Multimodal correlative imaging and modelling of phosphorus uptake from soil by hyphae of mycorrhizal fungi

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Summary

- Phosphorus (P) is essential for plant growth. Arbuscular mycorrhizal fungi (AMF) aid its uptake by acquiring sources distant from roots in return for carbon. Little is known about how AMF colonise soil pore-space, and models of AMF-enhanced P-uptake are poorly validated.
- We used synchrotron X-ray computed tomography (SXRCT) to visualize mycorrhizae in soil, and synchrotron X-ray fluorescence (XRF/XANES) elemental mapping for phosphorus (P), sulphur (S) and aluminium (Al), in combination with modelling.
- We found that AMF inoculation had a suppressive effect on colonisation by other soil fungi and identified differences in structure and growth rate between hyphae of AMF and nonmycorrhizal fungi. Results showed that AMF co-locate with areas

of high P and low Al, andpreferentially associate with organic-type P species in preference to Al-rich inorganic P.

- We discovered that AMF avoid Al-rich areas as a source of P. S-rich regions correlated with higher hyphal density and an increased organic-associated P-pool, whilst oxidized S-species were found close to AMF hyphae. Increased S oxidation close to AMF suggested the observed changes were microbiome-related. Our experimentally-validated model led to an estimate of P-uptake by AMF hyphae that is an order of magnitude lower than rates previously estimated; a result with significant implications for modelling of plant-soil-AMF interactions.

Keywords

Mycorrhizae, X-ray Computed Tomography, X-ray fluorescence, rhizosphere modelling, plant phosphorus uptake, synchrotron

Abbreviations

SXRCT: synchrotron X-ray computed tomography, XRF: X-ray fluorescence, XANES: X-ray absorption near edge structure, AMF: arbuscular mycorrhizal fungus

Word Counts

Intro: 758
Materials and Methods: 2579
Results and conclusions: 3975
Total: 7312

Introduction

Mineral phosphorus (P) resources across the world are sparse and unevenly distributed (Gross, 2017). Arbuscular mycorrhizal fungi (AMF) play an important role in mediating plant uptake of P (Smith *et al.*, 2003), which is often considered to be a growth-limiting soil resource (Vaccari, 2009). Mycorrhizal plants supply carbon (C) to AMF mycelia to drive hyphal growth, while in return the hyphae provide P and other nutrients back to the plant. Recent studies have found that in some cases a reduced plant C allocation to AMF does not alter AMF P supply to the plant (Charters *et al.*, 2020), which shows promise for crop production when P supply is limiting to growth. Much is known about plant-AMF symbioses on the soil bulk-scale, but little is known about the spatial distribution of AMF hyphae in soil due to difficulties in visualizing soil pore-space *in situ*. However, it is important to know how AMF hyphae interact with soil as P is strongly bound to soil surfaces, and pore-scale processes thus govern AMF-mediated P uptake. While we can modify certain pore-scale processes using chemical and microbial soil treatments, it is important to establish the mechanistic basis of symbiont behaviour within the soil matrix.

Sulphur (S), like P, is an essential macronutrient required by plants, and can also promote hyphal growth (Hepper, 1984). It has the potential to be a surrogate marker for the soil microbial community. S exists in soils in a wide variety of environmentally-dependent oxidation states ranging from -2 to +6, with organo-S comprising >90% of the total S pool in most soils (Gene *et al.*, 2002; Prietzel *et al.*, 2007; Prietzel *et al.*, 2011). AMF may play a role in plant S metabolism through uptake and up-regulation of plant sulphate carriers, and through their interaction with organo-S mobilizing microbes (Gahan & Schmalenberger, 2014; Berruti *et al.*, 2015). AMF mycelia are surrounded by complex bacterial and fungal communities that both interact with and sustain their metabolic function. Therefore, similar to the area affected by roots (*i.e.* the rhizosphere), a 'hyphosphere' of AMF can be a zone of increased bacterial abundance, and a site of localised biochemical activity (Rozmos *et al.*, 2021).

Several groups have modelled the growth of hyphal networks; as reviewed by Boswell and Davidson (2012). These models have addressed cell physics aspects (Bartnicki-Garcia *et al.*, 2000), physiological population growth (Edelstein, 1982), and biochemical processes controlling hyphal growth (Tlalka *et al.*, 2003). However, a persitient limitation in such models is a lack of validation due to experimental constraints. As soil is opaque to visible light, direct observation of hyphal morphology without disturbing the soil matrix is challenging. Boswell and Davidson (2012) reviewed models of mycelial development, while also identifying the need for model validation with experimental data. Simard *et al.* (2012) reviewed the ecology and modelling of mycorrhizal fungi and highlighted that a special challenge is the lack of techniques for observing hyphae in soil at a suitable spatial resolution. Some of the first models to specifically include AMF growth in soil and uptake of P were developed by our group (Schnepf & Roose, 2006; Schnepf *et al.*, 2008a). However, these models were only validated against data on the bulk soil and plant scales, and the soil P status was not monitored spatiotemporally. This lack of knowledge regarding AMF mycelial architecture on the soil pore-scale has impeded further investigation of the significance of soil-AMF-plant interactions for P uptake. In this paper we make the first necessary step towards building a fully validated plant-AMF P-uptake model. This is achieved by imaging AMF structures in the soil pore-space *in situ* using synchrotron X-ray computed tomography (SXRCT) in combination with traditional hyphal length measurements as set out by Jakobsen *et al.* (1992b), and correlating these data with spatial profiles of P, S and Al on the same soil samples using X-ray fluorescence (XRF/XANES) imaging. Al profiles are used to distinguish soil-mineral-associated P from the

organic pool. The structural and chemical imaging results are integrated into a plant-AMF mathematical model of P uptake (Schnepf & Roose, 2006; Schnepf *et al.*, 2008a) to further estimate hyphal uptake rates, and predict how the plant-AMF symbiosis benefits plant P acquisition. This enables us to image and quantify AMF in soil in three dimensions for the first time, apply finer-scale limits on P uptake rates by AMF hyphae, and provide the first calibrated mathematical model for AMF hyphae P uptake in soil. The comparison of hyphal length density measurements via SXRCT and a destructive approach also allows us to quantify the differences between two mycelial detection techniques.

Materials and Methods

Plant and Fungal Growth Assay

The growth medium was a sand-textured Eutric Cambisol soil collected from a surface plot at Abergwyngregyn, North Wales, UK (53°14′N, 4°01′W), for which the soil organic matter content was 7%. This soil corresponds to 'soil B' in Lucas and Jones (2006). A split-compartment system was designed to produce soil samples of small cross-section (∅<5 mm) for hyphal compartment, allowing SXRCT imaging at a sufficient spatial resolution to observe hyphal structures (see Figures S1.1 and S1.2). The system enabled the growth and maintenance of mycorrhizal wheat plants under controlled conditions for up to 4 weeks. A bespoke growth box was designed to maintain both hyphal and root compartments in dark conditions under stable and externally applied water potentials whilst allowing gas exchange and aerial plant growth (see SI-1 for full design details). A mesh barrier maintained separation between root and the hyphal compartments, while permitted transfer of hyphae (Faber *et al.*, 1991; Carminati *et al.*, 2009) and water between the two. In *P+* treatments, a small solid pellet of triple super phosphate (mean mass 0.0757 ± 0.007 g) was added to each hyphal compartment. In *P-* treatments, no supplemental P was added.

Plant-inoculation treatments. A wheat cultivar *Triticum aestivum* L. cv. Apache (WBCDB0003-PG-1) provided by the Germplasm Resources Unit of the John Innes Centre (UK) was selected due to its indicated high affinity for colonization by AMF (Leake, 2016). Seeds were surface sterilised and germinated in the dark for 96 h at 23 °C. Seedlings were selected for uniform radicle length and transplanted to each of 12 root compartments (2 P treatments × 2 time points × 3 replicates).

Root compartments were inoculated with *Rhizophagus irregularis* (BEG72, PlantWorks Ltd., UK)[a]. The inoculum contained hyphae, spores and colonized root fragments at 1.6×10^6 propagules L^{-1} of an inert zeolite carrier substrate. A uniform inoculating paste was achieved by concentrating the substrate according to manufacturer recommendations (SI-1.3). During filling of each root compartment[b] 1.5 g of inoculating paste and 0.5 g of raw inoculum (see also SI-1 section 1.3) was added at 25, 65 and 105 mm (d_1, d_2 and d_3, respectively) from the top of each root compartment (Figure S1.1).

The growth boxes were maintained in a climate-controlled growth chamber (Conviron A1000) with a 14 h light period at 23 °C and 75% humidity under full light (700 µmol m^{-2} s^{-1}), and a 10 h dark period at 18 °C and 75% humidity. For each treatment, three plant replicates (R_1, R_2, R_3) were grown to two and four weeks after transplantation, *i.e.* a total of six plants per P condition.

