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POPULATION BIO-CULTURAL HISTORY IN THE SOUTH AEGEAN DURING THE BRONZE AGE

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APPENDIX A
ENVIRONMENTAL FRAMEWORK TO THE RESEARCH

This section reviews information on the climatic conditions of the Bronze Age South Aegean, as well as the geological composition of the Argolid (Mainland), Naxos (Cyclades), and Crete.

A.1 Climate

The climatic conditions, suggested to have been prevailing in the South Aegean during the Bronze Age, are reviewed in order to explore, whether it is legitimate to expect sufficient differences between the Mainland, Crete and the island of Naxos, concerning the selection pressures operating on the skeletal biology of the three populations that would have accounted for inter-population heterogeneity of skeletal morphology.

Concerning the climatic conditions in the Bronze Age South Aegean, these would have been slightly warmer and drier than at present. The climate was semi-arid Mediterranean, characterized by long, hot, dry summers and cool largely frost-free winters, when the highest percentage of rainfall is recorded. In the northernmost parts of the Mainland, the climate is more continental (Dickinson 1994). The present climatic conditions suggest significant temperature differences between different parts of Greece, due to its geographical position, the particular variegated landscape and the proximity of the sea, which creates milder conditions (Mariolopoulos 1953). Mountain ranges partition the landscape and account for considerable inter-regional differences in climatic and vegetational conditions. A relatively recent incident in the north-eastern Peloponnese suffices to appreciate how inaccurate it is to infer the climatic conditions prevailing in one region on the basis of pollen data derived from another, regardless of how close they may be. In the summer of 1986, a sudden and severe hailstorm destroyed much of the vine crop in the Asopus valley. Surprisingly, the Nemea valley, to the east, suffered much less destruction. Further east, in the Logopotamus valley, there was no documentation of damage. These three valleys are within a distance of 20Km east to west (Hansen 1988).

During the autumn, winter and spring seasons, the layout of the isothermals accords well with the geographical latitude. Nevertheless, this layout is not preserved during the summer, when the sea and the proximity of the sites to it is the prime determinant of the temperature. Focusing on the South Aegean, the isothermal of 18.5^{0}C passes through the north part of the Peloponnese (Mycenae, Prosymna, Asine, Lerna) and Attica, whereas the island of Naxos is crossed by the 19^{0}C isothermal. Galatas in the Argolid, appears to occupy a marginal position between these two isothermals. Further to the south, Crete is crossed by the 19.5^{0}C isothermal, with an average annual temperature of 19^{0}C at Herakleion (5km to the North of Knossos). The values for the average annual temperature for the Argolid and the island of Naxos are lower, calculated as 18,1^{0}C and 18,4^{0}C respectively (Mariolopoulos 1953).

The highest annual temperature, 45.7/6^{0}C, is recorded at Herakleion (Crete), whereas at Nafplion (Argolid) is measured as 42,5/7^{0}C and on Naxos is significantly lower, measured as 36.8/7^{0}C. The discrepancy in the temperature values between the three regions becomes less pronounced, when the average highest temperature is considered. This was calculated as 23^{0}C, 22.9^{0}C and 21^{0}C for Herakleion, Nafplion and Naxos respectively. Moreover, the values for the average lowest annual temperature are estimated as 13.1^{0}C at Nafplion, 15^{0}C at Herakleion and 15.8^{0}C on Naxos. Inter-regional difference in temperature is exacerbated, if we consider the absolute lowest temperature in the three regions. This is recorded at -4.0^{0}C, 1.2^{0}C and 0.1^{0}C at Nafplion, Naxos and Herakleion respectively.

The humidity levels in the South Aegean show an unusual fluctuation throughout the course of the year. The annual average relative humidity at Herakleion is calculated as 65% and is lower than that for Nafplion and Naxos, calculated as 68% and 70% respectively. On Crete, during the summer period, the high temperature in combination with the dry atmosphere, and the strong wind do not permit to any plant that is not irrigated or supplied with ground water or deeply rooted to thrive (Rackham & Moody 1996).

The rainfall is dispersed unequally across the Aegean. The pattern of this dispersal is largely conditioned by the proximity of the land to the sea and the high relief. Across the Aegean, the amount of rainfall decreases from the north to south and from west to east. This is particularly interesting on Crete, where the amount of winter rain decreases along the north coast from circa 700mm to 350mm from West to East and from North to South (Dickinson 1994). It increases from the coast towards the interior of the island. Crete has a very broad range of rainfall that is compatible with that of the United States of America (Rackham & Moody 1996). The south coast, in contact with the Libyan sea, is in general terms hotter. The sirocco or south-eastern wind, blowing off the Sahara mainly in spring and autumn, can bring intense heat.

When the annual levels of precipitation for the three regions are comparatively examined, the calculated values for Nafplion and Herakleion do not differ greatly; these are 514.5mm and 521.8mm respectively. On the other hand, the island of Naxos appears to be drier. Its annual height of precipitation is recorded at 418.6mm. The rainfall, principally in the form of violent storms, is suggested to exacerbate the denudation of the high mountains on Crete (Nevros & Zvorykin 1939). Likewise, in the Argos plain rainfalls are brief violent episodes and thus of little value to agriculture (De Vooys & Piket 1958).

To conclude, although inter-regional variation in climatic conditions is not expected to have been conspicuously great, the selection pressures operating upon the Argolid, Naxos and Central Crete populations, introduced by the variety of microclimates and ecosystems in the South Aegean, must have been different enough between the three regions examined in this study, so as not to promote a biological homogeneity between the respective populations.

A.2 Geological Composition of the South Aegean

The ability to use strontium isotopes to track population movement relies on variation in the Sr isotopic composition in the local geology (Graustein 1989). Differences in the geological composition of the Argolid, Central Crete and Naxos will allow the analysis of strontium isotope ratio ($^{87}Sr/^{86}Sr$) to detect, if there was any migration (movement and settlement) of individuals between the three regions. More information on the principals of strontium isotope ratio analysis and its contribution to the more accurate answering of questions related to population movement and residential change is provided in Chapter 5.

A.2.1 Tectonic Zones in the South Aegean

The geological composition of the South Aegean is strongly affected by tectonic activity producing complex geology. Tectonic/isopic zones are groups of rocks that share a common history (Figure 2. 3) (Higgins & Higgins 1996:17).

Argolid-Mainland: The Argolid (Mainland) is the most geologically varied region of the South Aegean compared to Crete and the island of Naxos. It is crossed by the Pindus and the Parnassus tectonic zones, and at its easternmost part by the Sub-Pelagonian Zone. Focusing on the sites for which skeletal material was studied for the present project, Lerna belongs to the Pindus zone, whereas Apatheia-Galatas falls within the borders of the Sub-pelagonian Zone (Ager 1986). Mycenae, where the people who settled and dominated politically Knossos at the end of LMIB are suggested to have come from, also belongs to the Pindus zone.

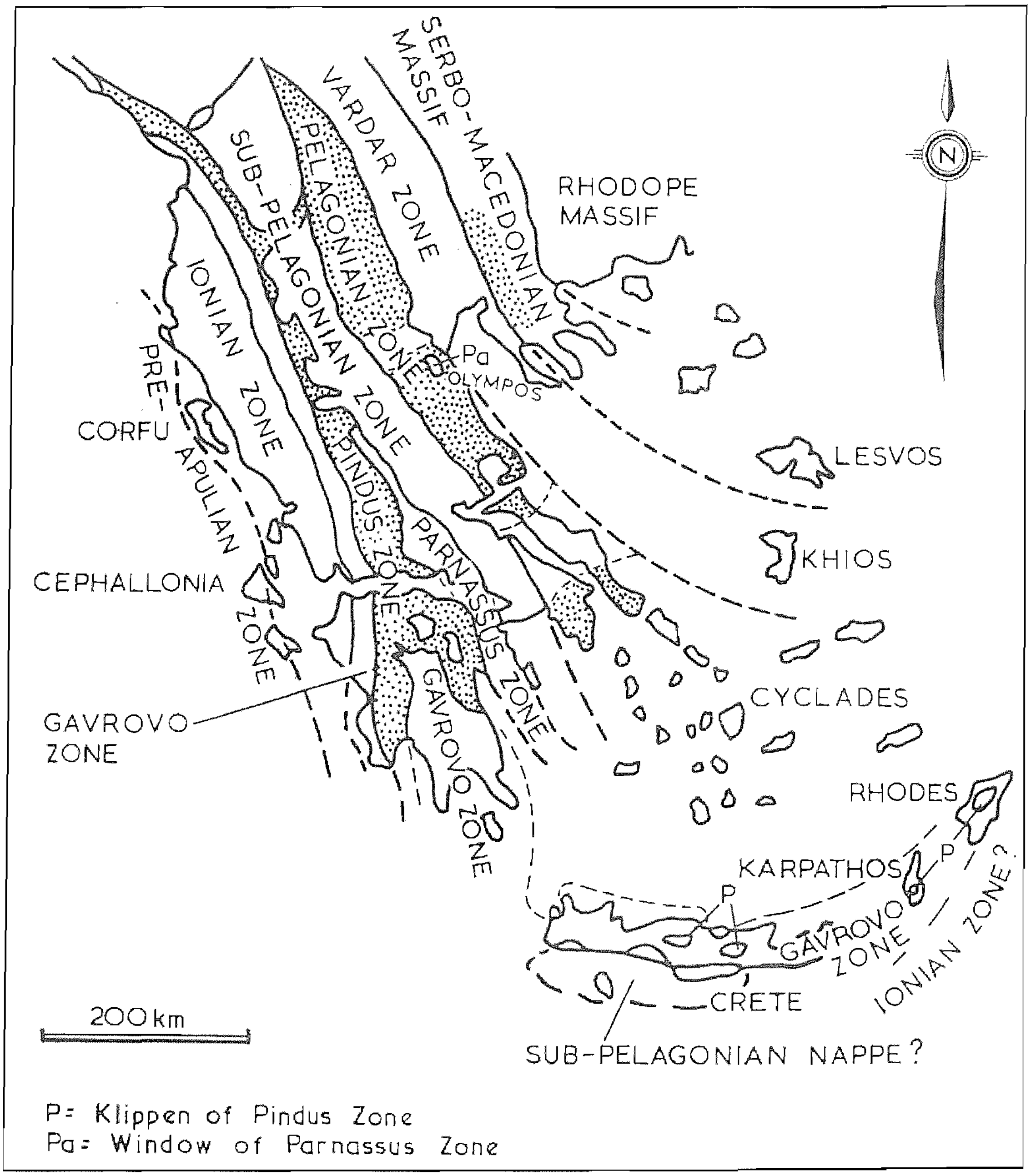


Figure 2.3 Sketch-map of the main tectonic zones in the Aegean (after Aubouin *et al.* 1963, cited by Ager 1986:502).

The Pindus Zone is allochthon and constitutes a distinct palaeogeographical entity between the shallow-water carbonates of the Parnassus Zone on one side, and the Gavrovo Zone on the other. Structurally, this zone is extremely complex. The whole Mesozoic amounts to just a thousand meters of sediment and has probably been diverted from its original depositional trough. It consists mainly of deep-water facies, with some intercalated micro-breccias of shallow-water material indicating the margins of the original trough. The Trias consists of thin-bedded siliceous limestones and shales succeeded by multicoloured radiolarites, often with mangenese of the Jurassic and the Earliest Cretaceous. Postdating that, flysch deposition interceded for a short period, succeeded by more pelagic limestones in the later Cretaceous and a main flysch episode beginning before the end of the Cretaceous and lasting until the Eocene (Ager 1986).

Crete: The Gavrovo Zone represents the autochthon in the South Aegean. Here, the sedimentary record begins in the late Jurassic with algal limestones, which continue to the early Cretaceous. This is significantly thick and consists of reef limestones with masses of in situ rudists and other horizons with nerineid gastropods. In the Lower and Middle Eocene, it is characterized by massive carbonates, whereas flysch sedimentation took place in the late Eocene. On Crete, this zone forms the rugged topography of the island, the White Mountains. However, the fertile lowlands of the Aegean region represent Neogene and Quaternary deposits (Ager 1986).

Naxos-Cyclades: Naxos falls within the Pelagonian/Attic-Cycladic metamorphic belt. The oldest rocks are schists, gneisses, and granites some of which date to the Palaeozoic. The age for some of the rocks is similar to that of the adjacent zones (Sub-Pelagonian and Vardar Zones). Moreover, the Alpine compressions created metamorphic conditions (Higgins & Higgins 1996).

A.2.2 Surface Geology and Soil Typology

Studies of Greek soils group them in two main categories, the Inter-zonal Limestone and the Zonal Non-Limestone soils. The two types are differentiated in terms of their sensitivity to the local climate. The first are suggested to be very little affected by the local climate, in contrast to the Non-Limestone ones, which reflect the

Greek climate and its regional variations (Bintliff 1977). Terra rossa produced by the weathering of pure limestone or marble, rich in iron oxides and clay, is a fertile soil, but usually too thin on the hill sides for successful agriculture. Redzina on the other hand, produced by marl and flysch, is considered to be of the most productive Aegean soils (Higgins & Higgins 1996).

Crete: On Crete, the project focuses principally on the Knossos district. This is an area of lowland, where non-alluvial arable land is predominant. This land is of good quality, but difficult to irrigate. Geologically, it consists of marls, sandstones and conglomerates, which belong to the late Tertiary or Neogene, extending into the Quaternary. These dense, soft white marls (known locally as kouskouras) were favoured during the Bronze Age for the construction of rock-cut tombs. It is from the kouskouras bedrock that the redzina soils derive, which are particularly suited to the cultivation of vine and olive trees, due to their high potassium consistency (Hood & Taylor 1981). Redzina soils are good at retaining water during the dry summer period, contain nitrogen from humus and other organic debris, and are easy to cultivate (Bintliff 1977). Aeolianite, a sandstone dating from the glacial periods, was employed from the Bronze Age onwards in the construction of buildings. Conglomerates, on the other hand, are natural concretes, composed of weathered river pebbles cemented together and can be of any date (Rackham & Moody 1996).

At the south-eastern coast of Crete, the settlement of Myrtos Pyrgos occupies a flat-topped hill on the east side of the mouth of the river Myrtos. The hill is composed predominantly of poorly cemented fossiliferous marl strata interbedded with laminated sands, collectively named by Fortuin (1977) as the "Makrilia" formation. Deposition of the "Makrilia" sediments dates to the late Miocene. At certain points the foundation walls at Myrtos-Pyrgos lay on carbonate-rich sandstone and beach-rock strata. This constitutes a thin layer of the so-called "Ammoudares", which overlies the "Makrilia" formation on the hill (*ibid.*). Selection of the site must have been determined by the river valley and the fertile soils provided by the narrow strip of alluvium on both sides of it (Myers *et al.* 1992). On southern Crete, the cemetery of Moni Odigitria is situated on a the flat hilltop of Chatzinas Liophyto, between flysch deposits of the Pindos-Ethia nappe and the overlying upper Miocene marine and brackish-water marls. The tholos tombs were constructed on a small ridge of Mio-Pliocene limestone (Myers *et al.* 1992).

Chania, in western Crete, lies in a small, asymmetrical graben between the White Mountains to the south and the hills of the peninsula of Akrotiri to the north-east that consist of mainly Triassic-Cretaceous limestones. The modern city occupies the site of the Bronze Age Kydonia and is constructed on Miocene marls and Pleistocene alluvial deposits (Higgins & Higgins 1996).

Argolid-Mainland: In the Argolid, the citadel of Mycenae occupies a hill of hard Late Triassic to Middle Jurassic limestone, with small steep hills to the north and south, and a mountain range to the east, of similar geological composition. The limestone terraces of the mountains to the east are covered by Older Fill among rocky slopes. The major deposition of the Older Fill took place during the Würm Glacial period and ceased c. 30 /40,000 to 12,000 years ago. The Older Fill constitutes a poor cereal and, to a lesser extent, olive land. It has been suggested to have had supported woodland vegetation, with open savannah woodland in the South-east. This is in contrast to the dense forests of Mediterranean oaks and lowland pines found in the north of Greece (Bintliff 1977). The greatest part of the valley between the hills, on which Mycenae lies, and the ridge that runs from west of the citadel towards the south, consist of marls and conglomerates. These were deposited by rivers and streams during the Late Pliocene to Pleistocene (Higgins & Higgins 1996). Further to the east, is the watershed area leading to the basin of Berbati, which consists mainly of bare limestone and scrub. On basis of these data, the local soils at Mycenae appear to have been relatively infertile (Bintliff 1977). In the Late Helladic period, the spring at the foot of Mt Ayios Elias, 200m east of the citadel, was channelled underground to the "Secret Cistern" beside the fortification walls. This spring is supplied by rainwater that falls on the hill. Its underground passage is blocked by Pleistocene marls, whereas on its way to the surface, it passes through scree deposited from the hill in ancient times (Higgins & Higgins 1996).

Likewise, the prosperity enjoyed by the settlement of Lerna during the Bronze Age cannot be adequately justified on the basis of the fertility of its soils. The tell on which it was founded, occupies most of the narrow pass between Mt. Pontinos and the sea. Mt. Pontinos and the other ridges to the west are steep and barren, hard limestone formations. Older Fill alluvium is deposited to its west and south, where it is interrupted by areas of marine sands and marls of Plio-Pleistocene age. This marine formation was probably the best soil, exploited by the population of

Lerna for the cultivation of cereals and olives. To the east of the settlement, recent alluvium overlies the colluvium. The recent alluvium deposited along the shore and on the southern part of the Argos plain (to the north of the settlement) dates to the historical times. In the Bronze age, it is suggested that this area was sea and brackish delta marsh. Despite the great number of springs on the tell, their journey to the sea is too short to have allowed the practice of an extensive irrigated agriculture (Bintliff 1977).

At Apatheia Galatas, soil samples have been taken to determine its geological composition but to date there are no published results (Konsolaki, pers. comm. 2003). Research on the island of Poros and the Peninsula of Methana, in the immediate vicinity of Apatheia Galatas, revealed that Pliocene/Quaternary volcanic rocks contribute greatly to their geological constitution, particularly the large pebbles in bays. These rocks contain plentiful clay and soil nutrients, and in depressions they form a rather heavy, but moist soil (Bintliff 1977). Moreover, the greenish colour of stones and soil recovered from the interior of the tombs, where the studied collections derive from, suggests the presence of serpentine rock at the site. Serpentine contains many useful nutrients for a rich soil and in the lowland, its weathering products formed an invaluable arable resource (Bintliff 1977).

Naxos-Cyclades: The island of Naxos in the Cyclades is a metamorphic core complex. The eastern part of the island is dominated by marble and dolomite with layers of schist, and its centre, by schist with layers of marble. These rocks, which were originally limestones and shales, deposited on the Hercynian basement, underwent several metamorphic events, notably 45 and 25 million years ago. The bauxite layers in the original limestones were transformed into emery, which contains the hard mineral corundum together with magnetite, haematite and margarite mica. Following metamorphism, granite was emplaced about 12 million years ago and it underlies the greatest part of the western side of the island. During the Tertiary period, sedimentary rocks were deposited on parts of both the granite and metamorphic rocks. Later on, these sediments were transferred on the granite by the low-angle normal faulting. The area of the two LC cemeteries from which skeletal material was studied is underlain by granite. Granite in the South Aegean breaks down easily to produce fine gravel that weathers to form a fertile soil. The geological composition of the settlement that most probably used the two LC cemeteries, despite being in the vicinity of the two cemetery sites, is different. It consists of well-cemented Pleistocene conglomerates (Higgins & Higgins 1996).

To conclude, on the basis of the differences between the local geology of the Argolid, Naxos and Crete, this study assumes differences between the three regions in terms of the range of the local biologically available strontium isotope ratio and applies the strontium isotope ratio analysis to explore hypotheses for the movement of people from the Argolid to Knossos (Crete) and Naxos. The assumption is tested by determining the local at the three regions biologically available strontium isotope ratio from archaeological and modern animal tissue and archaeological human bone.

APPENDIX B

LISTS OF METRIC AND NON-METRIC DATA

RECORDED FROM EACH SKELETON:

Description of measurements and Definition of non-metrics

Table 5.1 Recorded cranial metric variables and abbreviations used in the text.

CRANIAL MEASUREMENTS - CODES			
MEASUREMENT DESCRIPTION	CODE	MEASUREMENT DESCRIPTION	CODE
Glabello-Occipital Length	GOL	Simotic Chord	WNB
Nasio-Occipital Length	NOL	Simotic Subtense	SS
Basion-Nasion Length	BNL	Malar Length Inferior	IML
Basion-Bregma Height	BBH	Malar Length Maximum	XML
Maximum Cranial Breadth	XCB	Malar Subtense	MSUB
Maximum Frontal Breadth	XFB	Cheek Height	WMH
Bistephanic Breadth	STB	Supraorbital Projection	SOS
Bizygomatic Breadth	ZYB	Glabella Projection	GLS
Biauricular Breadth	AUB	Foramen Magnum Length	FOL
Minimum Cranial Breadth	WCB	Nasion-Bregma Chord	FRC
Biasterionic Breadth	ASB	Nasion-Bregma Subtense	FRS
Basion-Prosthion Length	BPL	Nasion-Subtense Fraction	FRF
Nasion-Prosthion Height	NPH	Bregma-Lambda Chord	PAC
Nasal Height	NLH	Bregma- Lambda Subtense	PAS
Orbit Height Left	OBH	Bregma-Subtense Fraction	PAF
Orbit Breadth Left	OBB	Lambda-Opisthion Chord	OCC
Bijugal Breadth	JUB	Lambda-Opisthion Subtense	OCS
Nasal Breadth	NLB	Lambda - Subtense Fraction	OCF
Palate Breadth (2nd M)	MAB	Vertex Radius	VRR
Mastoid Height	MDH	Nasion Radius	NAR
Mastoid Breadth	MDB	Subspinale Radius	SSR
Bimaxillary Breadth	ZMB	Prosthion Radius	PRR
Bimaxillary Subtense	SSS	Dacryon Radius	DKR
Bifrontal Breadth	FMB	Zygoorbitale Radius	ZOR
Nasio-Frontal Subtense	NAS	Frontomalare Radius	FMR
Biorbital Breadth	EKB	Ectoconchion Radius	EKR
Dacryon Subtense	DKS	Zygomaxillare Radius	ZMR
Interorbital Breadth	DKB	M1 Alveolus Radius	AVR
Naso-Dacryal Subtense	NS		

Table 5.2 Interpretation of Cranial Metric Variables (Zakrzewski 2001:101).

Cranial Metric Variables	**Cranial Region Described**
GOL, NOL, FRC, PAC, OCC	Cranial Length
XCB, AUB, ASB	Cranial Breadth a. Cranial
XFB, STB	b. Frontal
BBH, VRR	Cranial Height
BNL, NAR, FMR, EKR, ZOR, NAS	Facial Protrusion a. Upper/Midfacial
BPL, ZMR, SSR, PRR, AVR, SSS	b. Subnasal
NPH, NLH, OBH, WMH	Facial Height
ZYB, FMB, EKB, JUB	Facial Breadth a. Upper Facial Breadth
ZMB, MAB	b. Maxillary Breadth
NLB, WNB	c. Nasal Breadth
OBB, DKB	d. Orbital Region Breadth
IML, XML	Malar Morphology
FOL	Basi-cranium morphology
MDB, MDH	Mastoid morphology

Table 5.3 Definitions of the recorded cranial metric variables (Howells 1973).

CRANIAL MEASUREMENTS - DEFINITION	
MEASUREMENT	**DEFINITION**
Glabello-Occipital Length	The greatest cranial length in the median sagittal plane, from the glabellar region. If the nuchal crest is a sharp ridge protruding from the occipital surface, measurement should avoid it. It should be taken from immediately superior or inferior to the nuchal crest, wherever the maximum reading occurs.
Nasio-Occipital Length	The greatest cranial length in the median sagittal plane, measured from the nasion.
Basion-Nasion Length	The direct cranial length between nasion and basion.
Basion-Bregma Height	The direct cranial length between basion and bregma.
Maximum Cranial Breadth	The maximum cranial breadth perpendicular to the median saggital plane. It is measured on the parietals, normally above the supramastoid crest
Maximum Frontal Breadth	The maximum frontal breadth measured on the coronal suture, perpendicular to the median sagittal plane.
Bistephanic Breadth	The breadth on the coronal suture from right to left stephanion. It is the breadth between the intersections of the right and left halves of the coronal suture and the inferior temporal line, i.e. the attachment site for the temporal muscle.
Bizygomatic Breadth	The maximum breadth across the zygomatic arches, perpendicular to the median plane.
Biauricular Breadth	The breadth across the roots of the zygomatic processes, measured at the deapest point of the curvature of the right and left roots to give the minimum reading.
Minimum Cranial Breadth	The minimum breadth across the sphenoid at the base of the sphenoid fossa, at the infratemporal crests.
Biasterionic Breadth	The breadth from one asterior to the other.
Basion-Prosthion Length	The facial length from basion to prosthion.
Nasion-Prosthion Height	The upper facial height from nasion to prosthion.
Nasal Height	The average height from nasion to the lowest point on the border of the nasal aperture on right and left sides.
Orbit Height Left	The height between the upper and lower borders of the left orbit, measured perpendicular to the long axis of the orbit and bisecting it. Measurement is taken using the internal arms of a sliding calliper.
Bijugal Breadth	The external breadth across the malars at the jugalia, i.e. the deepest points in the curvature between the frontal and temporal process of the malars.
Nasal Breadth	The distance between the anterior edges of the nasal aperture at its widest extent.
Palate Breadth	The greatest breadth across the alveolar borders, perpenicular to the median plane. Most frequently it occurs between the right and left 2nd molars.
Mastoid Height	The length of the mastoid process inferior, and perpendicular to, the eye-ear plane, in the vertical plane.
Mastoid Breadth	The width of the mastoid process at its base, through its transerve axis.
Bimaxillary Breadth	The breadth across the maxillae, measured from right to left zygomaxillare.

MEASUREMENT	DEFINITION
Bimaxillary Subtense	The projection (subtense) from the subspinale to the bimaxillary breadth.
Bifrontal Breadth	The breadth across the frontal bone, measured from one frontomalare anterior to the other, i.e. the most anterior point on the fronto-malar suture.
Nasio-Frontal Subtense	The projection (subtense) from nasion to the bifrontal breadth
Biorbital Breadth	The breadth across the orbits, measured from right to left ectoconchion.
Dacryon Subtense	The mean (average of right and left sides) subtense from dacryon to the biorbital breadth.
Interorbital Breadth	The breadth across the nasal space from dacryon to dacryon.
Naso-Dacryal Subtense	The subtense from the deepest point on the midline curvature of the nasal bones to the interorbital breadth.
Simotic Chord	The minimum transverse breadth across the two nasal bones.
Simotic Subtense	The subtense from the deepest point on the midline curvature of the nasal bones to the simotic chord.
Malar Length Inferior	The direct distance from the zygomaxillare anterior to the lowest point of the zygotemporal suture on the lateral bone surface.
Malar Length Maximum	The direct length of the malar, measured in a diagonal direction from the most inferior point on the zygotemporal suture on the lateral bone surface to zygoorbitale, i.e. the most anterior point on the intersection of the zygomaxillare suture with the lower border of the orbit.
Malar Subtense	The maximum subtense from the convexity of the malar angle to the maximum malar length, at the level of the zygomaticofacial foramen.
Cheek Height	The minimum distance, in any direction, between the lower border of the orbit and the lower margin of the maxilla, mesial to the masseter attachment.
Supraorbital Projection	The maximum projection of the left supraorbital arch between the midline (in the region of the glabelle or superior to this) and the frontal bone just anterior to the temporal line. Measured as subtense.
Glabella Projection	The maximum projection of the frontal bone on midline between the nasion and supraglabellare, i.e. the point where the convex profile of the frontal bone changes to join the projection of the glabellar region.
Foramen Magnum Length	The length from prosthion to opisthion. It is measured using the internal arms of the sliding calliper, but with care not to sink them inside the foramen.
Nasion-Bregma Chord	The external frontal chord or direct distance from nasion to bregma taken in the midplane.
Nasion-Bregma Subtense	The maximum subtense, measured at the highest point on the convexity of the frontal bone in the midplane, to the nasion-bregma chord.
Nasion-Subtense Fraction	The distance along the nasion-bregma chord, recorded from nasion, at which the nasion-bregma subtense falls.
Bregma-Lambda Chord	The external parietal chord or direct distance from bregma to lambda, taken in the midplane.
Bregma- Lambda Subtense	The maximum subtense, at the highest point of the convexity of the parietal bones in the midplane, to the bregma-lambda chord.
Bregma-Subtense Fraction	The distance, measured from bregma, at which the bregma-lambda subtense falls.
Lambda-Opisthion Chord	The external occipital chord, or direct distance from lambda to opisthion, taken in the midplane.

MEASUREMENT	DEFINITION
Lambda-Opisthion Subtense	The maximum subtense, recorded on the most prominent point on the contour of the occipital bone in the midplane. The measurement should be taken with care to avoid the nuchal crest, if this has the form of a sharp rigde protruding from the surface of the occipital bone.
Lambda – Subtense Fraction	The distance along the lambda-opisthion chord, recorded from lambda, at which the lambda-opisthion subtense falls.
Vertex Radius	The perpendicular to the transmeatal axis from the most distant point on the parietals (including bregma or lambda), wherever it is recorded.
Nasion Radius	The perpendicular to the transmeatal axis from nasion.
Subspinale Radius	The perpendicular to the transmeatal axis from sub-spinale.
Prosthion Radius	The perpendicular to the transmeatal axis from prosthion.
Dacryon Radius	The perpendicular to the transmeatal axis from left dacryon.
Zygoorbitale Radius	The perpendicular to the transmeatal axis from left zygoorbitale.
Frontomalare Radius	The perpendicular to the transmeatal axis from left frontomalare.
Ectoconchion Radius	The perpendicular to the transmeatal axis from left ectoconchion.
Zygomaxillare Radius	The perpendicular to the transmeatal axis from left zygomaxillare.
M1 Alveolus Radius	The perpendicular to the transmeatal axis from the most anterior point on the alveolus of the left side 1st molar.

For definitions of cranial landmarks see Howells (1973).

Table 5.4 Recorded dental metric variables and abbreviations used in the text.

MAXILLARY TEETH	
Code	**Measurement Description**
I1LBL	Left 1st Incisor Bucco-lingual crown diameter
I1LMD	Left 1st Incisor Mesio-distal crown diameter
I2LBL	Left 2nd Incisor Bucco-lingual crown diameter
I2LMD	Left 2nd Incisor Mesio-distal crown diameter
CLBL	Left Canine Bucco-lingual crown diameter
CLMD	Left Canine Mesio-distal crown diameter
PM3LBL	Left 3rd Premolar Bucco-lingual crown diameter
PM3LMD	Left 3rd Premolar Mesio-distal crown diameter
PM4LBL	Left 4th Premolar Bucco-lingual crown diameter
PM4LMD	Left 4th Premolar Mesio-distal crown diameter
M1LBL	Left 1st Molar Bucco-lingual crown diameter
M1LMD	Left 1st Molar Mesio-distal crown diameter
M2LBL	Left 2nd Molar Bucco-lingual crown diameter
M2LMD	Left 2nd Molar Mesio-distal crown diameter
M3LBL	Left 3rd Molar Bucco-lingual crown diameter
M3LMD	Left 3rd Molar Mesio-distal crown diameter
I1RBL	Right 1st Incisor Bucco-lingual crown diameter
I1RMD	Right 1st Incisor Mesio-distal crown diameter
I2RBL	Right 2nd Incisor Bucco-lingual crown diameter
I2RMD	Right 2nd Incisor Mesio-distal crown diameter
CRBL	Right Canine Bucco-lingual crown diameter
CRMD	Right Canine Mesio-distal crown diameter
PM3RBL	Right 3rd Premolar Bucco-lingual crown diameter
PM3RMD	Right 3rd Premolar Mesio-distal crown diameter
PM4RBL	Right 4th Premolar Bucco-lingual crown diameter
PM4RMD	Right 4th Premolar Mesio-distal crown diameter
M1RBL	Right 1st Molar Bucco-lingual crown diameter
M1RMD	Right 1st Molar Mesio-distal crown diameter
M2RBL	Right 2nd Molar Bucco-lingual crown diameter
M2RMD	Right 2nd Molar Mesio-distal crown diameter
M3RBL	Right 3rd Molar Bucco-lingual crown diameter
M3RMD	Right 3rd Molar Mesio-distal crown diameter
MANDIBULAR TEETH	
Code	**Measurement Description**
LI1LBL	Left 1st Incisor Bucco-lingual crown diameter
LI1LMD	Left 1st Incisor Mesio-distal crown diameter
LI2LBL	Left 2nd Incisor Bucco-lingual crown diameter
LI2LMD	Left 2nd Incisor Mesio-distal crown diameter
LCLBL	Left Canine Bucco-lingual crown diameter
LCLMD	Left Canine Mesio-distal crown diameter
LPM3LBL	Left 3rd Premolar Bucco-lingual crown diameter
LPM3LMD	Left 3rd Premolar Mesio-distal crown diameter
LPM4LBL	Left 4th Premolar Bucco-lingual crown diameter
LPM4LMD	Left 4th Premolar Mesio-distal crown diameter
LM1LBL	Left 1st Molar Bucco-lingual crown diameter
LM1LMD	Left 1st Molar Mesio-distal crown diameter
LM2LBL	Left 2nd Molar Bucco-lingual crown diameter
LM2LMD	Left 2nd Molar Mesio-distal crown diameter
LM3LBL	Left 3rd Molar Bucco-lingual crown diameter

Code	Measurement Description
LM3LMD	Left 3rd Molar Mesio-distal crown diameter
LI1RBL	Right 1st Incisor Bucco-lingual crown diameter
LI1RMD	Right 1st Incisor Mesio-distal crown diameter
LI2RBL	Right 2nd Incisor Bucco-lingual crown diameter
LI2RMD	Right 2nd Incisor Mesio-distal crown diameter
LCRBL	Right Canine Bucco-lingual crown diameter
LCRMD	Right Canine Mesio-distal crown diameter
LPM3RBL	Right 3rd Premolar Bucco-lingual crown diameter
LPM3RMD	Right 3rd Premolar Mesio-distal crown diameter
LPM4RBL	Right 4th Premolar Bucco-lingual crown diameter
LPM4RRMD	Right 4th Premolar Mesio-distal crown diameter
LM1RBL	Right 1st Molar Bucco-lingual crown diameter
LM1RMD	Right 1st Molar Mesio-distal crown diameter
LM2RBL	Right 2nd Molar Bucco-lingual crown diameter
LM2RMD	Right 2nd Molar Mesio-distal crown diameter
LM3RBL	Right 3rd Molar Bucco-lingual crown diameter
LM3RMD	Right 3rd Molar Mesio-distal crown diameter

Table 5.5 Definitions of the recorded cranial non-metric traits.

