL-8 than age-matched controls (47), and it has been  
373 speculated that these cells could migrate to the macula to secrete additional IL-8.

374

375 The concept of AMD being a disease of systemic or local complement dysregulation was  
376 previously discussed by our group nearly eight years ago (48). Studies have suggested that  
377 SNPs associated with complement activation increase AMD risk by a combination of  
378 systemic activation of complement and dysregulation of complement activation in local  
379 tissues (49). It is unknown whether altered, systemic levels of complement in AMD are the  
380 result of AMD-associated gene variants whose effects are expressed in all tissues, or the  
381 result of circulating levels of complement that reach the choroid and retina to contribute to  
382 AMD pathogenesis. It is also thought that AMD pathogenesis is driven by a combination of  
383 locally-expressed complement factors (50, 51) in addition to systemic complement proteins  
384 which lead to local effects in tissues, e.g. the FH-related proteins (FHR) such as FHR-4 (52).  
385 This study did not demonstrate any statistically-significant functional effects, after  
386 Bonferroni correction, of elevated or reduced systemic complement pathway activity on  
387 outcomes of VEGF inhibition.

388

389 There were several limitations to this study, the most significant being the small cohort size  
390 of 65 participants. The primary reason for this is the challenge in recruiting a large number of  
391 patients for a study in which blood tests are taken every visit over seven months, in addition  
392 to an intravitreal injection where required. Fortunately, no participants dropped out of this  
393 study and all participant data from each visit was used in the analysis. Another limitation to  
394 this study was taking blood tests (for serum cytokine and complement pathway activity) after  
395 the loading dose of three, monthly intravitreal ranibizumab injections. The ethical regulations  
396 of this study meant that the priority was for patients to receive their loading dose of  
397 intravitreal anti-VEGF injections prior to enrolment in the study and for blood tests to be  
398 undertaken subsequently. Although the biggest gains in visual activity usually take place  
399 during the loading phase of intravitreal anti-VEGF injections, measurement of visual

400 outcomes took place in this study from the starting point of all participants having received  
401 their three, monthly intravitreal injections. Although there is emerging evidence of  
402 intravitreal ranibizumab injections affecting serum concentrations of pro-inflammatory  
403 cytokines (including a transient reduction of serum TNF- $\alpha$  levels) in patients with diabetic  
404 macular oedema (53), similar / significant evidence has not been demonstrated in the context  
405 of nAMD. Similarly, increased serum levels of TNF- $\alpha$  have been demonstrated in treatment-  
406 naïve patients with diabetic macular oedema (54), but not in nAMD patients (32).

407

408 In light of serum TNF- $\alpha$  levels being associated in this study with small, but significant  
409 effects on visual acuity after treatment with anti-VEGF intravitreal injections, it would be  
410 worth investigating this cytokine in larger cohorts to determine if this effect can be replicated.  
411 This would determine if systemic levels of TNF- $\alpha$  could be used to identify non-responders  
412 to anti-VEGF treatment.

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436 analyses during the study.

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438

439 **CONFLICT OF INTEREST**

440 None of the authors have any conflicts of interest to disclose.

441

442

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446

447 **AUTHOR CONTRIBUTION STATEMENT**

448 AHK contributed to research design, data analysis and interpretation and manuscript  
449 preparation. COP, GDS, HG, MN, AJC and GM contributed to research design, data  
450 acquisition and research execution. AJL contributed to research design, data analysis and  
451 interpretation, and manuscript preparation.

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454 **Supplementary information is available at *Eye's* website.**

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652 **TITLES AND LEGENDS TO FIGURES**

653

654 **Figure 1. A flowchart diagram summarising the investigation pathway of 65 study**  
655 **participants with a new diagnosis of neovascular Age-Related Macular Degeneration**  
656 **(AMD).** Peripheral blood samples were taken at seven visits (baseline visit and six follow-up  
657 visits), for serum and genotypic analysis.