[a] We also tried inoculation with slow growing fungus *Gigaspora rosea*, but as this fungus did not grow we discounted it from the further study (see SI-1).
[b] As advised by the manufacturer PlantWorks Ltd to maximise the symbiosis.

Control treatments. The soil was neither autoclaved nor gamma-irradiated, since this would dramatically alter the natural microbiome and biogeochemical cycling processes occurring within the soil. Hence, the soil was assumed to contain native fungal spores. To prevent AMF propagules from developing hyphae in the controls, these contained no plants. Therefore, the 'plant' compartments for the control replicates were filled as outlined above and were lightly sealed with rubber bungs; with the soil water potential of the controls being kept similar to that of the plant compartments.

Structural Imaging

Following a growth period of 14 or 28 days, depending on treatment, the hyphal compartments were imaged using two separate SXRCT beamlines: The I13 beamline at the Diamond Light Source, UK, and the TOMCAT beamline at the Swiss Light Source, Switzerland, using a setup described in previous studies (Keyes *et al.*, 2013; Koebernick *et al.*, 2017; Koebernick *et al.*, 2019). For each hyphal compartment, three vertical positions were imaged: an *intermediate* position (h_2) was set as the approximate P pellet position in the P+ samples, and a *near-root* (h_1) and *far* (h_3) position were set 2000 µm below and above this position respectively. Effective voxel size was ~1.6 µm. For data collected at I13 only absorption reconstruction was available. For data collected at SLS, both phase/Paganin (Paganin *et al.*, 2002) and absorption reconstruction were available. See SI-1 for details.

SXRCT Image Analysis

Unless specified otherwise, image analysis was carried out using custom scripts written in ImageJ/Fiji (Schindelin *et al.*, 2012). The soil phase was first segmented into three phases: air-filled pore-spaces (*pore*), primary mineral grains (*primary*), and mixed phase (*mixed*) (SI-1 Figures S1.13-S1.16) using a WEKA machine-learning approach (Daly *et al.*, 2015; Keyes *et al.*, 2017; Koebernick *et al.*, 2017). Hyphal classification was carried out using the *absorption* volumes due to the greater hypha-to-pore contrast (Figure S1.12), after the soil segmentation result from the *phase* reconstructed volumes had first been used to mask out mineral regions. Hyphal classification was carried out using a custom segmentation approach with morphology first filtered using a two-pass morphological filter to remove small noise artifacts while conserving long filamentous structures of largely consistent diameter, *i.e.* hyphae (SI-1).

The segmented and filtered hyphal structures were skeletonised and the following metrics were quantified using the BoneJ toolbox (Doube *et al.*, 2010): total hyphal length, number of discrete hyphal clusters, branch count per cluster, mean branch length per cluster, tortuosity of branches, and angle of orientation to the hyphal compartment midline (*i.e.* the vector normal to the barrier mesh). These metrics were computed in MATLAB 16b (Natick, MA, USA), along with the standard deviation and standard error in the mean across replicates for each measure (SI-1).

Traditional Measurements

Traditional hyphal counting. The fraction of hyphae captured by the SXRCT approach was compared against that derived from a destructive sampling method (Jakobsen *et al.*, 1992b; Jakobsen *et al.*, 1992a). Replicate hyphal compartments were cut into three equal sections at the same locations used for SXRCT imaging, and each soil sample was divided in two subsamples for counting (SI-1). Hyphae were counted using digitised images collected via light microscopy with Olympus BX41 microscope; 20× objective magnification and bright-field illumination. A total of 20 images were acquired in randomised locations across each sample. This produced a total of 40 images for each soil sample. These images were examined to assess the phenotypic diversity within the entire sample set (Figure S1.29). Based on these

classifications, a set of candidate AMF phenotypes was defined with reference to literature (Abbott & Robson, 1985; Friese & Allen, 1991; Giovannetti *et al.*, 1993; Giovannetti *et al.*, 2001; Giovannetti *et al.*, 2004), to which all hyphal structures were subsequently assigned manually during the counting stage. See SI-1 for full detailed methods and results.

Polymerase chain reaction (PCR). PCR analysis of soil double-stranded DNA (dsDNA) purified from soil was carried out to validate the colonisation of hyphal compartments by AMF. Four sets of primers were used: (a) AMF primers (Kruger *et al.*, 2009) designed to amplify Glomeromycota fungi; (b) a bespoke set of AMF ITS primers (referred as *Tkacz* primers); (c) a broad range primer (Buee *et al.*, 2009a; Buee *et al.*, 2009b), designed to amplify Asco- and Basidio- rather than Glomero-mycota, and (d) bacterial primers (515F and 806R) targeting the prokaryotic 16S rRNA V4 region (Caporaso *et al.*, 2011). See SI-1 for further details of primers. We first confirmed that primers a) and b) target AMF, including *R. irregularis*, while primers c) and d) target other fungal and non-fungal species, but not AMF (SI-1-7.3). For all primer sets, standard PCR conditions were used: 98°C for 3 min, 35 cycles of 98°C for 15 sec, 55°C for 30 sec and 72°C for 100 sec, followed by a final elongation step at 72°C for 7 min using Phusion high-fidelity polymerase (NEB M0530L) and a PCR master mix with GC buffer (NEB M0532L). Following PCR, electrophoresis gels were run to determine whether amplification had been successful.

Microbial diversity analysis. The 18S rRNA genes were amplified from soil DNA extracts from hyphal compartments using the PCR primers F-574 and R-962 (Hadziavdic *et al.*, 2014) and sequencing was performed on an Illumina MiSeq platform (Environmental Sequencing Facility, University of Southampton). Sequences were processed and analysed using the DADA2 pipeline (Callahan *et al.*, 2016). Sequences shorter than 320 bp were discarded. Amplicon sequence variants (ASVs) were classified using the Wang Bayesian classifier in DADA2 and the Silva taxonomy (Pruesse *et al.*, 2012). The taxonomy of selected ASVs that could not be identified using the classifier were individually analysed by BLAST (Altschul *et al.*, 1990) against the non-redundant (nr) NCBI database; the taxonomy of closest BLAST match was used as an indication of the identity of the ASV. The ASV counts were subjected to a Hellinger transformation with the *decostand* function in the *vegan* (Oksanen *et al.*, 2013) package. Principal component analysis (PCA) was performed using the R-function *prcomp*. To focus on the ASVs that best explained the difference between these samples, the 50 ASVs with the largest absolute loadings in the first and second components, were selected. A heatmap representation of the relative abundance of these ASVs was constructed using *pheatmap* (Kolde, 2019). Sequence data were deposited in the NCBI Sequence Read Archive (SRA) under accession number PRJNA498673.

Root colonization analysis. To independently confirm AMF colonisation; cleared and stained roots were imaged via microscopy for signs of AMF colonisation based on the visible presence of intraradical hyphae, vesicles, arbuscules and external hyphae.

Plant Biomass. Fresh roots and shoots from the plant compartments were weighed and dried at 70°C for 48 hr. Dry samples were digested using a H_2SO_4/H_2O_2 digestion method (Novozamsky *et al.*, 1983) and prepared for HR-ICP-MS analyses to determine the total P content.

Chemical mapping of P and S. The SXRCT-imaged hyphal compartment replicates were freeze-dried (to preserve organic residues, *e.g.* hyphae and bacteria) and fixed in epoxy resin

(Epotek-301 diluted with ethanol); Figure S1.32. The samples were prepared using standard geological thin-sectioning procedures (Camuti & McGuire, 1999; Lanzirotti *et al.*, 2010).

At the DLS I18 beamline hyphal compartment, longitudinal thin-sections of hyphal compartments were mounted on an 3-axis-stage. The beamline comprises Kirkpatrick-Baez mirrors producing a spot size of 20 µm, utilising a Si(111) monochromator to scan the incident beam energy. Chemical maps were acquired under a helium atmosphere at 3 and 2.7 keV photon energies (to mitigate artifacts from chlorine comprised in the epoxy resin). For each pixel, a full energy-dispersive spectrum was recorded using a 4-element Vortex silicon drift detector positioned normal to the incident beam and 45° from the sample. Flux was estimated to be between 10^{10} to 10^{11} photons s^{-1}. SLS had the same setup, except that elemental maps were obtained under vacuum (10^{-6} mbar) and X-rays were detected using a single element Ketek silicon drift detector. All maps were fitted using the PyMCA package in batch fitting mode (Sole, 2020). Concentrations (in µg g^{-1}) of Al, Si, P and S were obtained by fitting maps obtained under identical experimental parameters using a spessartine garnet and Durango apatite mineral standards with known elemental concentrations (Table S1.2 and S1.3).

X-ray Absorption Near Edge Structure (XANES) spectra were collected in fluorescence mode to constrain changes in P (K-edge) and S (K-edge) speciation. The K-edge positions of P and S were calibrated against the first derivative of plots obtained using apatite [$Ca_5(PO_4)_3(OH)$] and sulphate [$ZnSO_4$] standards. Spectra from a series of P and S standards (Figure 1) were collected to perform a linear combination fit.