CRANIAL NON-METRIC TRAIT	DEFINITION
Highest Nuchal Line present (Berry & Berry 1967)	The highest line, running horizontally across the occipital bone superior to the inferior and superior nuchal lines. It arises with the superior nuchal line at the external occipital protruberance and arches anterior-laterally.
Ossicle at Lambda (Berry & Berry 1967)	A bone (ossicle) at the intersenction of the sagittal and lambdoid sutures. This is the position of the posterior fontanelle.
Lambdoid ossicle present (Berry & Berry 1967)	A bone (ossicle) in the lambdoid suture. (If this occurs at the intersection of the sagittal and lambdoid sutures or at asterion, then it is scored as ossicle at lambda and ossicle at asterion respectively).
Parietal foramen present (Berry & Berry 1967)	A foramen that pierces the parietal bone laterally to the sagittal suture and anterior to lambda. It transmits an emissary vein and sometimes a small branch of the occipital artery.
Bregmatic bone present (Berry & Berry 1967)	A sutural bone at the intersection of the sagittal with the coronal suture. This is the position of the anterior fontanelle.
Metopic suture (Berry & Berry 1967)	The medio-frontal suture fuses within the first two years of the life of the individual. In few individuals, howerer, it is retained (remains unfused) in adulthood. In the present study it was scored as present, if at least one quarter of the suture was unfused.
Coronal ossicle present (Berry & Berry 1967)	A bone (ossicle) in the coronal suture. If it occurs at the intersection of the sagittal and coronal sutures, it is scored as bregmatic bone.
Epipteric bone present (Berry & Berry 1967)	A sutural bone inserted between the anterior-inferior angle of the parietal bone and greater wing of the sphenoid. When this is large, it may also articulate with the squamous part of the temporal bone.
Fronto-temporal articulation (Berry & Berry 1967)	Normally the frontal bone does not articulate with the temporal bone. Frontal and temporal bones are separated by the greater wing of the sphenoid and the anterior-inferior angle of the parietal. Occasionally, however, the frontal and termporal bones articulate directly.
Parietal notch bone present (Berry & Berry 1967)	A bone (ossicle) present in the parietal notch. The parietal notch is the part of the parietal bone between the squamous and mastoid portions of the temporal bone.
Ossicle at asterion (Berry & Berry 1967)	A sutural bone (ossicle) at the intersection of the posterior-inferior angle of the parietal bone, the occipital and the mastoid portion of the temporal bone.
Auditory torus present (Berry & Berry 1967)	A bony ridge or torus found on the floor of the external auditory meatus.
Foramen of Huschke present (Berry & Berry 1967)	A foramen in the floor of the external auditory meatus. It is present in children, but only occasionally persists after the 5th year of life of the individual.
Mastoid foramen exsutural (Berry & Berry 1967)	If present, the mastoid foramen usually lies in the suture between the mastoid portion of the temporal bone and the occipital. Less frequently it lays exsuturally, on the mastoid part of the temporal bone, or more rarely, on the occipital.
Mastoid foramen absent (Berry & Berry 1967)	When it is not present in the portion of the lambdoid suture between the mastoid part of the temporal bone and the occipital, it is scored as absent.
Pharyngeal tubercle (Berry & Berry 1967)	It occurs on the inferior surface of the basioccipital, in front of the the foramen magnum. It gives attachment to the fibrous raphe of the pharynx.
Posterior condylar canal patent (Berry & Berry 1967)	The posterior condylar canal usually pierces the condylar fossa, which lies immediately posterior to the occipital condyle. Sometimes the canal ends blindly in the bone. When a seeker can be passed through it, it is scored as patent.

CRANIAL NON-METRIC TRAIT	DEFINITION
Condylar facet double (Berry & Berry 1967)	Occasionally the articular surface of the occipital condyle is divided into two distinct faces.
Precondylar tubercle present (Berry & Berry 1967)	A bony tubercle that occurs immediately anterior-medially to the occipital condyle.
Anterior condylar canal double (Berry & Berry 1967)	The condylar canal (foramen hypoglossi) pierces the anterior part of the occipital condyle and transmits the hypoglossal nerve. When it is divided, it is scored as double.
Foramen ovale incomplete (Berry & Berry 1967)	The posterior wall of the foramen ovale is incomplete so that the foramen is continuous with the foramen spinosum.
Foramen spinosum open (Berry & Berry 1967)	The posterior wall of the foramen spinosum is deficient (incomplete).
Accessory lesser palatine foramen present (Berry & Berry 1967)	The lesser palatine foramina lie on the right and left sides of the posterior border of the hard palate, immediately posterior to the greater palatine foramen. When more than one lesser palatine foramen is present, they are scored as accessory.
Palatine torus present (Berry & Berry 1967)	A bony ridge running longitudinally down the midline of the hard palate.
Maxillary torus present (Berry & Berry 1967)	A bony ridge running on the alveolus along the lingual aspects of the roots of the molar teeth.
Supraorbital foramen complete (Berry & Berry 1967)	A foramen on the frontal bone, superior to the orbit. It transmits the supraorbital vessels and nerve.
Frontal notch / foramen present (Berry & Berry 1967)	When the supraorbital foramen is incomplete (open), it is scored as frontal notch. Frontal foramen is a well-defined secondary foramen in the vicinity to the supraorbital foramen (usually laterally). Smaller foramina might be present, but these should be ignored.
Anterior ethmoid foramen exsutural (Berry & Berry 1967)	The anterior ethmoid foramen pierces the medial wall of the orbit. It normally lies between the mesial edge of the orbital plates of the frontal and the ethmoid bones. When it does not occur in the suture, but superior to it, it is scored as exsutural.
Posterior ethmoid foramen absent (Berry & Berry 1967)	The posterior ethmoid foramen occurs just posterior to the anterior ethmoid foramen on the same suture line.
Accessory infraorbital foramen present (Berry & Berry 1967)	A second foramen immediately adjacent to the infraorbital foramen.
Inca bone (Ossenberg 1970)	The portion of the occipital squama inferior to the highest nuchal line is ossified in cartilage, and the superior portion in membrane. When the two parts fail to fuse and remain separated by the transverse occipital suture, the upper portion is known as the Os Incae or Inca bone. Partial and complete Inca bones were not scored separately.
Infraorbital suture (Berry & Berry 1967)	A suture originating from the inferior margin of the orbit that normally reaches the infra-orbital foramen. It is usually obliterated a few years after birth, but may persist into adulthood.
Nasal foramen (Williams & Warwick 1980)	A small foramen on the nasal bone that transmits small veins.
Trochlear spur (Ossenberg 1970)	A small spine on the orbital plate, midway between the supraorbital notch and the lacrimal suture. It is formed by the ossification of part of the fibro-cartilaginous trochlea of the superior oblique muscle.
Trochlear fossa (Williams & Warwick 1980)	A small rounded depression (fossa) that occurs near the intersection of the superior and mesial walls of the orbit, and close to the orbital opening. It marks the attachment for the fibrous loop for the tendon of the superior oblique muscle of the eye ball.

CRANIAL NON-METRIC TRAIT	DEFINITION
Squamo-parietal ossicles (Ossenberg 1970)	A bone (ossicle) in the temporo-parietal suture, where the squamous portion of the temporal bone articulates with the parietal.
Processus marginalis (Czarnetzki 1971)	Just inferior to the fronto-malare suture, the posterior border of the malar bone frequently presents a small rounded projection, the marginal tubercle.
Zygomatico-temporal foramen (Williams & Warwick 1980)	A foramen that occurs on the temporal surface of the zygomatic bone, near the base of the frontal process. It transmits brances derived from the zygomatic nerve. Only well-defined foramina are scored as present.
Zygomatico-facial foramen (Williams & Warwick 1980)	A foramen that occurs on the anterior (facial) surface of the zygomatic bone. It transmits branches derived from the zygomatic nerve and it may be single, multiple of absent.
Occipito-mastoid ossicle (Ossenberg 1970)	A bone (ossicle) in the portion of the labmdoid suture between the mastoid part of the temporal bone and the occipital.
Intermediate condylar canal (Ossenberg 1970)	It is formed by the bridging of a gutter which lies immediately laterally to the occipital condyle.
Post condylar tubercle (Corruccini 1974)	A tubercle on the posterior rim of the foramen magnum. It occurs bilaterally, posterior to the right and left occipital condyles.
Jugular foramen bridge (Corruccini 1974)	A superiorly placed spur that divides the jugular fossa.
Foramen of vesalius (Ossenberg 1970)	The least common of the emissary foramina, situated anterior-medially to the foramen ovale.
Pterygo-basal bridge (Corruccini 1974)	A bony bridge that connects the inferior surface of the greater wing of the sphenoid to the lateral surface of the lateral pterygoid plate near its root. The bridge is formed by the ossification of a ligament and usually occurs laterally to the foramen ovale.
Pterygo-spinous bridge (Corruccini 1974)	A bony bridge that connects the posterior border of the lateral pterygoid plate to, or to some point near, the sphenoid spine. The bridge is formed by ossification into the pterygo-spinous ligament and usually occurs medially to the foramen ovale.
Spino-basal bridge (Corruccini 1974)	A bridge formed over the foramen spinosum.
Foramen ovale spine (Corruccini 1974)	A spine inside the foramen ovale, on its lateral wall.
Accessory foramen spinosum (Corruccini 1974)	When the foramen spinosum is multiple, it is scored as accessory foramen spinosum.
Lateral pterygoid perforated (Corruccini 1974)	A foramen near the posterior edge of the pterygoid that appears to be formed by the coalescence of two pterygoid spurs.
Pterygoid spurs (Corruccini 1974)	Small spurs on the posterior edge of the lateral pterygoid plate.
Palatine bridge (Corruccini 1974)	A bridge forming over the lateral palatal sulci. Bridges over the accessory lateral canaliculi are also scored as palatine bridges.
Zygomatico-facial foramen multiple (Corrucini 1974)	The zygomatico-facial foramen that transmits nerves and vessels may be single or multiple. Accessory foramina relate to nerve branching and ossification around the branches.

Table 5.6 Dental Non-metric Traits: equivalence in scores for presence of non-metrics between the recording system employed in this thesis and the ASUDAS (Turner *et al.* 1991).

DENTAL NON-METRIC TRAITS		
MAXILLARY TEETH	**Equivalence in scores for presence of non-metrics between the 2 systems**	
	System employed in this thesis	**ASUDAS (grades of trait expression in ASUDAS recorded as presence in this thesis)**
Incisor 1st Shoveling	1	2-7
Incisor 2nd Shoveling	1	2-7
Canine Mesial ridge	1	1-3
Canine Distal Accessory ridge	1	2-5
Molar 1st Metaconule (cusp 5)	1	1-5
Molar 1st Carabelli's trait	1	2-7
Molar 1st Enamel extension	1	1-3
Molar 1st Enamel Pearl (Risnes 1989)	1	-
Molar 2nd Hypocone (cusp 4)	1	1-5
Molar 3rd Metacone (cusp 3)	1	1-5
Molar 3rd Parastyle	1	2-6
Molar 3rd Congenital Absence	1	1
MANDIBULAR TEETH		
Incisor 1st Shoveling	1	2-7
Incisor 2nd Shoveling	1	2-7
Canine Mesial Ridge	1	1-3
Canine Distal Accessory Ridge	1	2-5
Premolar 1st Number of cusps (>1)	1	2-9 (Premolar lingual cusp variation)
Premolar 2nd Number of cusps (>1)	1	2-9 (Premolar ligual cusp variation)
Molar 1st Number of cusps (>4)	1	5-6 (Cusp Number)
Molar 1st Protostylid	1	2-7
Molar 2nd Number of cusps (>3)	1	-[1] (Cusp Number)
Molar 3rd Protostylid	1	2-7
Molar 3rd Congenital Absence	1	1

Key: 1= Presence

[1] In ASUDAS, when scoring for Cusp Number in lower molars, grade 4 is assigned if cusps 1-4 are present (1, protoconid; 2, metaconid; 3; hypoconid; 4, entoconid). In this thesis I scored the 2nd molar for the presence of more than 3 cusps. ASUDAS does not score the presence of the 4th cusp in the lower 2nd molar separately from the other three cusps. Therefore, there is no direct equivalence between the ASUDAS and this thesis' scoring system in terms of recording the number of cusps for the lower 2nd molar. For all other traits, direct equivalence between the two methods exists.

Table 5.7 Recording form designed and used for this research.

CRANIAL MEASUREMENTS			
Glabello-Occipital Length		Simotic Chord	
Nasio-Occipital Length		Simotic Subtense	
Basion-Nasion Length		Malar Length Inferior	
Basion-Bregma Height		Malar Length Maximum	
Maximum Cranial Breadth		Malar Subtense	
Maximum Frontal Breadth		Cheek Height	
Bistephanic Breadth		Supraorbital Projection	
Bizygomatic Breadth		Glabella Projection	
Biauricular Breadth		Foramen Magnum Length	
Minimum Cranial Breadth		Nasion-Bregma Chord	
Biasterionic Breadth		Nasion-Bregma Subtense	
Basion-Prosthion Length		Nasion-Subtense Fraction	
Nasion-Prosthion Height		Bregma-Lambda Chord	
Nasal Height		Bregma- Lambda Subtense	
Orbit Height Left		Bregma-Subtense Fraction	
Orbit Breadth Left		Lambda-Opisthion Chord	
Bijugal Breadth		Lambda-Opisthion Subtense	
Nasal Breadth		Lambda – Subtense Fraction	
Palate Breadth (2nd M)		Vertex Radius	
Mastoid Height		Nasion Radius	
Mastoid Width		Subspinale Radius	
Bimaxillary Breadth		Prosthion Radius	
Bimaxillary Subtense		Dacryon Radius	
Bifrontal Breadth		Zygoorbitale Radius	
Nasio-Frontal Subtense		Frontomalare Radius	
Biorbital Breadth		Ectoconchion Radius	
Dacryon Subtense		Zygomaxillare Radius	
Interorbital Breadth		M1 Alveolus Radius	
Naso-Dacryal Subtense			

CRANIAL NON - METRIC TRAITS	L	R
Highest Nuchal Line present		
Ossicle at Lambda		
Lambdoid ossicle present		
Parietal foramen present		
Bregmatic bone present		
Metopic suture		
Coronal ossicle present		
Epipteric bone present		
Fronto-temporal articulation		
Parietal notch bone present		
Ossicle at asterion		
Auditory torus present		
Foramen of Huschke present		
Mastoid foramen exsutural		
Mastoid foramen absent		
Posterior condylar canal patent		
Condylar facet double		
Pharyngeal tubercle present		
Precondylar tubercle present		
Anterior condylar canal double		
Foramen ovale incomplete		
Foramen spinosum open		
Accessory lesser palatine foramen present		
Palatine torus present		
Maxillary torus present		
Maxillary bridge present		
Zygomatic – facial foramen absent		
Supraorbital foramen complete		
Frontal notch / foramen present		
Anterior ethmoid foramen exsutural		
Posterior ethmoid foramen absent		
Accessory infraorbital foramen present		
Inca bone		
Infraorbital suture		
Nasal foramen		
Trochlear spur		
Trochlear fossa		
Squamo- parietal ossicles		
Processus marginalis		
Zygomatico-temporal foramen		
Occipito mastoid ossicle		
Intermediate condylar canal		
Post condylar tubercle		
Jugular foramen bridge		
Foramen of vesalius		
Pterygo-basal bridge		
Pterygo-spinous bridge		
Spino-basal bridge		
Foramen ovale spine		
Accessory foramen spinosum		
Lateral pterygoid perforated		
Pterygoid spurs		
Palatine bridge		
Zygomatico- facial foramen multiple		

DENTAL MEASUREMENTS

Maxillary Teeth L side	Bucco-Lingual	Mesio-Distal	Wear (Molnar 1971)	Wear (Brothwell 1981)	Lost PM	Lost AM
I 1st						
I 2nd						
C						
PM3						
PM4						
M1						
M2						
M3						
Maxillary Teeth Rside						
I 1st						
I 2nd						
C						
PM3						
PM4						
M1						
M2						
M3						
Mandibular Teeth Lside						
I 1st						
I 2nd						
C						
PM3						
PM4						
M1						
M2						
M3						
Mandibular Teeth R side						
I 1st						
I 2nd						
C						
PM3						
PM4						
M1						
M2						
M3						

Dental Non Metric Traits

Maxillary teeth_ L side	Present	Absence	Tooth missing	
			PM	AM
Incisor 1st Shoveling				
Incisor 2nd Shoveling				
Canine mesial ridge				
Canine distal accessory ridge				
Molar 1st Metaconule				
Molar 1st Carabelli's trait				
Molar 1st Enamel extension				
Molar 1st Enamel Pearl				
Molar 2nd Hypocone				
Molar 3rd Metacone				
Molar 3rd Parastyle				
Molar 3rd Congenital Absence				
Maxillary teeth_ R side				
Incisor 1st Shoveling				
Incisor 2nd Shoveling				
Canine mesial ridge				
Canine distal accessory ridge				
Molar 1st Metaconule				
Molar 1st Carabelli's trait				
Molar 1st Enamel extension				
Molar 1st Enamel Pearl				
Molar 2nd Hypocone				
Molar 3rd Metacone				
Molar 3rd Parastyle				
Molar 3rd Congenital Absence				
Mandibular teeth _ L side				
Incisor 1st Shoveling				
Incisor 2nd Shoveling				
Canine mesial ridge				
Canine distal accessory ridge				
Premolar 1st number of l cusps (>1)				
Premolar 2nd number of cusps (>1)				
Molar 1st number of cusps (>4)				
Molar 1st Protostylid				
Molar 2nd number of cusps (>3)				
Molar 3rd Protostylid				
Molar 3rd congenital absence				
Mandibular teeth _ R side				
Incisor 1st Shoveling				
Incisor 2nd Shoveling				
Canine mesial ridge				
Canine distal accessory ridge				
Premolar 1st number of l cusps (>1)				
Premolar 2nd number of l cusps (>1)				
Molar 1st number of cusps (>4)				
Molar 1st Protostylid				
Molar 2nd number of cusps (>3)				
Molar 3rd Protostylid				
Molar 3rd congenital absence				

PRESERVATION	VERY GOOD	GOOD	MODERATE	POOR

COMPLETENESS (%)	CRANIUM	MANDIBLE	POST CRANIAL SKELETON

SEX	SKULL MORPHOLOGY	OS COXAE MORPHOLOGY

AGE AT DEATH	DENTAL	OS COXAE MORPHOLOGY	OTHER

COMMENTS:

APPENDIX C

INFORMATION ABOUT THE SEX[2] AND AGE[3] AT DEATH OF THE ANALYSED INDIVIDUALS

Table 6.1 Lerna sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
LER 72	M	Y
LER 66	F	Y
LER 3	F	Y
LER 13	F	Y
LER 42	F	Y
LER 2	M	Y
LER 95	M	Y
LER 56	F	Y
LER 69	M	Y
LER 59	F	Y
LER 52	M	Y
LER 48	M	Y
LER 178	F	Y
LER 181	M	Y
LER 189	M	Y
LER 239	M	Y
LER 91	M	Y
LER 97	F	Y
LER 182	F	Y
LER 115	M	Y
LER 125	M	Y
LER 132	M	Y
LER 174	M	Y
LER 20	F	Y
LER 33	F	Y
LER 18	M	Y
LER 45	F	Y
LER 50	M	Y
LER 19	M	Y
LER 40	M	Y
LER 23	M	Y
LER 38	F	Y
LER 28	M	Y
LER 31	F	Y
LER 1	M	Y
LER 37	M	Y
LER 43	M	Y

KEY: M= MALE, F=FEMALE, Y=PRESENT

[2] The methodology of sex determination is outlined in section 5.1.2.2.
[3] The methodology of age determination is outlined in section 5.1.2.1.

Table 6.2 Apatheia sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
A1.z	F	Y
A1.b	F	Y
A1.c	M	Y
A1.f	F	Y
A3.a	M	Y
A3.b	F	Y
A3.c	F	Y
A3.d	M	Y
A2.b	M	Y
A2.a	M	Y
A5.a	M	Y
A5.17	M	Y
A5.8	M	Y
A5.15	M	Y
A5.14	F	Y
A5.24	M	Y
A5.9	F	Y
A5.C	F	Y
A5.6	M	Y
A5.18	F	Y
A5.11	F	Y
A5.5	F	Y
A5.5a	M	Y
A5.3	M	Y
A6.1	M	Y

KEY: M= MALE, F=FEMALE, Y=YES

Table 6.3 Palaikastro sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
PK 56	F	Y
PK C	F	Y
PK M7	M	Y
PK 101	M	Y
PK 110	F	Y
PK 67	F	Y
PK 36	M	Y
PK 107	M	Y
PK 22	F	Y
PK 60	F	Y
PK 131	M	Y
PK 123	F	Y
PK 12	F	Y
PK 118	F	Y
PK 130	M	Y
PK 17	F	Y
PK 51	F	Y
PK 6	M	Y
PK 108	F	Y
PK 7	M	Y
PK 4	M	Y
PK 100	M	Y
PK 46	M	Y
PK 13	M	Y
PK 143	M	Y
PK 53	F	Y
PK 100b	M	Y

KEY: M= MALE, F=FEMALE, Y=YES

Table 6.4 Moni Odigitria sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
MO 10	M	Y
MO 15	M	Y
MO K20	M	Y
MO 7	M	Y
MO K13	F	Y
MO 71	M	Y
MO 18	M	Y
MO 2	M	Y
MO 6	M	Y
MO 67	F	Y
MO 22	M	Y
MO 12	F	Y
MO 17	M	Y
MO 8	F	Y
MO 19	M	Y
MO 4	M	Y
MO 21	M	Y
MO 5	F	Y
MO 9	F	Y
MO 25	F	Y

KEY: M= MALE, F=FEMALE, Y=YES

Table 6.5 Ailias sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
AILT94	M	Y
AILT21	M	Y
AILT10B	M	Y
AILT92	M	Y
AILT30	M	Y
AILT8	F	Y
AILT9	M	Y
AILT107	F	Y
AILT68	M	Y
AILT108	F	Y
AIL T3	M	Y
AILT1	M	Y
AILT23	M	Y
AILT7	M	Y
AILT106	F	Y
AILT103	F	Y

KEY: M= MALE, F=FEMALE, Y=YES

Table 6.6 Ailias sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
AILT2	M	Y
AILT4	M	Y
AILT15	M	Y
AILT6	M	Y
AILT16	F	Y
AILT20	M	Y
AILT109	F	Y
AILT24	M	Y
AILT36	F	Y
AILT93	M	Y
AILT111	F	Y
AILT29	M	Y
AILT95	M	Y
AILT105	F	Y
AILT78	M	Y
AILT12	M	Y
AIL T71	M	Y
AIL T167	F	Y
AIL T64	M	Y
AIL T14	M	Y
AIL T90	F	Y
AIL T19	M	Y
AIL T17	M	Y
AIL T113	F	Y
AIL T26	M	Y
AIL T112	F	Y
AIL T89	M	Y
AIL T87	F	Y
AIL T102	F	Y
AIL T99	M	Y
AIL T91	F	Y
AIL T86	M	Y
AIL T85	M	Y
AIL T88	M	Y
AIL T74	M	Y
AIL T5	M	Y
AILT10	M	Y
AILT166	F	Y
AIL T172	F	Y
AIL T76	M	Y
AIL T72	M	Y
AILT165	F	Y
AILT97	M	Y
AILT96	M	Y

INDIVIDUAL	SEX	AGE (17-45)
AILT73	M	Y
AILT95	F	Y
AILT110	F	Y

KEY: M= MALE, F=FEMALE, Y=YES

Table 6.6 Myrtos Pyrgos sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
MP SK.D	M	Y
MP SK 35	M	Y
MP SK 34	M	Y
MP SK H	M	Y
MP SK A	M	Y
MP SK C	M	Y
MP SK 33	F	Y
MP SK E	M	Y
MP SK 17	F	Y
MP SK 22	M	Y
MP SK 30	F	Y
MP SK 130-31	F	Y
MP SK AJ	F	Y
MP SK 24	F	Y
MP SK 35	M	Y
MP SK 11C	F	Y
MP SK 26	F	Y
MP SK G	M	Y
MP SK 7	F	Y
MP SK 23	M	Y
MP SK 20A	F	Y
MP SK 25A	M	Y
MP SK 4	M	Y
MP SK 11A	M	Y
MP SK 13	F	Y
MP SK 5	M	Y
MP SK 2	M	Y

KEY: M= MALE, F=FEMALE, Y=YES

Table 6.7 Gypsades sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
LGI 21	M	Y
LGI 24	M	Y
LGI LARNAX	F	Y
LGI F5.19	M	Y
LGI F2.12	F	Y
GXVIII.VI	M	Y
GXVIII.VII	M	Y
LGI F7.33	F	Y
GXVIII.VIIB	F	Y
LG F5.23	F	Y
LGI F5.25	M	Y
LGI F7.31	M	Y
LGI F5.28	M	Y
LGI F5.13	F	Y
LGI F5.18	M	Y
LGI A-B.2	M	Y
LGI DOORWAY THOLOS	F	Y
LGI 29	M	Y
LGI 11	F	Y
LGI F5.22	M	Y
LGI F5.26	M	Y
LGI F7.34	F	Y

KEY: M= MALE, F=FEMALE, Y=YES

Table 6.8 Sellopoulo sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
SELI, IV	F	Y
SEL4, 2	F	Y
SELLII, N.CNR.2	M	Y
SELI, SEB1	M	Y
SELII, NB1	F	Y
SEL3, 3	M	Y

KEY: M= MALE, F=FEMALE, Y=YES

Table 6.9 Mavrospelio sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
MSP 6	F	Y
MSP 5	M	Y
MSP 4	M	Y
MSP 7	F	Y
MSP 2	M	Y
MSP 8	F	Y
MSP 1	M	

KEY: M= MALE, F=FEMALE, Y=YES

Table 6.10 Kastelos sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
KAS 6	M	Y
KAS 5	F	Y
KAS 3	M	Y
KAS 2	F	Y

KEY: M= MALE, F=FEMALE, Y=YES

Table 6.11 Palama sample used in cranial metric analysis.

INDIVIDUAL	SEX	AGE (17-45)
PAL11	F	Y
PAL8	F	Y
PALI	M	Y
PAL3	F	Y
PAL IA	F	Y
PAL8, 2	M	Y
PAL15, C	M	Y
PAL7, 1	M	Y
PAL7, 2	F	Y
PAL16, 2	M	Y
PAL16, 1	M	Y

KEY: M= MALE, F=FEMALE, Y=YES

APPENDIX D

STATISTICAL TESTS AND GRAPHS FOR CHAPTER 6

D.1 STATISTICAL TESTS

Table 6.1 Sexual Dimorphism in Lerna. Test of Homogeneity of Variances, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Levene Statistic	Significance
GOL	24	.440	.513
NOL	26	.433	.517
BNL	11	.043	.840
BBH	13	1.794	.207
XCB	27	.509	.482
XFB	24	.097	.758
STB	8	2.950	.137
ZYB	10	8.392	.020
AUB	21	2.617	.122
WCB	18	.034	.856
ASB	27	1.130	.298
BPL	11	.707	.422
NPH	15	.267	.614
NLH	16	.068	.798
OBH	18	1.094	.311
OBB	19	1.115	.306
JUB	11	7.102	.026
NLB	10	.026	.877
MAB	21	.128	.724
MDH	28	.094	.761
MDB	27	2.305	.142
ZMB	11	1.413	.265
SSS	11	.924	.362
FMB	26	.894	.354
NAS	13	.236	.636
EKB	15	.018	.895
DKS	5	49.503	.006
IML	9	.097	.765
XML	9	.002	.963
WMH	23	.234	.634
SOS	28	8.700	.007
GLS	27	.005	.942
FOL	8	.339	.582
FRC	30	1.883	.181
FRS	29	2.361	.136
FRF	29	1.560	.222
PAC	28	2.834	.104
PAS	27	.002	.967
PAF	27	.132	.719

Cranial Metric Variables	N	Levene Statistic	Significance
OCC	23	.081	.779
OCS	21	.056	.815
OCF	21	5.849	.026
VRR	22	.629	.437
NAR	20	.772	.391
SSR	16	.025	.878
PRR	11	.517	.490
DKR	13	.646	.439
ZOR	18	.452	.511
FMR	22	1.120	.302
EKR	20	.717	.408
ZMR	17	.036	.852
AVR	11	.000	.990

Table 6.2 Sexual Dimorphism in Lerna. Robust Tests of Equality of Means, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Tests of Equality of Means	Statistic	Significance
GOL	24	Welch	20.791	<0.001
		Brown-Forsythe	20.791	<0.001
NOL	26	Welch	15.582	.001
		Brown-Forsythe	15.582	.001
BNL	11	Welch	8.601	.021
		Brown-Forsythe	8.601	.021
BBH	13	Welch	3.044	.111
		Brown-Forsythe	3.044	.111
XCB	27	Welch	.596	.448
		Brown-Forsythe	.596	.448
XFB	24	Welch	1.530	.241
		Brown-Forsythe	1.530	.241
STB	8	Welch	24.957	.003
		Brown-Forsythe	24.957	.003
ZYB	10	Welch	1.342	.445
		Brown-Forsythe	1.342	.445
AUB	21	Welch	.915	.372
		Brown-Forsythe	.915	.372
WCB	18	Welch	.702	.416
		Brown-Forsythe	.702	.416
ASB	27	Welch	1.488	.250
		Brown-Forsythe	1.488	.250
BPL	11	Welch	1.535	.247
		Brown-Forsythe	1.535	.247
NPH	15	Welch	.049	.830
		Brown-Forsythe	.049	.830
NLH	16	Welch	.233	.640
		Brown-Forsythe	.233	.640
OBH	18	Welch	.724	.409
		Brown-Forsythe	.724	.409
OBB	19	Welch	1.565	.232
		Brown-Forsythe	1.565	.232
JUB	11	Welch	3.419	.305
		Brown-Forsythe	3.419	.305
NLB	10	Welch	45.095	.039
		Brown-Forsythe	45.095	.039
MAB	21	Welch	.445	.515
		Brown-Forsythe	.445	.515
MDH	28	Welch	3.836	.068
		Brown-Forsythe	3.836	.068
MDB	27	Welch	4.623	.042
		Brown-Forsythe	4.623	.042
ZMB	11	Welch	.889	.395
		Brown-Forsythe	.889	.395

Cranial Metric Variables	N	Tests of Equality of Means	Statistic	Significance
SSS	11	Welch	1.403	.270
		Brown-Forsythe	1.403	.270
FMB	26	Welch	2.247	.162
		Brown-Forsythe	2.247	.162
NAS	13	Welch	.732	.501
		Brown-Forsythe	.732	.501
EKB	15	Welch	1.934	.256
		Brown-Forsythe	1.934	.256
DKS	5	Welch	4.356	.272
		Brown-Forsythe	4.356	.272
IML	9	Welch	2.830	.155
		Brown-Forsythe	2.830	.155
XML	9	Welch	4.846	.089
		Brown-Forsythe	4.846	.089
WMH	23	Welch	2.133	.164
		Brown-Forsythe	2.133	.164
SOS	28	Welch	.616	.455
		Brown-Forsythe	.616	.455
GLS	27	Welch	17.737	.001
		Brown-Forsythe	17.737	.001
FOL	8	Welch	.005	.947
		Brown-Forsythe	.005	.947
FRC	30	Welch	18.016	.001
		Brown-Forsythe	18.016	.001
FRS	29	Welch	.002	.968
		Brown-Forsythe	.002	.968
FRF	29	Welch	19.989	.001
		Brown-Forsythe	19.989	.001
PAC	28	Welch	1.137	.296
		Brown-Forsythe	1.137	.296
PAS	27	Welch	.353	.561
		Brown-Forsythe	.353	.561
PAF	27	Welch	.664	.425
		Brown-Forsythe	.664	.425
OCC	23	Welch	8.142	.010
		Brown-Forsythe	8.142	.010
OCS	21	Welch	1.881	.196
		Brown-Forsythe	1.881	.196
OCF	21	Welch	4.290	.052
		Brown-Forsythe	4.290	.052
VRR	22	Welch	6.997	.020
		Brown-Forsythe	6.997	.020
NAR	20	Welch	16.441	.002
		Brown-Forsythe	16.441	.002
SSR	16	Welch	13.711	.012
		Brown-Forsythe	13.711	.012

Cranial Metric Variables	N	Tests of Equality of Means	Statistic	Significance
PRR	11	Welch	11.863	.016
		Brown-Forsythe	11.863	.016
DKR	13	Welch	9.716	.032
		Brown-Forsythe	9.716	.032
ZOR	18	Welch	3.274	.095
		Brown-Forsythe	3.274	.095
FMR	22	Welch	3.947	.071
		Brown-Forsythe	3.947	.071
EKR	20	Welch	3.566	.084
		Brown-Forsythe	3.566	.084
ZMR	17	Welch	10.056	.012
		Brown-Forsythe	10.056	.012
AVR	11	Welch	7.065	.064
		Brown-Forsythe	7.065	.064

Table 6.3 Sexual Dimorphism in Apatheia Galatas. Test of Homogeneity of Variances, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Levene Statistic	Significance
GOL	15	.826	.380
NOL	15	.812	.384
XCB	15	1.630	.224
XFB	11	.510	.493
STB	12	.265	.618
AUB	5	44.757	.007
WCB	6	.392	.565
ASB	16	7.349	.017
MDH	14	.512	.488
MDB	15	.475	.503
FMB	11	1.033	.336
NAS	8	.636	.455
WMH	5	89.762	.002
SOS	19	.141	.712
GLS	18	.399	.536
FRC	18	1.235	.283
FRS	18	.000	.985
FRF	18	1.050	.321
PAC	20	.018	.895
PAS	20	.007	.936
PAF	19	2.908	.106
OCC	13	.199	.664
OCS	13	1.572	.236
OCF	13	2.852	.119
VRR	8	3.950	.094
NAR	7	.391	.559
FMR	6	.236	.652

Table 6.4 Sexual Dimorphism in Apatheia Galatas. Robust Tests of Equality of Means, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Tests of Equality of Means	Statistic	Significance
GOL	15	Welch	6.044	.031
		Brown-Forsythe	6.044	.031
NOL	15	Welch	7.904	.016
		Brown-Forsythe	7.904	.016
XCB	15	Welch	.778	.396
		Brown-Forsythe	.778	.396
XFB	11	Welch	.093	.782
		Brown-Forsythe	.093	.782
STB	12	Welch	.000	1.000
		Brown-Forsythe	.000	1.000
AUB	5	Welch	3.550	.295
		Brown-Forsythe	3.550	.295
WCB	6	Welch	.033	.867
		Brown-Forsythe	.033	.867
ASB	16	Welch	.029	.869
		Brown-Forsythe	.029	.869
MDH	14	Welch	.988	.342
		Brown-Forsythe	.988	.342
MDB	15	Welch	.823	.381
		Brown-Forsythe	.823	.381
FMB	11	Welch	5.530	.043
		Brown-Forsythe	5.530	.043
NAS	8	Welch	3.574	.113
		Brown-Forsythe	3.574	.113
WNB	4	Welch	1.231	.467
		Brown-Forsythe	1.231	.467
WMH	5	Welch	.667	.557
		Brown-Forsythe	.667	.557
SOS	19	Welch	4.386	.052
		Brown-Forsythe	4.386	.052
GLS	18	Welch	.599	.451
		Brown-Forsythe	.599	.451
FRC	18	Welch	.405	.537
		Brown-Forsythe	.405	.537
FRS	18	Welch	.472	.503
		Brown-Forsythe	.472	.503
FRF	18	Welch	3.672	.075
		Brown-Forsythe	3.672	.075
PAC	20	Welch	7.477	.014
		Brown-Forsythe	7.477	.014
PAS	20	Welch	2.260	.150
		Brown-Forsythe	2.260	.150
PAF	19	Welch	4.653	.046
		Brown-Forsythe	4.653	.046