658

659 **Figure 2. Change in visual acuity score (VAS) and central macular thickness (CMT)**  
660 **associated with serum concentration of alternative complement pathway components**  
661 **and inflammatory proteins.** Study participants were stratified into quartiles according to  
662 average serum concentration of an investigated inflammatory protein (including pro-  
663 inflammatory cytokine) or complement pathway-specific components over seven study visits.  
664 The change in VAS from baseline at each visit is plotted for all study patients who had a  
665 mean serum concentration of inflammatory protein or complement pathway component  
666 below the first quartile and above the third quartile. The percentage change in CMT from  
667 baseline at each visit is also plotted for the same study participants. Shown in parts **A-B** are  
668 the results for alternative complement pathway components and change in VAS or CMT  
669 from baseline at each study visit for patients below or above the indicated quartiles;  
670 \* $P=0.048$ . Shown in parts **C-D** are the results for C-Reactive Protein (CRP) and change in  
671 VAS or CMT from baseline at each study visit for patients below or above the indicated  
672 quartiles; \* $P=0.029$ . Shown in parts **E-F** are the results for Tumour Necrosis Factor- $\alpha$  (TNF-  
673  $\alpha$ ) and change in VAS or CMT from baseline at each study visit. \*\* $P=0.0024$ . The unpaired  $t$   
674 test, two-tailed, with Welch's correction, was used to determine whether there was a  
675 statistically significant difference in VA or CMT change from baseline between groups.

676



677 **Figure 3. Classical or alternative complement pathway activity associated with single**  
678 **nucleotide polymorphisms (SNPs) in study participants.** Study participants underwent  
679 DNA sequencing for the detection of six SNPs associated with the complement pathway and  
680 AMD risk. Assessment of serum levels of classical or alternative pathway complement  
681 components was undertaken on the same participants. The bar graphs **A-F** show the  
682 measurement of classical or alternative pathway complement activity on participants who  
683 express no SNP, are heterozygous, or homozygous for the following SNPs: *CFH* region:  
684 rs1061170 (**A-B**). *SERPING1 / C1-INH*: rs2511989 (**C-D**). *CFB*: rs641153 (**E-F**).  
685 Measurement of activated end components specific for the classical or alternative  
686 complement pathway in serum samples is expressed as a percentage relative to the activity of  
687 the positive control. The unpaired *t* test, two-tailed, with Welch's correction, was used to  
688 determine whether there was a statistically significant difference in classical or alternative  
689 pathway components between groups who had no SNP, were heterozygous for the indicated  
690 SNP, or homozygous for the indicated SNP. \* $P < 0.05$ . \*\* $P < 0.005$ . \*\*\* $P < 0.0001$ .  
691

**Patients with a new diagnosis of neovascular  
Age-Related Macular Degeneration (AMD) in one eye recruited by  
University Hospital Southampton NHS Foundation Trust**

Total patients recruited to study = 65

Patient with a new diagnosis of Neovascular AMD is treated with  
an initial loading dosing of three, monthly  
ranibizumab intravitreal injections

Patient becomes eligible to be a participant  
of the study on their 3<sup>rd</sup> intravitreal ranibizumab  
injection and invited to a baseline visit

**Baseline visit:**

Consent  
Demographic details  
Medical history  
Visual Acuity Score (VAS)  
Central Macular Thickness (CMT)  
Blood test:  
Serum inflammatory proteins and Genotype analysis

**Each of Six Monthly Follow-up Visits:**

Visual Acuity Score (VAS)  
Central Macular Thickness (CMT)  
Blood test:  
Serum inflammatory proteins and Genotype analysis  
Review by study investigator

Participant with active neovascular AMD  
receives intravitreal ranibizumab injection  
at visit