P concentration with distance from the root compartment was estimated from the XRF P intensity maps. Average P concentrations and standard deviations were calculated for the mixed-phase pixels. Since both P and S are in the tender X-ray region, and the attenuation length of P in quartz is *ca.* 2 µm, these maps are assumed to be 2D representations of concentration. Thus, the P, S and Al XRF maps were aligned via visual inspection to the corresponding SXRCT data of the same physical samples to correlate the chemical and structural information. The correlated data were used to determine if the presence of hyphae correlated with local variation in elemental concentrations of P, S and Al. A 3D Euclidean distance transform of the SXRCT-derived segmented hyphal geometries for each hyphal compartment was used to determine distance from hyphae to each position on the XRF maps. Pixels on the XRF maps were separated into two distance classes: pixels within 50 µm of the nearest hyphal surface ('close to hyphae') and further than 200 µm ('far from hyphae'). Pixels on the XRF maps were labelled as mixed phase if they were determined to be neither air (low total XRF signal) nor primary mineral (high Si signal). Excluding pixels that indicated high Si also limited overflow of Si signal into the neighbouring P or Al signal. Mean XRF counts of P, S and Al in the mixed phase were measured for each distance class of pixels over all treatments.

Mathematical Modelling

The AMF hyphae length density data at T_2 and T_4 (2 and 4 weeks of plant growth) were fitted with a model developed by Schnepf *et al.* (2008a) which included the simplest linear net hyphal branching term. See SI-2 for full mathematical modelling description. The model fitted was $\partial_t n + v\partial_x n = bn$, $\partial_t \rho = n|v| - d\rho$, where n is the hyphal tip density, v is the hyphal tip growth rate, b is the net hyphal branching rate, ρ is the hyphal length density, d is the net hyphal length destruction rate, t is the time and x is the distance from the root compartment along the hyphal compartment midline. The model was solved with a zero initial condition, *i.e.,* n=0 and $\rho = 0$ at t=0, assuming constant k tip production at the root surface, *i.e.*, v=k on x=0 for t>0.

We estimated values using the Matlab R2017a *fmincon* routine for k, d, b and v, by fitting the model to the length-density data from both SXRCT and destructive (Jakobsen *et al.* (1992b)) measurements. This gave us the lower and upper bound for the parameter values, since SXRCT only detects hyphae in the pore spaces and Jakobsen *et al.* (1992b) detects all hyphae.

XRF data for P was fitted with the model of Schnepf and Roose (2006), which links the fitted hyphal length density results for ρ to the soil P profiles. The model consists of a soil P movement equation that accounts for first order binding of P to the soil mineral surfaces and P diffusion in the soil pore-space, *i.e.*, $\partial_t c_{TOT} = D_{eff}\partial_{xx} c_{TOT} - 2\pi r_m \lambda_h \rho c_{TOT}$, where $c_{TOT} = c_s + \theta c_l = (b+\theta)c_l$ is the total amount of P in the soil, c_l is the concentration of P in the soil fluid/mixed phase, θ is the volume fraction of the mixed phase, b is the first order equilibrium binding buffer power of P in soil, λ_h characterizes the rate of P uptake per unit of hyphal surface areaper unit volume of total soil P. The model was solved setting the boundary condition at the root/AMF hyphae compartment boundary as $D_{eff}\partial_x c_{TOT} = F_{max} c_{TOT}$ on x=0 and $c_l \to c_\infty$ as $x \to \infty$; c_∞ is the P farfield concentration assumed to be the initial condition of P at t=0. We used the value for $D_{eff} = \frac{D\theta f}{\theta + b} = 1.05 \times 10^{-8}$ cm^2 s^{-1} as estimated by McKay Fletcher *et al.* (2017) and used the fittied results of our hyphal length density measurements ρ from the growth model above. We estimated the parameter λ_h against the XRF data by minimizing the sum of squares between the data points and the model using *fmincon* in Matlab R2017a.

Statistics

To analyse the data throughout this paper we used the Matlab Statistics Toolbox. We used a t-test at a significance of $p<0.05$, using *ttest2,* for means, and the Kolmogorov-Smirnov (KS) test at $p<0.05$, using *kstest2,* for the distributions throughout this paper.

Results and Discussion

Imaging of hyphal networks

Observations from SXRCT data revealed significant differences in hyphal morphology between treatments (Figures 2a&b). Hyphae in the control samples are more heterogeneous, have many more branches and have spherical features at some termini, which are assumed to be spores or spore-like structures. *Rhizophagus irregularis* inoculated samples (Figure 2b) are characterized by more linear/straight hyphae, less branching and fewer spherical/spore-like structures.

SXRCT imaging is non-invasive and provides richer information than destructive bulk length-density measures, enabling estimation of the number of branches per hyphal cluster, branch length, angle and tortuosity. However, as hyphae were only detectable in the pore space due to the contrast limitations of SRXCT, these measures might not be fully representative of the entire soil hyphal population. Control *P+* measurements showed significantly higher (t-test) and differently distributed (KS-test) branch numbers per cluster (when defining a single cluster to be all hyphae that are continuously connected on the images) than control *P-* and both *P+* and *P-* inoculated treatments (see Figure 2), suggesting that in the soil used, non-AMF hyphae are more branched in the soil pore space than AMF hyphae. The differences in branch number and distribution between inoculated *P+* and *P-* treatments was not found to be statistically significant (t-test and KS-test) potentially indicating, consistent with previous work (Drew *et al.*, 2003), that AMF hyphal morphology is not significantly dependent on P availability. Whilst we cannot fully quantify the non-AMF contribution to the length density, it is logical to conclude that any correction of the length density downwards would correct the hyphal uptake

rate upwards. Further integrated studies which go hand in hand with more detailed genomic sequencing would be able to shed light on this.

SXRCT data enabled quantification of the alignment of hyphae with respect to the major axis of each hyphal compartment. On average, this hyphal alignment angle varied between 45° and 85°. The only significant difference was that the control *P*+ treatment had a lower mean angle than control *P*- and inoculated *P*+ and *P*- treatments (Figure S28, t-test). Further analysis indicated that at the fertilizer pellet location (h_2), statistically significant differences were recorded in alignment of hyphae between control *P*+ (49°) and both *P*- treatments (CP- is 65° and IP- is 67°), and when pooling the data at the same location, hyphae in *P*+ treatments were found to be statistically more aligned to the compartment axis (51°) than in *P*- treatments (66°). One way of explaining these differences is in the context of the cost-benefit of the symbiosis: when the supply of P is comparatively more scarce, the hyphae 'search' for P by deviating more from the primary growth direction as defined by the geometry of the growth assay. The SXRCT data also allowed analysis of the (normalised) tortuosity of hyphal branches. Means of tortuosity in the control *P*+ and *P*- treatments were not significantly different (t-test), but the distributions were significantly different (KS-test). This could be explained by the larger variation seen in control P+ (see Fig 2c). The opposite was found for inoculated treatments, where *P*+ displayed a significantly different/wider (normalised) hyphal tortuosity distribution (KS-test) than *P*-. However, when comparing the inoculated and control *P*+ treatments, the inoculated *P*+ treatment had a significantly different/wider tortuosity distribution than the control *P*+ treatment, supporting the hypothesis that AMF fungi might be searching out P sources more aggressively than the non-mycorrhizal strains present in the control samples.

In the control samples with added P (Figure 2c CP+), the overall mean hyphal length density was higher (t-test) and the distribution was different (KS-test) compared to inoculated samples with plants with and without P addition (*P*+/-) and control samples with no P (*P*-). Since the soil was non-sterile, this suggests that AMF inoculation had a suppressive/allelopathic effect on the development of other soil fungi. Adding localised P also resulted in higher overall hyphal length in both control (CP+) and inoculated (IP+) samples. Figures 2d & e show the differences in the observed hyphal length densities in time as a function of distance (h_1, h_2, h_3) from the mesh interface with the plant compartment. Hyphal length density in this context means the total length of observed hyphae – of any length - per unit volume of soil. Based on these measurements we found that the initial growth of native soil fungi in the *P*+ control treatment (Figure 2d CP+) was reduced such that the hyphal length density in control (CP+) and inoculated treatments (IP-) was roughly the same by week 4. The inoculated P+ treatment (IP+) had higher hyphal length density at all three positions at the 2-week stage, but by week 4, the inoculated P- treatment (IP-) had a higher hyphal length density than IP+ closest to the root compartment (h1). See SI-1-5.4 and Figure S1.28 for supplementary analysis of hyphal branch distance, branch alignment and tortuosity as detected by SXRCT.