Cranial Metric Variables	N	Tests of Equality of Means	Statistic	Significance
OCC	13	Welch	.004	.948
		Brown-Forsythe	.004	.948
OCS	13	Welch	.497	.500
		Brown-Forsythe	.497	.500
OCF	13	Welch	.591	.463
		Brown-Forsythe	.591	.463
VRR	8	Welch	1.727	.263
		Brown-Forsythe	1.727	.263
NAR	7	Welch	3.172	.136
		Brown-Forsythe	3.172	.136
FMR	6	Welch	.011	.920
		Brown-Forsythe	.011	.920

Table 6.5 Sexual Dimorphism in Apatheia Galatas. One-way ANOVA, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	F	Significance
GOL	15	6.243	.027
NOL	15	8.172	.013
BNL	4	.645	.506
BBH	4	.073	.812
XCB	15	.727	.409
XFB	11	.137	.719
STB	12	.000	1.000
AUB	5	5.891	.094
WCB	6	.033	.866
ASB	16	.029	.868
NLH	3	.043	.870
MDH	14	.988	.340
MDB	15	.814	.383
FMB	11	5.206	.048
NAS	8	3.060	.131
WNB	4	1.231	.383
WMH	5	1.126	.366
SOS	19	4.274	.054
GLS	18	.599	.450
FOL	4	9.936	.088
FRC	18	.405	.533
FRS	18	.472	.502
FRF	18	3.672	.073
PAC	20	7.340	.014
PAS	20	2.229	.153
PAF	19	3.955	.063
OCC	13	.004	.948
OCS	13	.532	.481
OCF	13	.631	.444
VRR	8	1.727	.237
NAR	7	2.739	.159
PRR	3	386643.000	.001
DKR	3	30.177	.115
FMR	6	.011	.920

Table 6.6 Sexual Dimorphism in the Argolid population (samples pooled). One-way ANOVA, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	F	Significance
GOL	41	17.486	<0.001
NOL	39	16.184	<0.001
BNL	15	7.851	.015
BBH	17	2.141	.164
XCB	42	.143	.707
XFB	35	3.818	.059
STB	20	4.556	.047
ZYB	10	4.779	.060
AUB	26	.044	.836
WCB	24	.901	.353
ASB	43	2.574	.116
BPL	13	.721	.414
NPH	17	.007	.936
NLH	19	.109	.745
OBH	21	.067	.799
OBB	21	.818	.377
JUB	13	11.096	.007
NLB	10	44.752	<0.001
MAB	23	.463	.504
MDH	42	6.029	.019
MDB	42	4.708	.036
ZMB	12	1.701	.221
SSS	11	.289	.604
FMB	37	9.438	.004
NAS	21	7.718	.012
EKB	17	5.133	.039
DKS	6	1.032	.367
DKB	4	.019	.904
WNB	14	.707	.417
IML	10	.307	.595
XML	10	2.010	.194
WMH	28	1.858	.185
SOS	47	.572	.453
GLS	45	14.651	<0.001
FOL	12	.351	.567
FRC	48	15.562	<0.001
FRS	47	.431	.515
FRF	47	27.467	<0.001

Cranial Metric Variables	N	F	Significance
PAC	48	3.850	.056
PAS	47	.140	.710
PAF	46	3.011	.090
OCC	36	4.120	.050
OCS	34	.774	.386
OCF	34	2.433	.129
VRR	30	7.019	.013
NAR	27	14.126	.001
SSR	18	6.214	.024
PRR	14	3.334	.093
DKR	16	4.979	.043
ZOR	20	3.352	.084
FMR	28	3.543	.071
EKR	22	3.480	.077
ZMR	18	6.573	.021
AVR	13	3.705	.080

Table 6.7 Sexual Dimorphism in the Argolid population (samples pooled). Test of Homogeneity of Variances, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Levene Statistic	Significance
GOL	41	.149	.701
NOL	39	.049	.825
BNL	15	.348	.565
BBH	17	1.689	.213
XCB	42	.545	.465
XFB	35	.388	.538
STB	20	3.157	.092
ZYB	10	8.392	.020
AUB	26	.042	.839
WCB	24	.021	.886
ASB	43	.523	.474
BPL	13	1.164	.304
NPH	17	.257	.620
NLH	19	.157	.697
OBH	21	1.514	.234
OBB	21	1.582	.224
JUB	13	15.300	.002
NLB	10	.026	.877
MAB	23	.352	.559
MDH	42	.309	.581
MDB	42	2.806	.102
ZMB	12	.415	.534
SSS	11	.924	.362
FMB	37	2.205	.146
NAS	21	.257	.618
EKB	17	.202	.660
DKS	6	2.800	.170
WNB	14	.010	.922
IML	10	.387	.551
XML	10	.868	.379
WMH	28	3.364	.078
SOS	47	4.566	.038
GLS	45	.280	.599
FOL	12	.217	.651
FRC	48	.004	.953
FRS	47	.758	.389
FRF	47	.315	.577
PAC	48	2.427	.126
PAS	47	.065	.800
PAF	46	1.172	.285
OCC	36	.046	.832
OCS	34	.110	.743
OCF	34	3.509	.070
VRR	30	2.514	.124
NAR	27	1.618	.215

Cranial Metric Variables	N	Levene Statistic	Significance
SSR	18	.315	.582
PRR	14	.192	.669
DKR	16	1.688	.215
ZOR	20	.688	.418
FMR	28	1.997	.169
EKR	22	1.306	.267
ZMR	18	.086	.773
AVR	13	.492	.498

Table 6.8 Sexual Dimorphism in the Argolid population (samples pooled). Robust Tests of Equality of Means, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Tests of Equality of Means	Statistic	Significance
GOL	41	Welch	16.967	<0.001
		Brown-Forsythe	16.967	<0.001
NOL	39	Welch	15.843	<0.001
		Brown-Forsythe	15.843	<0.001
BNL	15	Welch	8.110	.014
		Brown-Forsythe	8.110	.014
BBH	17	Welch	2.251	.156
		Brown-Forsythe	2.251	.156
XCB	42	Welch	.157	.694
		Brown-Forsythe	.157	.694
XFB	35	Welch	3.834	.059
		Brown-Forsythe	3.834	.059
STB	20	Welch	4.093	.066
		Brown-Forsythe	4.093	.066
ZYB	10	Welch	1.342	.445
		Brown-Forsythe	1.342	.445
AUB	26	Welch	.044	.838
		Brown-Forsythe	.044	.838
WCB	24	Welch	.969	.337
		Brown-Forsythe	.969	.337
ASB	43	Welch	2.551	.120
		Brown-Forsythe	2.551	.120
BPL	13	Welch	.799	.395
		Brown-Forsythe	.799	.395
NPH	17	Welch	.009	.927
		Brown-Forsythe	.009	.927
NLH	19	Welch	.173	.683
		Brown-Forsythe	.173	.683
OBH	21	Welch	.087	.771
		Brown-Forsythe	.087	.771
OBB	21	Welch	1.656	.220
		Brown-Forsythe	1.656	.220
JUB	13	Welch	5.740	.086
		Brown-Forsythe	5.740	.086
NLB	10	Welch	45.095	.039
		Brown-Forsythe	45.095	.039
MAB	23	Welch	.635	.435
		Brown-Forsythe	.635	.435
MDH	42	Welch	6.452	.016
		Brown-Forsythe	6.452	.016
MDB	42	Welch	5.576	.023
		Brown-Forsythe	5.576	.023
ZMB	12	Welch	2.225	.203
		Brown-Forsythe	2.225	.203

Cranial Metric Variables	N	Tests of Equality of Means	Statistic	Significance
SSS	11	Welch	1.403	.270
		Brown-Forsythe	1.403	.270
FMB	37	Welch	8.007	.010
		Brown-Forsythe	8.007	.010
NAS	21	Welch	7.051	.023
		Brown-Forsythe	7.051	.023
EKB	17	Welch	5.120	.056
		Brown-Forsythe	5.120	.056
DKS	6	Welch	1.032	.412
		Brown-Forsythe	1.032	.412
WNB	14	Welch	.726	.452
		Brown-Forsythe	.726	.452
IML	14	Welch	.261	.631
		Brown-Forsythe	.261	.631
XML	10	Welch	1.802	.233
		Brown-Forsythe	1.802	.233
WMH	28	Welch	2.139	.156
		Brown-Forsythe	2.139	.156
SOS	47	Welch	.356	.559
		Brown-Forsythe	.356	.559
GLS	45	Welch	15.822	<0.001
		Brown-Forsythe	15.822	<0.001
FOL	12	Welch	.306	.598
		Brown-Forsythe	.306	.598
FRC	48	Welch	16.626	<0.001
		Brown-Forsythe	16.626	<0.001
FRS	47	Welch	.414	.524
		Brown-Forsythe	.414	.524
FRF	47	Welch	28.616	<0.001
		Brown-Forsythe	28.616	<0.001
PAC	48	Welch	4.633	.037
		Brown-Forsythe	4.633	.037
PAS	47	Welch	.141	.709
		Brown-Forsythe	.141	.709
PAF	46	Welch	3.579	.065
		Brown-Forsythe	3.579	.065
OCC	36	Welch	4.043	.053
		Brown-Forsythe	4.043	.053
OCS	34	Welch	.799	.379
		Brown-Forsythe	.799	.379
OCF	34	Welch	2.743	.107
		Brown-Forsythe	2.743	.107
VRR	30	Welch	9.764	.004
		Brown-Forsythe	9.764	.004
NAR	27	Welch	21.213	<0.001
		Brown-Forsythe	21.213	<0.001

Cranial Metric Variables	N	Tests of Equality of Means	Statistic	Significance
SSR	18	Welch	9.623	.009
		Brown-Forsythe	9.623	.009
PRR	14	Welch	3.869	.074
		Brown-Forsythe	3.869	.074
DKR	16	Welch	12.926	.003
		Brown-Forsythe	12.926	.003
ZOR	20	Welch	3.963	.064
		Brown-Forsythe	3.963	.064
FMR	28	Welch	4.568	.044
		Brown-Forsythe	4.568	.044
EKR	22	Welch	4.161	.056
		Brown-Forsythe	4.161	.056
ZMR	18	Welch	6.631	.027
		Brown-Forsythe	6.631	.027
AVR	11	Welch	3.427	.103
		Brown-Forsythe	3.427	.103

Table 6.9 Inter-sample analysis of cranial metric variables using Independent Samples Test, Argolid samples, sexes pooled (p<0.05).

Cranial Metric Variables		Levene's Test for Equality of Variances		T-test for equality of means		
		F	Sig.	t	df	Sig. (2-tailed)
GOL	Equal variances assumed	7.269	.010	1.392	39	.172
NOL	Equal variances assumed	6.842	.013	1.089	37	.283
BNL	Equal variances assumed	4.329	.058	-.367	13	.720
BBH	Equal variances assumed	.024	.879	-1.179	15	.257
XCB	Equal variances assumed	2.116	.154	-.865	40	.392
XFB	Equal variances assumed	3.935	.056	-1.746	33	.090
STB	Equal variances assumed	.023	.880	-3.298	18	.004
AUB	Equal variances assumed	.213	.649	-2.848	24	.009
WCB	Equal variances assumed	.117	.736	-1.660	22	.111
ASB	Equal variances assumed	6.213	.017	-1.683	41	.100
BPL	Equal variances assumed	1.505	.245	.285	11	.781
NPH	Equal variances assumed	.035	.855	-.180	15	.860
NLH	Equal variances assumed	.760	.396	.156	17	.878
OBH	Equal variances assumed	.461	.505	-.629	19	.537
OBB	Equal variances assumed	.269	.610	-.444	19	.662
JUB	Equal variances assumed	1.180	.301	-1.273	11	.229

Cranial Metric Variables		Levene's Test for Equality of Variances		T-test for equality of means		
				t	df	Sig. (2-tailed)
MDH	Equal variances assumed	.002	.966	-2.288	40	.028
MDB	Equal variances assumed	.080	.778	-1.459	40	.152
FMB	Equal variances assumed	.003	.956	-3.597	35	.001
NAS	Equal variances assumed	.316	.581	-2.481	19	.023
EKB	Equal variances assumed	.007	.935	-1.896	15	.077
DKS	Equal variances assumed	.	.	-1.073	4	.344
DKB	Equal variances assumed	.	.	-1.709	2	.230
WNB	Equal variances assumed	1.042	.328	-1.507	12	.158
IML	Equal variances assumed	.	.	1.684	8	.131
XML	Equal variances assumed	.	.	.791	8	.452
WMH	Equal variances assumed	4.887	.036	-1.853	26	.075
SOS	Equal variances assumed	2.028	.161	-1.071	45	.290
GLS	Equal variances assumed	2.214	.144	-1.443	43	.156
FOL	Equal variances assumed	.042	.842	-1.087	10	.302
FRC	Equal variances assumed	.339	.563	-1.920	46	.061
FRS	Equal variances assumed	.701	.407	.798	45	.429
FRF	Equal variances assumed	.759	.388	-2.470	45	.017

Cranial Metric Variables		Levene's Test for Equality of Variances		T-test for equality of means t	df	Sig. (2-tailed)
PAC	Equal variances assumed	5.250	.027	.738	46	.464
PAS	Equal variances assumed	.521	.474	.452	45	.654
PAF	Equal variances assumed	.868	.357	-.883	44	.382
OCC	Equal variances assumed	.131	.720	.231	34	.819
OCS	Equal variances assumed	1.850	.183	2.485	32	.018
OCF	Equal variances assumed	2.640	.114	.528	32	.601
VRR	Equal variances assumed	3.779	.062	-.584	28	.564
NAR	Equal variances assumed	2.700	.113	-1.499	25	.146
SSR	Equal variances assumed	.000	.987	-2.101	16	.052
PRR	Equal variances assumed	.107	.749	-1.424	12	.180
DKR	Equal variances assumed	1.126	.307	-.972	14	.347
ZOR	Equal variances assumed	1.156	.296	-.296	18	.770
FMR	Equal variances assumed	.969	.334	-1.637	26	.114
EKR	Equal variances assumed	1.166	.293	-.507	20	.618
AVR	Equal variances assumed	3.988	.071	.355	11	.730

Table 6.10 Inter-sample analysis, Argolig samples, males. Test of Homogeneity of variances, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	Levene Statistic	N	Significance
GOL	2.980	25	.098
NOL	4.802	24	.039
XCB	2.280	25	.145
XFB	.320	19	.579
STB	1.460	9	.266
AUB	2.245	18	.154
WCB	.005	15	.944
ASB	.116	27	.736
MDH	.394	26	.536
MDB	.081	26	.779
FMB	.001	23	.980
NAS	.225	15	.644
WNB	2.606	11	.141
WMH	.589	17	.455
SOS	2.547	29	.122
GLS	.038	28	.848
FRC	1.351	29	.255
FRS	.818	29	.374
FRF	.310	29	.582
PAC	4.658	29	.040
PAS	.200	29	.659
PAF	.355	29	.556
OCC	.008	20	.932
OCS	2.864	20	.108
OCF	5.826	20	.027
VRR	.413	20	.528
NAR	2.068	19	.169
FMR	.104	19	.751

Table 6.11 Inter-sample analysis, Argolid samples, males. Robust tests of Equality of Means, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Test of Equality of Means	Statistic	Significance
GOL	25	Welch	.449	.509
		Brown-Forsythe	.449	.509
NOL	24	Welch	.343	.564
		Brown-Forsythe	.343	.564
XCB	25	Welch	2.550	.124
		Brown-Forsythe	2.550	.124
XFB	19	Welch	1.916	.253
		Brown-Forsythe	1.916	.253
STB	9	Welch	10.620	.026
		Brown-Forsythe	10.620	.026
AUB	18	Welch	63.895	<0.001
		Brown-Forsythe	63.895	<0.001
WCB	15	Welch	2.043	.226
		Brown-Forsythe	2.043	.226
ASB	27	Welch	3.435	.083
		Brown-Forsythe	3.435	.083
MDH	26	Welch	2.155	.175
		Brown-Forsythe	2.155	.175
MDB	26	Welch	1.336	.266
		Brown-Forsythe	1.336	.266
FMB	23	Welch	4.294	.075
		Brown-Forsythe	4.294	.075
NAS	15	Welch	.642	.469
		Brown-Forsythe	.642	.469
WNB	11	Welch	1.930	.202
		Brown-Forsythe	1.930	.202
WMH	17	Welch	4.138	.234
		Brown-Forsythe	4.138	.234
SOS	29	Welch	1.100	.304
		Brown-Forsythe	1.100	.304
GLS	28	Welch	5.228	.035
		Brown-Forsythe	5.228	.035
FRC	29	Welch	4.236	.067
		Brown-Forsythe	4.236	.067
FRS	29	Welch	.005	.946
		Brown-Forsythe	.005	.946
FRF	29	Welch	6.883	.021
		Brown-Forsythe	6.883	.021
PAC	29	Welch	1.349	.256
		Brown-Forsythe	1.349	.256
PAS	29	Welch	1.767	.197
		Brown-Forsythe	1.767	.197
PAF	29	Welch	.045	.834

Cranial Metric Variables	N	Test of Equality of Means	Statistic	Significance
PAF		Brown-Forsythe	.045	.834
OCC	20	Welch	.416	.536
		Brown-Forsythe	.416	.536
OCS	20	Welch	2.305	.174
		Brown-Forsythe	2.305	.174
OCF	20	Welch	.185	.681
		Brown-Forsythe	.185	.681
VRR	20	Welch	.279	.616
		Brown-Forsythe	.279	.616
NAR	19	Welch	5.628	.040
		Brown-Forsythe	5.628	.040
FMR	19	Welch	2.292	.229
		Brown-Forsythe	2.292	.229

Table 6.12 Inter-sample analysis, Argolig samples, females. Test of Homogeneity of variances, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	Levene Statistic	N	Significance
GOL	.424	16	.525
NOL	.718	15	.412
BNL	7.761	7	.039
BBH	4.965	8	.067
XCB	.003	17	.957
XFB	1.772	16	.204
STB	2.367	11	.158
AUB	.000	8	.991
WCB	.729	9	.422
ASB	6.460	16	.023
BPL	6.315	6	.066
NPH	.033	6	.864
NLH	1.212	6	.333
OBH	.007	8	.938
OBB	.003	7	.957
JUB	164084107819675.000	4	<0.001
MAB	1.278	8	.301
MDH	.272	16	.610
MDB	.094	16	.764
FMB	.000	14	.991
NAS	.642	7	.459
EKB	.062	5	.820
WMH	10.192	11	.011
SOS	4.593	18	.048
GLS	.422	17	.526
FOL	1.266	7	.312
FRC	2.055	19	.170
FRS	.038	18	.849
FRF	2.548	18	.130
PAC	.372	19	.550
PAS	.145	18	.708
PAF	4.736	17	.046
OCC	.756	16	.399
OCS	.163	14	.693
OCF	2.854	14	.117
VRR	4.607	10	.064
NAR	1.975	8	.210
PRR	29.745	6	.005
FMR	.003	9	.955
EKR	.281	8	.615
AVR	2.031	5	.249

Table 6.13 Inter-sample analysis, Argolid samples, females. Robust tests of Equality of Means, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Test of Equality of Means	Statistic	Significance
GOL	16	Welch	12.805	.003
		Brown-Forsythe	12.805	.003
NOL	15	Welch	7.905	.016
		Brown-Forsythe	7.905	.016
BNL	7	Welch	3.089	.166
		Brown-Forsythe	3.089	.166
BBH	8	Welch	.014	.915
		Brown-Forsythe	.014	.915
XCB	17	Welch	.042	.841
		Brown-Forsythe	.042	.841
XFB	16	Welch	.239	.636
		Brown-Forsythe	.239	.636
STB	11	Welch	1.223	.298
		Brown-Forsythe	1.223	.298
AUB	8	Welch	.009	.937
		Brown-Forsythe	.009	.937
WCB	9	Welch	.560	.513
		Brown-Forsythe	.560	.513
ASB	16	Welch	.027	.873
		Brown-Forsythe	.027	.873
BPL	6	Welch	3.507	.134
		Brown-Forsythe	3.507	.134
NPH	6	Welch	.092	.791
		Brown-Forsythe	.092	.791
NLH	6	Welch	.254	.647
		Brown-Forsythe	.254	.647
OBH	8	Welch	2.419	.183
		Brown-Forsythe	2.419	.183
OBB	7	Welch	.089	.799
		Brown-Forsythe	.089	.799
JUB	4	Welch	.090	.794
		Brown-Forsythe	.090	.794
MAB	8	Welch	.021	.896
		Brown-Forsythe	.021	.896
MDH	16	Welch	1.337	.268
		Brown-Forsythe	1.337	.268
MDB	16	Welch	.301	.595
		Brown-Forsythe	.301	.595
FMB	14	Welch	6.212	.029
		Brown-Forsythe	6.212	.029
NAS	7	Welch	2.306	.244
		Brown-Forsythe	2.306	.244
EKB	5	Welch	.594	.521

Cranial Metric Variables	N	Test of Equality of Means	Statistic	Significance
EKB		Brown-Forsythe	.594	.521
WMH	11	Welch	.578	.468
		Brown-Forsythe	.578	.468
SOS	18	Welch	1.069	.331
		Brown-Forsythe	1.069	.331
GLS	17	Welch	1.928	.188
		Brown-Forsythe	1.928	.188
FOL	7	Welch	.029	.872
		Brown-Forsythe	.029	.872
FRC	19	Welch	.421	.526
		Brown-Forsythe	.421	.526
FRS	18	Welch	.811	.381
		Brown-Forsythe	.811	.381
FRF	18	Welch	.001	.977
		Brown-Forsythe	.001	.977
PAC	19	Welch	.019	.892
		Brown-Forsythe	.019	.892
PAS	18	Welch	.640	.436
		Brown-Forsythe	.640	.436
PAF	17	Welch	1.548	.236
		Brown-Forsythe	1.548	.236
OCC	16	Welch	1.934	.192
		Brown-Forsythe	1.934	.192
OCS	14	Welch	4.578	.054
		Brown-Forsythe	4.578	.054
OCF	14	Welch	.631	.449
		Brown-Forsythe	.631	.449
VRR	10	Welch	.598	.465
		Brown-Forsythe	.598	.465
NAR	8	Welch	.370	.566
		Brown-Forsythe	.370	.566
PRR	6	Welch	3.151	.174
		Brown-Forsythe	3.151	.174
DKR	4	Welch	.801	.521
		Brown-Forsythe	.801	.521
FMR	9	Welch	.109	.756
		Brown-Forsythe	.109	.756
EKR	8	Welch	.134	.742
		Brown-Forsythe	.134	.742
AVR	5	Welch	7.805	.105
		Brown-Forsythe	7.805	.105

Table 6.14 Sexual Dimorphism in the Palaikastro sample. One-way ANOVA, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	F	Significance
GOL	22	23.170	<0.001
NOL	21	20.625	<0.001
XCB	21	1.432	.246
XFB	20	1.328	.264
AUB	15	.080	.781
WCB	10	.629	.451
ASB	20	.229	.638
MDH	16	.297	.594
MDB	17	2.347	.146
FMB	11	5.667	.041
SOS	20	1.317	.266
GLS	17	15.296	.001
FRC	25	2.087	.162
FRS	22	1.418	.248
FRF	22	9.154	.007
PAC	24	19.948	<0.001
PAS	22	6.722	.017
PAF	22	18.383	.000
OCC	10	.070	.798
OCS	9	.007	.938
OCF	9	.909	.372
VRR	16	2.473	.138
NAR	14	7.212	.020
FMR	9	5.096	.059
EKR	5	2.855	.190

Table 6.15 Sexual Dimorphism in the Palaikastro sample. Test of Homogeneity of Variances, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Levene Statistic	Significance
GOL	22	4.574	.045
NOL	21	4.462	.048
XCB	21	.061	.808
XFB	20	.646	.432
AUB	15	.122	.732
WCB	10	1.774	.220
ASB	20	4.076	.059
MDH	16	.299	.593
MDB	17	.141	.713
FMB	11	.942	.357
SOS	20	3.047	.098
GLS	17	.248	.626
FRC	25	.636	.433
FRS	22	.426	.522
FRF	22	.298	.591
PAC	24	.184	.672
PAS	22	.061	.807
PAF	22	.173	.682
OCC	10	.026	.875
OCS	9	7.128	.032
OCF	9	.493	.505
VRR	16	.231	.638
NAR	14	.313	.586
FMR	9	1.502	.260
EKR	5	4.114	.136

Table 6.16 Sexual Dimorphism in the Palaikastro sample. Robust Tests of Equality of Means, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Test of Equality of means	Statistic	Significance
GOL	22	Welch	21.100	<0.001
		Brown-Forsythe	21.100	<0.001
NOL	21	Welch	19.583	.001
		Brown-Forsythe	19.583	.001
XCB	21	Welch	1.371	.261
		Brown-Forsythe	1.371	.261
XFB	20	Welch	1.263	.279
		Brown-Forsythe	1.263	.279
AUB	15	Welch	.105	.751
		Brown-Forsythe	.105	.751
WCB	10	Welch	1.421	.271
		Brown-Forsythe	1.421	.271
ASB	20	Welch	.253	.621
		Brown-Forsythe	.253	.621
MDH	16	Welch	.280	.607
		Brown-Forsythe	.280	.607
MDB	17	Welch	2.350	.146
		Brown-Forsythe	2.350	.146
FMB	11	Welch	5.322	.054
		Brown-Forsythe	5.322	.054
SOS	20	Welch	1.621	.231
		Brown-Forsythe	1.621	.231
GLS	17	Welch	15.013	.002
		Brown-Forsythe	15.013	.002
FRC	25	Welch	2.045	.168
		Brown-Forsythe	2.045	.168
FRS	22	Welch	1.542	.230
		Brown-Forsythe	1.542	.230
FRF	22	Welch	9.703	.006
		Brown-Forsythe	9.703	.006
PAC	24	Welch	19.948	<0.001
		Brown-Forsythe	19.948	<0.001
PAS	22	Welch	6.722	.017
		Brown-Forsythe	6.722	.017
PAF	22	Welch	18.383	<0.001
		Brown-Forsythe	18.383	<0.001
OCC	10	Welch	.070	.798
		Brown-Forsythe	.070	.798
OCS	9	Welch	.008	.932
		Brown-Forsythe	.008	.932
OCF	9	Welch	.949	.363
		Brown-Forsythe	.949	.363
VRR	16	Welch	2.640	.131
		Brown-Forsythe	2.640	.131

Cranial Metric Variables	N	Test	Statistic	Significance
NAR	14	Welch	6.110	.041
		Brown-Forsythe	6.110	.041
FMR	9	Welch	9.759	.049
		Brown-Forsythe	9.759	.049
EKR	5	Welch	2.127	.327
		Brown-Forsythe	2.127	.327

Table 6.17 Sexual Dimorphism in the Moni Odigitria sample. One-way ANOVA, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	F	Significance
GOL	13	.357	.562
NOL	12	.161	.696
BNL	3	.001	.976
BBH	3	1.333	.454
XCB	2	.126	.731
XFB	14	.269	.613
AUB	5	.772	.444
WCB	4	1.003	.422
ASB	14	1.033	.330
OBH	3	44.725	.094
OBB	3	.108	.798
MDH	14	3.218	.098
MDB	14	.373	.553
FMB	6	.005	.947
WMH	8	1.380	.285
SOS	13	6.033	.032
GLS	13	9.138	.012
FOL	3	.162	.756
FRC	15	.388	.544
FRS	15	2.923	.111
	2	2.923	.111
FRF	15	.971	.343
PAC	18	3.977	.063
PAS	18	.539	.473
PAF	18	.369	.552
OCC	10	.665	.439
OCS	9	.259	.627
OCF	9	.143	.717
VRR	7	.003	.959
NAR	5	1.602	.295

Table 6.18 Sexual Dimorphism in the Moni Odigitria sample. Test of Homogeneity of Variances, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Levene Statistic	Significance
GOL	13	.096	.762
NOL	12	.279	.609
XCB	11	.254	.626
XFB	14	.659	.433
AUB	5	1.494	.309
WCB	4	9277583482804090.000	<0.001
ASB	14	.001	.977
MDH	14	.412	.533
MDB	14	.316	.584
FMB	6	13.726	.021
WMH	8	.204	.667
SOS	13	.018	.896
GLS	13	7.238	.021
FRC	15	1.269	.280
FRS	15	2.260	.157
FRF	15	.969	.343
PAC	18	4.491	.050
PAS	18	3.394	.084
PAF	18	.239	.632
OCC	10	.326	.584
OCS	9	.011	.919
OCF	9	2.301	.173
VRR	7	.098	.767
NAR	5	.632	.485

Table 6.19 Sexual Dimorphism in the Moni Odigitria sample. Tests of Equality of Means, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Test	Statistic	Significance
GOL	13	Welch	.370	.556
		Brown-Forsythe	.370	.556
NOL	12	Welch	.161	.697
		Brown-Forsythe	.161	.697
XCB	11	Welch	.139	.720
		Brown-Forsythe	.139	.720
XFB	14	Welch	.242	.635
		Brown-Forsythe	.242	.635
AUB	5	Welch	1.201	.370
		Brown-Forsythe	1.201	.370
WCB	4	Welch	1.003	.489
		Brown-Forsythe	1.003	.489
ASB	14	Welch	1.063	.330
		Brown-Forsythe	1.063	.330
MDH	14	Welch	4.311	.072
		Brown-Forsythe	4.311	.072
MDB	14	Welch	.425	.537
		Brown-Forsythe	.425	.537
FMB	6	Welch	.005	.950
		Brown-Forsythe	.005	.950
WMH	8	Welch	1.470	.359
		Brown-Forsythe	1.470	.359
SOS	13	Welch	5.764	.039
		Brown-Forsythe	5.764	.039
FRC	15	Welch	.420	.530
		Brown-Forsythe	.420	.530
FRS	15	Welch	2.796	.123
		Brown-Forsythe	2.796	.123
FRF	15	Welch	1.024	.331
		Brown-Forsythe	1.024	.331
PAC	18	Welch	3.205	.107
		Brown-Forsythe	3.205	.107
PAS	18	Welch	.391	.550
		Brown-Forsythe	.391	.550
PAF	18	Welch	.354	.563
		Brown-Forsythe	.354	.563
OCC	10	Welch	.739	.416
		Brown-Forsythe	.739	.416
OCS	9	Welch	.257	.629
		Brown-Forsythe	.257	.629
OCF	9	Welch	.163	.699
		Brown-Forsythe	.163	.699
VRR	7	Welch	.003	.960
		Brown-Forsythe	.003	.960

Cranial Metric Variables	N	Test	Statistic	Significance
NAR	5	Welch	1.349	.382
		Brown-Forsythe	1.349	.382

Table 6.20 Sexual Dimorphism in the Ailias sample. Tests of Equality of Means, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Test	Statistic	Significance
GOL	51	Welch	29.356	<0.001
		Brown-Forsythe	29.356	<0.001
NOL	40	Welch	25.556	<0.001
		Brown-Forsythe	25.556	<0.001
BNL	10	Welch	19.613	.018
		Brown-Forsythe	19.613	.018
BBH	11	Welch	6.994	.051
		Brown-Forsythe	6.994	.051
XCB	45	Welch	4.086	.053
		Brown-Forsythe	4.086	.053
XFB	31	Welch	4.477	.051
		Brown-Forsythe	4.477	.051
AUB	25	Welch	10.067	.019
		Brown-Forsythe	10.067	.019
WCB	16	Welch	.191	.673
		Brown-Forsythe	.191	.673
ASB	42	Welch	15.003	<0.001
		Brown-Forsythe	15.003	<0.001
NLH	6	Welch	.032	.866
		Brown-Forsythe	.032	.866
OBH	12	Welch	.037	.854
		Brown-Forsythe	.037	.854
OBB	12	Welch	4.188	.073
		Brown-Forsythe	4.188	.073
JUB	4	Welch	69.444	.018
		Brown-Forsythe	69.444	.018
MAB	7	Welch	.360	.575
		Brown-Forsythe	.360	.575
MDH	37	Welch	10.046	.005
		Brown-Forsythe	10.046	.005
MDB	37	Welch	8.403	.006
		Brown-Forsythe	8.403	.006
ZMB	6	Welch	2.388	.245
		Brown-Forsythe	2.388	.245
FMB	30	Welch	7.406	.011
		Brown-Forsythe	7.406	.011
NAS	5	Welch	4.800	.117
		Brown-Forsythe	4.800	.117
EKB	6	Welch	11.449	.029
		Brown-Forsythe	11.449	.029
IML	6	Welch	.237	.672
		Brown-Forsythe	.237	.672
XML	6	Welch	4.914	.095
		Brown-Forsythe	4.914	.095

Cranial Metric Variables	N	Test	Statistic	Significance
WMH	13	Welch	3.148	.104
		Brown-Forsythe	3.148	.104
SOS	35	Welch	2.070	.166
		Brown-Forsythe	2.070	.166
GLS	37	Welch	29.702	.000
		Brown-Forsythe	29.702	.000
FOL	8	Welch	.006	.942
		Brown-Forsythe	.006	.942
FRC	47	Welch	8.252	.006
		Brown-Forsythe	8.252	.006
FRS	45	Welch	1.023	.317
		Brown-Forsythe	1.023	.317
FRF	45	Welch	1.284	.266
		Brown-Forsythe	1.284	.266
PAC	57	Welch	7.861	.008
		Brown-Forsythe	7.861	.008
PAS	55	Welch	.426	.519
		Brown-Forsythe	.426	.519
PAF	55	Welch	9.375	.004
		Brown-Forsythe	9.375	.004
OCC	23	Welch	.725	.404
		Brown-Forsythe	.725	.404
OCS	23	Welch	2.263	.150
		Brown-Forsythe	2.263	.150
OCF	23	Welch	.378	.546
		Brown-Forsythe	.378	.546
VRR	24	Welch	.874	.367
		Brown-Forsythe	.874	.367
NAR	20	Welch	9.900	.019
		Brown-Forsythe	9.900	.019
SSR	5	Welch	.405	.639
		Brown-Forsythe	.405	.639
DKR	6	Welch	1.891	.366
		Brown-Forsythe	1.891	.366
ZOR	9	Welch	7.605	.077
		Brown-Forsythe	7.605	.077
FMR	18	Welch	1.137	.337
		Brown-Forsythe	1.137	.337
EKR	9	Welch	1.163	.381
		Brown-Forsythe	1.163	.381
ZMR	9	Welch	.605	.494
		Brown-Forsythe	.605	.494
AVR	6	Welch	6.051	.075
		Brown-Forsythe	6.051	.075