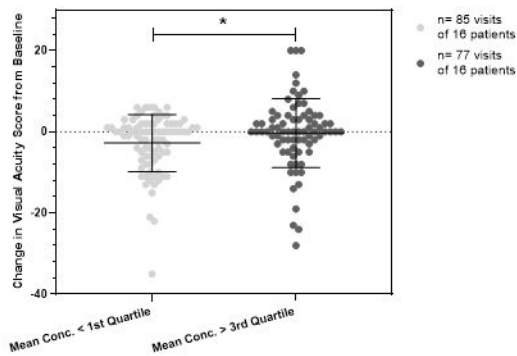
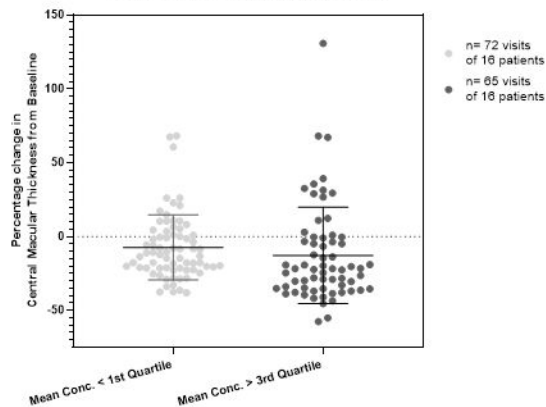
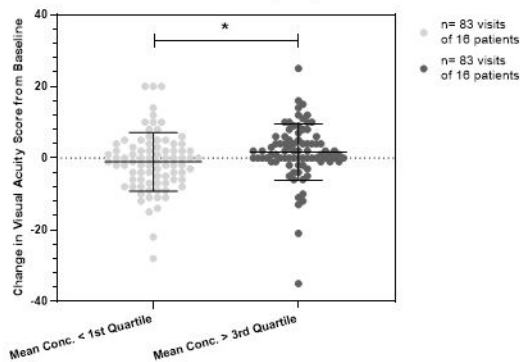
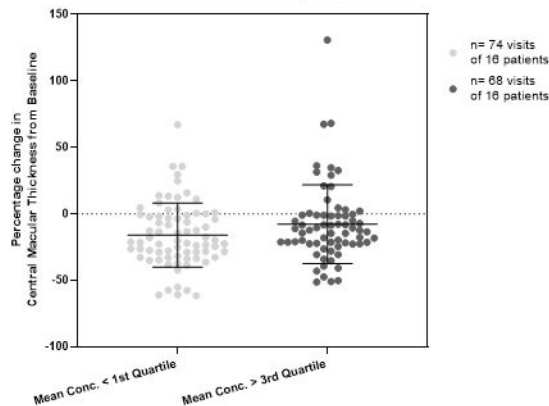
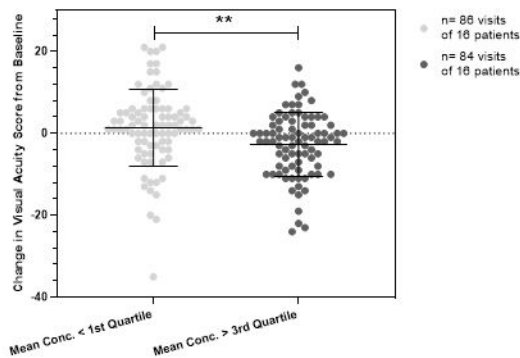
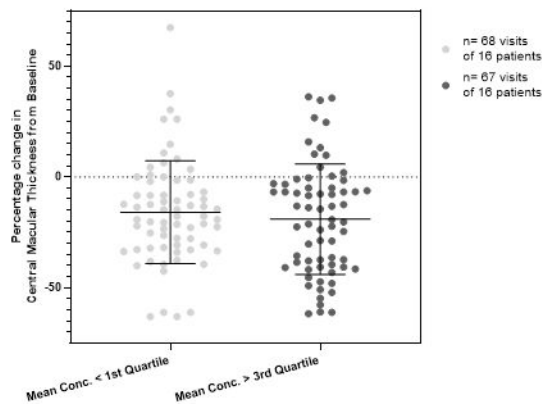
**End of follow-up / analysis period:**

1. Gene sequencing for six complement SNPs associated with AMD risk.
2. Participants stratified into quartiles according to average serum inflammatory protein concentration over total number of visits.