Figure 2c indicates that the P+ treatments in both control and inoculated treatments led to higher hyphal length density and different distribution compared to *P*- treatments (t-test and KS-test) when summing over all positions in the hyphal compartments; this is consistent with previous studies (Abbott *et al.*, 1984; Olsson *et al.*, 1997). Mean hyphal length density (t-test) and distribution (KS-test) for control *P*+ was significantly higher/different compared to the inoculated *P*+ treatment (Figure 2c). Similarly, the hyphal length density closest to the root compartment (h1, Figure S1.1) for control samples had a significantly higher mean and a different distribution than inoculated samples at h2 and h3, but the differences were not found when comparing at the closest (h1) distance from the root compartment (Figure S1.1, t-test and

KS-test). However, the incoluated samples indicated AMF colonisation whereas hyphae in the control samples were likely from saprothophic fungal strains. Similar analysis of the results for the Jakobsen measurements revealed an equivalent suppressive/allelopathic effect of inoculum/plant presence (Figure S1.30). Whilst there are many studies that describe plant root architectural responses to P conditions/heterogeneities in soil, there are few studies that show such results for AMF or other soil fungi (Olsson *et al.*, 1997; Cavagnaro *et al.*, 2005).

Destructively-derived hyphal length densityies were significantly higher than those determined via SXRCT (Figure S1.30). This difference can be explained by the majority of hyphae residing in the soil mixed-phase and/or at the soil-particle/air interfaces, where they cannot be detected by SXRCT. We conclude that 5% of control and 2% of inoculated sample hyphae were present in the soil pore-space and 95% of control and 98% of inoculated hyphae were in the inter-soil aggregate/mixed space. This is to be expected as the soil mixed-phase has higher P and S bioavailable concentrations and greater water availability. An alternative explanation is that the addition of P as a fertilizer, rather than a substrate with higher naturally-occurring P, might affect the extent to which hyphae penetrate into the soil mixed phase.

PCR, sequencing and other traditional validation measurements

PCR analysis indicated that one out of three inoculated samples (*P*+/-) tested at the 2-week stage were positive with the AMF-specific Kruger and Tkacz primers. At the 4-week stage, all inoculated samples were positive to Kruger primers and 2 out of 3 inoculated samples were positive to Tkacz primers. None of the inoculated samples were positive to broad primers (Buee *et al.*, 2009a; Buee *et al.*, 2009b). None of the control samples at weeks 2 or 4 were positive to Kruger, Tkacz, or broad primers, indicating a lack of AMF colonisation as expected.

Amplicon sequencing of eukaryote-specific 18S rRNA genes verified the presence of AMF in the inoculated samples. One sample (IP+, 4 weeks) failed to sequence properly and was excluded. Two amplicon sequence variants affiliated to AMF were detected in four of the inoculated samples with combined relative abundances of 0.34 – 1.34% whereas only one was detected in an uninoculated control sample at a relative abundance of 0.02%; see SI-1. The reverse PCR primer for amplification had a single mismatch to the gene in *Rhizophagus* species in the penultimate (3') position, which may explain the low detection rate.

The total combined relative abundance of sequences assigned to fungi detected in the samples (*i.e*. Ascomycota, Basidiomycota, and Mucoromycota) was higher in the uninoculated (32.3%) than inoculated treatments (24.7%). Although the sequencing only shows relative and not absolute abundances, when integrated with SRXCT and destructive imaging results this suggests the growth of native fungi was suppressed by AMF inoculation. The profiles between the inoculated and uninoculated treatments were distinct; see PCA analysis in Figure 3. These differences were spread across diverse phylogenetic groups of eukaryotes including nematodes and protozoa. The relative abundance of 18S rRNA genes in the original soil clustered together with the profiles from the hyphal compartments of the inoculated microcosms, implying that the eukaryotic community in the uninoculated hyphal compartments underwent a greater degree of change. On the basis of the genomic analyses, we conclude that the imaged inoculated samples contained AMF hyphae and control samples did not. Moreover, root staining showed clear mycorrhizal colonization (Figure 4a) in all inoculated treatments.

Elemental mapping

SXRCT images were correlated with elemental maps of P, S and Al for the same samples using XRF, allowing hyphae and soil morphology to be correlated with soil chemistry at the pore-

scale. Previous work has suggested hyphal length density and soil P content should, theoretically, show strong negative correlation (Joner *et al.*, 1995; Schnepf *et al.*, 2005; Schnepf & Roose, 2006; Schnepf *et al.*, 2008b; Schnepf *et al.*, 2008a; Schnepf *et al.*, 2011; Thonar *et al.*, 2011). This study did not find this strong correlation. However, previous studies carried out no chemical mapping or hyphal P-uptake calibration, hence, the hyphal P uptake parameters were highly uncertain. In this study, simultaneous measurement of hyphal length density and P gradients allowed more accurate estimation of hyphal P uptake rates. Further analysis indicated that organic S is the main S fraction in soil, making up 95-98% of total soil S, due to its being a major component of soil microorganisms, plants and animals (Scherer, 2009). Since the majority of observed soil S can be assumed to be associated with soil microorganisms due to exclusion of animals and roots from hyphal compartments, we treated S as a "surrogate marker" for microbe-derived soil organic matter. The sequencing analysis enables attribution of this S signal to a wide diversity of prokaryotes (>2000 amplicon sequence variants - ASVs) and microeukaryotes (>3000 ASVs).

The effect of organic matter on hyphal densities was also observed, and demonstrated how P and S counts were correlated against the hyphal distance maps (Figures 5(a-c)). These images show a clear positive correlation between soil organic material and fungal hyphae, in that hyphae are clearly oriented towards a structure which is high in S and relatively low in P. Figure 5(e) shows relatively uniform S counts across the mixed phase, with the exception of an S-rich fragment of organic matter which is apparently low in P. Figure 5(f) shows an SXRCT-derived map of 3D distance from hyphal surfaces indicating high hyphal presence in the S-rich regions. Hyphal correlation with organic material (Hodge, 2014) was also observed in other samples (SI-1, Figure S1.26). It must be noted that since SXRCT does not exclusively detect AMF hyphae, there is a possibility that some of the structures observed may be saprophytic in origin.

Figures 6(a-c) show distributions of P, S and Al XRF counts for pixels in the mixed-phase, both close to hyphae (within 50 µm of SXRCT measured hyphae) and far from hyphae (further than 200 µm from SXRCT measured hyphea) over all inoculated treatments. The mean P XRF concentration close to hyphae (1.03×10^4 XRF-counts) was significantly higher ($p<0.05$) than far from hyphae (8.3×10^3 XRF-counts). Figure 6(a) shows the shapes of the two distributions are similar, but close to the hyphae, the distribution is shifted to the right. A possible explanation is preferential hyphal growth close to areas of higher P concentration. Similar calculations for S showed no significant difference between the means (t-test) or distributions (KS-test) of the two S distributions (Figure 6(b)). Figure 6(c) shows the corresponding aluminium (Al) distributions, with the Al mean concentration far from the hyphae being significantly higher than that close to the hyphae (t-test).

The Al concentration close to the hyphae is generally lower and has a narrow distribution of concentration compared to the Al distribution far away from the hyphae (Figure 6(c)). This could suggest that AMF hyphae are less likely to grow in proximity to Al-rich clay minerals, which is consistent with previous findings (Vosátka *et al.*, 1999; Göransson, 2008; Zhang *et al.*, 2015). This leads to the question: do AMF hyphae seek P from Al-rich clay minerals if they contain high P? To test this hypothesis, XRF pixels were classified into two elemental classes: (1) *high P and low Al*, and (2) *high P and high Al,* and mean distance from hyphae for these pixels was calculated. '*High P*' pixels were defined as having a higher P concentration than the mean concentration of pixels within 50 µm of hyphae *i.e.* 1.03×10^4. '*Low Al*' pixels were defined as having lower Al concentration than the mean concentration of pixels within 50 µm of hyphae, *i.e.*, 3.23×10^3. The same computation was performed for Al , *i.e.*, 8×10^3

(Figure 6(c)). The mean distance from hyphae for the '*highP/lowAl'* pixels was 884 µm, which was significantly lower (t-test $p<0.05$) than the '*highP/highAl*' mean of 1100 µm. The distribution of distance from hyphae for the two elemental classes is shown in Figure 6(d), '*HighP/lowAl*' has many more pixels that are close to hyphae (within 500 µm). Interestingly, only 0.05% of '*highP/highAl'* pixels are within 100 µm of hyphae, compared to 1.52% of '*highP/lowAl*' pixels. These results suggest that AMF might be avoiding Al-rich clay minerals as a source of P. Such a result, if confirmed to be AMF-specific, could have significant and detrimental implications on the ecological effectiveness of the AMF-plant symbiosis in clay soils. Previous research has also highlighted the importance of soil texture in the role of AMF in N cycling (Zhai *et al.*, 2021) and the establishment of different AMF species (Mathimaran *et al.*, 2005; Verbruggen *et al.*, 2012).