Table 6.21 Sexual Dimorphism in the Ailias sample. Test of Homogeneity of Variances, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Levene Statistic	Significance
GOL	51	2.886	.096
NOL	40	1.998	.166
BNL	10	.267	.619
BBH	11	.780	.400
XCB	45	.706	.406
XFB	31	1.141	.294
AUB	25	4.690	.041
WCB	16	.488	.496
ASB	42	1.337	.254
NLH	6	4.401	.104
OBH	12	.041	.844
OBB	12	.326	.581
MAB	7	.381	.564
MDH	37	.788	.381
MDB	37	1.819	.186
ZMB	6	3.613	.130
FMB	30	1.950	.174
NAS	5	1.800	.272
EKB	6	.319	.603
IML	6	9.991	.034
XML	6	.437	.545
WMH	13	.699	.421
SOS	35	.117	.734
GLS	37	.205	.654
FOL	8	.007	.934
FRC	47	1.136	.292
FRS	45	1.279	.264
FRF	45	.386	.538
PAC	57	.269	.606
PAS	55	4.703	.035
PAF	55	.054	.818
OCC	23	.497	.488
OCS	23	.124	.728
OCF	23	.663	.425
VRR	24	.981	.333
NAR	20	.433	.519
SSR	5	2059.516	<0.001
DKR	6	3.613	.130
ZOR	9	1.538	.255
FMR	18	3.237	.091
EKR	9	2.319	.172
ZMR	9	.481	.510
AVR	6	3.597	.131

Table 6.22 Sexual Dimorphism in the Myrtos Pyrgos sample. One-way ANOVA, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	F	Significance
GOL	17	5.614	.032
NOL	15	7.124	.019
BNL	6	2.533	.187
BBH	7	2.031	.213
XCB	16	.446	.515
XFB	15	5.071	.042
STB	10	7.392	.026
AUB	11	.018	.897
WCB	9	.022	.887
ASB	15	.300	.593
MAB	6	1.827	.248
MDH	17	3.506	.081
MDB	17	2.430	.140
FMB	17	.515	.484
WMH	11	1.781	.215
SOS	16	.129	.725
GLS	16	1.176	.297
FRC	18	3.426	.083
FRS	16	1.510	.239
FRF	16	3.223	.094
PAC	21	7.694	.012
PAS	19	5.345	.034
PAF	19	2.874	.108
OCC	8	2.551	.161
OCS	7	.130	.734
OCF	7	.942	.376
VRR	9	1.743	.228
NAR	8	8.122	.029
FMR	9	7.272	.031
EKR	4	2.439	.259

Table 6.23 Sexual Dimorphism in the Myrtos Pyrgos sample. Test of Homogeneity of Variances, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Levene Statistic	Significance
GOL	17	.019	.893
NOL	15	3.980	.067
XCB	16	1.344	.266
XFB	15	2.791	.119
STB	10	.008	.930
AUB	11	6.758	.029
WCB	9	34.362	.001
ASB	15	2.601	.131
MDH	17	.020	.890
MDB	17	1.376	.259
FMB	17	1.324	.268
SOS	16	.068	.798
GLS	16	3.458	.084
FRC	18	1.058	.319
FRS	16	1.853	.195
FRF	16	.031	.862
PAC	21	.393	.538
PAS	19	.083	.777
PAF	19	2.209	.156
OCC	8	.302	.602
OCS	7	.535	.497
OCF	7	2.594	.168
VRR	9	.001	.982
FMR	9	2.763	.140

Table 6.24 Sexual Dimorphism in the Myrtos Pyrgos sample. Tests of Equality of Means, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Test	Statistic	Significance
GOL	17	Welch	5.325	.071
		Brown-Forsythe	5.325	.071
NOL	15	Welch	48.606	.000
		Brown-Forsythe	48.606	.000
XCB	16	Welch	.778	.393
		Brown-Forsythe	.778	.393
XFB	15	Welch	7.731	.016
		Brown-Forsythe	7.731	.016
STB	10	Welch	7.542	.029
		Brown-Forsythe	7.542	.029
AUB	11	Welch	.007	.938
		Brown-Forsythe	.007	.938
WCB	9	Welch	.005	.953
		Brown-Forsythe	.005	.953
ASB	15	Welch	.227	.651
		Brown-Forsythe	.227	.651
MDH	17	Welch	2.972	.132
		Brown-Forsythe	2.972	.132
MDB	17	Welch	4.110	.062
		Brown-Forsythe	4.110	.062
FMB	17	Welch	.282	.626
		Brown-Forsythe	.282	.626
SOS	16	Welch	.122	.751
		Brown-Forsythe	.122	.751
GLS	16	Welch	.511	.542
		Brown-Forsythe	.511	.542
FRC	18	Welch	7.709	.017
		Brown-Forsythe	7.709	.017
FRS	16	Welch	2.150	.184
		Brown-Forsythe	2.150	.184
FRF	16	Welch	3.682	.105
		Brown-Forsythe	3.682	.105
PAC	21	Welch	8.333	.012
		Brown-Forsythe	8.333	.012
PAS	19	Welch	4.625	.062
		Brown-Forsythe	4.625	.062
PAF	19	Welch	3.645	.078
		Brown-Forsythe	3.645	.078
OCC	8	Welch	3.460	.186
		Brown-Forsythe	3.460	.186
OCS	7	Welch	.087	.807
		Brown-Forsythe	.087	.807
OCF	7	Welch	.470	.601
		Brown-Forsythe	.470	.601

Cranial Metric Variables	N	Test	Statistic	Significance
VRR	9	Welch	1.675	.267
		Brown-Forsythe	1.675	.267
FMR	9	Welch	14.536	.009
		Brown-Forsythe	14.536	.009

Table 6.25 Sexual Dimorphism in the Gypsades (Upper and Lower) sample. Test of Homogeneity of Variances, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Levene Statistic	Significance
GOL	11	5.006	.052
NOL	10	.927	.364
XCB	12	.251	.627
XFB	9	.070	.799
ASB	10	1.999	.195
MDH	11	.699	.425
MDB	11	1.053	.332
FMB	12	1.110	.317
SOS	9	.289	.608
GLS	11	3.705	.086
FRC	17	.527	.479
FRS	13	.869	.371
FRF	13	7.837	.017
PAC	17	.685	.421
PAS	16	.053	.822
PAF	16	.272	.610

Table 6.26 Sexual Dimorphism in the Gypsades (Upper and Lower) sample. One-way ANOVA, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	F	Significance
GOL	11	.630	.448
NOL	10	3.309	.106
XCB	12	5.191	.046
XFB	9	19.977	.003
STB	10	3.372	.164
ASB	11	6.275	.037
MDH	11	.065	.805
MDB	12	12.012	.007
FMB	9	4.843	.052
SOS	11	3.036	.125
GLS	17	6.770	.029
FRC	13	.401	.536
FRS	13	.913	.360
FRF	17	1.137	.309
PAC	16	.031	.862
PAS	16	2.075	.172
PAF	11	.017	.898

Table 6.27 Sexual Dimorphism in the Lower Gypsades sample. One-way ANOVA, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	F	Significance
GOL	9	1.857	.215
NOL	8	15.078	.008
XCB	11	4.267	.069
XFB	7	11.954	.018
STB	5	3.372	.164
ASB	9	4.822	.064
MDH	10	.086	.777
MDB	10	11.564	.009
FMB	10	3.764	.088
SOS	7	17.286	.009
GLS	9	9.016	.020
FRC	14	.296	.596
FRS	10	.258	.625
FRF	10	.071	.796
PAC	14	.000	.992
PAS	14	1.732	.213
PAF	14	.045	.835

Table 6.28 Sexual Dimorphism in the Lower Gypsades sample. Test of Homogeneity of Variances, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Levene Statistic	Significance
GOL	9	.637	.451
NOL	8	.973	.362
XCB	11	.108	.750
XFB	7	.004	.952
ASB	5	1.419	.272
MDH	9	.473	.511
MDB	10	1.348	.279
FMB	10	1.336	.281
SOS	10	.357	.576
GLS	7	6.254	.041
FRC	9	.265	.616
FRS	14	3.337	.105
FRF	10	3.150	.114
PAC	10	.136	.719
PAS	14	.043	.840
PAF	14	.187	.673

Table 6.29 Sexual Dimorphism in the Lower Gypsades sample. Tests of Equality of Means, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Test	Statistic	Significance
GOL	9	Welch	1.681	.251
		Brown-Forsythe	1.681	.251
NOL	8	Welch	18.760	.005
		Brown-Forsythe	18.760	.005
XCB	11	Welch	3.744	.108
		Brown-Forsythe	3.744	.108
XFB	7	Welch	11.441	.027
		Brown-Forsythe	11.441	.027
ASB	9	Welch	4.069	.114
		Brown-Forsythe	4.069	.114
MDH	10	Welch	.071	.806
		Brown-Forsythe	.071	.806
MDB	10	Welch	19.371	.003
		Brown-Forsythe	19.371	.003
FMB	10	Welch	5.577	.057
		Brown-Forsythe	5.577	.057
SOS	7	Welch	17.286	.011
		Brown-Forsythe	17.286	.011
GLS	9	Welch	10.601	.018
		Brown-Forsythe	10.601	.018
FRC	14	Welch	.340	.573
		Brown-Forsythe	.340	.573
FRS	10	Welch	.258	.632
		Brown-Forsythe	.258	.632
FRF	10	Welch	.071	.798
		Brown-Forsythe	.071	.798
PAC	14	Welch	.000	.992
		Brown-Forsythe	.000	.992
PAS	14	Welch	1.793	.206
		Brown-Forsythe	1.793	.206
PAF	14	Welch	.043	.840
		Brown-Forsythe	.043	.840

Table 6.30 Sexual Dimorphism in the Sellopoulo sample. Tests of Equality of Means, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Test	Statistic	Significance
GOL	5	Welch	1.207	.382
		Brown-Forsythe	1.207	.382
XCB	5	Welch	.046	.849
		Brown-Forsythe	.046	.849
STB	4	Welch	.077	.820
		Brown-Forsythe	.077	.820
ASB	4	Welch	1.053	.477
		Brown-Forsythe	1.053	.477
FMB	4	Welch	1.484	.369
		Brown-Forsythe	1.484	.369
PAC	6	Welch	1.364	.310
		Brown-Forsythe	1.364	.310
PAS	6	Welch	1.482	.310
		Brown-Forsythe	1.482	.310
PAF	6	Welch	.000	.995
		Brown-Forsythe	.000	.995
OCS	4	Welch	18.000	.051
		Brown-Forsythe	18.000	.051
OCF	4	Welch	1.434	.416
		Brown-Forsythe	1.434	.416

Table 6.31 Sexual Dimorphism in Mavrospelio sample. Test of Homogeneity of Variances, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Levene Statistic	Significance.
GOL	7	.457	.529
NOL	7	.258	.633
XCB	6	.092	.777
XFB	6	1.667	.266
ASB	6	1.788	.252
SOS	6	3.303	.143
GLS	7	18.687	.008
FRC	7	1.709	.248
FRS	7	.817	.408
FRF	7	1.880	.229
PAC	7	3.262	.131
PAS	7	15.060	.012
PAF	7	2.273	.192
VRR	4	34206406440090160.000	<0.001

Table 6.32 Sexual Dimorphism in the Mavrospelio sample. One-way ANOVA, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	F	Significance
GOL	7	1.146	.333
NOL	7	1.240	.316
XCB	7	1.624	.272
XFB	6	.024	.885
AUB	4	.672	.499
ASB	6	1.670	.266
MDH	3	29.940	.115
MDB	3	.133	.777
FMB	3	.255	.702
WNB	3	.035	.882
SOS	6	.417	.554
GLS	7	5.576	.065
FRC	7	.204	.670
FRS	7	4.124	.098
FRF	7	1.429	.286
PAC	7	.194	.678
PAS	7	1.923	.224
PAF	7	.031	.868
OCC	3	16.333	.154
OCS	3	16.333	.154
OCF	3	.083	.821
VRR	4	4.169	.178
NAR	4	1.581	.336
FMR	3	.197	.734

Table 6.33 Sexual Dimorphism in the Mavrospelio sample. Tests of Equality of Means, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Test	Statistic	Significance
GOL	7	Welch	1.054	.367
		Brown-Forsythe	1.054	.367
NOL	7	Welch	1.169	.341
		Brown-Forsythe	1.169	.341
XCB	7	Welch	1.382	.376
		Brown-Forsythe	1.382	.376
XFB	6	Welch	.051	.835
		Brown-Forsythe	.051	.835
AUB	4	Welch	.672	.561
		Brown-Forsythe	.672	.561
ASB	6	Welch	3.181	.151
		Brown-Forsythe	3.181	.151
SOS	6	Welch	.854	.413
		Brown-Forsythe	.854	.413
FRC	7	Welch	.173	.705
		Brown-Forsythe	.173	.705
FRS	7	Welch	4.880	.080
		Brown-Forsythe	4.880	.080
FRF	7	Welch	1.742	.249
		Brown-Forsythe	1.742	.249
PAC	7	Welch	.219	.659
		Brown-Forsythe	.219	.659
PAS	7	Welch	2.519	.191
		Brown-Forsythe	2.519	.191
PAF	7	Welch	.037	.856
		Brown-Forsythe	.037	.856
VRR	4	Welch	4.169	.251
		Brown-Forsythe	4.169	.251
NAR	4	Welch	1.581	.416
		Brown-Forsythe	1.581	.416

Table 6.34 Sexual Dimorphism in the Kastelos sample. One-way ANOVA, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	F	Significance
XCB	4	.431	.579
XFB	4	.862	.451
STB	3	.037	.879
ASB	4	3.167	.217
MDH	3	6.550	.237
MDB	3	6.001	.247
FMB	4	3.200	.216
PAC	4	2.390	.262
PAS	4	2.596	.248
PAF	4	.021	.898

Table 6.35 Sexual Dimorphism in the Kastelos sample. Tests of Equality of Means, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Test	Statistic	Significance
XCB	4	Welch	.431	.605
		Brown-Forsythe	.431	.605
XFB	4	Welch	.862	.492
		Brown-Forsythe	.862	.492
ASB	4	Welch	3.167	.222
		Brown-Forsythe	3.167	.222
FMB	4	Welch	3.200	.259
		Brown-Forsythe	3.200	.259
PAC	4	Welch	2.390	.314
		Brown-Forsythe	2.390	.314
PAS	4	Welch	2.596	.255
		Brown-Forsythe	2.596	.255
PAF	4	Welch	.021	.903
		Brown-Forsythe	.021	.903

Table 6.36 Sexual Dimorphism in the Kastelos sample. Test of Homogeneity of Variances, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Levene Statistic	Significance
XFB	4	5975406588806000.000	<0.001
FMB	4	2166636607201028.000	<0.001
PAC	4	2438038896020118.000	<0.001
PAF	4	7837275396484740.000	<0.001

Table 6.37 Sexual Dimorphism in the Palama sample. One-way ANOVA, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	F	Significance
GOL	9	.074	.793
NOL	7	.530	.499
BBH	3	1.215	.469
XCB	8	1.372	.286
XFB	6	4.721	.096
STB	5	8.620	.061
AUB	3	.746	.546
ASB	5	.885	.416
MAB	3	.020	.910
MDH	5	.132	.741
MDB	7	.132	.732
FMB	7	.284	.617
WNB	3	5.058	.266
WMH	4	2.274	.271
SOS	7	9.545	.027
GLS	5	.481	.538
FRC	8	1.392	.283
FRS	8	.106	.756
FRF	8	1.964	.211
PAC	10	4.050	.079
PAS	10	.870	.378
PAF	10	.489	.504
OCC	7	.254	.636
OCS	7	.159	.706
OCF	7	1.128	.337

Table 6.38 Sexual Dimorphism in the Palama sample. Test of Homogeneity of Variances, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	Levene Statistic	Significance
GOL	9	.215	.657
NOL	7	.558	.489
XCB	8	.007	.938
XFB	6	.441	.543
STB	5	1.338	.331
ASB	5	4.499	.124
MDH	5	42.774	.007
MDB	7	1.052	.352
FMB	7	4.823	.079
SOS	7	.854	.398
GLS	5	2.019	.250
FRC	8	5.236	.062
FRS	8	.275	.619
FRF	8	.359	.571
PAC	10	.274	.615
PAS	10	.116	.742
PAF	10	.351	.570
OCC	7	.370	.570
OCS	7	19.688	.007
OCF	7	.079	.790

Table 6.39 Sexual Dimorphism in the Palama sample. Tests of Equality of Means, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Test	Statistic	Significance
GOL	9	Welch	.071	.798
		Brown-Forsythe	.071	.798
NOL	7	Welch	.647	.461
		Brown-Forsythe	.647	.461
XCB	8	Welch	1.372	.287
		Brown-Forsythe	1.372	.287
XFB	6	Welch	6.601	.076
		Brown-Forsythe	6.601	.076
STB	5	Welch	10.482	.050
		Brown-Forsythe	10.482	.050
ASB	5	Welch	1.461	.347
		Brown-Forsythe	1.461	.347
MDH	5	Welch	.081	.819
		Brown-Forsythe	.081	.819
MDB	7	Welch	.173	.700
		Brown-Forsythe	.173	.700
FMB	7	Welch	.207	.691
		Brown-Forsythe	.207	.691
SOS	7	Welch	7.737	.076
		Brown-Forsythe	7.737	.076
GLS	5	Welch	.771	.464
		Brown-Forsythe	.771	.464
FRC	8	Welch	2.004	.207
		Brown-Forsythe	2.004	.207
FRS	8	Welch	.132	.730
		Brown-Forsythe	.132	.730
FRF	8	Welch	2.161	.202
		Brown-Forsythe	2.161	.202
PAC	10	Welch	4.050	.079
		Brown-Forsythe	4.050	.079
PAS	10	Welch	.870	.379
		Brown-Forsythe	.870	.379
PAF	10	Welch	.489	.506
		Brown-Forsythe	.489	.506
OCC	7	Welch	.356	.596
		Brown-Forsythe	.356	.596
OCS	7	Welch	.443	.542
		Brown-Forsythe	.443	.542
OCF	7	Welch	1.380	.345
		Brown-Forsythe	1.380	.345

Table 6.40 Sexual Dimorphism in the Cretan population (samples pooled). One-way ANOVA, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	N	F	Significance
GOL	141	44.843	<0.001
NOL	121	39.387	<0.001
BNL	28	11.090	<0.001
BBH	32	10.179	<0.001
XCB	133	8.332	.005
XFB	115	11.979	.001
STB	32	5.227	.029
ZYB	11	.431	.528
AUB	71	10.191	.002
WCB	48	3.038	.088
ASB	126	14.842	.000
BPL	8	.386	.557
NPH	10	.287	.607
NLH	20	1.926	.182
OBH	30	.916	.347
OBB	30	.273	.606
JUB	15	.643	.437
NLB	8	.065	.807
MAB	23	.145	.707
MDH	111	20.183	.000
MDB	113	14.551	.000
ZMB	20	4.579	.046
SSS	13	.058	.814
FMB	98	7.502	.007
NAS	17	2.709	.121
EKB	20	2.378	.140
DKS	10	2.120	.184
DKB	8	1.189	.317
WNB	20	.061	.808
IML	17	.878	.364
XML	15	1.503	.242
WMH	47	10.585	.002
SOS	117	3.290	.072
GLS	117	51.342	<0.001
FOL	22	2.811	.109
FRC	151	14.300	.000
FRS	139	6.045	.015
FRF	139	13.014	<0.001
PAC	170	25.863	<0.001
PAS	163	7.868	.006

Cranial Metric Variables	N	F	Significance
PAF	163	16.813	<0.001
OCC	73	5.248	.025
OCS	69	5.412	.023
OCF	69	2.767	.101
VRR	72	7.926	.006
NAR	61	12.725	.001
SSR	14	1.719	.214
PRR	12	2.274	.162
DKR	18	3.538	.078
ZOR	22	11.183	.003
FMR	52	10.958	.002
EKR	27	6.973	.014
ZMR	21	2.795	.111
AVR	17	7.106	.018

Table 6.41 Sexual Dimorphism in the Cretan population (samples pooled). Test of Homogeneity of Variances, Cranial Metric Variables (p<0.05).

Cranial Metric Variables	N	Levene Statistic	Significance
GOL	141	.860	.355
NOL	121	1.151	.285
BNL	28	.002	.968
BBH	32	.491	.489
XCB	133	.674	.413
XFB	115	.301	.585
STB	32	.006	.937
ZYB	11	3.926	.079
AUB	71	.302	.584
WCB	48	.356	.554
ASB	126	.439	.509
BPL	8	.946	.368
NPH	10	.164	.696
NLH	20	1.650	.215
OBH	30	.008	.932
OBB	30	4.246	.049
JUB	15	.819	.382
NLB	8	.095	.768
MAB	23	.592	.450
MDH	111	.574	.450
MDB	113	3.444	.066
ZMB	20	15.461	.001
SSS	13	2.317	.156
FMB	98	.004	.950
NAS	17	3.338	.088
EKB	20	1.529	.232
DKS	10	.002	.966
DKB	8	9.373	.022
WNB	20	.179	.677
IML	17	9.129	.009
XML	15	.165	.691
WMH	47	.022	.884
SOS	117	.811	.370
GLS	117	1.644	.202
FOL	22	.492	.491
FRC	151	2.055	.154
FRS	139	4.000	.047
FRF	139	.037	.849
PAC	170	.035	.853
PAS	163	1.903	.170
PAF	163	.025	.874
OCC	73	.565	.455
OCS	69	3.877	.053
OCF	69	1.924	.170

Cranial Metric Variables	N	Levene Statistic	Significance
VRR	72	.041	.840
NAR	61	.420	.519
SSR	14	.535	.478
PRR	12	3.387	.096
DKR	18	2.989	.103
ZOR	22	1.005	.328
FMR	52	.019	.891
EKR	27	.078	.782
ZMR	21	.150	.703
AVR	17	4.525	.050

Table 6.42 Sexual Dimorphism in the Cretan population (samples pooled). Robust tests of equality of means, Cranial Metric Variables ($p<0.05$).

Cranial Metric Variables	Tests of Equality of Means	Statistic	Df	Significance
GOL	Welch	46.288	131.643	<0.001
	Brown-Forsythe	46.288	131.643	<0.001
NOL	Welch	41.026	107.411	<0.001
	Brown-Forsythe	41.026	107.411	<0.001
BNL	Welch	10.407	9.805	.009
	Brown-Forsythe	10.407	9.805	.009
BBH	Welch	10.406	17.961	.005
	Brown-Forsythe	10.406	17.961	.005
XCB	Welch	8.919	107.824	.003
	Brown-Forsythe	8.919	107.824	.003
XFB	Welch	11.474	89.352	.001
	Brown-Forsythe	11.474	89.352	.001
STB	Welch	5.082	24.651	.033
	Brown-Forsythe	5.082	24.651	.033
ZYB	Welch	.484	7.739	.507
	Brown-Forsythe	.484	7.739	.507
AUB	Welch	9.830	41.539	.003
	Brown-Forsythe	9.830	41.539	.003
WCB	Welch	4.340	41.951	.043
	Brown-Forsythe	4.340	41.951	.043
ASB	Welch	15.421	108.718	<0.001
	Brown-Forsythe	15.421	108.718	<0.001
BPL	Welch	.767	3.981	.431
	Brown-Forsythe	.767	3.981	.431
NPH	Welch	.314	1.634	.642
	Brown-Forsythe	.314	1.634	.642
NLH	Welch	2.931	15.906	.106
	Brown-Forsythe	2.931	15.906	.106
OBH	Welch	.945	22.022	.342
	Brown-Forsythe	.945	22.022	.342
OBB	Welch	.361	27.999	.553
	Brown-Forsythe	.361	27.999	.553
JUB	Welch	.798	6.702	.403
	Brown-Forsythe	.798	6.702	.403
NLB	Welch	.092	2.448	.785
	Brown-Forsythe	.092	2.448	.785
MAB	Welch	.124	9.775	.732
	Brown-Forsythe	.124	9.775	.732
MDH	Welch	19.505	74.318	<0.001
	Brown-Forsythe	19.505	74.318	<0.001
MDB	Welch	17.653	103.417	<0.001
	Brown-Forsythe	17.653	103.417	<0.001
ZMB	Welch	2.525	6.344	.160

Cranial Metric Variables	Tests of Equality of Means	Statistic	Df	Significance
ZMB	Brown-Forsythe	2.525	6.344	.160
SSS	Welch	.034	3.693	.863
	Brown-Forsythe	.034	3.693	.863
FMB	Welch	7.458	83.141	.008
	Brown-Forsythe	7.458	83.141	.008
NAS	Welch	5.036	14.916	.040
	Brown-Forsythe	5.036	14.916	.040
EKB	Welch	2.976	12.498	.109
	Brown-Forsythe	2.976	12.498	.109
DKS	Welch	2.121	6.586	.191
	Brown-Forsythe	2.121	6.586	.191
DKB	Welch	3.820	5.369	.104
	Brown-Forsythe	3.820	5.369	.104
WNB	Welch	.061	17.335	.808
	Brown-Forsythe	.061	17.335	.808
IML	Welch	.717	8.567	.420
	Brown-Forsythe	.717	8.567	.420
XML	Welch	1.220	6.304	.310
	Brown-Forsythe	1.220	6.304	.310
WMH	Welch	10.665	27.732	.003
	Brown-Forsythe	10.665	27.732	.003
SOS	Welch	5.504	88.876	.021
	Brown-Forsythe	5.504	88.876	.021
GLS	Welch	52.175	95.993	.000
	Brown-Forsythe	52.175	95.993	.000
FOL	Welch	3.160	17.362	.093
	Brown-Forsythe	3.160	17.362	.093
FRC	Welch	15.333	140.139	<0.001
	Brown-Forsythe	15.333	140.139	<0.001
FRS	Welch	6.900	136.733	.010
	Brown-Forsythe	6.900	136.733	.010
FRF	Welch	12.619	111.771	.001
	Brown-Forsythe	12.619	111.771	.001
PAC	Welch	27.117	151.953	.000
	Brown-Forsythe	27.117	151.953	.000
PAS	Welch	7.292	117.503	.008
	Brown-Forsythe	7.292	117.503	.008
PAF	Welch	17.397	142.154	<0.001
	Brown-Forsythe	17.397	142.154	<0.001
OCC	Welch	5.638	70.666	.020
	Brown-Forsythe	5.638	70.666	.020
OCS	Welch	5.894	66.791	.018
	Brown-Forsythe	5.894	66.791	.018
OCF	Welch	2.950	65.943	.091
	Brown-Forsythe	2.950	65.943	.091
VRR	Welch	8.065	44.086	.007

Cranial Metric Variables	Tests of Equality of Means	Statistic	Df	Significance
	Brown-Forsythe	8.065	44.086	.007
NAR	Welch	9.989	26.859	.004
	Brown-Forsythe	9.989	26.859	.004
SSR	Welch	1.354	4.513	.302
	Brown-Forsythe	1.354	4.513	.302
PRR	Welch	2.664	9.921	.134
	Brown-Forsythe	2.664	9.921	.134
DKR	Welch	3.821	15.844	.068
	Brown-Forsythe	3.821	15.844	.068
ZOR	Welch	14.888	16.972	.001
	Brown-Forsythe	14.888	16.972	.001
FMR	Welch	10.898	28.671	.003
	Brown-Forsythe	10.898	28.671	.003
EKR	Welch	7.414	20.824	.013
	Brown-Forsythe	7.414	20.824	.013
ZMR	Welch	2.497	10.496	.144
	Brown-Forsythe	2.497	10.496	.144
AVR	Welch	10.984	14.543	.005
	Brown-Forsythe	10.984	14.543	.005

Table 6.43 Inter-population analysis, Cretan samples, males. One-way ANOVA, Dental Metric Variables ($p<0.05$).

Dental Metric Variables	N	F	Significance
I1LBL	9	.176	.939
I1LMD	10	7.612	.024
I2LBL	12	.387	.812
I2LMD	14	4.027	.038
CLBL	14	.248	.904
CLMD	15	1.242	.354
PM3LBL	16	.166	.951
PM3LMD	17	1.888	.177
PM4LBL	14	.449	.771
PM4LMD	16	.742	.583
M1LBL	15	.737	.588
M1LMD	15	1.299	.334
M2LBL	13	1.623	.259
M2LMD	14	.705	.608
I1RBL	5	1.200	.455
I1RMD	5	14.453	.065
I2RBL	12	1.208	.387
I2RMD	11	1.617	.285
CRBL	11	1.512	.309
CRMD	11	.842	.546
PM3RBL	19	.789	.552
PM3RMD	19	1.407	.282
PM4RBL	14	.495	.741
PM4RMD	14	2.462	.120
M1RBL	13	.243	.906
M1RMD	16	2.565	.097
M2RBL	16	1.890	.182
M2RMD	15	2.328	.127
M3RBL	7	2.255	.261
M3RMD	7	1.599	.355
LI1LBL	7	.437	.674
LI1LMD	9	1.107	.428
LI2LBL	8	.397	.763
LI2LMD	12	1.969	.204
LCLBL	18	2.415	.102
LCLMD	18	2.347	.109
LPM3LBL	15	3.736	.041
LPM3LMD	17	3.083	.058
LPM4LBL	14	.341	.844
LPM4LMD	15	1.399	.303
LM1LBL	18	1.311	.317
LM1LMD	19	2.187	.123
LM2LBL	17	1.608	.236

LM2LMD	17	4.862	.015
LM3LBL	10	.183	.836
LM3LMD	10	.437	.662
LI1RBL	8	.766	.570
LI1RMD	11	.750	.593
LI2RBL	7	.147	.948
LI2RMD	11	3.603	.079
LCRBL	12	1.147	.409
LCRMD	12	3.323	.079
LPM3RBL	13	1.044	.442
LPM3RMD	16	4.141	.028
LPM4RBL	12	2.120	.181
LPM4RMD	13	2.059	.179
LM1RBL	15	.469	.758
LM1RMD	15	.096	.981
LM2RBL	16	.257	.899
LM2RMD	16	.816	.541
LM3RBL	12	.129	.967
LM3RMD	12	.260	.895

Table 6.44 Inter-population analysis, Cretan samples, females. One-way ANOVA, Dental Metric Variables (p<0.05).

Dental Metric Variables	N	F	Significance
I1LBL	7	.838	.402
I1LMD	7	13.869	.014
I2LBL	6	1.557	.414
I2LMD	6	2.719	.280
CLBL	7	.192	.896
CLMD	8	1.970	.302
PM3LBL	8	.945	.541
PM3LMD	7	.703	.659
PM4LBL	7	4.730	.117
PM4LMD	8	1.573	.328
M1LBL	14	.173	.947
M1LMD	15	1.806	.204
M2LBL	10	3.217	.116
M2LMD	10	1.395	.356
M3LBL	6	.262	.850
M3LMD	6	.797	.598
I1RBL	4	2.052	.443
I1RMD	4	301.500	.041
I2RBL	5	.918	.521
I2RMD	5	3.589	.218
CRBL	8	2.291	.261
CRMD	10	.481	.751
PM3RBL	13	.403	.802
PM3RMD	15	.344	.843
PM4RBL	12	.228	.914
PM4RMD	14	.133	.966
M1RBL	15	1.239	.355
M1RMD	16	.305	.869
M2RBL	14	1.998	.179
M2RMD	14	1.240	.360
M3RBL	7	7.537	.066
M3RMD	7	3.074	.191
LI1LMD	4	.429	.580
LI2LBL	3	.083	.821
LI2LMD	9	.645	.619
LCLBL	11	1.423	.315
LCLMD	12	1.243	.357
LPM3LBL	12	.708	.574
LPM3LMD	11	.235	.870
LPM4LBL	10	1.790	.249
LPM4LMD	10	.784	.545
LM1LBL	11	1.369	.328
LM1LMD	9	.574	.656

LM2LBL	11	.224	.877
LM2LMD	12	.926	.471
LM3LBL	7	1.438	.386
LM3LMD	9	.425	.744
LI1RBL	3	.037	.879
LI1RMD	8	.017	.984
LI2RBL	7	.298	.758
LI2RMD	9	.142	.870
LCRBL	7	.496	.642
LCRMD	9	.478	.642
LPM3RBL	9	2.029	.229
LPM3RMD	11	3.691	.070
LPM4RBL	11	1.618	.270
LPM4RMD	11	.076	.971
LM1RBL	15	.420	.742
LM1RMD	15	.393	.761
LM2RBL	13	1.688	.238
LM2RMD	16	1.259	.332
LM3RBL	7	1.849	.270
LM3RMD	7	1.701	.292

Table 6.45 Inter-population analysis, Cretan samples, unsexed individuals. One-way ANOVA, Dental Metric Variables ($p<0.05$).

Dental Metric Variables	N	F	Significance
CLBL	4	.152	.734
CLMD	4	.045	.852
PM3LBL	6	1.966	.285
PM3LMD	7	3.478	.133
PM4LBL	11	.252	.783
PM4LMD	10	6.711	.024
M1LBL	7	3.742	.121
M1LMD	8	.712	.535
M2LBL	10	2.086	.195
M2LMD	9	1.049	.407
M3LBL	6	2.506	.189
M3LMD	6	.299	.613
I1RBL	5	.257	.647
I1RMD	5	7.305	.074
I2RBL	4	12.500	.072
I2RMD	3	120.333	.058
CRBL	7	.653	.456
CRMD	7	.331	.590
PM3RBL	10	10.238	.013
PM3RMD	10	1.140	.317
PM4RBL	11	.110	.747
PM4RMD	11	.335	.577
M1RBL	6	.132	.735
M1RMD	5	62.422	.004
M2RBL	7	1.706	.248
M2RMD	7	.049	.834
LI2LMD	4	.195	.702
LCLBL	6	.368	.720
LCLMD	7	.276	.772
LPM3LBL	8	3.174	.129
LPM3LMD	8	1.346	.341
LPM4LBL	8	.927	.455
LPM4LMD	7	.853	.491
LM1LBL	7	.066	.937
LM1LMD	7	.906	.474
LM2LBL	8	.404	.688
LM2LMD	8	1.752	.265
LI2RMD	3	.196	.735
LCRBL	5	.375	.584
LCRMD	5	.298	.623
LPM3RBL	10	.241	.637
LPM3RMD	10	1.301	.287
LPM4RBL	4	.465	.566
LPM4RMD	4	1.142	.397

LM1RBL	5	.662	.602
LM1RMD	5	.119	.894
LM2RBL	8	.697	.541
LM2RMD	8	.327	.735
LM3RBL	5	1.939	.258
LM3RMD	5	.521	.523

Table 6.46 Inter-population analysis, Cretan samples, males. Robust tests of Equality of Means, Dental Metric Variables ($p<0.05$).

Dental Metric Variables	N	Tests of Equality of Means	Statistic	Significance
CLBL	14	Welch	.512	.738
		Brown-Forsythe	.278	.876
CLMD	15	Welch	1.494	.368
		Brown-Forsythe	1.948	.189
PM4LBL	14	Welch	.347	.834
		Brown-Forsythe	.421	.789
PM4LMD	16	Welch	2.429	.230
		Brown-Forsythe	.641	.671
PM3RBL	19	Welch	1.845	.292
		Brown-Forsythe	1.141	.454
PM3RMD	19	Welch	3.535	.120
		Brown-Forsythe	1.119	.456
PM4RBL	14	Welch	.494	.747
		Brown-Forsythe	.555	.717
PM4RMD	14	Welch	2.582	.241
		Brown-Forsythe	2.416	.258
M1RMD	16	Welch	2.347	.219
		Brown-Forsythe	2.486	.193
LM3LBL	10	Welch	.292	.762
		Brown-Forsythe	.224	.808
LM3LMD	10	Welch	.518	.636
		Brown-Forsythe	.555	.601

Table 6.47 Inter-population analysis, Cretan samples, unsexed individuals. Robust tests of Equality of Means, Dental Metric Variables ($p<0.05$).