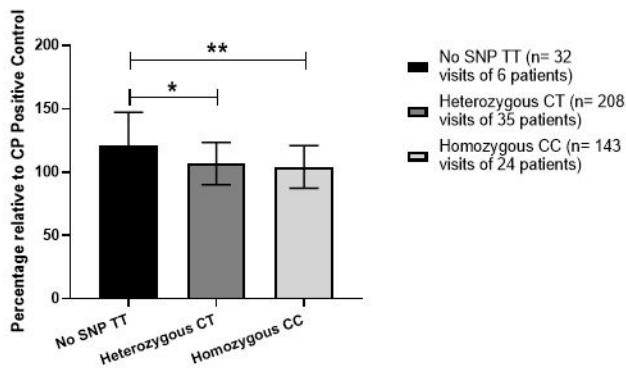
<b>Table 1. Study Participant Demographics and Summary Table</b>						
<b>Demographics</b>						
Number of patients recruited to study	65					
Patient sex	48% (n=31) male : 52% (n=34) female					
Patient age (mean, SD)	79.7 (SD = 8.6)					
Smoking Status	Current Smoker: 11% (n=7) Ex-Smoker: 46% (n=30) Non-smoker: 43% (n=28)					
Body Mass Index (BMI) (mean, SD)	26.8 (SD = 3.94)					
Co-morbidities	Hypertension: 47.7% (n=31) Asthma or COPD: 13.8% (n=9) Hyperlipidaemia: 13.8% (n=9) Diabetes (Type I or II): 12.3% (n=8) Nil co-morbidities declared: 16.9% (n=11)					
Mean Ranibizumab intravitreal injections over six, monthly follow-ups (mean, SD)	1.94 (SD = 0.98)					
<b>Serum Inflammatory Protein; Units</b>	<b>Serum Concentration (Cohort): 1<sup>st</sup> Quartile; Median; 3<sup>rd</sup> Quartile; Variance</b>					
C-Reactive Protein (CRP)	1.23 mg/L; 2.81 mg/L; 7.82 mg/L; 571.21 mg/L					
Tumour Necrosis Factor- $\alpha$ (TNF- $\alpha$ )	0.098 pg/ml; 0.136 pg/ml; 0.179 pg/ml; 0.078 pg/ml					
Interleukin-1 $\beta$ (IL-1 $\beta$ )	0.004 pg/ml; 0.010 pg/ml; 0.023 pg/ml; 0.012 pg/ml					
Interleukin-2 (IL-2)	0.009 pg/ml; 0.017 pg/ml; 0.033 pg/ml; 0.010 pg/ml					
Interleukin-6 (IL-6)	0.180 pg/ml; 0.300 pg/ml; 0.492 pg/ml; 2.723 pg/ml					
Interleukin-8 (IL-8)	0.964 pg/ml; 1.462 pg/ml; 2.067 pg/ml; 61.87 pg/ml					
<b>Complement Pathway Activity*</b>	<b>Percentage activity relative to positive control (Cohort) 1<sup>st</sup> Quartile; Median; 3<sup>rd</sup> Quartile</b>					
Classical Complement Pathway (CP)	95.57%; 102.2%; 117.1%;					
Alternative Complement Pathway (AP)	89.29%; 97.27%; 115.7%;					
<b>Gene sequencing analysis undertaken on patient cohort</b>						
<b>Gene / DNA region</b>	<b>Reference SNP</b>	<b>Chromosome and Position (bp)</b>	<b>Major / Minor Allele</b>	<b>MAF</b>	<b>OR</b>	<b>Reference</b>
<i>CFH</i> region	rs1061170	Chr 1; 196,659,237	T / C	0.61	2.41	11
<i>CFI</i> region	rs10033900	Chr 4; 110,659,067	C / T	0.52	1.18	11
<i>SERPING1 / CI-INH</i>	rs2511989	Chr 11; 57,134,901	G / A	0.45	0.63	15
<i>CFB</i>	rs641153	Chr 6; 31,914,180	C / T	0.05	0.54	11
<i>C2</i>	rs9332739	Chr 6; 31,903,804	G / C	0.02	0.46	11
<i>C3</i>	rs2230199	Chr 19; 6,718,387	G / C	0.24	1.53	11

\* Measurement of classical or alternative complement pathway activity is expressed as a percentage relative to the fluorescence intensity of a positive control.