Results for '*highS/highAl*' and '*highS/lowAl'* are shown in Figure 6(e), indicating hyphae are closer to regions of high-S and low-Al. This is consistent with the assumption that a high-S signal could be associated with microbial presence in the soil and that much of the microbial population chooses other sources to mine, (*e.g*. Fe-rich minerals (redox active) and organic material). There were no significant differences between the mean concentrations of P, S or Al of pixels (t-test) close and far from the hyphae measured in the control samples.

When interpreting these results, it should be recalled that SXRCT only detects hyphae in the soil pore-space, and as set out above, the destructive Jakobsen measurement suggests the majority of the hyphae are in the clay-water mixture phase (see Figure S1.30). Hence, mixed-phase pixels classified as far from hyphae may in fact be near to hyphae that were undetected by SXRCT. Nevertheless, it is likely that regions of mixed-phase surrounded by hyphae in the pore-space also contain a large number of hyphae (*i.e*. hyphal presence in pores is a proxy for hyphal presence in proximate mixed phase regions and soil/air interfaces). Figure 6(f) disregards pixel distance from hyphae, and shows the S XRF concentration distribution between the '*highP/highAl'* pixels and '*highP/lowAl'* pixels, demonstrating the '*highP/lowAl'* pixels have a significantly higher mean S concentration and a larger tail. Under the assumption that S is a surrogate marker for microbial presence, a similar conclusion as to Figure 6(e) could be made: the microbial community avoids Al and P rich clay minerals. Local biotoxicity and low bioavailability are both possible explanations for this observed behaviour.

Speciation of P and S via XANES

The collected XANES spectra of standard samples and a selection of spectra of the hyphal compartment samples are presented in Figure 1: see features (a-f). For S, thiol peaks (~2473 eV) and sulfonate (~2480.5 eV) and sulphate (~2482 eV) ratios are clearly observable which suggest differences in organic (cysteine, glutathione, sulfoxide, and sulfone) and inorganic S (sulfate cysteic, and sulfone) pools. For P, these spectral changes are subtle, with the largest changes observed around the fertilizer pellet in the *P+* samples - which showed signs of dissolution – said changes being indicated by a shift in the white line and disappearance of the shoulder after the white line (features d-f on Figure 1), suggesting adsorption of P to soil mineral surfaces. Spectral changes with increasing distance from fungal hyphae did not show significant differences.

Linear combination fits of the P and S XANES spectra (Figure 1) were performed in order to constrain the compositions. Results suggest that there is an increased organic-associated pool of P and oxidized S species close to the interface with the root compartment (Figures 7c and 8c&d). These results are in accordance with a previous study by our group (van Veelen *et al.*, 2020). These changes in speciation appear to be the result of an increased microbial community

and related biochemical activity in either the rhizosphere or hyphosphere. The increased microbial biomass resulting from C input by roots (*i.e.* rhizodeposits) or mycorrhizal fungi is known to play an important role in the S cycle (Leustek & Saito, 1999). The rate at which mineralisation of organo-sulphur in soils takes place is dependent on both the proportions of sulphate-esters and C-bonded S, and by the type of crop (Bertin *et al.*, 2003). (Vong, Phuy-Chhoy, 2003; Vong, P-C., 2003)Previous studies by Vong, P-C. (2003) found that addition of glucose or other C compounds, such as organic acids, to soil encourages rapid bacterial growth which transforms inorganic S into organic S. Similarly, significant organic S increase has been observed when cellulose was added as a C source (Chowdhury *et al.*, 2000). An increase in microbial biomass and organic matter will also increase the concentration of organic-associated P. There is also evidence that soil microbes actively compete with plants for available sulphate in the rhizosphere (Kertesz & Mirleau, 2004).

The presence of AMF caused an opposite trend in S speciation, with increased oxidized sulphur closer to hyphae (Figure 8a&b). These changes were not observed in the controls (SI-1 Figure S1.37), which indicates that the observed changes are microbiome related. The S composition of the organic structure presented in Figure 8 was determined to comprise amino acid thiols (64.1%), sulfonate (12%) and sulphate (23.9%). This suggests that the hyphae, together with bacteria, preferentially mine organic S (amino acids and sulphate esters). In addition, this suggests that AMF hyphae are actively involved in the conversion and transport of S to the plant roots (Allen & Shachar-Hill, 2009). This is another indication that S could be used as a surrogate marker for hyphal abundance, providing a proxy for protein, which is abundant within hyphae (Rak *et al.*, 2014). Finally, P speciation in both controls (SI-1 Figure S1.36) and inoculated treatments (SI-1 Figure S1.34) did not reveal clear trends with increasing distance from hyphae. Such a lack of clear changes in P speciation around hyphae might have been caused by their preferential mining of P from organic material which has a low overall P concentration. In addition, the competition and mutualism between microbes and AMF also influence the speciation of P in the soil microbiome. Finally, the different dynamics (*i.e.* competition between organisms, and mineral mining), as shown in Figure 6, will likely affect the P speciation.

The addition of a fertilizer pellet had an overall effect on the speciation of both P and S in the surrounding soil via dissolution and diffusion (see Figure 7). However, these effects were limited to the area directly adjacent to the pellet due to the absence of a significant mass flow of water and low P diffusion in soil. Consequently, it did not influence P speciation closer to the root compartment.

Modelling results

The results of the hyphal growth model fitted to the traditional destructive and SRXCT hyphal measurements (see SI-2) is shown on Table 1 together with a previous estimation of the same parameters using a different dataset by Schnepf *et al.* (2008a). We observed that the hyphal elongation rate $v \sim 10^{-5}$ cm s^{-1} estimated using the Jakobsen and SXRCT data are similar, as the pattern of soil colonisation is similar, but both are one order of magnitude faster than was estimated by Schnepf *et al.* (2008a). The root hyphal infection rate estimates based on our two methods of hyphal observation for $k \sim 10^{-4}$ $cm^2\ s^{-1}$ are an order of magnitude lower than estimated by Schnepf *et al.* (2008a). These differences between our hyphal parameter estimates and those estimated in Schnepf *et al.* (2008a) might be due to the fact that the previous estimates were based on experimental measurements (Jakobsen *et al.*, 1992b; Jakobsen *et al.*, 1992a) gathered using a much larger-scale assay, and whilst the fungal inoculum was the same,

602 the plant used in past studies (Jakobsen *et al.*, 1992b; Jakobsen *et al.*, 1992a) was not a wheat
603 plant, but clover (*Trifolium subterraneum* L.). Furthermore, a different soil was used between
604 the two experiments, and this would be expected to lead to different dynamics of AMF
605 colonisation.
606
607 When the SXRCT-based hyphal growth model was used to estimate the hyphal P uptake rate
608 λ, the values ranged between 10^{-11} and 1.86×10^{-5} cm^{-2} s^{-1} with a mean of 3.78×10^{-6} cm^{-2} s^{-1}
609 (Figure 9) which is in agreement with the value 3.26×10^{-6} cm^{-2} s^{-1} estimated by (Schnepf &
610 Roose, 2006). Estimates of λ based on data from destructive hyphal analysis ranged from 4.5
611 $\times 10^{-14}$ to 2.19×10^{-7} cm^{-2} s^{-1} with a mean of 1.05×10^{-7} cm^{-2} s^{-1}. These are much lower than
612 both the SXRCT estimate and that of Schnepf and Roose (2006). The destructive length density
613 measure is probably an overestimation of the true AMF hyphal length density, as it contains
614 non-AMF fungi, resulting in an underestimation of the true hyphal P uptake rate.
615

616 ## Conclusions

617 In this study we showed for the first time that AMF networks can be visualised in 3D within
618 soil using SXRCT, albeit as yet only in the soil pore spaces. We found that fungal structures
619 and morphology respond to local P levels (Cavagnaro *et al.*, 2005), *i.e.*, *P+* treatments had
620 many more hyphal structures than *P-* treatments. Whilst it has previously been shown that root
621 architectures respond to soil conditions in this manner (Shen *et al.*, 2011), this is possibly one
622 of the first studies to visualise *in situ* that AMF hyphae respond similarly to differences in soil
623 P conditions.
624
625 The modelling results unequivocally show that hyphal P uptake rates are an order of magnitude
626 lower than previously thought (Schnepf & Roose, 2006; Schnepf *et al.*, 2008a; Schnepf *et al.*,
627 2008b; Schnepf *et al.*, 2011) and an increase in hyphal length density does not lead to a
628 significant decrease in soil P content. Correlation of SXRCT and XRF/XANES data suggested
629 colocation of hyphal length density and P and S species in the clay/water soil phase, and
630 suggested S is a candidate proxy marker for hyphae in this soil phase. Our results also hint that
631 AMF hyphae might preferentially mine P from organic structures such as organic-matter
632 fragments, and confirm the hypothesis of microbial-fungal mutualism. As such, this would
633 suggest that AMF fungi (or organisms that they support in the hyphosphere) acquire nutrients
634 from more reactable labile and/or organic sources over clay minerals, even when soil is high
635 in P, supporting our observation of preferential P acquisition.
636
637 Finally, whilst many challenges remain, our study provides strong motivation for
638 interdisciplinary groups to apply fully-integrated experimental and modelling approaches to
639 the characterisation of plant-soil-fungi systems. Our study represents a necessary step forward
640 in the integration of different techniques to unravel the processes underpinning plant-AMF
641 interaction across different spatiotemporal scales (Ferlian *et al.*, 2018), demonstrating that
642 closer integration of theoretical and experimental techniques is important in gaining deeper
643 insights.
644

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References

Abbott LK, Robson AD. 1985. Formation of external hyphae in soil by four species of vesicular–arbuscular mycorrhizal fungi. *New Phytologist* **99**(2): 245-255.