Dental Metric Variables	N	Tests of Equality of Means	Statistic	Significance
M3LBL	6	Welch	1.323	.432
		Brown-Forsythe	1.323	.432
M3LMD	6	Welch	.580	.490
		Brown-Forsythe	.580	.490
I1RBL	5	Welch	.200	.710
		Brown-Forsythe	.200	.710
I1RMD	5	Welch	9.308	.056
		Brown-Forsythe	9.308	.056
I2RBL	4	Welch	12.500	.072
		Brown-Forsythe	12.500	.072
PM3RMD	10	Welch	2.496	.155
		Brown-Forsythe	2.496	.155
LI2LMD	4	Welch	.195	.734
		Brown-Forsythe	.195	.734
LPM3RBL	10	Welch	.373	.561
		Brown-Forsythe	.373	.561
LPM3RMD	10	Welch	3.061	.125
		Brown-Forsythe	3.061	.125
LM3RBL	5	Welch	2.316	.229
		Brown-Forsythe	2.316	.229
LM3RMD	5	Welch	.796	.449
		Brown-Forsythe	.796	.449

Table 6.48 Inter-population analysis, Cretan samples, females. Robust tests of Equality of Means, Dental Metric Variables (p<0.05).

Dental MetricVariables	N	Tests of Equality of Means	Statistic	Significance
I1LBL	7	Welch	.868	.397
		Brown-Forsythe	.868	.397
I1LMD	7	Welch	19.253	.020
		Brown-Forsythe	19.253	.020
PM4LMD	8	Welch	1.854	.358
		Brown-Forsythe	1.573	.367
M1LMD	15	Welch	.571	.701
		Brown-Forsythe	1.213	.525
PM3RBL	13	Welch	.435	.781
		Brown-Forsythe	.298	.860
PM3RMD	15	Welch	.219	.913
		Brown-Forsythe	.287	.873
PM4RBL	12	Welch	.228	.906
		Brown-Forsythe	.188	.928
PM4RMD	14	Welch	.046	.994
		Brown-Forsythe	.097	.975
M1RBL	15	Welch	3.827	.115
		Brown-Forsythe	1.294	.374
M1RMD	16	Welch	.360	.827
		Brown-Forsythe	.302	.861
LPM3LBL	12	Welch	.493	.711
		Brown-Forsythe	.764	.557
LPM3LMD	11	Welch	.260	.851
		Brown-Forsythe	.243	.863
LPM4LBL	10	Welch	86.248	.007
		Brown-Forsythe	1.127	.577
LPM4LMD	10	Welch	3.089	.213
		Brown-Forsythe	1.088	.469
LM1LBL	11	Welch	4.564	.100
		Brown-Forsythe	1.913	.236
LM1LMD	9	Welch	3.061	.220
		Brown-Forsythe	.746	.590
LM2LBL	11	Welch	.177	.905
		Brown-Forsythe	.239	.865
LM2LMD	12	Welch	3.273	.147
		Brown-Forsythe	1.005	.501
LPM3RMD	11	Welch	10.517	.055
		Brown-Forsythe	2.280	.403
LPM4RBL	11	Welch	3.486	.135
		Brown-Forsythe	1.886	.252
LPM4RMD	11	Welch	.043	.986
		Brown-Forsythe	.081	.967
LM3RMD	7	Welch	1.391	.397
		Brown-Forsythe	2.110	.247

Table 6.49 Robustness Index, right and left sides as separate variables, polar teeth: Sample size

Sample size (N)	Robustness Index															
	I1L	CL	PM3L	M1L	I1R	CR	PM3R	M1R	LI1L	LCL	LPM3L	LM1L	LI1R	LCR	LPM3R	LM1R
AILIAS	1	4	4	5	0	4	4	3	0	1	1	1	1	1	1	1
MYRTOS PYRGOS	5	9	11	12	6	12	17	10	2	10	15	10	0	6	12	14
GYPSADES	5	5	7	9	3	5	10	10	1	11	9	10	3	6	9	8
SELLOPOULO	2	3	3	6	2	3	5	5	3	8	5	7	3	4	3	8
PALAMA	6	8	8	9	3	6	10	8	5	6	4	6	6	7	8	5
LERNA	13	23	19	18	13	15	17	18	7	18	22	9	7	18	24	14
APATHEIA	5	6	13	13	1	5	9	12	6	10	11	10	4	11	8	13

Table 6.50 Crown Index, right and left sides pooled, polar teeth: Sample size

Sample size (N)	Crown Index							
	UI1	UC	UPM3	UM1	LI1	LC	LPM3	LM1
AILIAS	1	4	6	5	1	1	1	1
MYRTOS PYRGOS	7	15	21	16	2	14	22	18
GYPSADES	7	7	13	14	3	15	15	13
SELLOPOULO	2	5	6	7	3	9	6	10
LERNA	15	24	23	24	12	26	30	18
APATHEIA	5	10	18	20	7	16	15	18

Table 6.51 Crown Index, right and left sides as separate variables, polar teeth: Sample size

Sample size (N)	Crown Index															
	I1L	CL	PM3L	M1L	I1R	CR	PM3R	M1R	LI1L	LCL	LPM3L	LM1L	LI1R	LCR	LPM3R	LM1R
AILIAS	1	4	4	5	0	4	4	3	0	1	1	1	1	1	1	1
MYRTOS PYRGOS	5	9	11	12	6	12	17	10	2	10	14	10	0	6	12	14
GYPSADES	5	5	6	9	3	5	10	10	1	11	9	10	3	6	9	8
SELLOPOULO	2	3	3	6	2	3	5	5	3	8	5	7	3	4	3	8
LERNA	13	23	19	18	13	15	17	18	7	18	22	9	7	18	25	14
APATHEIA	5	6	13	13	1	5	9	12	6	10	11	10	4	11	8	13

Table 6.52 Robustness Index, right and left sides pooled, polar teeth: Sample size

Sample size (N)	Robustness Index							
	UI1	UC	UPM3	UM1	LI1	LC	LPM3	LM1
AILIAS	1	4	6	5	1	1	1	1
MYRTOS PYRGOS	7	15	21	16	2	14	22	18
GYPSADES	7	7	13	14	3	15	15	13
SELLOPOULO	2	5	6	7	3	9	6	10
LERNA	16	24	23	24	12	26	30	18
APATHEIA	5	10	18	20	7	16	15	18

D.2 BOX AND WHISKER PLOTS OF TOOTH CROWN MEASUREMENTS

Maxillary teeth, Right and Left sides separately

Figure 1 Box and whisker plot of Bucco-Lingual (BL) crown diameter of 1st Incisor, Upper, Left side, Cretan and Argolid samples.

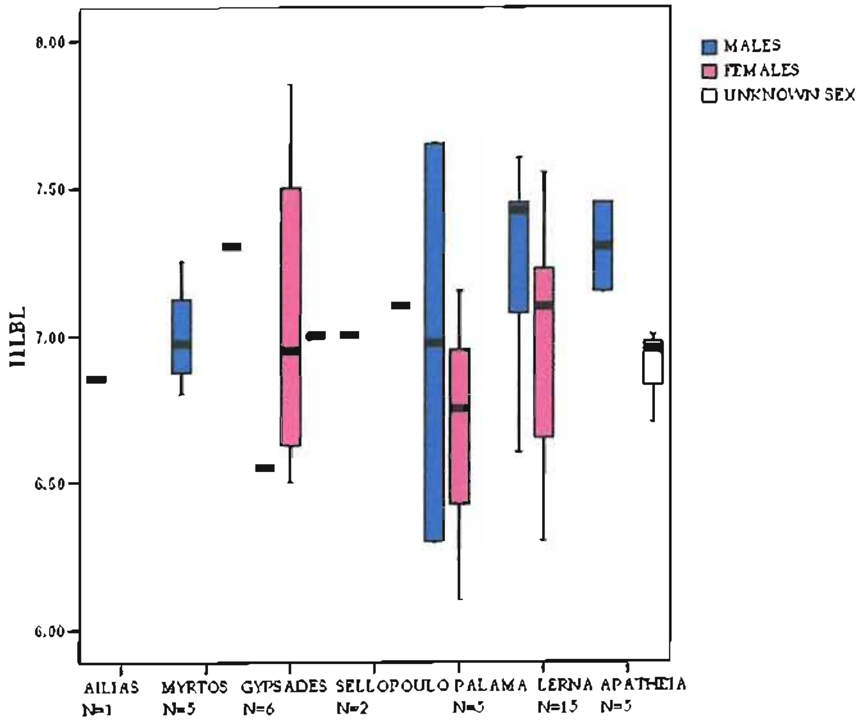


Figure 2 Box and whisker plot of Mesio-Distal (MD) crown diameter of 1st Incisor, Upper, Left side, Cretan and Argolid samples.

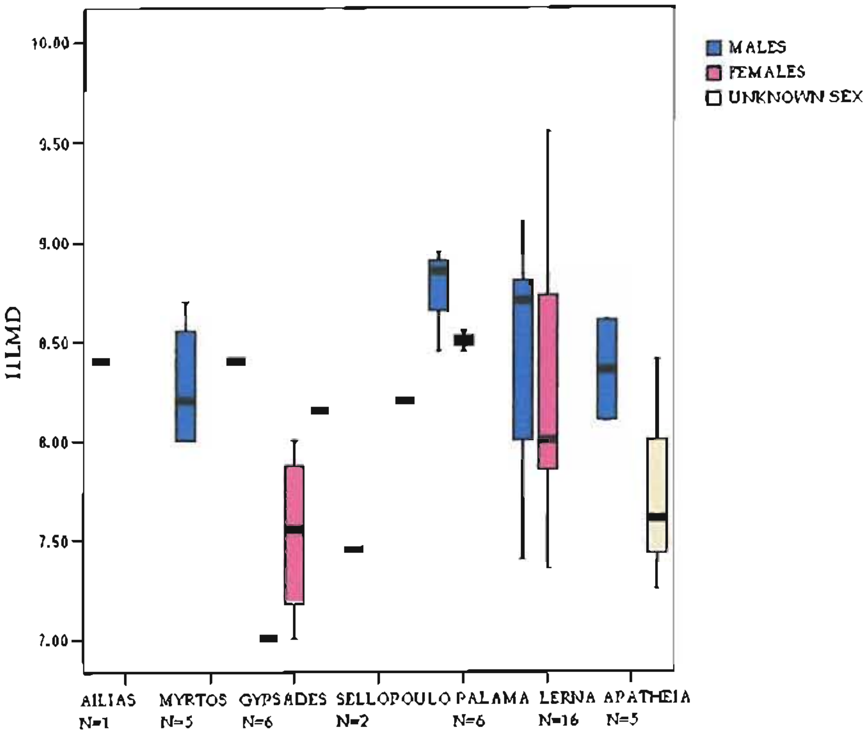


Figure 3 Box and whisker plot of BL crown diameter of 2nd Incisor, Upper, Left side, Cretan and Argolid samples.

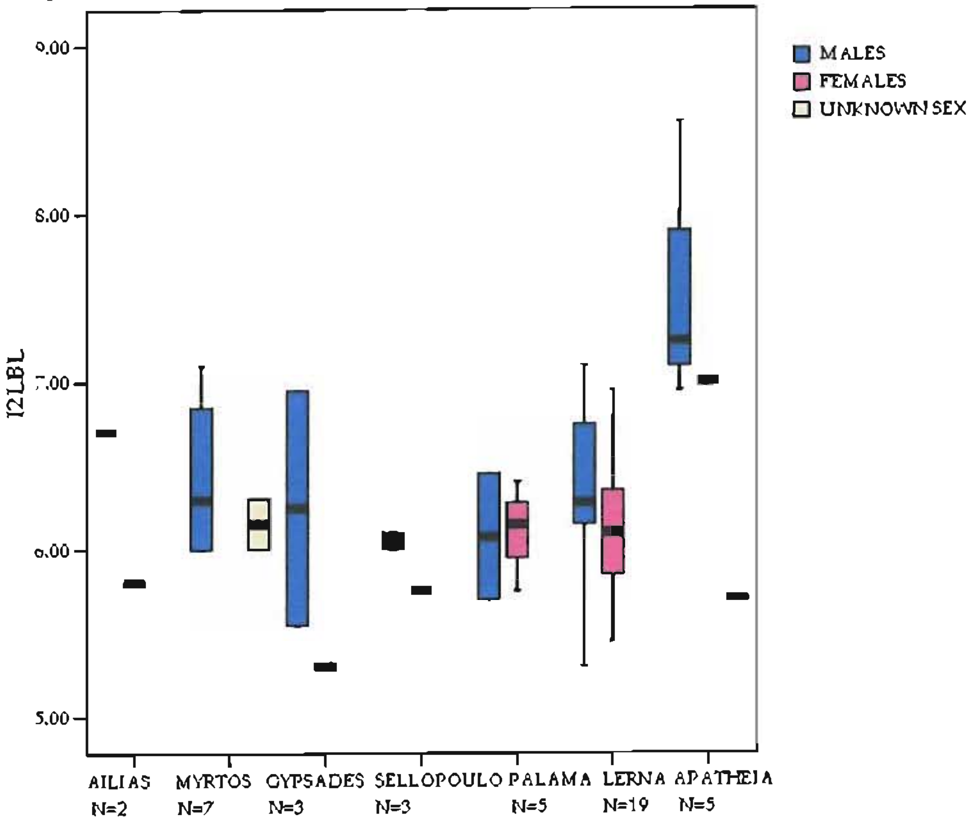


Figure 4 Box and whisker plot of MD crown diameter of 2nd Incisor, Upper, Left side, Cretan and Argolid samples.

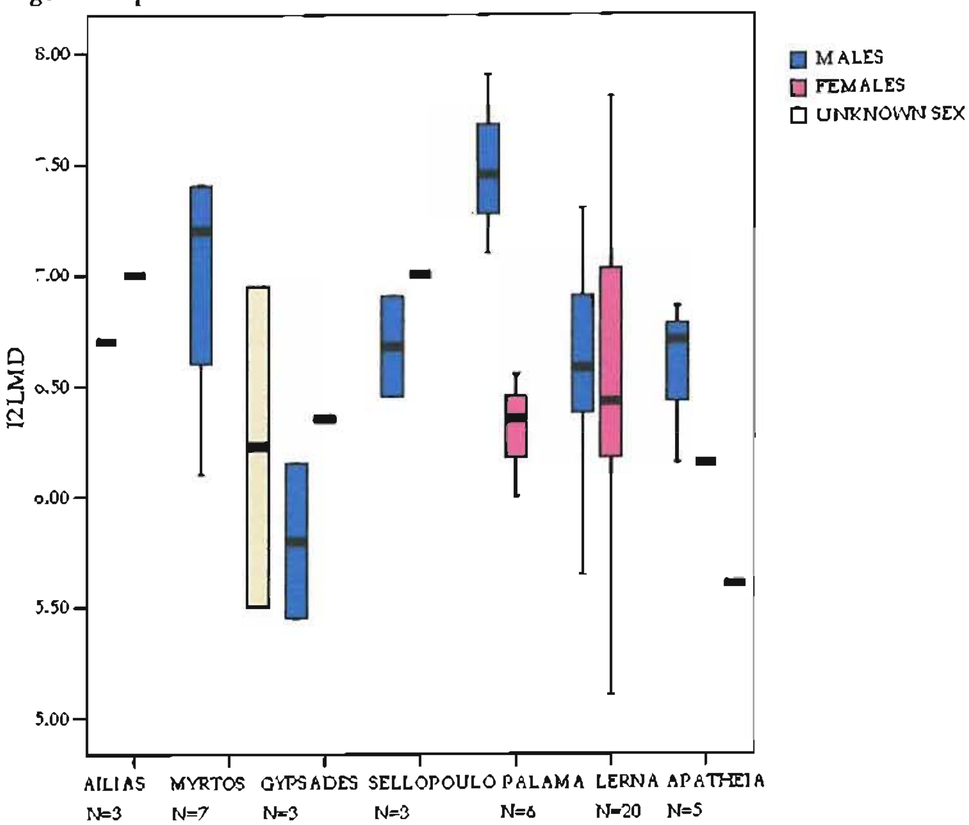


Figure 5 Box and whisker plot of BL crown diameter of Canine, Upper, Left side, Cretan and Argolid samples.

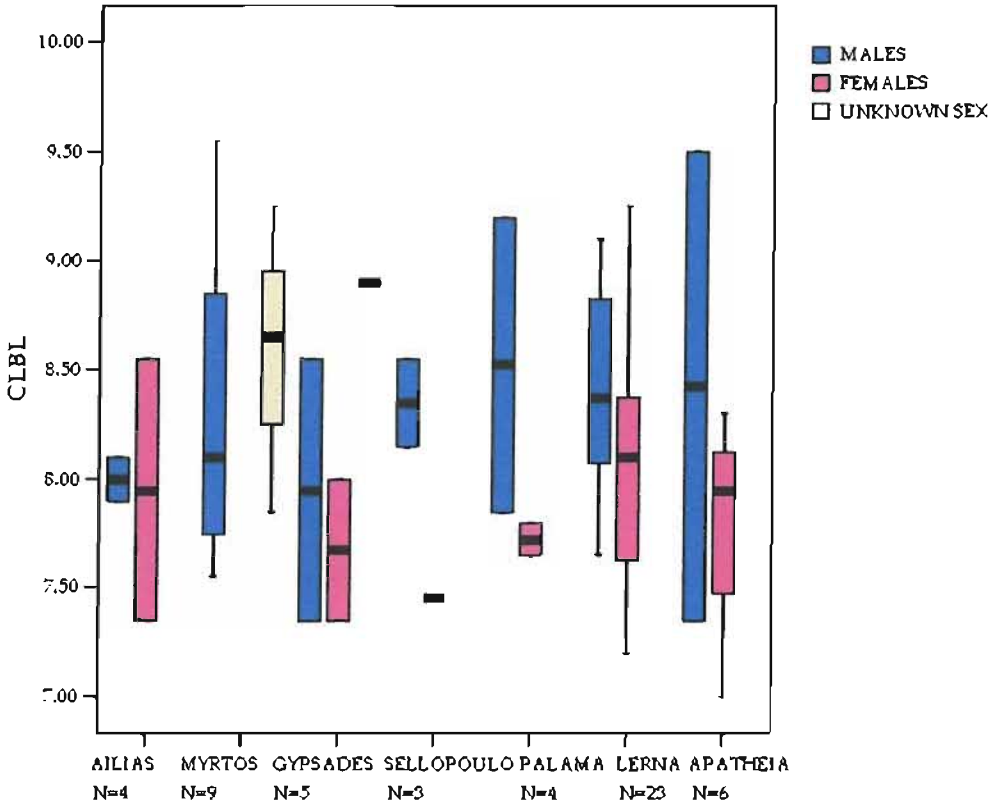


Figure 6 Box and whisker plot of MD crown diameter of Canine, Upper, Left side, Cretan and Argolid samples.

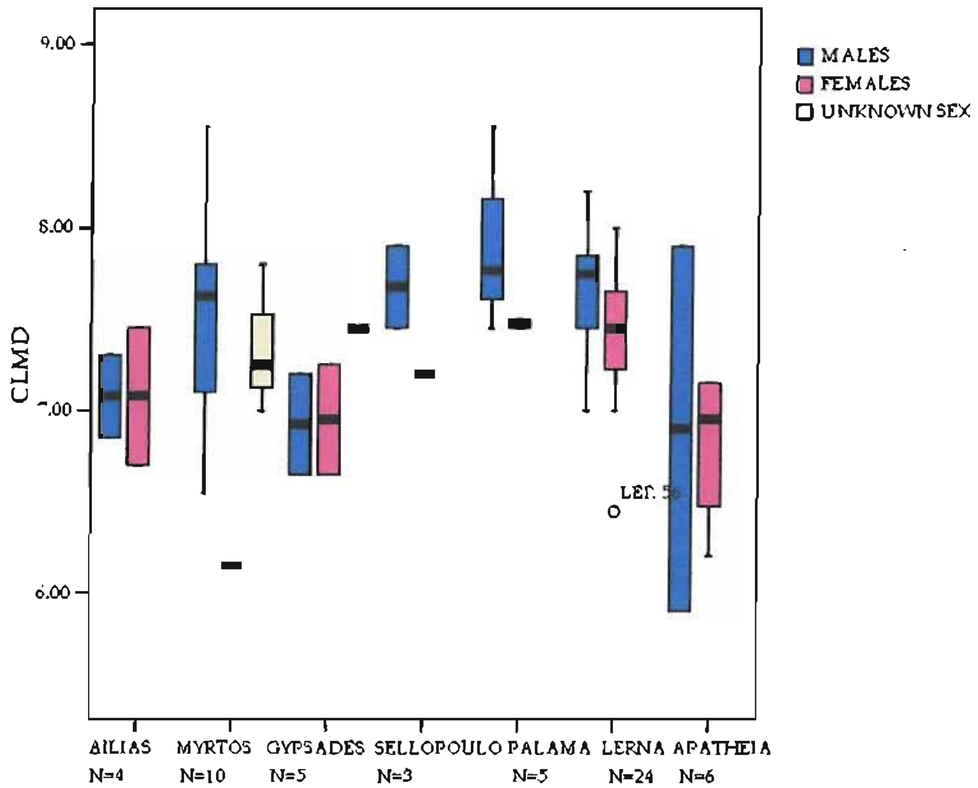


Figure 7 Box and whisker plot of BL crown diameter of 3rd Premolar, Upper, Left side, Cretan and Argolid samples.

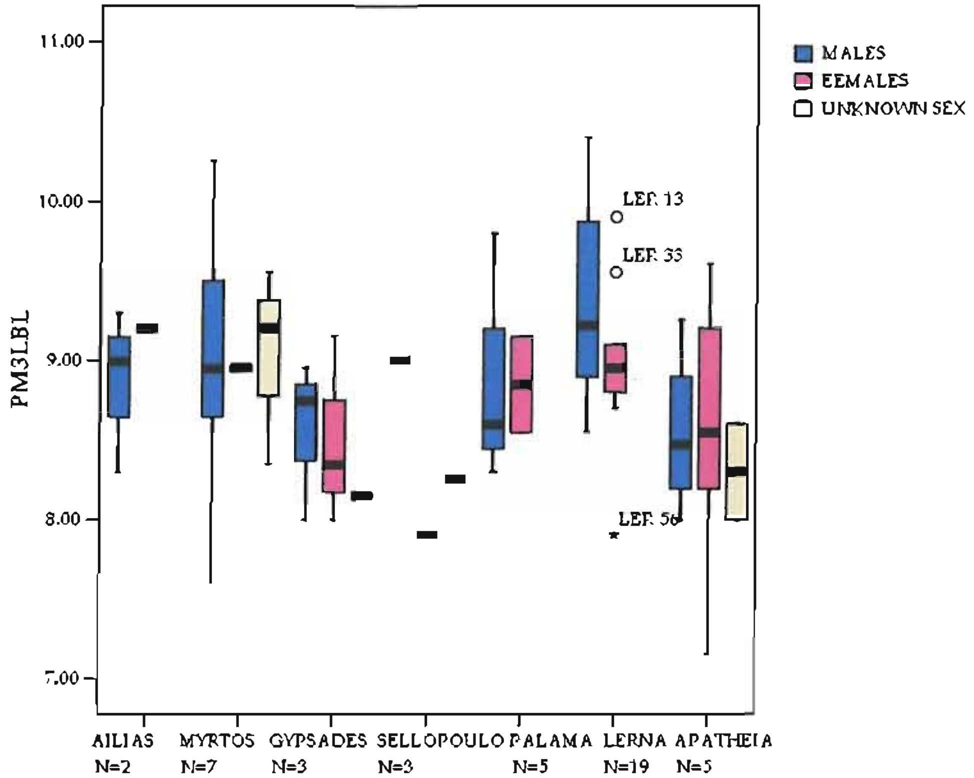


Figure 8 Box and whisker plot of MD crown diameter of 3rd Premolar, Upper, Left side, Cretan and Argolid samples.

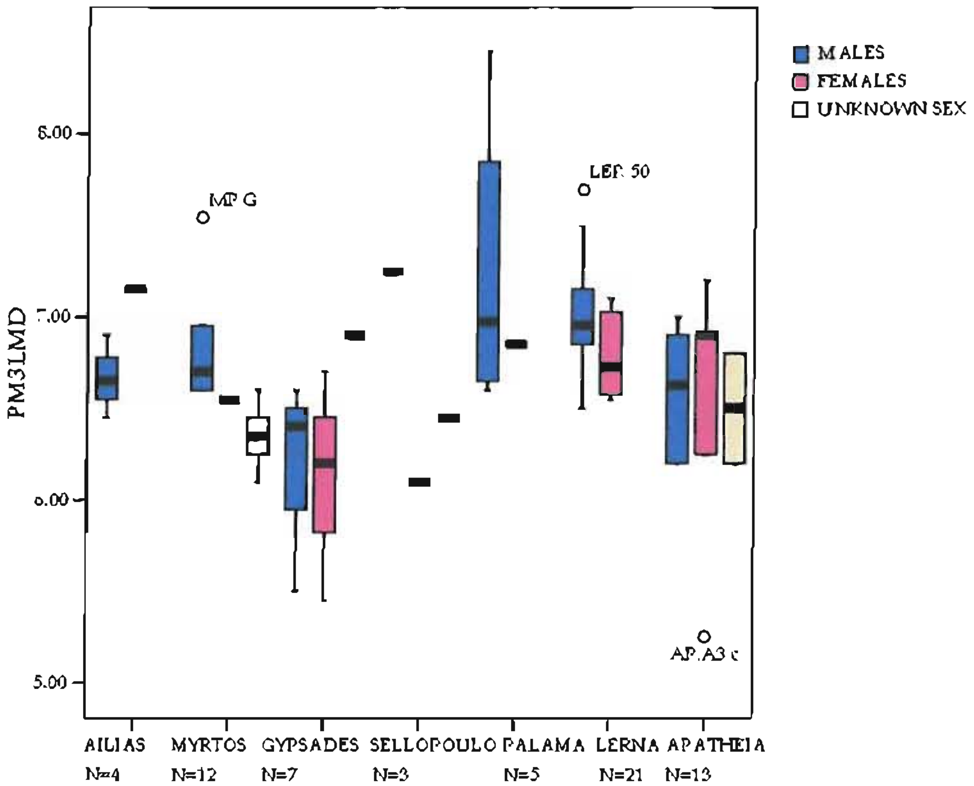


Figure 9 Box and whisker plot of BL crown diameter of 4th Premolar, Upper, Left side, Cretan and Argolid samples.

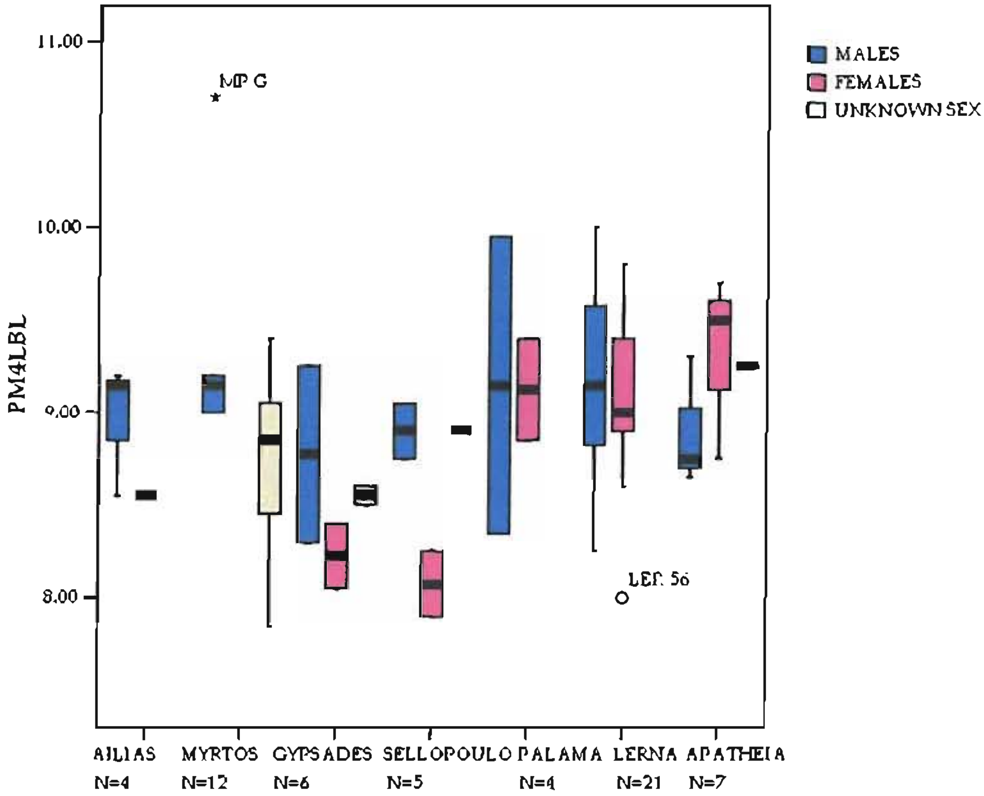


Figure 10 Box and whisker plot of MD crown diameter of 4th Premolar, Upper, Left side, Cretan and Argolid samples.

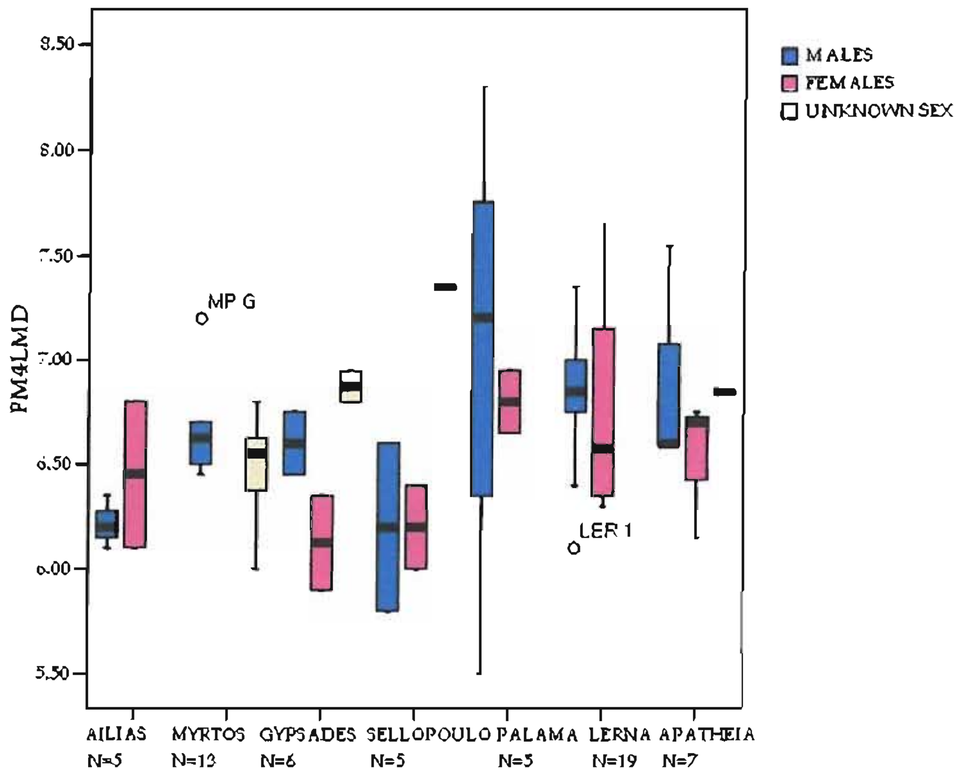


Figure 11 Box and whisker plot of BL crown diameter of 1st Molar, Upper, Left side, Cretan and Argolid samples.

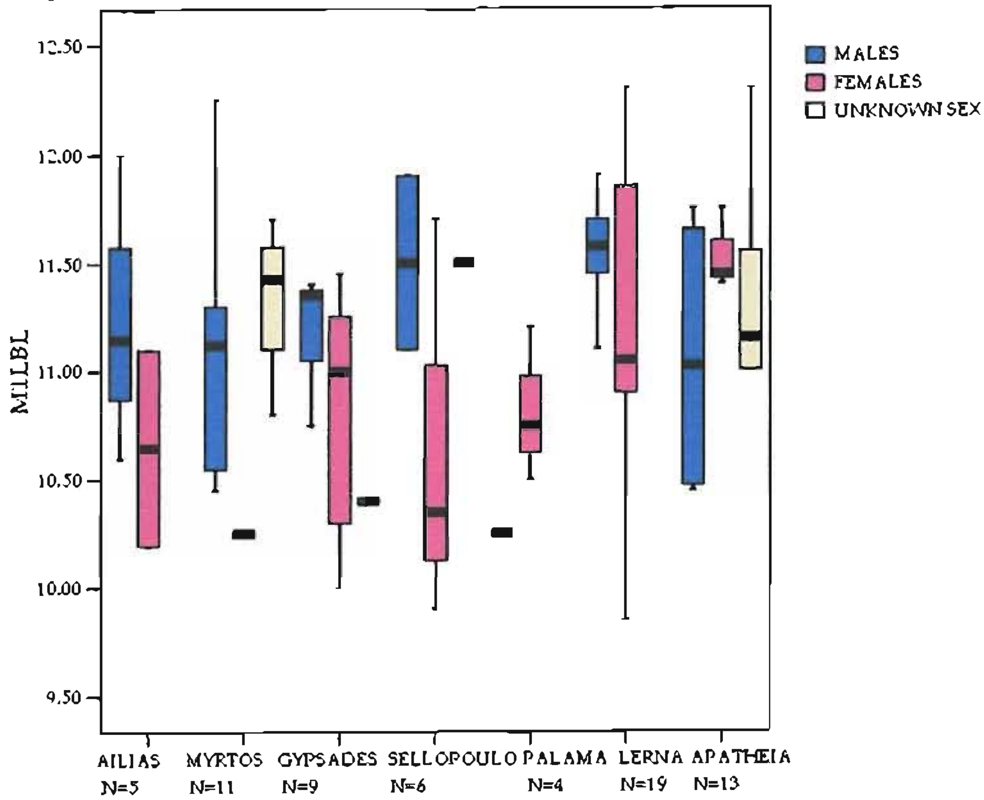


Figure 12 Box and whisker plot of MD crown diameter of 1st Molar, Upper, Left side, Cretan and Argolid samples.