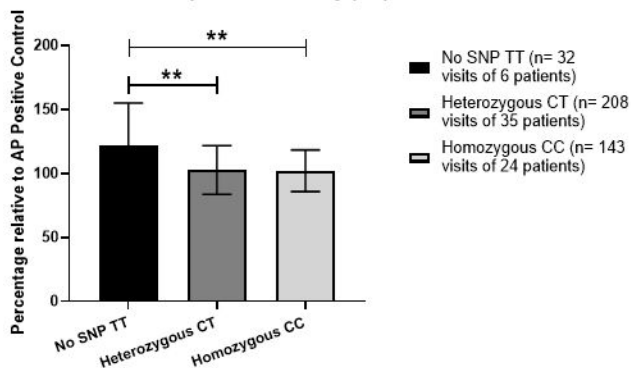
SD= Standard Deviation. SNP = Single Nucleotide Polymorphism. MAF = Minor Allele Frequency. OR = Odds Ratio of AMD.

**A** Alternative Complement Pathway (AP)**B** Alternative Complement Pathway (AP)**C** C-Reactive Protein (CRP)**D** C-Reactive Protein (CRP)**E** Tumour Necrosis Factor- $\alpha$  (TNF- $\alpha$ )**F** Tumour Necrosis Factor- $\alpha$  (TNF- $\alpha$ )

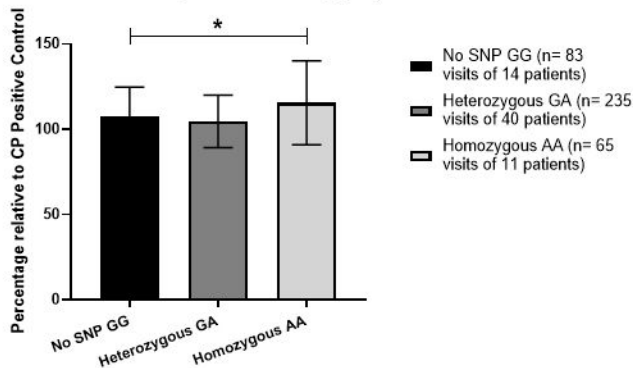
**A** *CFH* rs1061170 and Classical Complement Pathway (CP)



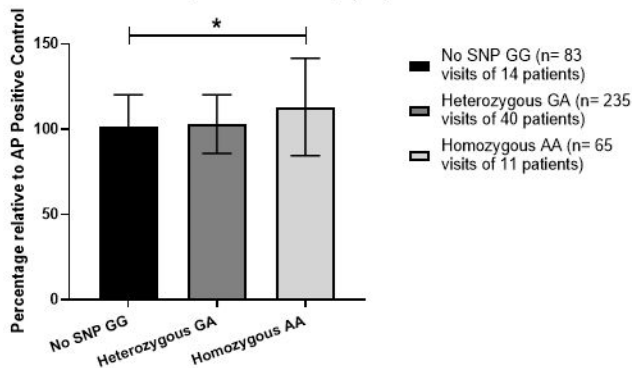
**B** *CFH* rs1061170 and Alternative Complement Pathway (AP)



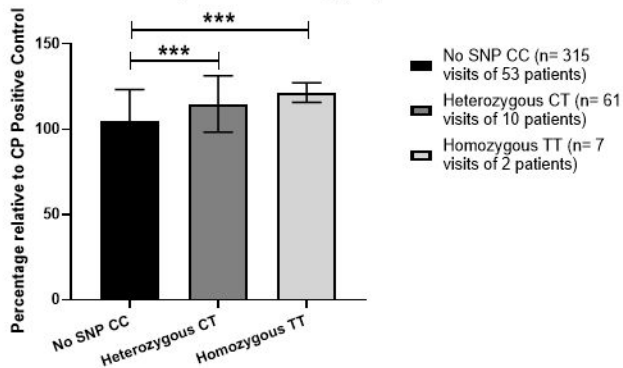
**C** *SERPING1 / C1-INH* rs2511989 and Classical Complement Pathway (CP)



**D** *SERPING1 / C1-INH* rs2511989 and Alternative Complement Pathway (AP)



**E** *CFB* rs641153 and Classical Complement Pathway (CP)



**F** *CFB* rs641153 and Alternative Complement Pathway (AP)