Abbott LK, Robson AD, De Boer G. 1984. The effect of phosphorus on the formation of hyphae in soil by the vesicular-arbuscular mycorrhizal fungus, Glomus fasciculatum. *New Phytologist* **97**: 437-446.

Allen JW, Shachar-Hill Y. 2009. Sulfur transfer through an arbuscular mycorrhiza. *Plant Physiology* **149**: 549-560.

Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *Journal of Molecular Biolology* **215**: 403-410.

Bartnicki-Garcia S, Bracker CE, Gierz G, Lopez-Franco R, Lu HS. 2000. Mapping the growth of fungal hyphae: Orthogonal cell wall expansion during tip growth and the role of turgor. *Biophysical Journal* **79**(5): 2382-2390.

Berruti A, Lumini E, Balestrini R, Bianciotto V. 2015. Arbuscular Mycorrhizal Fungi as Natural Biofertilizers: Let's Benefit from Past Successes. *Front Microbiol* **6**: 1559.

Bertin C, Yang X, Weston LA. 2003. The role of root exudates and allelochemicals in the rhizosphere. *Plant and Soil* **256**(1): 67-83.

Boswell GP, Davidson FA. 2012. Modelling hyphal networks. *Fungal Biology Reviews* **26**: 30-38.

Buee M, De Boer W, Martin F, van Overbeek L, Jurkevitch E. 2009a. The rhizosphere zoo: An overview of plant-associated communities of microorganisms, including phages, bacteria, archaea, and fungi, and of some of their structuring factors. *Plant and Soil* **321**(1-2): 189-212.

Buee M, Reich M, Murat C, Morin E, Nilsson RH, Uroz S, Martin F. 2009b. 454 Pyrosequencing analyses of forest soils reveal an unexpectedly high fungal diversity. *New Phytologist* **184**(2): 449-456.

Callahan BJ, McMurdie PJ, Rosen MJ, Han AW, Johnson AJA, Holmes SP. 2016. DADA2: High-resolution sample inference from Illumina amplicon data. *Nature Methods* **13**(7): 581-+.

Camuti KS, McGuire PT. 1999. Preparation of polished thin sections from poorly consolidated regolith and sediment materials. *Sedimentary Geology* **128**(1): 171-178.

Caporaso JG, Lauber CL, Walters WA, Berg-Lyons D, Lozupone CA, Turnbaugh PJ, Fierer N, Knight R. 2011. Global patterns of 16S rRNA diversity at a depth of millions of sequences per sample. *Proceedings of the National Academy of Sciences of the United States of America* **108**: 4516-4522.

Carminati A, Vetterlein D, Weller U, Vogel HJ, Oswald SE. 2009. When Roots Lose Contact. *Vadose Zone Journal* **8**(3): 805-809.

Cavagnaro TR, Smith FA, Smith SE, Jakobsen I. 2005. Functional diversity in arbuscular mycorrhizas: exploitation of soil patches with different phosphate enrichment differs among fungal species. *Plant, Cell and Environment* **28**: 642-650.

Charters MD, Sait SM, Field KJ. 2020. Aphid Herbivory Drives Asymmetry in Carbon for Nutrient Exchange between Plants and an Arbuscular Mycorrhizal Fungus. *Curr Biol* **30**(10): 1801-1808 e1805.

Chowdhury MAH, Kouno K, Ando T. 2000. Critical sulphur concentration and sulphur requirement of microbial biomass in a glucose and cellulose-amended regosol. *Biology and Fertility of Soils* **32**(4): 310-317.

Daly KR, Keyes SD, Masum S, Roose T. 2015. Image based modeling of nutrient movement in and around the rhizosphere. Journal of Experimental Botany (In Press). *Journal of Experimental Botany (In Press).*

Doube M, Kłosowski MM, Arganda-Carreras I, Cordelieres FP, Dougherty RP, Jackson JS, Schmid B, Hutchinson JR, Shefelbine SJ. 2010. BoneJ: free and extensible bone image analysis in ImageJ. *Bone* **47**: 1076–1079.

Drew EA, Murray RS, Smith SE, Jakobsen I. 2003. Beyond the rhizosphere: growth and function of adbuscular mycorrhizal external hyphae in sands of varying pore sizes. *Plant and Soil* **251**: 105-114.

Edelstein L. 1982. The Propagation of Fungal Colonies - a Model for Tissue-Growth. *Journal of Theoretical Biology* **98**(4): 679-701.

Faber BA, Zasoski RJ, Munns DN, Shackel K. 1991. A Method for Measuring Hyphal Nutrient and Water-Uptake in Mycorrhizal Plants. *Canadian Journal of Botany-Revue Canadienne De Botanique* **69**(1): 87-94.

Ferlian O, Biere A, Bonfante P, Buscot F, Eisenhauer N, Fernandez I, Hause B, Herrmann S, Krajinski-Barth F, Meier IC, et al. 2018. Growing Research Networks on Mycorrhizae for Mutual Benefit. *Trends in Plant Science* **23**(11): 975-984.

Friese CF, Allen MF. 1991. The spread of VA mycorrhizal fungal hyphae in the soil: inoculum types and external hyphal architecture. *Mycologia* **83**(4): 409-418.

Gahan J, Schmalenberger A. 2014. The role of bacteria and mycorrhiza in plant sulfur supply. *Frontiers in Plant Science* **5**(723): 1-7.

Gene EL, Charles TD, Buso DC, Myron JM, Gary ML, Scott WB, Siccama TG, Reiners WA, Alewell C. 2002. The biogeochemistry of sulfur at Hubbard Brook. *Biogeochemistry* **60**.

Ghani A, McLaren RG, Swift RS. 1992. Sulphur mineralisation and transformations in soils as influenced by additions of carbon, nitrogen and sulphur. *Soil Biology and Biochemistry* **24**(4): 331-341.

Giovannetti M, Fortuna P, Citernesi AS, Morini S, Nuti MP. 2001. The occurrence of anastomosis formation and nuclear exchange in intact arbuscular mycorrhizal networks. *New Phytologist* **151**(3): 717-724.

Giovannetti M, Sbrana C, Avio L, Citernesi AS, Logi C. 1993. Differential hyphal morphogenesis in arbuscular mycorrhizal fungi during pre‐infection stages. *New Phytologist* **125**(3): 587-593.

Giovannetti M, Sbrana C, Avio L, Strani P. 2004. Patterns of below‐ground plant interconnections established by means of arbuscular mycorrhizal networks. *New Phytologist* **164**(1): 175-181.

Göransson P, Olsson, P.A., Postma, J., Falkengren-Grerup, U. 2008. Colonisation by arbuscular mycorrhizal and fine endophytic fungi in four woodland grasses - variation in relation to pH and aluminium. *Soil Biology & Biochemistry* **40**: 2260-2265.

Gross M. 2017. Where is all the phosphorus? *Current Biology* **27**: R1141-R1155.

Hadziavdic K, Lekang K, Lanzen A, Jonassen I, Thompson EM, Troedsson C. 2014. Characterization of the 18S rRNA Gene for Designing Universal Eukaryote Specific Primers. *PLoS One* **9**(2).

Hepper CM. 1984. Inorganic Sulfur Nutrition of the Vesicular Arbuscular Mycorrhizal Fungus Glomus-Caledonium. *Soil Biology & Biochemistry* **16**(6): 669-671.

Hodge A. 2014. Interactions between adbuscular mycorrhizal fungi and organic material substrates. *Advances in Applied Microbiology* **89**: 47—99.

Jakobsen I, Abbott LK, Robson AD. 1992a. External Hyphae of Vasicular-Arbuscular Mycorrhizal Fungi Associated with Trifolium-Subtarraneum L. 1. Spread of Hyphae and Phosphorus Inflow Into Roots. *New Phytologist* **120**(3): 371-380.

Jakobsen I, Abbott LK, Robson AD. 1992b. External Hyphae of Vesicular Arbuscular Mycorrhizal Fungi Associated with Trifolium-Subterraneum L .2. Hyphal Transport of P-32 over Defined Distances. *New Phytologist* **120**(4): 509-516.