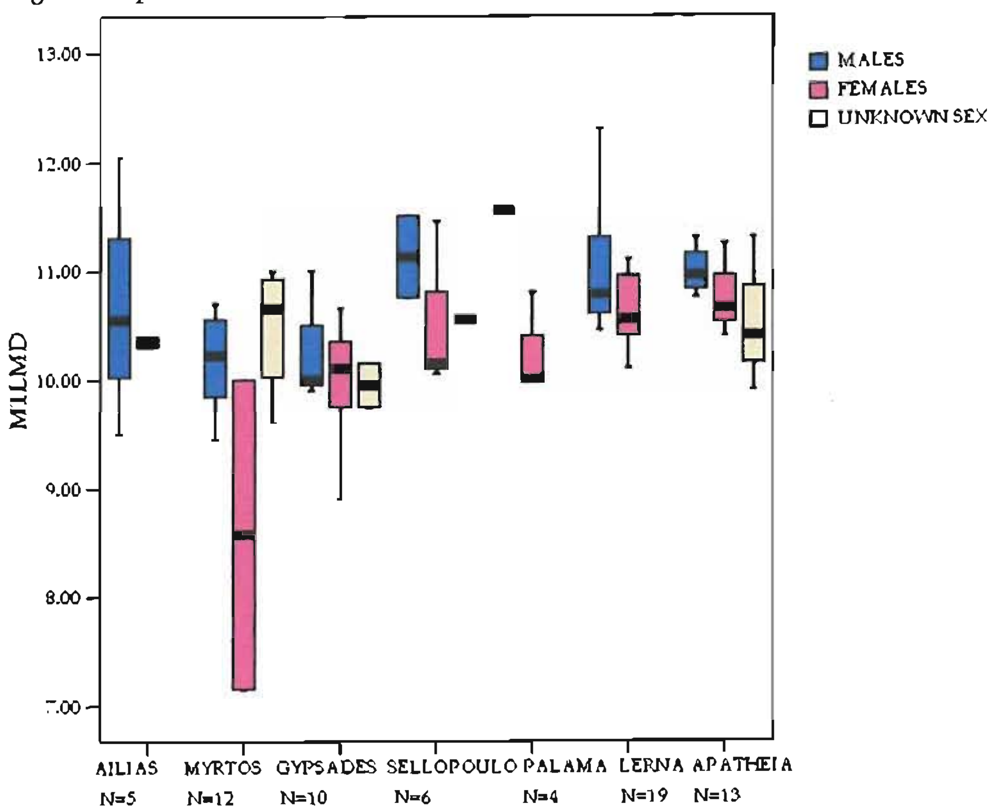


Figure 13 Box and whisker plot of BL crown diameter of 2nd Molar, Upper, Left side, Cretan and Argolid samples.

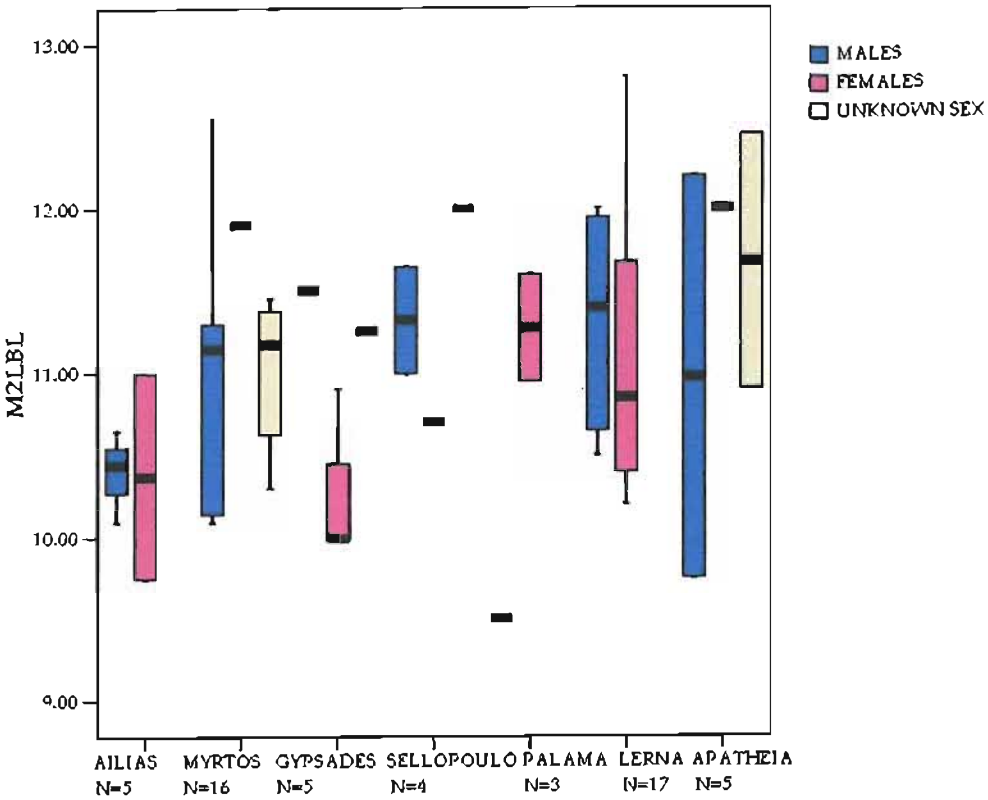


Figure 14. Box and whisker plot of MD crown diameter of 2nd Molar, left side, Cretan and Argolid samples.

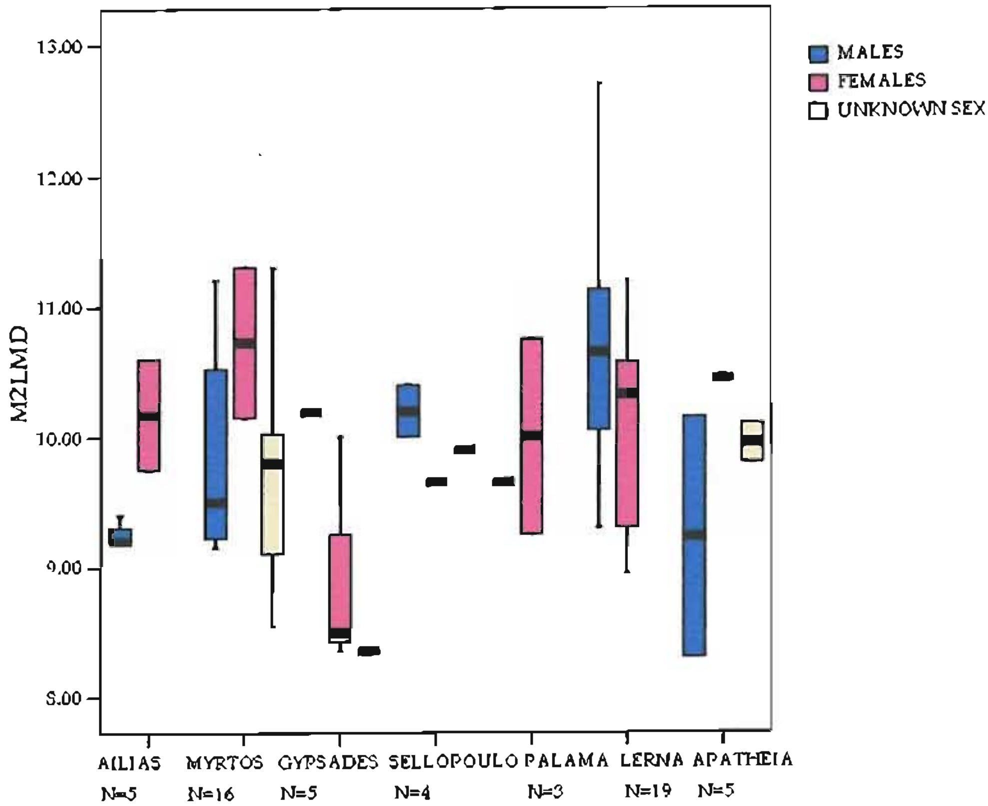


Figure 15 Box and whisker plot of BL crown diameter of 3rd Molar, Upper, Left side, Cretan and Argolid samples.

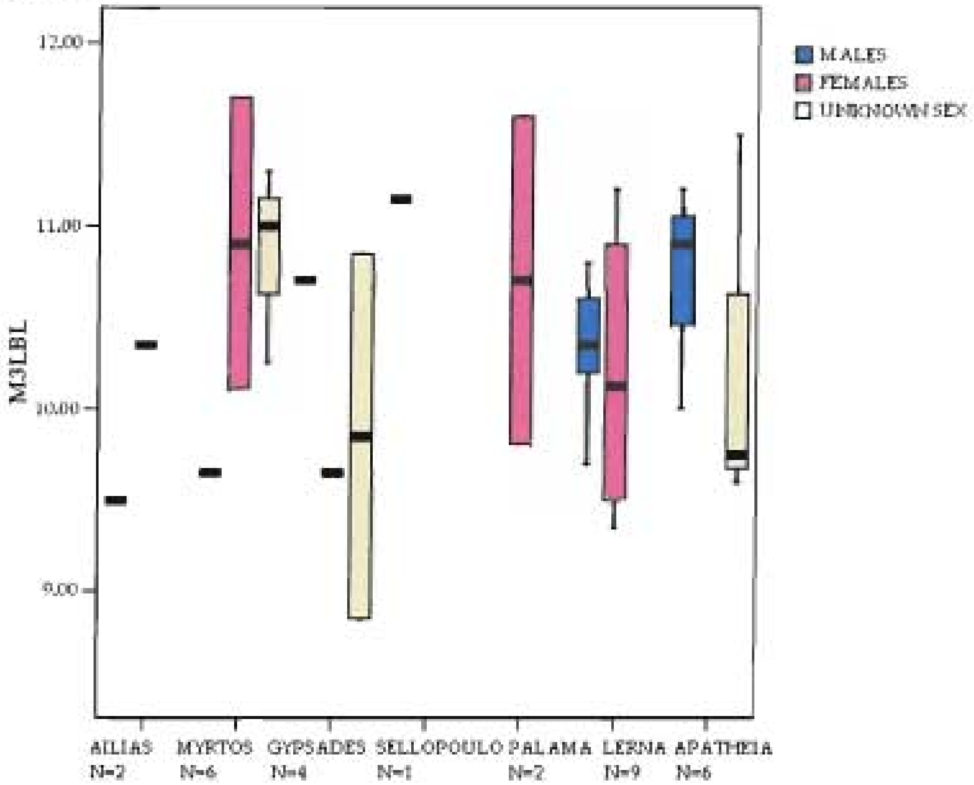


Figure 16 Box and whisker plot of MD crown diameter of 3rd Molar, Upper, Left side, Cretan and Argolid samples.

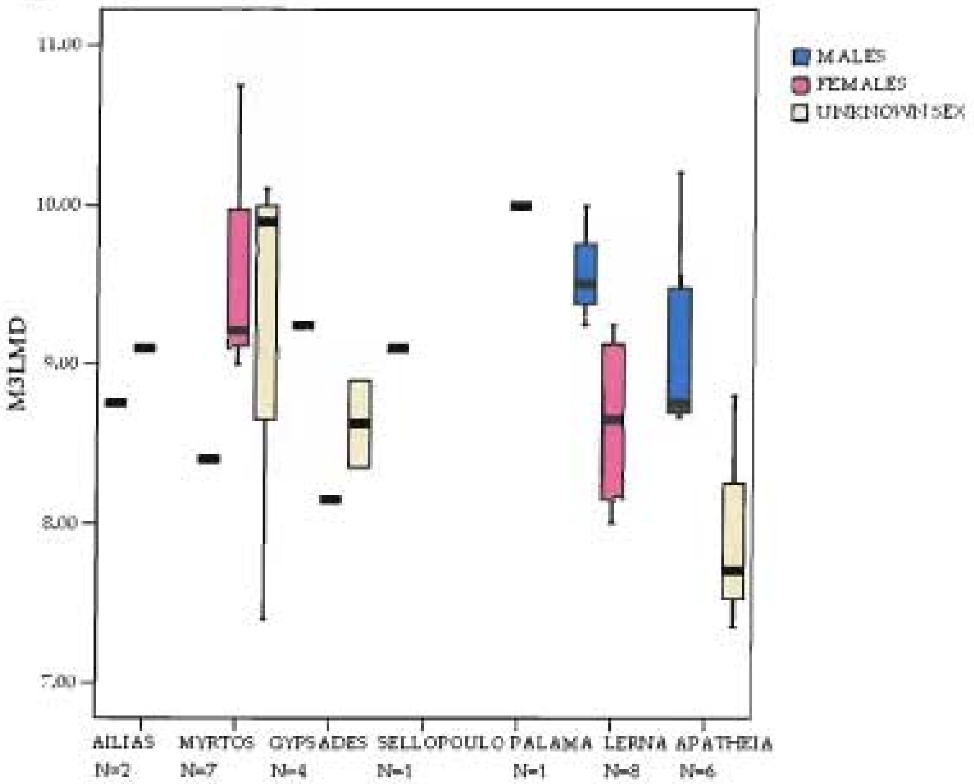


Figure 17 Box and whisker plot of BL crown diameter of 1st Incisor, Upper, Right side, Cretan and Argolid samples.

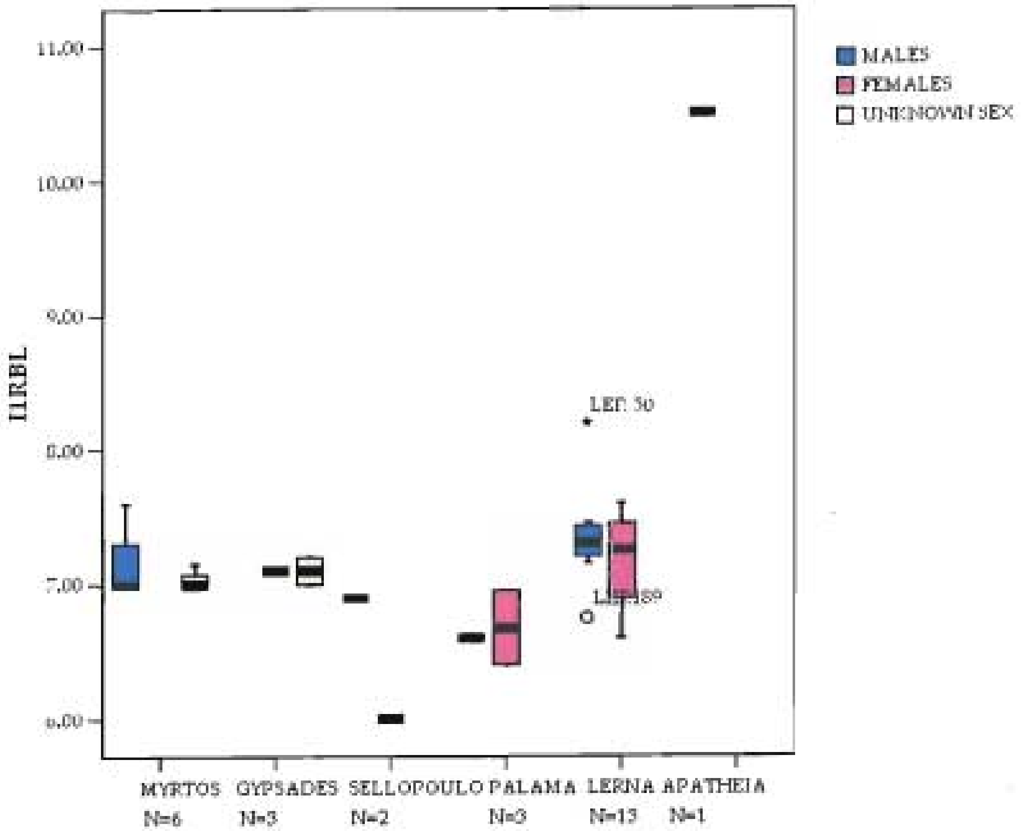


Figure 18 Box and whisker plot of MD crown diameter of 1st Incisor, right side, Cretan and Argolid samples.

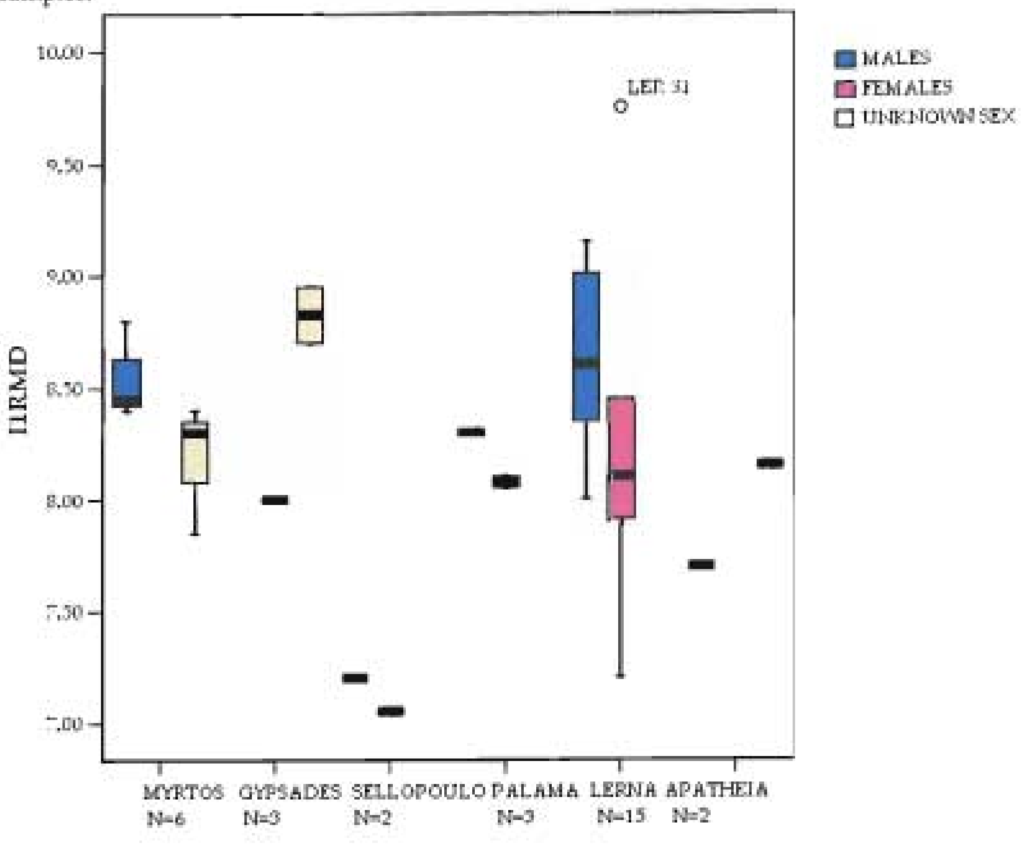


Figure 19 Box and whisker plot of BL crown diameter of 2nd Incisor, right side, Cretan and Argolid samples.

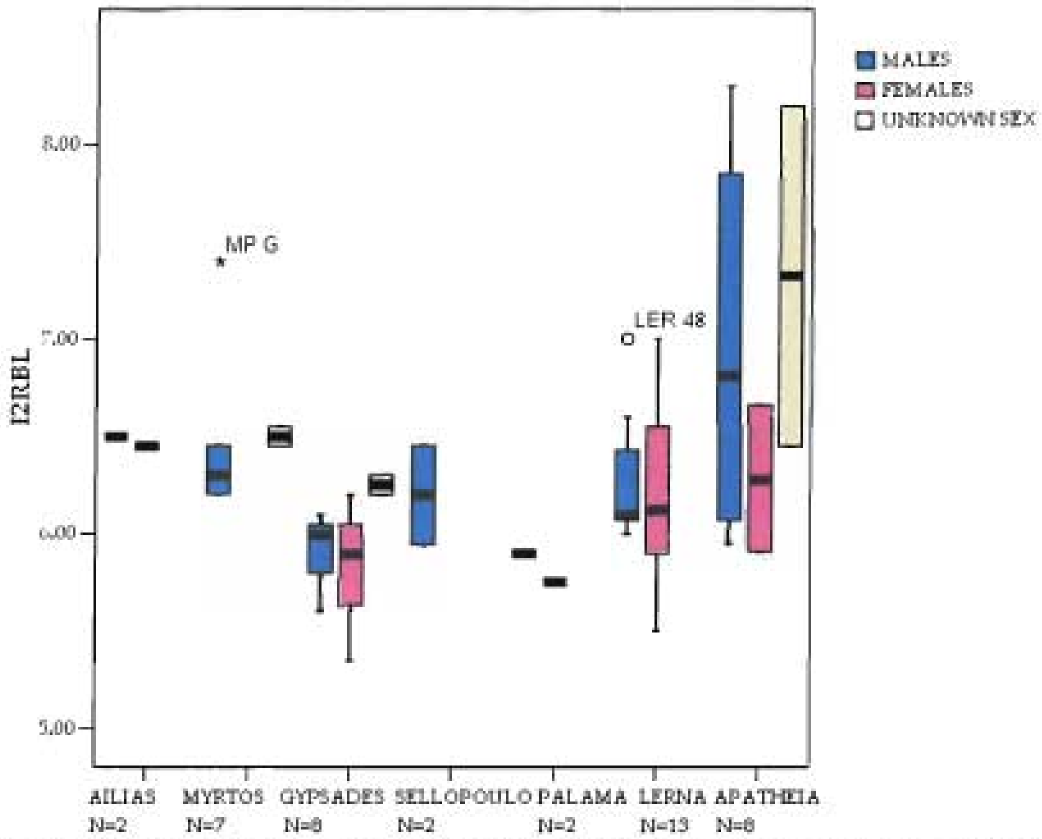


Figure 20 Box and whisker plot of MD crown diameter of 2nd Incisor, Upper, Right side, Cretan and Argolid samples.

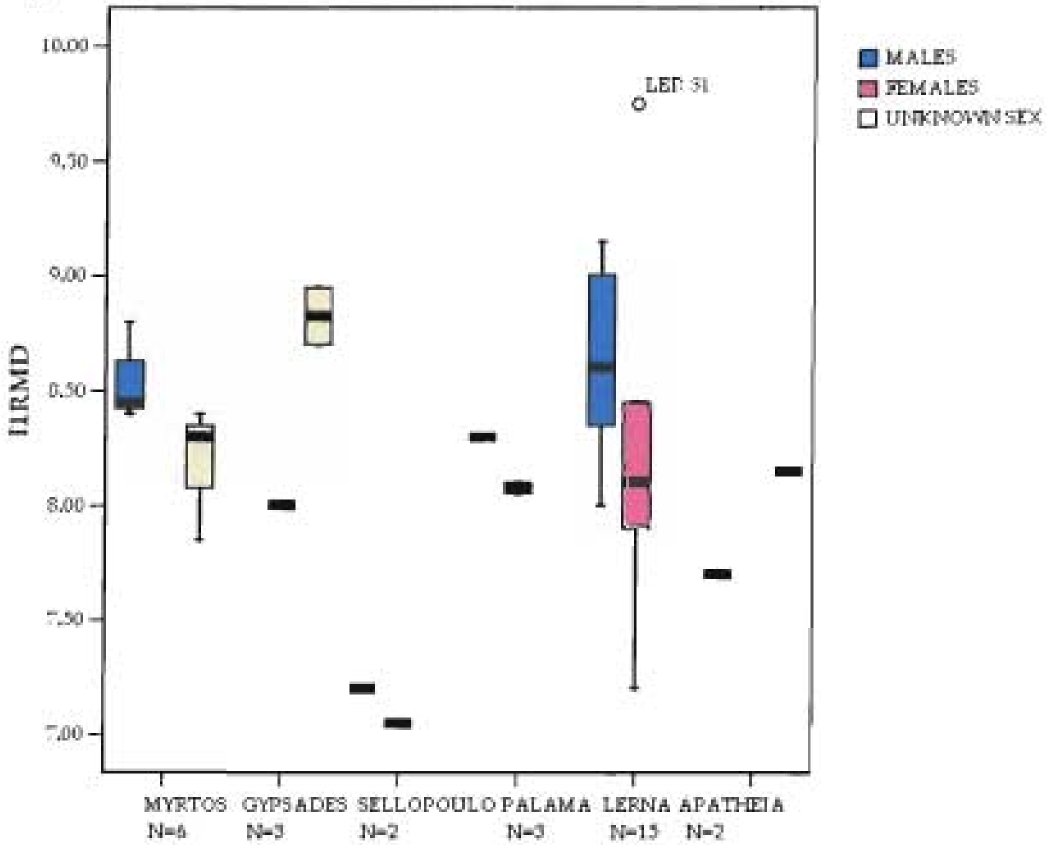


Figure 21 Box and whisker plot of BL crown diameter of Canine, right side, Cretan and Argolid samples.

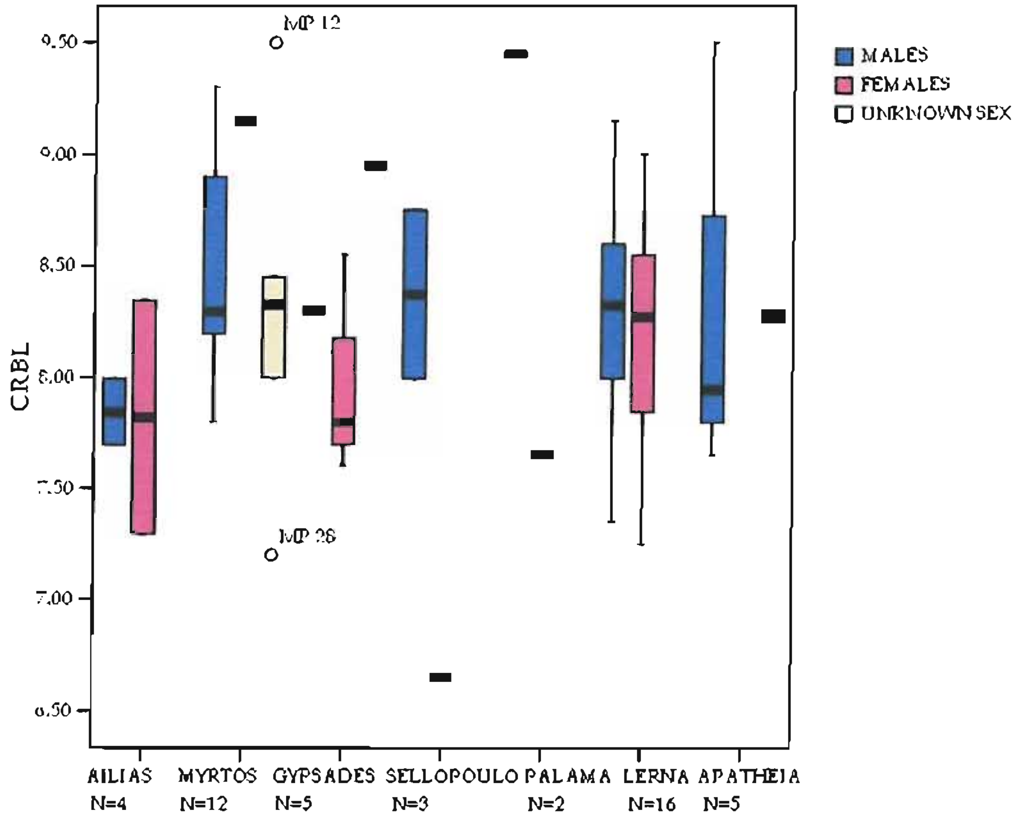


Figure 22 Box and whisker plot of MD crown diameter of Canine, Upper, Right side, Cretan and Argolid samples.

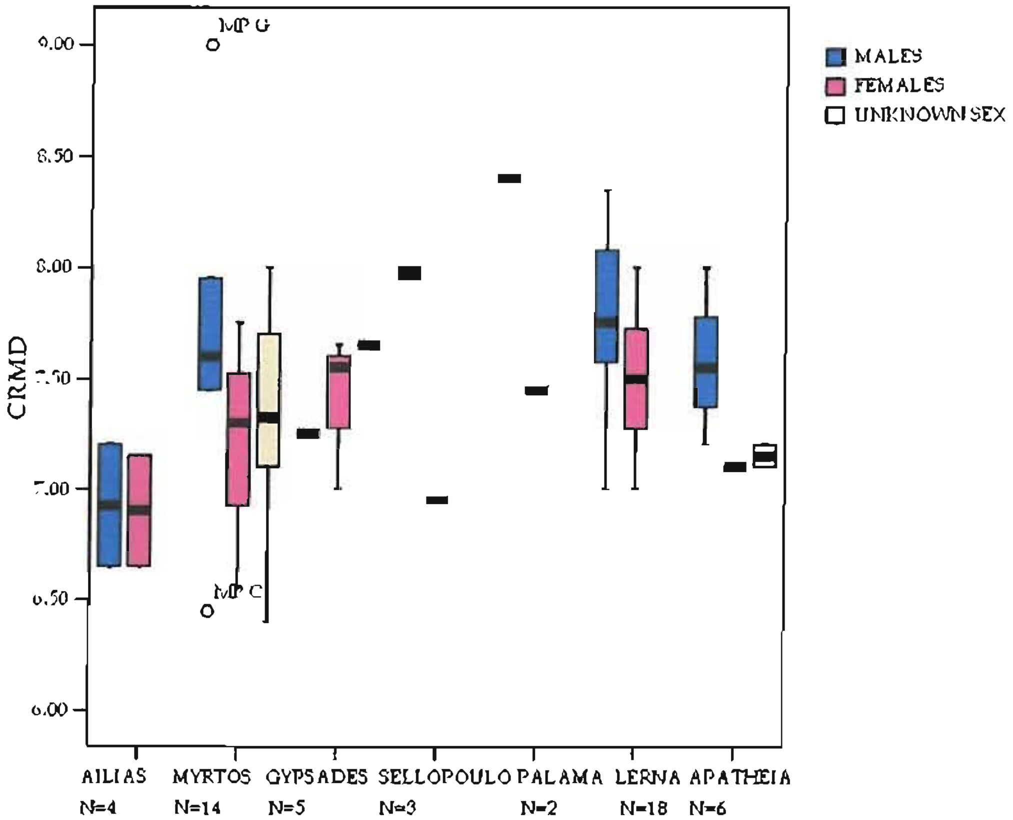


Figure 23 Box and whisker plot of BL crown diameter of 3rd Premolar, right side, Cretan and Argolid samples.

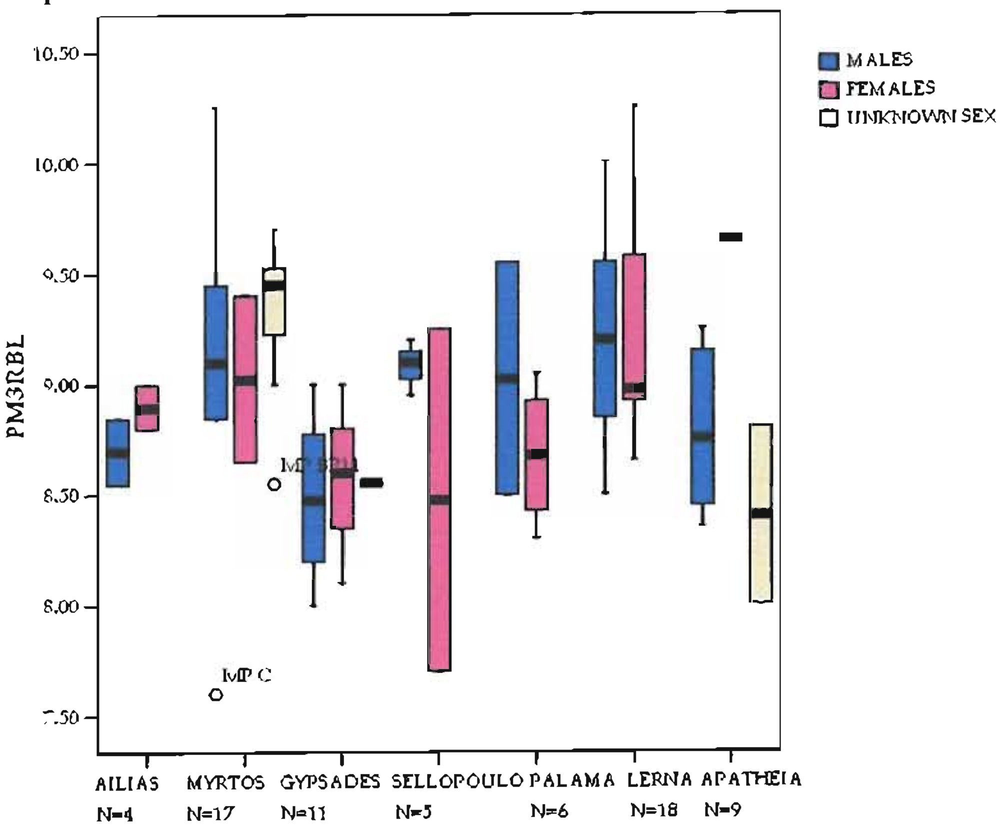


Figure 24 Box and whisker plot of MD crown diameter of 3rd Premolar, Upper, Right side, Cretan and Argolid samples.

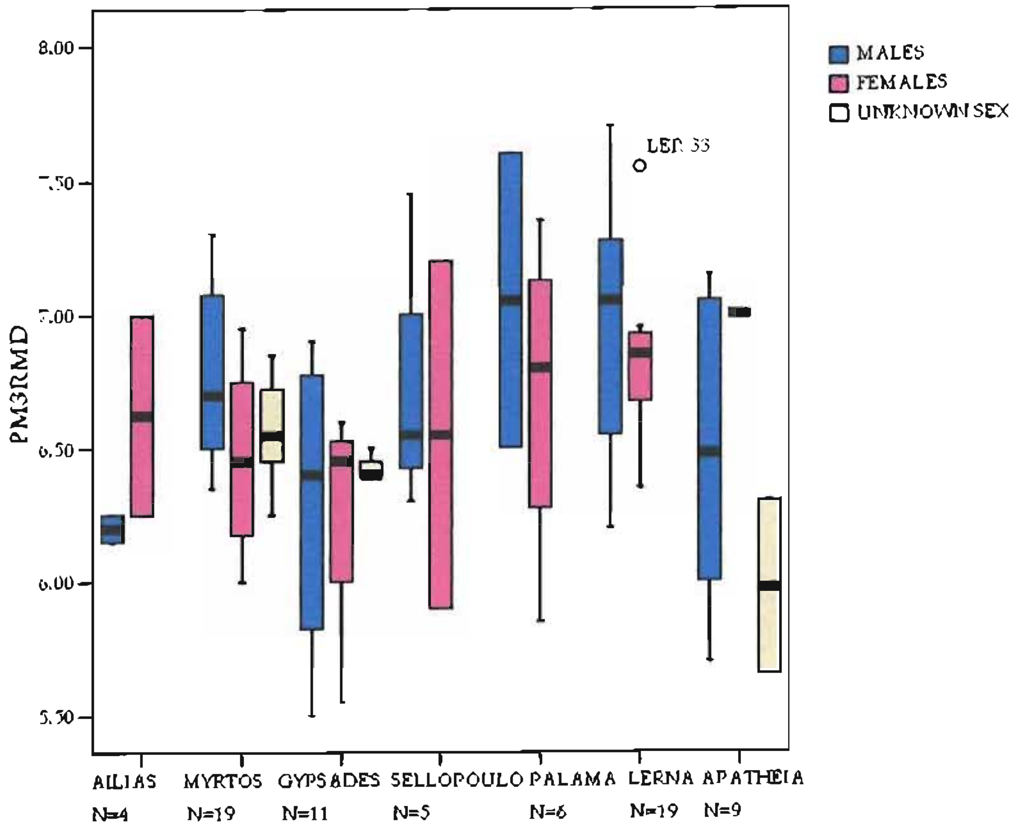


Figure 25 Box and whisker plot of BL crown diameter of 4th Premolar, Upper, Right side, Cretan and Argolid samples.

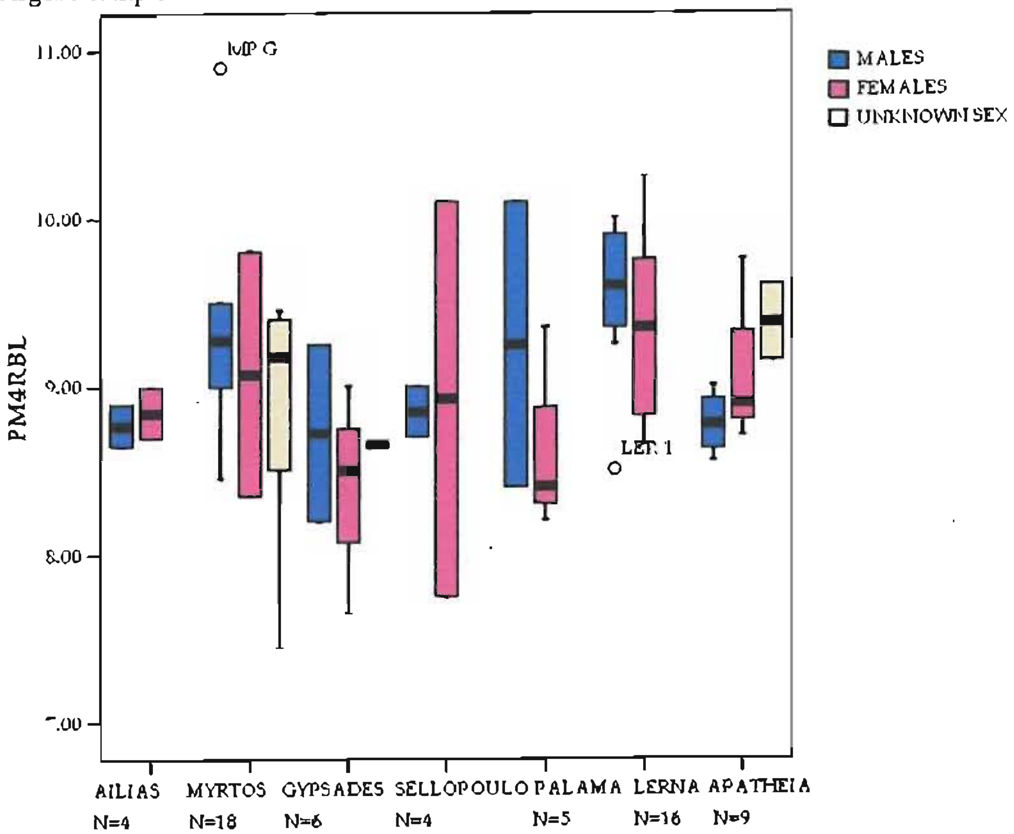


Figure 26 Box and whisker plot of MD crown diameter of 4th Premolar, Upper, Right side, Cretan and Argolid samples.

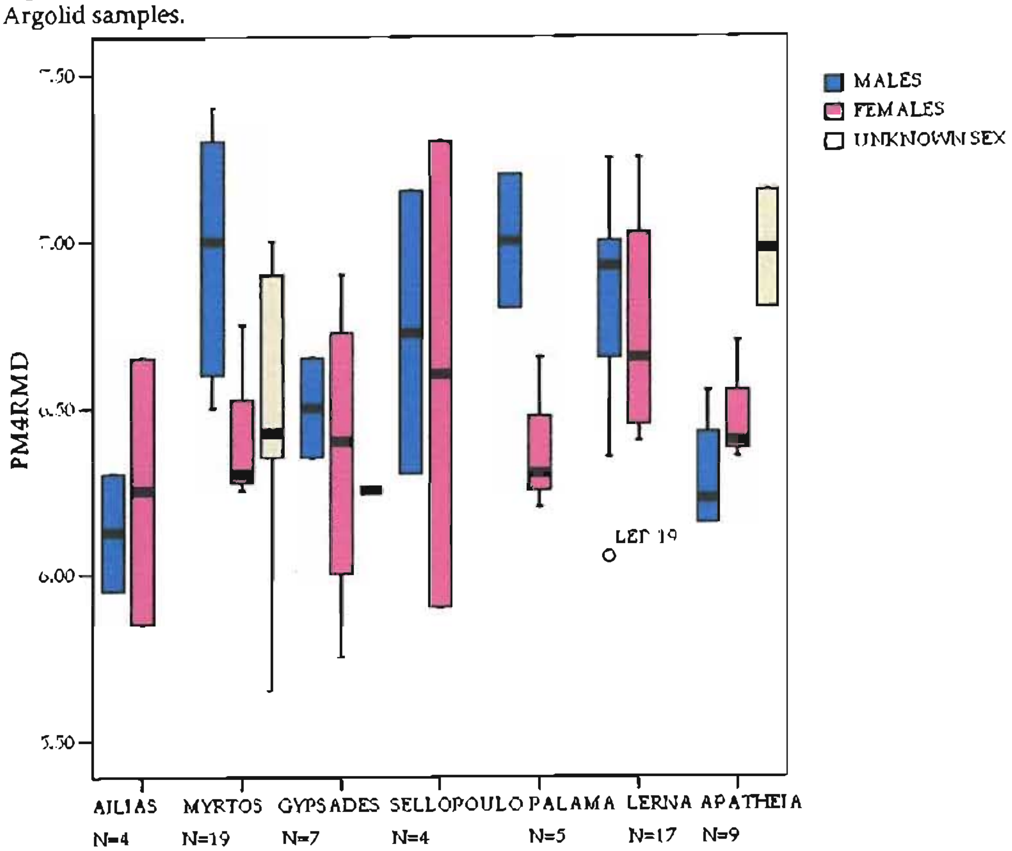


Figure 27 Box and whisker plot of BL crown diameter of 1st Molar, Upper, Right side, Cretan and Argolid samples.

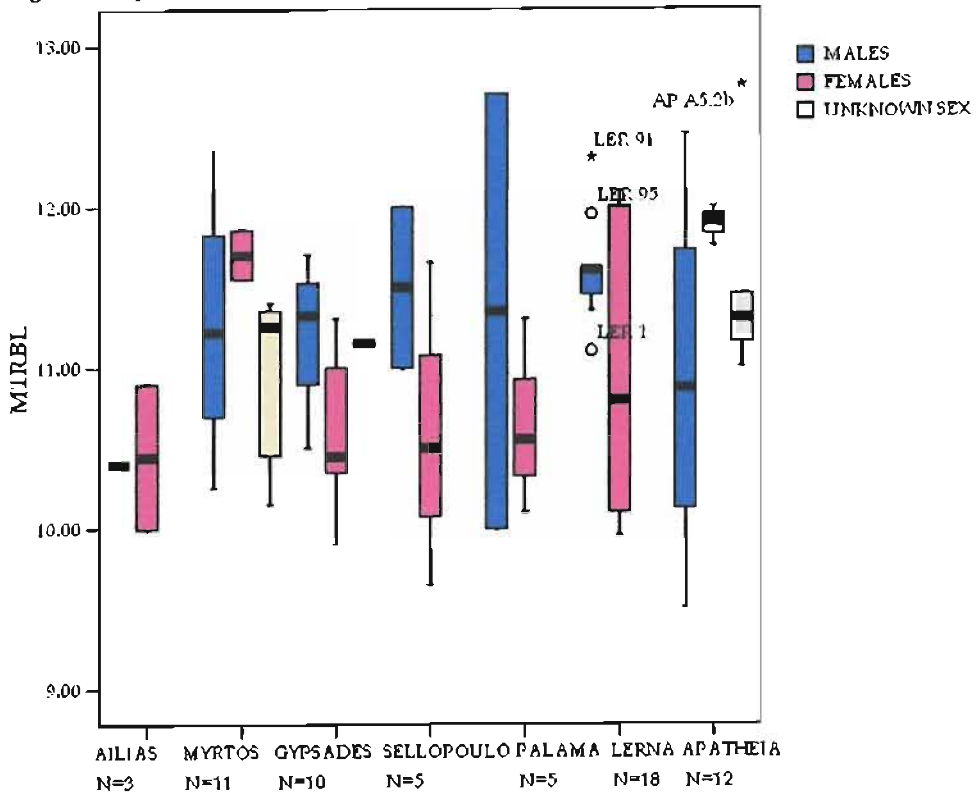


Figure 28 Box and whisker plot of MD crown diameter of 1st Molar, Upper, Right side, Cretan and Argolid samples.

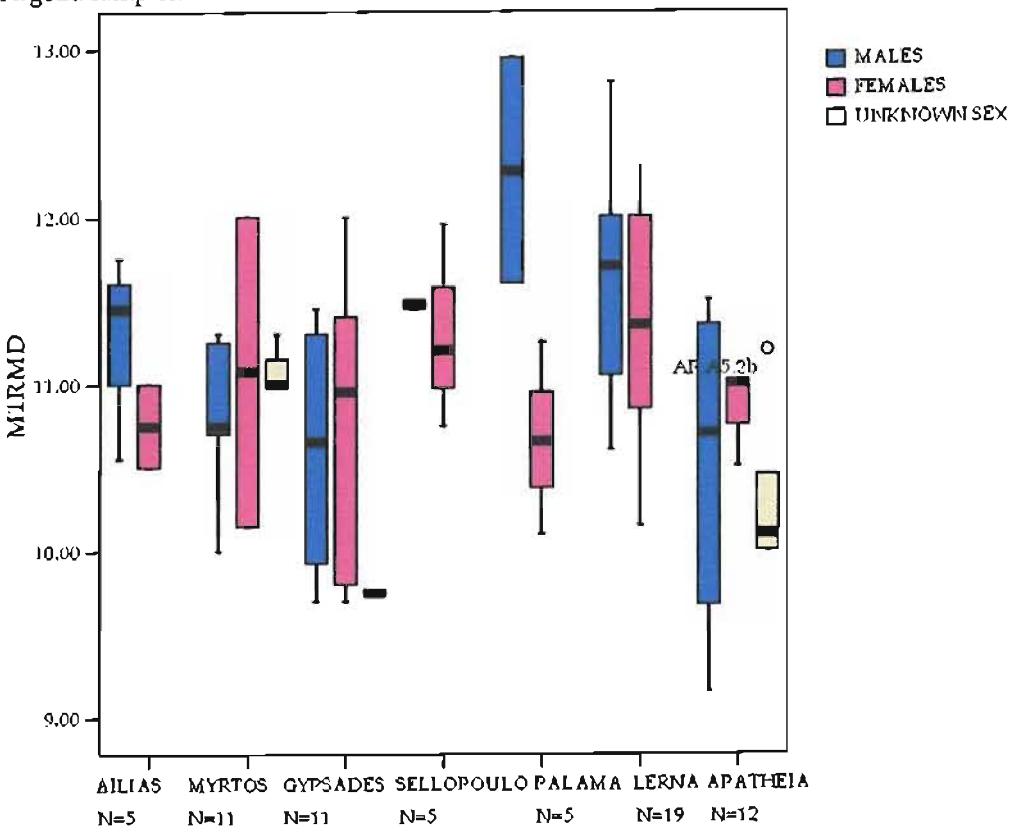


Figure 29 Box and whisker plot of BL crown diameter of 2nd Molar, Upper, Right side, Cretan and Argolid samples.

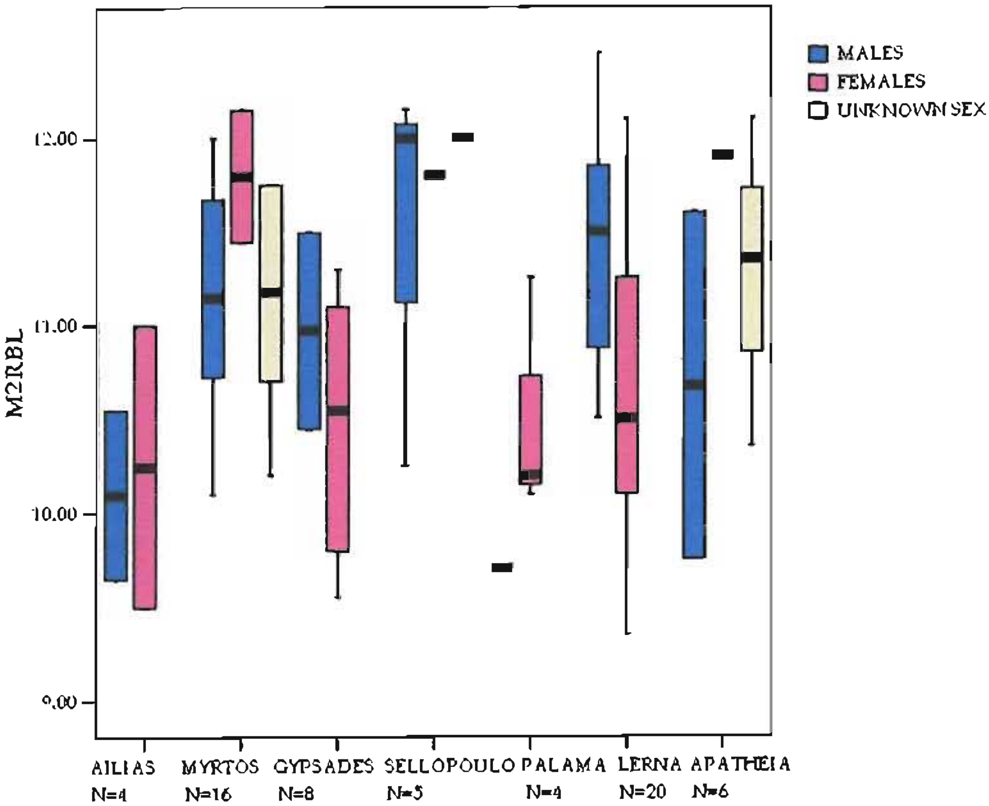


Figure 30 Box and whisker plot of MD crown diameter of 2nd Molar, Upper, Right side, Cretan and Argolid samples.

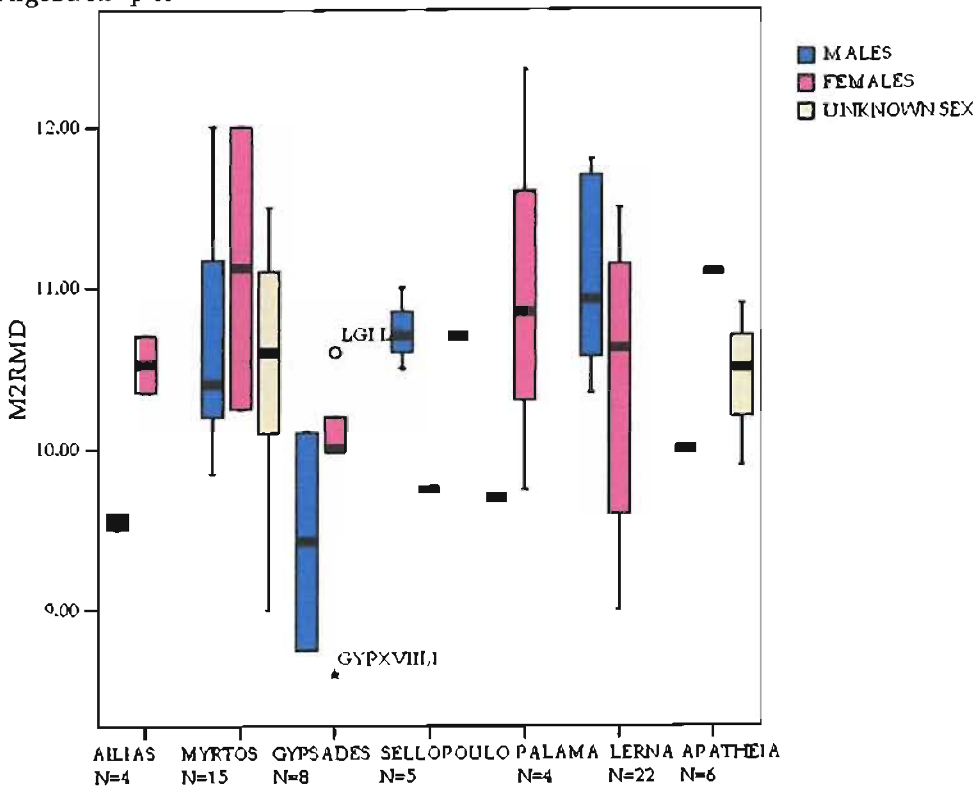


Figure 31 Box and whisker plot of BL crown diameter of 3rd Molar, Upper, Right side, Cretan and Argolid samples.

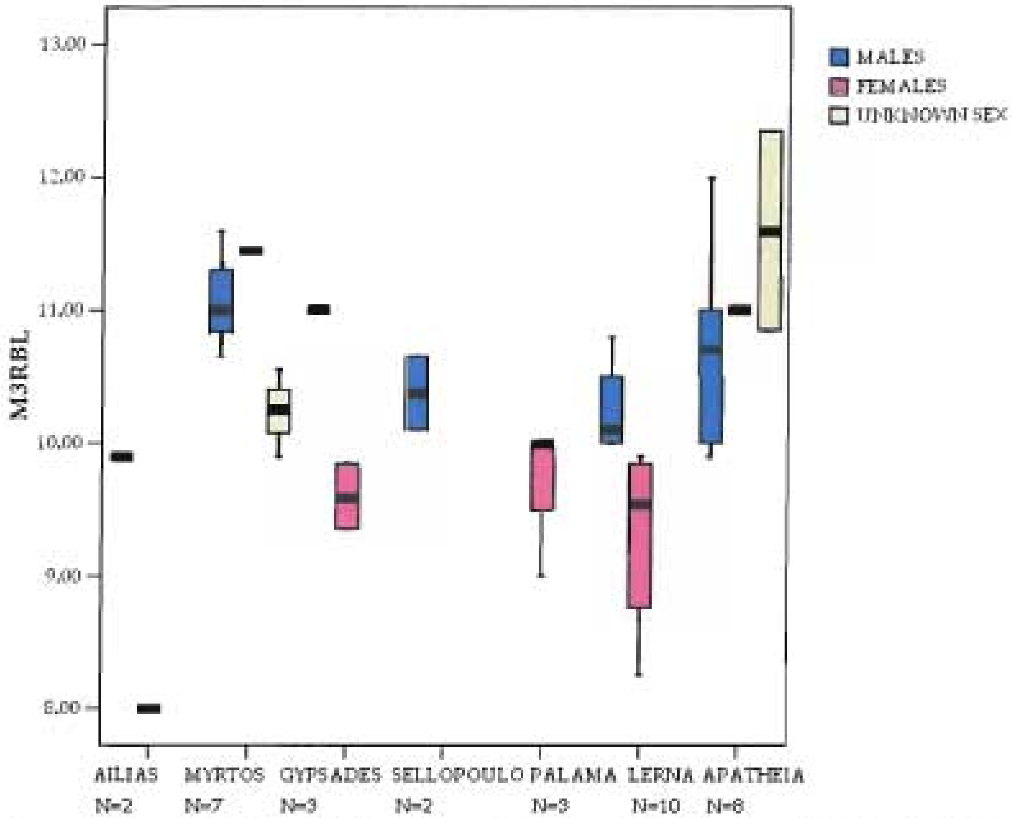
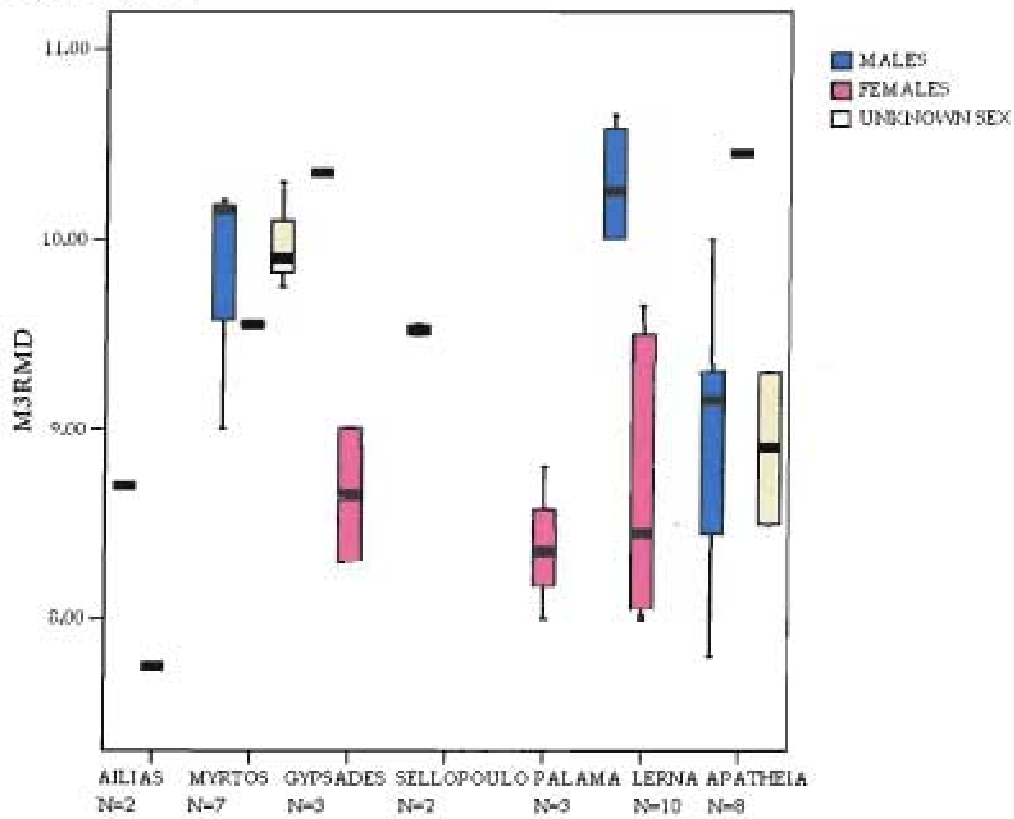


Figure 32 Box and whisker plot of MD crown diameter of 3rd Molar, Upper, Right side, Cretan and Argolid samples.



Mandibular teeth, Right and Left sides separately

Figure 33 Box and whisker plot of BL crown diameter of 1st Incisor, Lower, Left side, Cretan and Argolid samples.

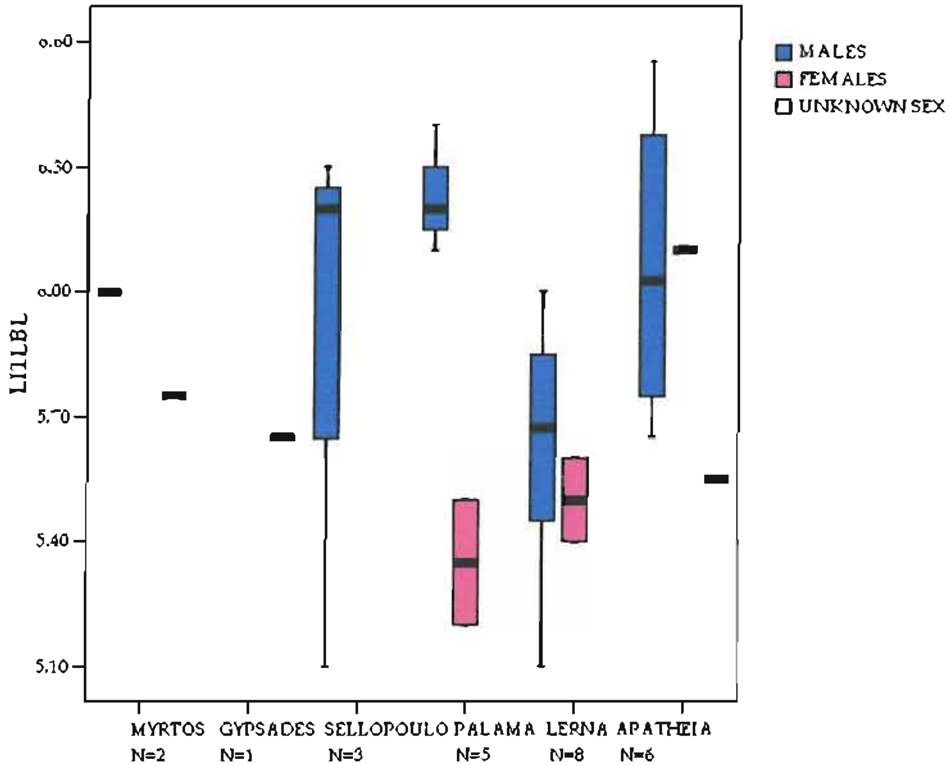


Figure 34 Box and whisker plot of MD crown diameter of 1st Incisor, Lower, Left side, Cretan and Argolid samples.

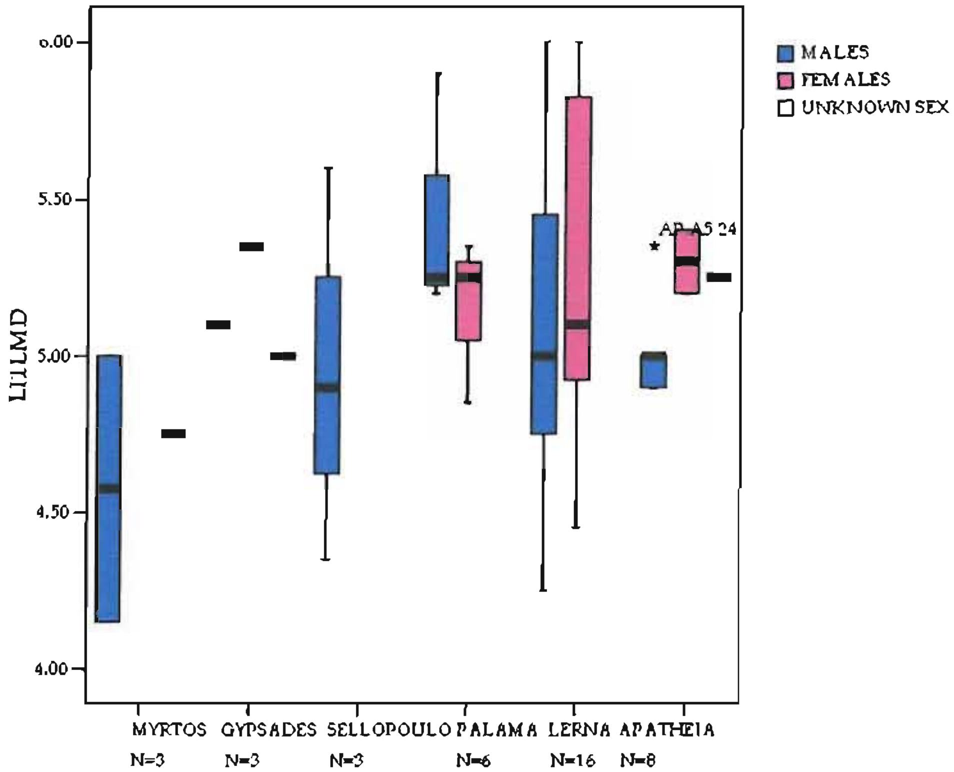


Figure 35 Box and whisker plot of BL crown diameter of 2nd Incisor, Lower, Left side, Cretan and Argolid samples.

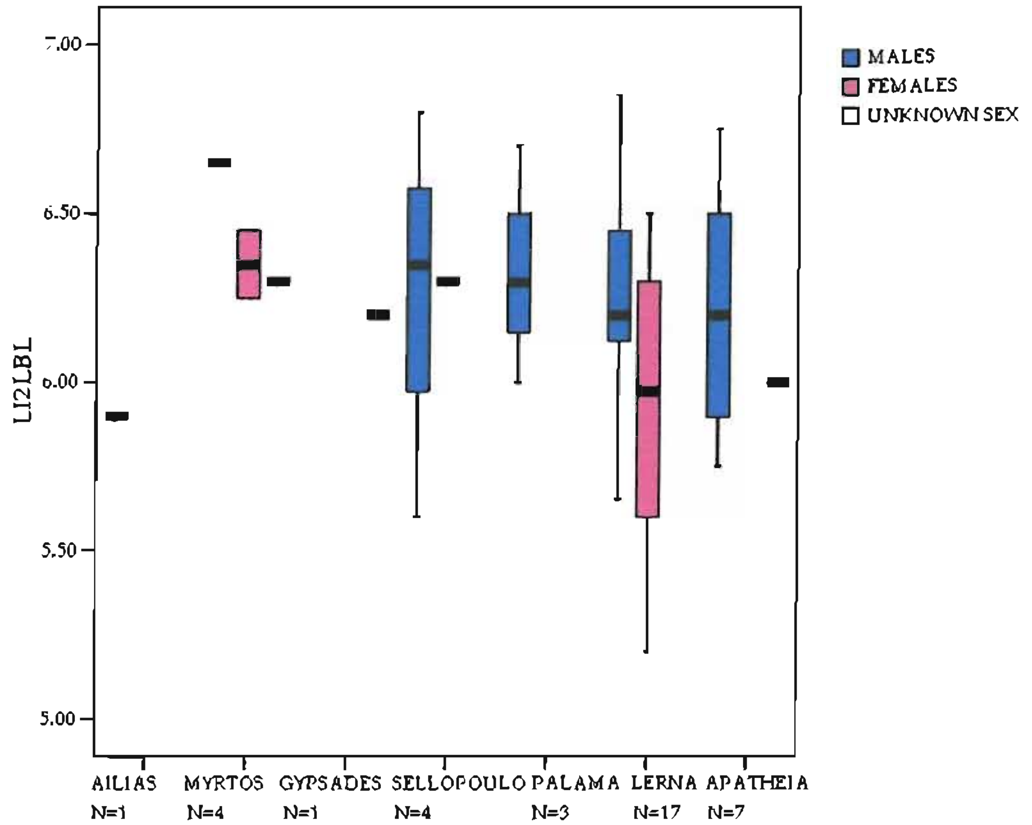


Figure 36 Box and whisker plot of MD crown diameter of 2nd Incisor, Lower, Left side, Cretan and Argolid samples.

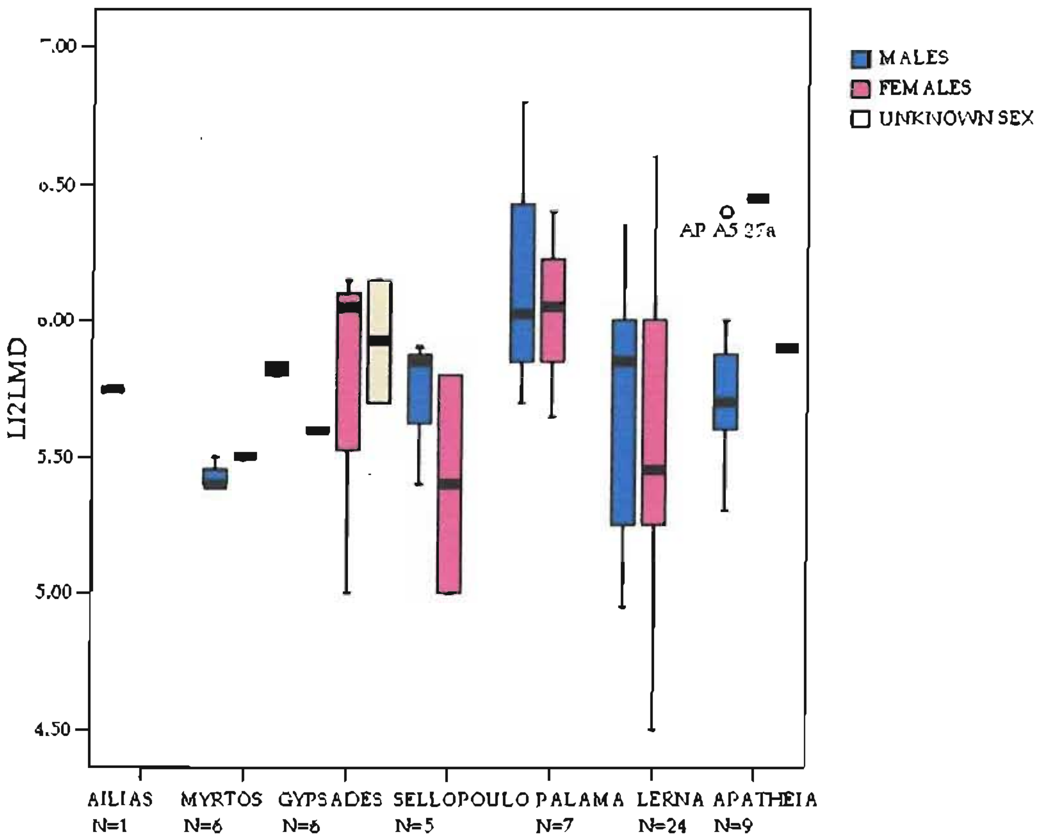


Figure 37 Box and whisker plot of BL crown diameter of Canine, Lower, Left side, Cretan and Argolid samples.

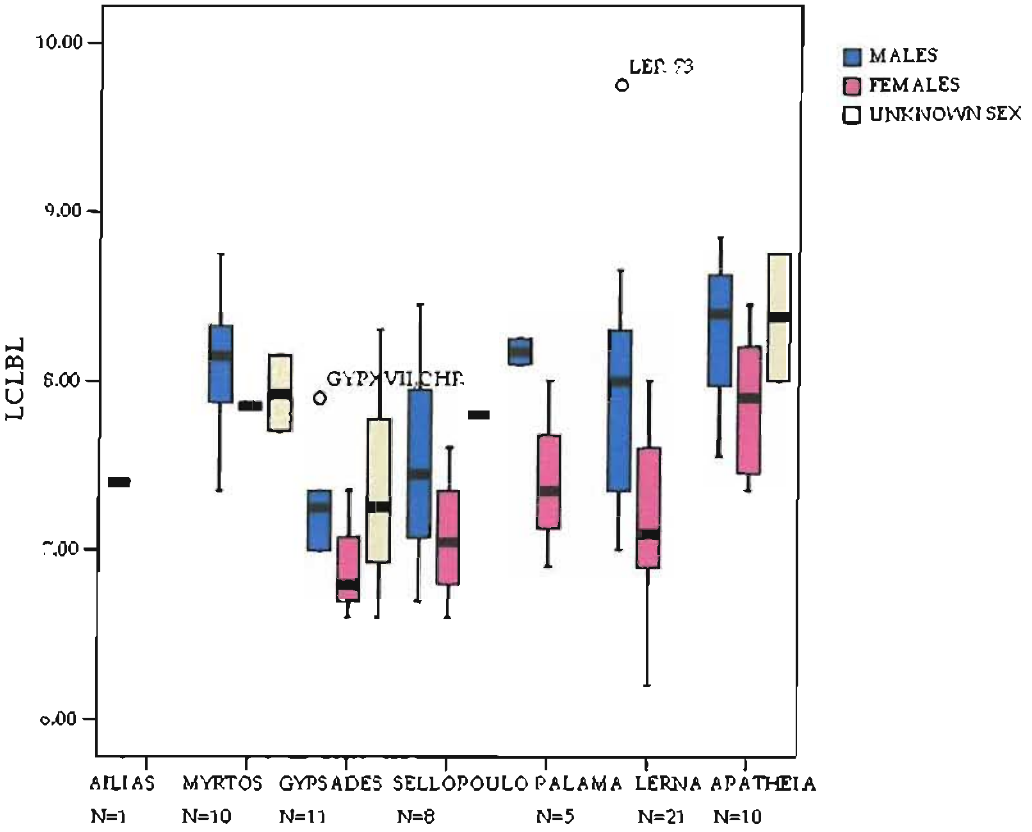


Figure 38 Box and whisker plot of MD crown diameter of Canine, Lower, Left side, Cretan and Argolid samples.