Joner EJ, Magid J, Gahoonia TS, Jakobsen I. 1995. P depletion and activity of phosphatases in the rhizosphere of mycorrhizal and non-mycorrhizal cucumber (cucumis sativus L.). *Soil Biology & Biochemistry* **27**(9): 1145-1151

Kertesz MA, Mirleau P. 2004. The role of soil microbes in plant sulphur nutrition. *Journal of Experimental Botany* **55**(404): 1939-1945.

Keyes SD, Boardman RP, Marchant A, Roose T, Sinclair I. 2013. A robust approach for determination of the macro-porous volume fraction of soils with X-ray computed tomography and an image processing protocol. *European Journal of Soil Science* **64**(3): 298-307.

Keyes SD, Cooper L, Duncan S, Koebernick N, Fletcher DMM, Scotson CP, van Veelen A, Sinclair I, Roose T. 2017. Measurement of micro-scale soil deformation around roots using four-dimensional synchrotron tomography and image correlation. *Journal of the Royal Society Interface* **14**(136).

Koebernick N, Daly KR, Keyes SD, Bengough AG, Brown LK, Cooper L, George TS, Hallett PD, Naveed M, Raffan A, et al. 2019. Imaging microstructure of the barley rhizosphere: particle packing and root hair influences. *New Phytologist* **221**(4): 1878-1889.

Koebernick N, Daly KR, Keyes SD, George TS, Brown LK, Raffan A, Cooper LJ, Naveed M, Bengough AG, Sinclair I, et al. 2017. High-resolution synchrotron imaging shows that root hairs influence rhizosphere soil structure formation. *New Phytologist* **216**(1): 124-135.

Kolde R 2019. Pheatmap v1.0.12.

Kruger M, Stockinger H, Kruger C, Schussler A. 2009. DNA-based species level detection of Glomeromycota: one PCR primer set for all arbuscular mycorrhizal fungi. *New Phytologist* **183**(1): 212-223.

Lanzirotti A, Tappero R, Schulze DG 2010. Chapter 2 - Practical Application of Synchrotron-Based Hard X-Ray Microprobes in Soil Sciences. In: Singh B, Gräfe M eds. *Developments in Soil Science*: Elsevier, 27-72.

Leake J 2016. MycoRhizaSoil (https://www.soilsecurity.org/presentations-archive-page/#MycoAC2015)).

Leustek T, Saito K. 1999. Sulfate Transport and Assimilation in Plants. *Plant Physiology* **120**(3): 637.

Lucas SD, Jones DL. 2006. Biodegradation of estrone and 17 beta-estradiol in grassland soils amended with animal wastes. *Soil Biology & Biochemistry* **38**(9): 2803-2815.

Mathimaran N, Ruh R, Vullioud P, Frossard E, Jansa J. 2005. Glomus intraradices dominates arbuscular mycorrhizal communities in a heavy textured agricultural soil. *Mycorrhiza* **16**(1): 61-66.

McKay Fletcher DM, Keyes SD, Daly KR, Van Veelen A, Roose T. 2017. A multi image based approach for plant fertilizer interaction *Rhizosphere* **10**: 100152.

Novozamsky I, Houba VJG, Vaneck R, Vanvark W. 1983. A Novel Digestion Technique for Multi-Element Plant Analysis. *Communications in Soil Science and Plant Analysis* **14**(3): 239-248.

Oksanen J, Blanchet FG, Kindt R, Legendre P, O'Hara RG 2013. Vegan: community ecology package. R package version 2.0–10. Available: http://CRAN.R-project.org/package = vegan.

Olsson PA, Baath E, Jakobsen I. 1997. Phosphorus effects on the mycelium and storgae structures of an adbuscular mycorrhizal fungus as studies in the soil and roots by analysis of fatty acid signatures. *Applied and Environmental Microbiology* **63**(9): 3531-3538.

Paganin D, Mayo SC, Gureyev TE, Miller PR, Wilkins SW. 2002. Simultaneous phase and amplitude extraction from a single defocused image of a homogeneous object. *Journal of Microscopy-Oxford* **206**: 33-40.

Prietzel J, Botzaki A, Tyufekchieva N, Brettholle M, Thieme J, Klysubun W. 2011. Sulfur Speciation in Soil by S K-Edge XANES Spectroscopy: Comparison of Spectral Deconvolution and Linear Combination Fitting. *Environmental Science & Technology* **45**(7): 2878-2886.

Prietzel J, Thieme J, Salomé M, Knicker H. 2007. Sulfur K-edge XANES spectroscopy reveals differences in sulfur speciation of bulk soils, humic acid, fulvic acid, and particle size separates. *Soil Biology and Biochemistry* **39**(4): 877-890.

Pruesse E, Peplies J, Glockner FO. 2012. SINA: accurate high-throughput multiple sequence alignment of ribosomal RNA genes. *Bioinformatics* **28**(14): 1823-1829.

Rak M, Salome M, Kaminskyj SG, Gough KM. 2014. X-ray microfluorescence (muXRF) imaging of Aspergillus nidulans cell wall mutants reveals biochemical changes due to gene deletions. *Anal Bioanal Chem* **406**(12): 2809-2816.

Rozmos M, Bukovska P, Hrselova H, Kotianova M, Dudas M, Gancarcikova K, Jansa J. 2021. Organic nitrogen utilisation by an arbuscular mycorrhizal fungus is mediated by specific soil bacteria and a protist. *ISME J*.

Scherer HW. 2009. Sulfur in soils. *Journal of Plant Nutrition and Soil Science-Zeitschrift Fur Pflanzenernahrung Und Bodenkunde* **172**(3): 326-335.

Schindelin J, Arganda-Carreras I, Frise E. 2012. Fiji: an open-source platform for biological-image analysis. *Nature Methods* **9**(7): 676-682.

Schnepf A, Himmelbauer M, Loiskandl W, Roose T. 2005. Modelling the rhizosphere. *Comparative Biochemistry and Physiology a-Molecular & Integrative Physiology* **141**(3): S222-S222.
Schnepf A, Jones D, Roose T. 2011. Modelling Nutrient Uptake by Individual Hyphae of Arbuscular Mycorrhizal Fungi: Temporal and Spatial Scales for an Experimental Design. *Bulletin of Mathematical Biology* **73**(9): 2175-2200.
Schnepf A, Roose T. 2006. Modelling the contribution of arbuscular mycorrhizal fungi to plant phosphate uptake. *New Phytologist* **171**(3): 669-682.
Schnepf A, Roose T, Schweiger P. 2008a. Growth model for arbuscular mycorrhizal fungi. *Journal of the Royal Society Interface* **5**(24): 773-784.
Schnepf A, Roose T, Schweiger P. 2008b. Impact of growth and uptake patterns of arbuscular mycorrhizal fungi on plant phosphorus uptake - a modelling study. *Plant and Soil* **312**(1-2): 85-99.
Shen J, Yuan L, Zhang J, Li H, Bai Z, Chen X, Zhang W, Zhang F. 2011. Phosphorus dynamics: from soil to plant. *Plant Physiol* **156**(3): 997-1005.
Simard SW, Beiler KJ, Bingham MA, Diselippe JR, Philip LJ, Teste FP. 2012. Mycorrhizal networks: Mechanisms, ecology and modelling. *Fungal Biology Reviews* **26**: 39-60.
Smith SE, Smith FA, Jakobsen I. 2003. Mycorrhizal fungi can dominate phosphate supply to plants irrespective of growth responses. *Plant Physiology* **133**(1): 16-20.
Sole A 2020. PyMca Software tool via http://pymca.sourceforge.net.
Thonar C, Schnepf A, Frossard E, Roose T, Jansa J. 2011. Traits related to differences in function among three arbuscular mycorrhizal fungi. *Plant and Soil* **339**(1-2): 231-245.
Tlalka M, Hensman D, Darrah PR, Watkinson SC, Fricker MD. 2003. Noncircadian oscillations in amino acid transport have complementary profiles in assimilatory and foraging hyphae of Phanerochaete velutina. *New Phytologist* **158**(2): 325-335.
Vaccari DA. 2009. Phosphorus: a looming crisis. *Sci Am* **300**(6): 54-59.
van Veelen A, Koebernick N, Scotson CS, McKay-Fletcher D, Huthwelker T, Borca CN, Mosselmans JFW, Roose T. 2020. Root-induced soil deformation influences Fe, S and P: rhizosphere chemistry investigated using synchrotron XRF and XANES. *New Phytologist* **225**(4): 1476-1490.
Verbruggen E, Van Der Heijden MG, Weedon JT, Kowalchuk GA, Roling WF. 2012. Community assembly, species richness and nestedness of arbuscular mycorrhizal fungi in agricultural soils. *Mol Ecol* **21**(10): 2341-2353.
Vong P-C. 2003. Immobilized-S, microbial biomass-S and soil arylsulfatase activity in the rhizosphere soil of rape and barley as affected by labile substrate C and N additions. *Soil biology & biochemistry* **v. 35**(no. 12): pp. 1651-1661-2003 v.1635 no.1612.
Vong P-C. 2003. Immobilized-S, microbial biomass-S and soil arylsulfatase activity in the rhizosphere soil of rape and barley as affected by labile substrate C and N additions. *Soil biology & biochemistry* **35**(12): 1651-1661.
Vosátka M, Batkhuugyin E, Albrechtová J. 1999. Response of Three Arbuscular Mycorrhizal Fungi to Simulated Acid Rain and Aluminium Stress. *Biologia Plantarum* **42**(2): 289-296.
Zhai S, Wu Y, Xu C, Chen W, Feng J, Zheng Q, Meng Y, Yang H. 2021. Symbiotic soil fungi suppress N2O emissions but facilitate nitrogen remobilization to grains in

sandy but not clay soils under organic amendments. *Applied Soil Ecology* **167**: 104012.

Zhang S, Zhou J, Wang G, Wang X, Liao H. 2015. The role of mycorrhizal symbiosis in aluminum and phosphorus interactions in relation to aluminum tolerance in soybean. *Applied Microbiology and Biotechnology* **99**: 10225—10235.

Tables

Table 1. Estimated best-fit model parameters for hyphal length density measured using traditional Jakobsen destructive soil sample testing (first column), and SRXCT data (middle column), in comparison to the values estimated in Schnepf et al. (2008) (last column).

		Model fitting parameters for hyphal length densities obtained by Jakobsen destructive testing method	Model fitting parameters for hyphal length densities obtained by SRXCT analysis method	Values from Schnepf et al. (Schnepf *et al.*, 2008a)
k	[$cm^{-2}\ s^{-1}$]	$4.66*10^{-4} \pm 1.758*10^{-4}$	$1.02*10^{-4} \pm 1.767*10^{-4}$	$2.89*10^{-3}$
b	[s^{-1}]	$2.07*10^{-6} \pm 1.386*10^{-6}$	$5.45*10^{-6} \pm 1.273*10^{-5}$	$5.79*10^{-9}$
v	[$cm\ s^{-1}$]	$1.64*10^{-5} \pm 9.047*10^{-6}$	$2.06*10^{-5} \pm 2.566*10^{-5}$	$2.89*10^{-6}$
R^2	-	$9.89*10^{-1} \pm 7.680*10^{-3}$	$2.34*10^{-1} \pm 2.53*10^{-1}$	-

Figureure Captions

Figure 1: S and P K-edge XANES spectra. The top set of spectra are from standards used for linear combination fits (LCF). The bottom set of spectra are point measurements as a function of distance from the segmented hyphae. The selection of collected spectra from the samples show spectral differences. For S, the ratios between different peaks are particularly important. The vertical dashed lines represent the energies of (a) L-cysteine, (b) sulfonate and (c) sulfate. Some spectra show contrasts, e.g. IP- T4 at the end of the compartment closest to roots (predominantly cysteine) and IP+ T2 at the fertilizer pellet location (predominantly sulfate). For P, the spectral differences are much more subtle. However, the differences are measurable. In particular, CaPO4-like species have a shifted white line (the sharp feature in the K-edge spectrum) towards lower energy (feature d), a shoulder (feature e) and oscillation (f) at higher energies. Interestingly, the shoulder for CaPO4-type species disappears with increasing distance from the fertilizer pellet, suggesting dissolution and re-adsorption to different mineral surfaces. Spectral changes with increasing distance from fungal hyphae do not show significant differences.

Figure 2: SXRCT imaging and image processing protocols enabled extraction of air-based hyphal morphology. (a) and (b) show sample images of 3D hyphal morphology for exemplary samples from c*ontrol* and *inoculated* treatments*;* (c) shows the mean (red line) hyphal length density, the blue box describes the inter quartile range (i.e. 25%-75% of data falls within this interval), red dots indicate outliers, and black whiskers show $\pm 2.7\boldsymbol{\sigma}$ where $\boldsymbol{\sigma}$ is the standard deviation; (d) shows the average hyphal length density for all different treatments at t=2 weeks, along with the standard deviation, (e) shows the average hyphal length density for all different treatments at t=4 weeks along with the standard deviation. On (d) and (e) h1=13.5 mm, h2= 28.5 mm, and h3=43.5 mm from the interface of he hyphal compartment to the adjoining root compartment along the hyphal compartment midline.

Figure 3: Principal component analysis (PCA) of microbial sequencing results.

Figure 4: (a) Trypan stained mycorrhizal structures (*Rhizosphagus Irregularis*) in wheat roots from root compartments. Staining confirms that plants from all inoculated samples were successfully colonised by mycorrhizal fungi. (b) Microscope image of mycorrhizal structures from a root compartment showing hyphae emerging from a root into the soil at the interface with a hyphal compartment. (c) Root and shoot dry weight and P content measurements. The significant differences (t-test, $p<0.05$) are detected for: P+ v P- root and shoot weights at t=2 weeks, shoot and root P content at T=2 weeks and T=4 weeks for P+ v P-, and T=4 weeks P+ v P-, and dry root weight.

Figure 5: P, S and distance from hyphae in a two week time point (T2) IP+ treatment shown in their 3D environment. The green tubes are the hyphae segmented from the SRXCT data and the brown (red-grey'ish opaque) shows organic structure segmented from the SRXCT data. (a) The P heat map correlated to SRXCT. (b) S heat map correlated to XCT. (c) The distance transform of the segmented hyphae correlated to the P and S XRF maps and XCT data. Bottom row (d-f) show the P, S and distance transform from hyphae maps used in the correlation with SRXCT data. (d) The P heat map shows P concentration. (e) The S heat map shows S concentration. (f) The distance from hyphae, heat map shows distance from nearest segmented hyphae in 3D in microns. The images show a clear preferential relationship between organic

material in the soil and fungal hyphae. The hyphae are pointing towards this structure which is high in S and relatively low in P. Concentration a, b, d and e s are expressed in ppm (mg kg^{-1}).

Figure 6: Histograms of XRF P, S and Al concentrations in the clay-water mixture phase over all inoculated samples and planting times separated by distance from hyphae. (a) –(c) P,S and Al respectively distributions of XRF pixels within 50 µm of a hyphae ('Close to hyphae') and further than 200 µm of a hyphae ('Far from hyphae'). The distance class means, shown in the legend, are significantly different for only P and Al. (d) Histogram of distances from hyphae of XRF pixels with both High P (>9.7 g kg^{-1}) and Low Al (25 g kg^{-1}) mean=884 µm and High P and High Al (>90 g kg^{-1}) mean =1100 µm. The means are significantly different with $p<<0.01$. (e) Histogram of distances from hyphae of XRF pixels with both High S (>780 mg kg^{-1}) and Low Al mean=840 µm and High S and High Al mean =1055 µm. The means are significantly different with $p<<0.01$. (f) Histogram of S XRF counts with both High P Low Al, mean 980 mg kg^{-1} and High P High Al, mean 595 mg kg^{-1} Means are significantly different with $p<<0.01$. Distances were calculated using the correlated SRXCT images. The bin width was selected as 1000 raw XRF units for (a), (b) and (c) and 100 µm for (d) and (e). The bin width for (f) was 100 raw XRF units for High P Low Al and 50 raw XRF units for High P High Al. The number of pixels used to create the (a), (b) and (c) histogram was 1207 for those within 50 µm and 603410 for those further than 200 µm. Each histogram was normalized by the total counts.

Figure 7: Linear combination fitted XANES results of P K-edge with respect to distance from the root compartment, showing (a) apatite-like and soil mineral P fractions, (b) organic associated PO4, (c) the collected elemental maps which show how the distances are related, and (d) the postulated P interactions based on the XANES results. The blue highlighted ellipsoids (a and b) and vertical blue bar (c) denotes the area affected by the TSP fertilizer. The data shows no clear trends for apatite-like and mineral-associated P (a). However, the organic-associated P fraction is increased closer to the roots as depicted in (d). Units are in mass fraction (0-1) with 1 meaning 100%. The residual P fractions of (a) and (b) add up to 1. Error bars represent standard deviation.

Figure 8: Linear combination fitted XANES results of S K-edge. (a) represents the reduced (i.e. amino acid thiols) and oxidized (i.e. sulfonate/sulfate) S species distribution versus distance from segmented hyphae derived from SRXCT data. The data shows that closer to hyphal surfaces, more amino acid thiols (e.g. cysteine) and fewer oxidized S species (e.g. sulfonate and sulfate) are detected. (b) represents the reduced and oxidized S species distributions with respect to distance from the interface of the hyphal compartment with the associated root compartment. Here data show an opposite trend: more oxidized S species closer to the roots and less amino acid thiols. Units are in are mass fraction (0-1) with 1 meaning 100%. Error bars represent standard deviation.

Figure 9: Model-fitted hyphal P uptake rate λ for the inoculated samples with (IP+) and without (IP-) fertiliser treatments using both the SXRCT and destructively measured hyphal parameters. Error bars represent the standard deviation.
Error bars represent the standard deviation