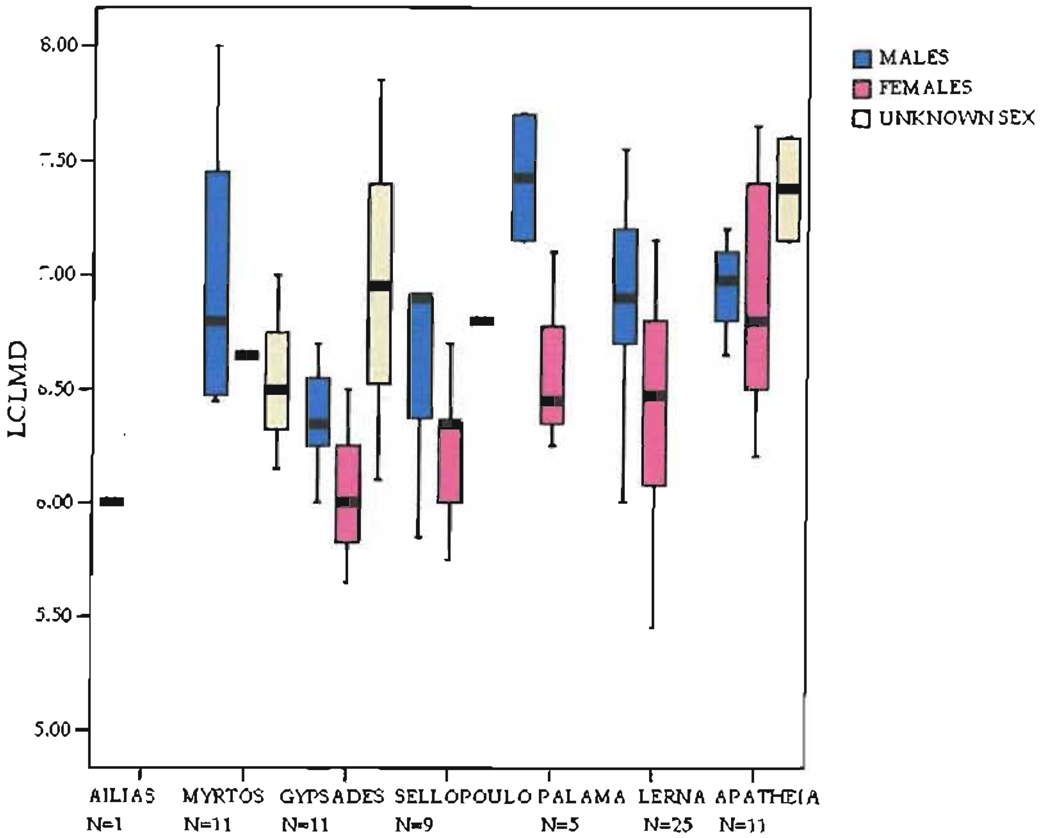


Figure 39 Box and whisker plot of BL crown diameter of 3rd Premolar, Lower, Left side, Cretan and Argolid samples.

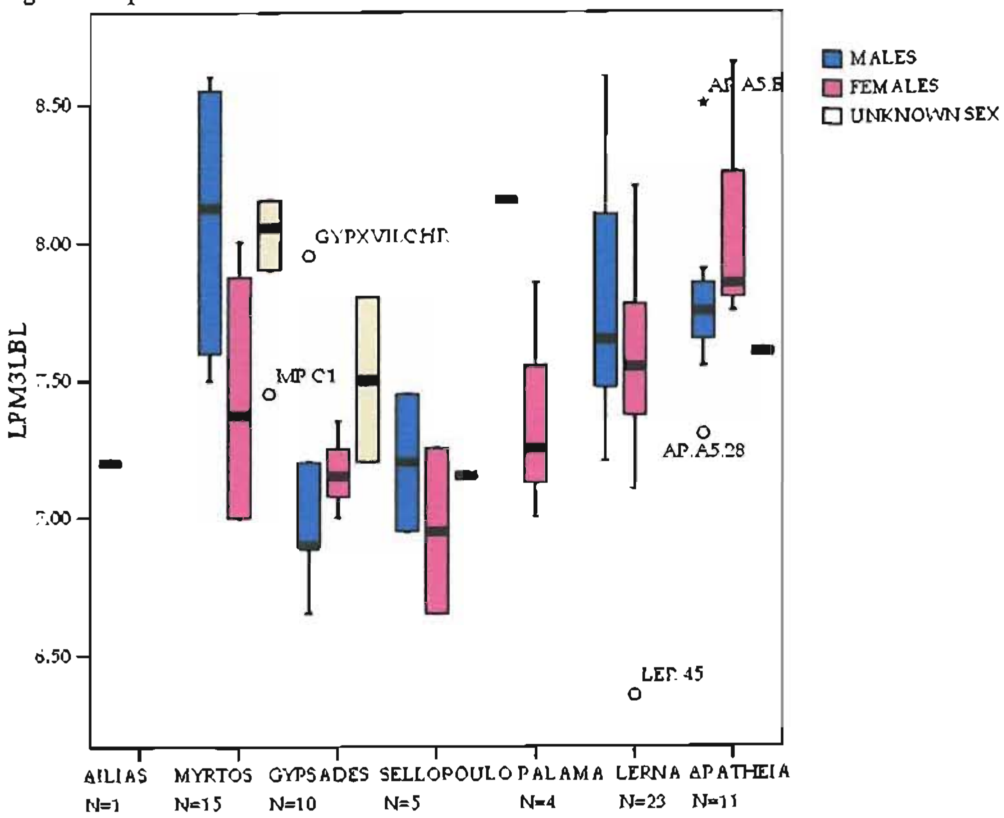


Figure 40 Box and whisker plot of MD crown diameter of 3rd Premolar, Lower, Left side, Cretan and Argolid samples.

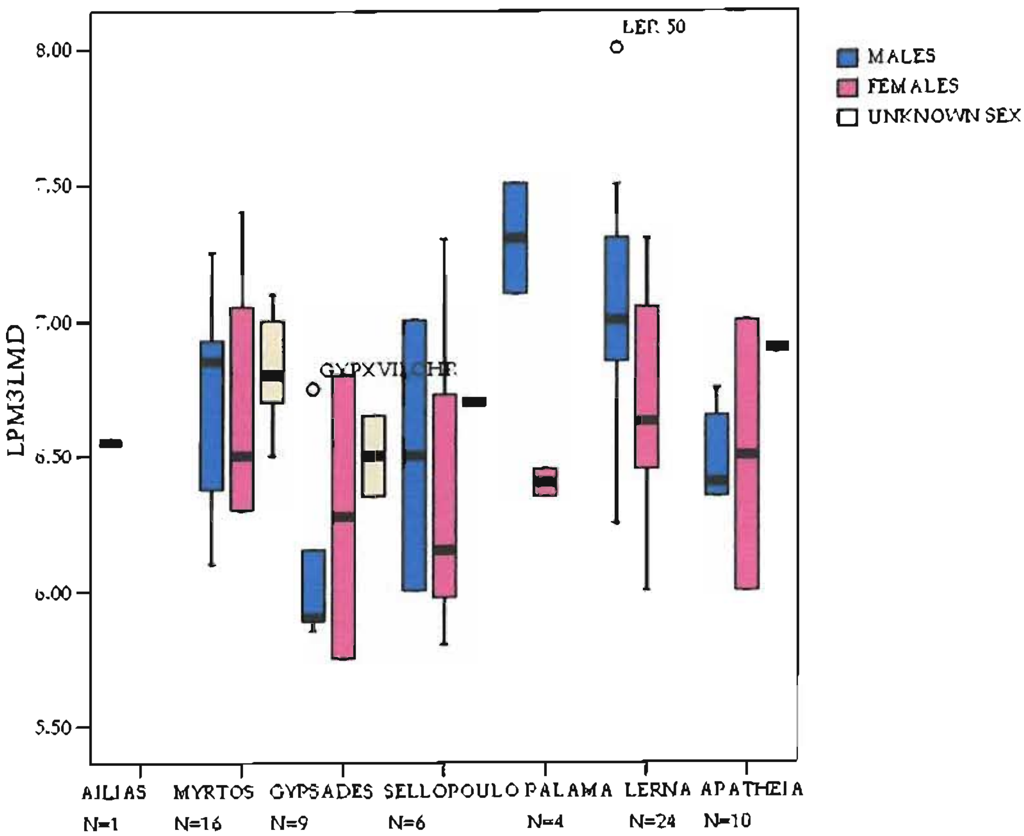


Figure 41 Box and whisker plot of BL crown diameter of 4th Premolar, Lower, Left side, Cretan and Argolid samples.

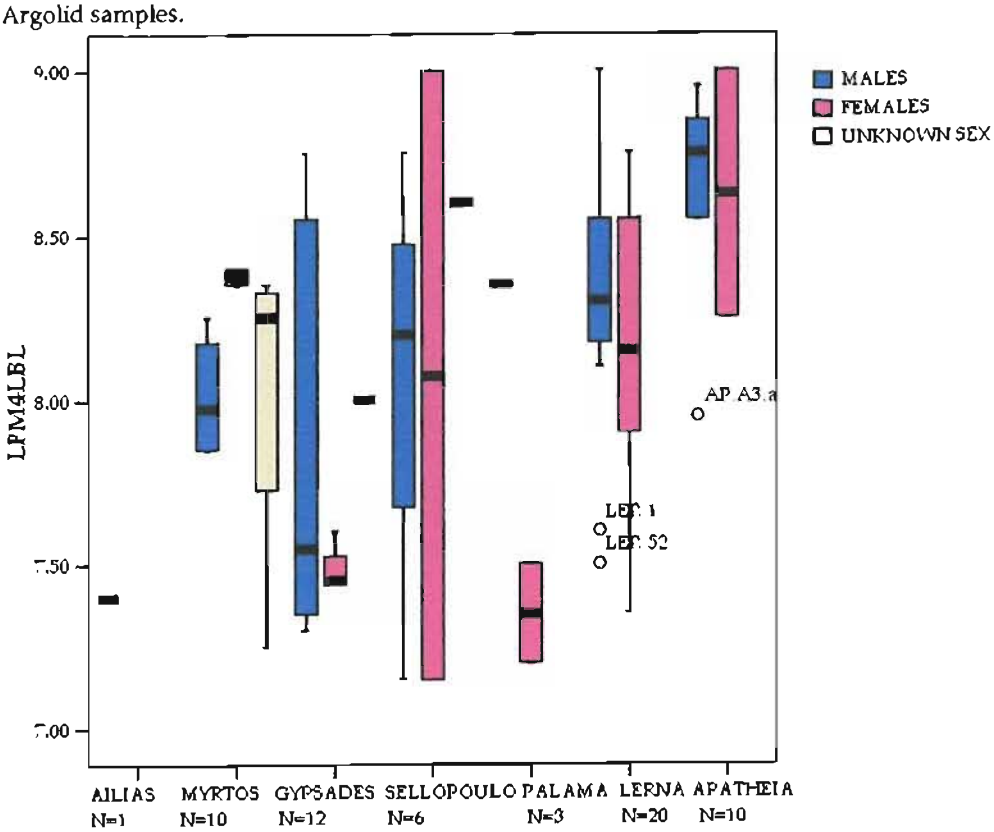


Figure 42 Box and whisker plot of MD crown diameter of 4th Premolar, Lower, Left side, Cretan and Argolid samples.

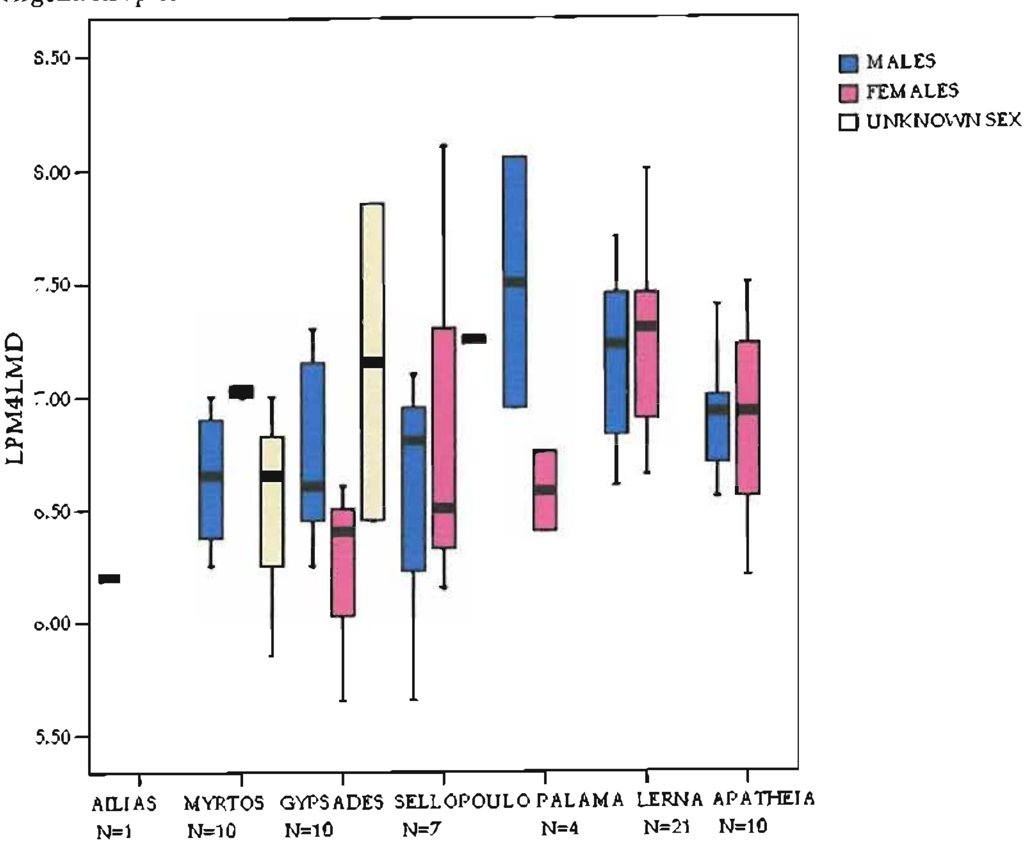


Figure 43 Box and whisker plot of BL crown diameter of 1st Molar, Lower, Left side, Cretan and Argolid samples.

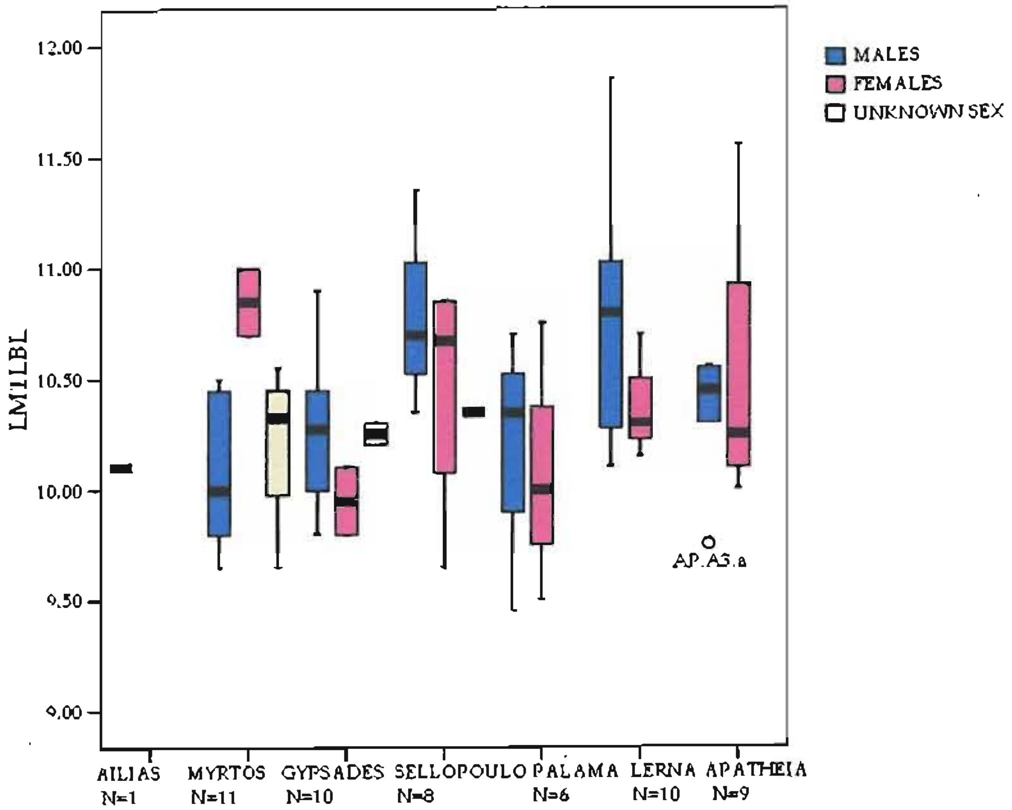


Figure 44 Box and whisker plot of MD crown diameter of 1st Molar, Lower, Left side, Cretan and Argolid samples.

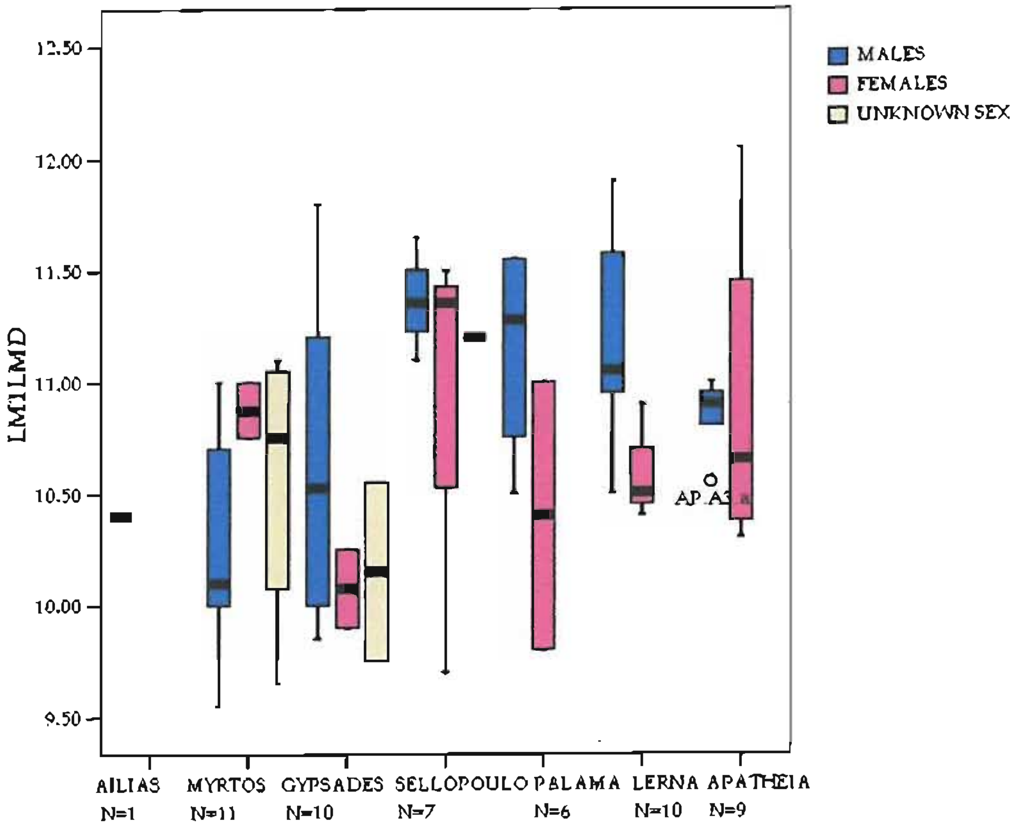


Figure 45 Box and whisker plot of BL crown diameter of 2nd Molar, Lower, Left side, Cretan and Argolid samples.

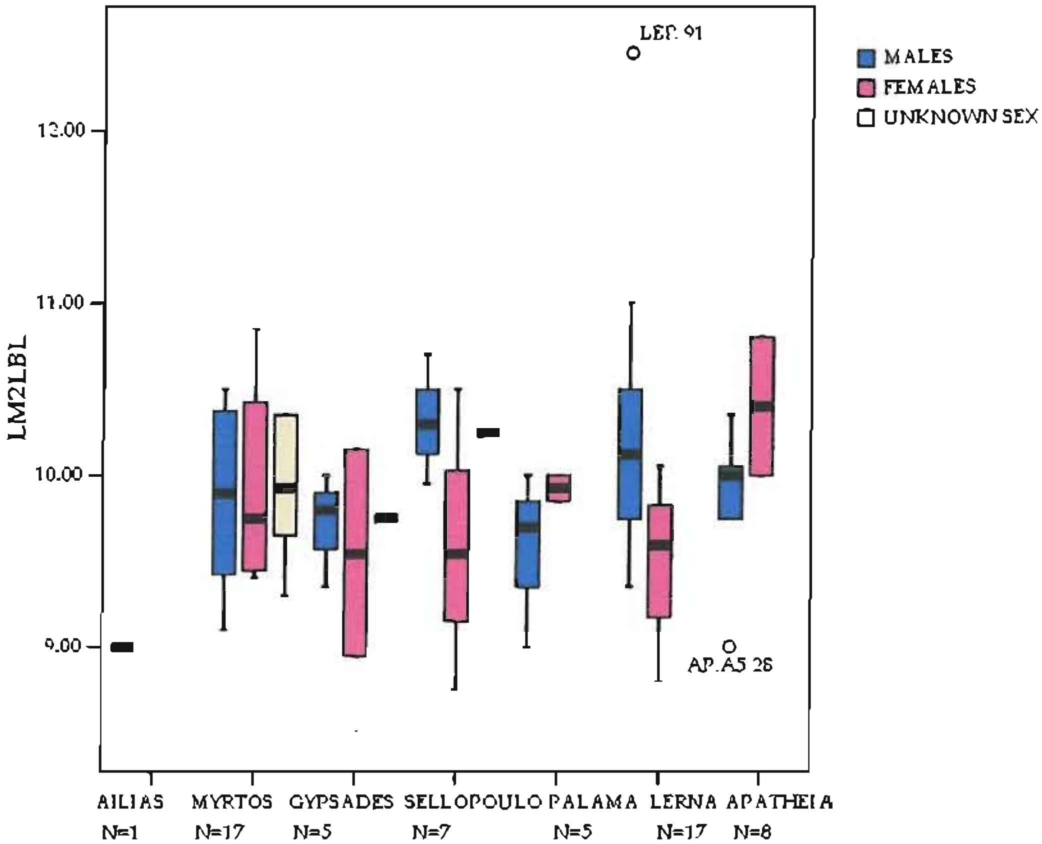


Figure 46 Box and whisker plot of MD crown diameter of 2nd Molar, Lower, Left side, Cretan and Argolid samples.

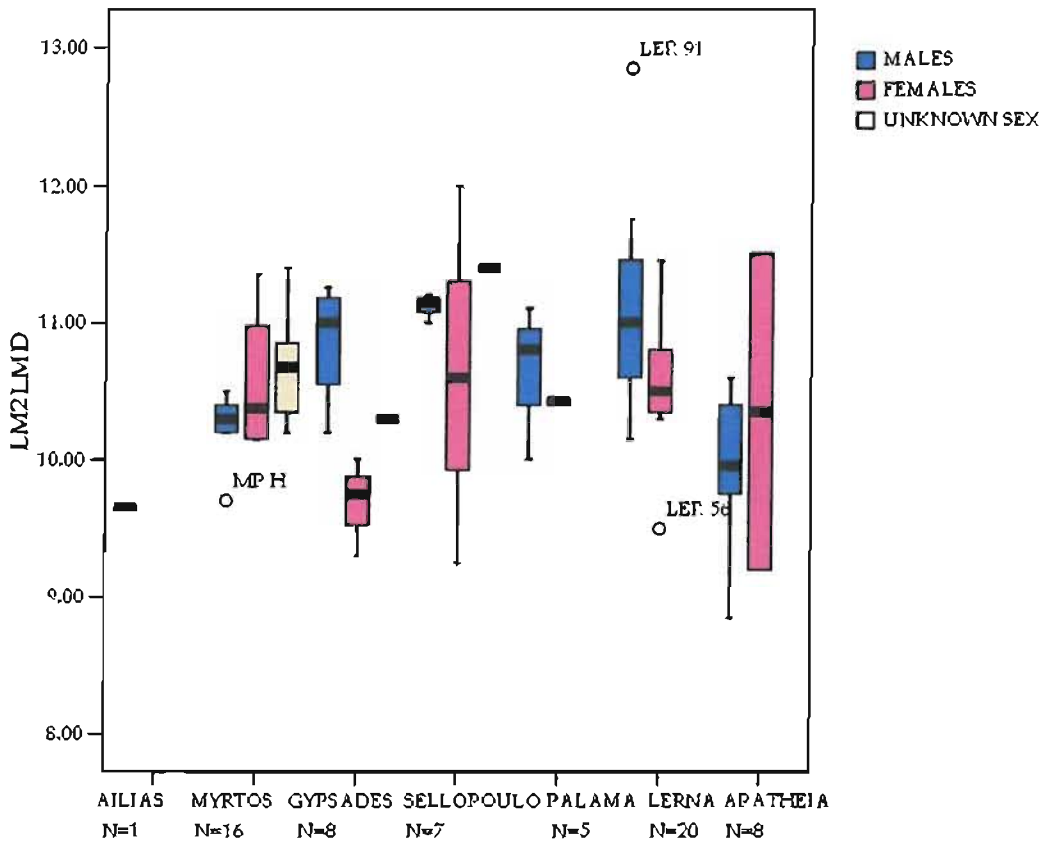


Figure 47 Box and whisker plot of BL crown diameter of 3rd Molar, Lower, Left side, Cretan and Argolid samples.

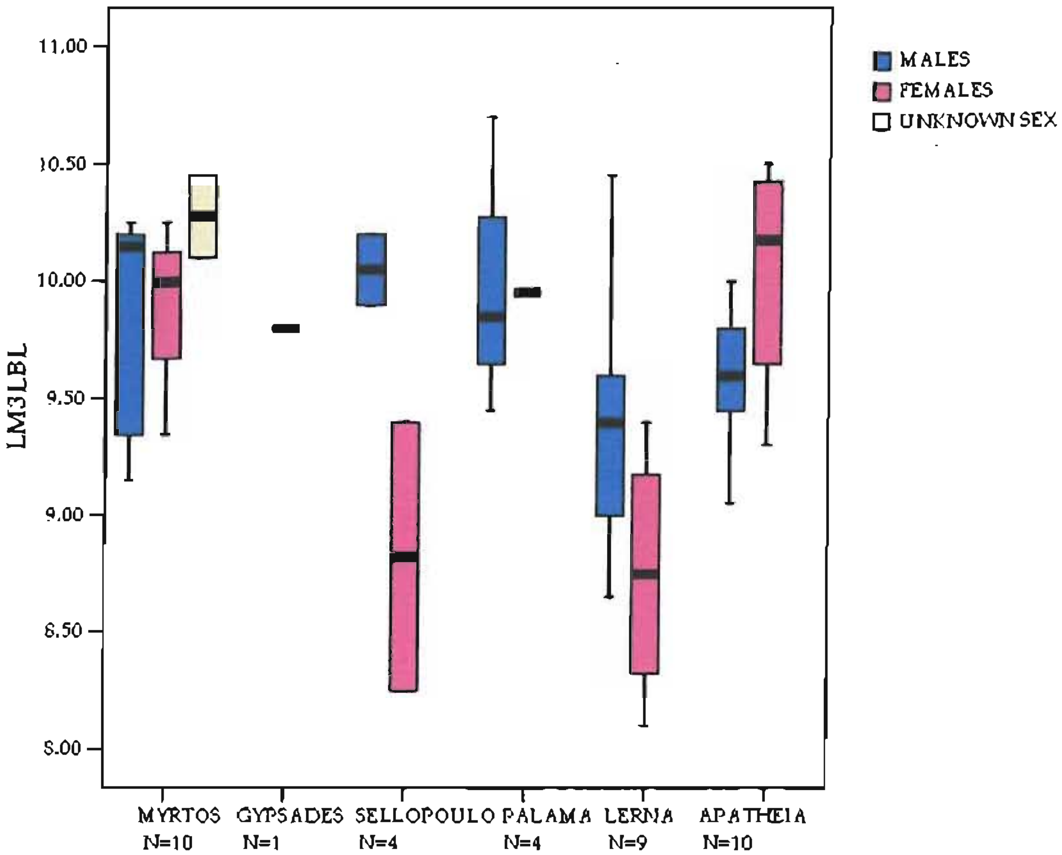


Figure 48 Box and whisker plot of MD crown diameter of 3rd Molar, Lower, Left side, Cretan and Argolid samples.

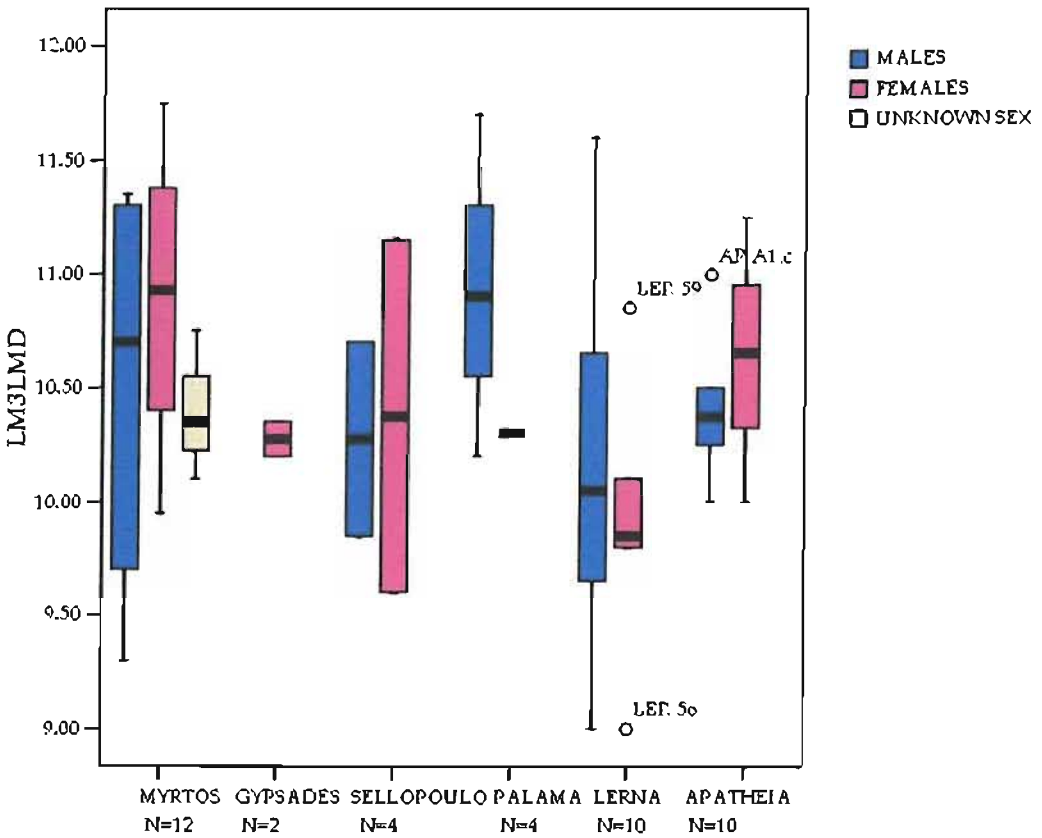


Figure 49 Box and whisker plot of BL crown diameter of 1st Incisor, Lower, Right side, Cretan and Argolid samples.

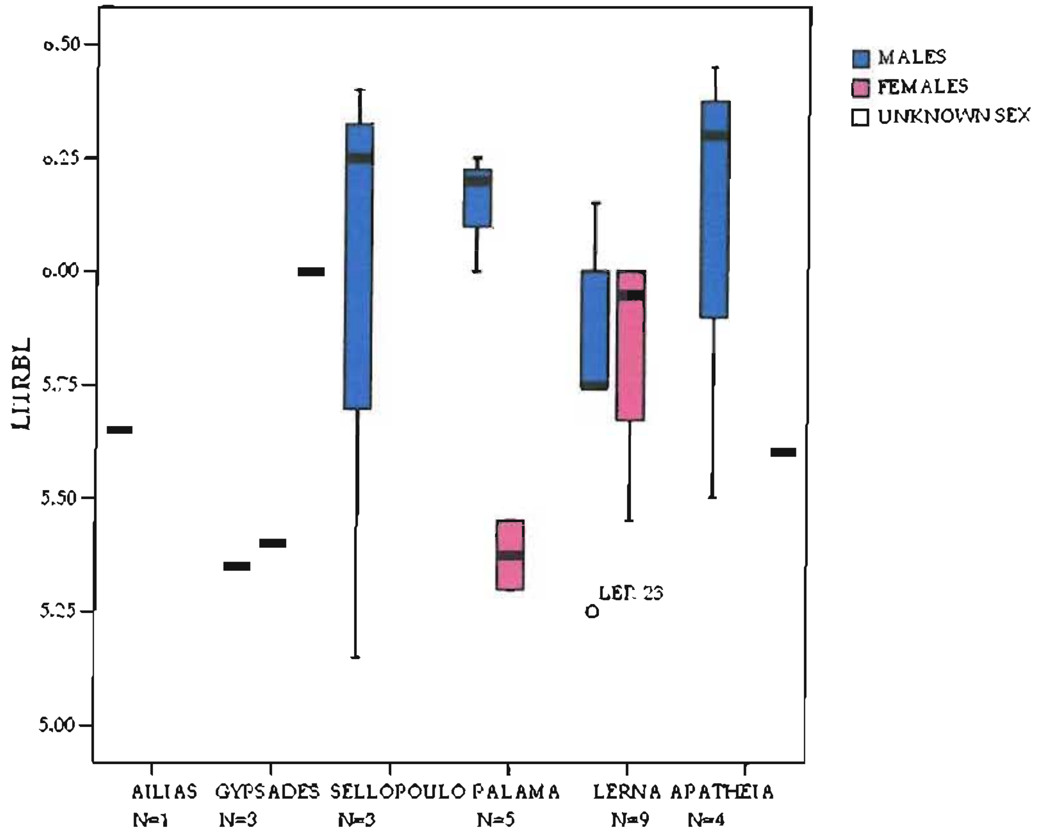


Figure 50 Box and whisker plot of MD crown diameter of 1st Incisor, Lower, Right side, Cretan and Argolid samples.

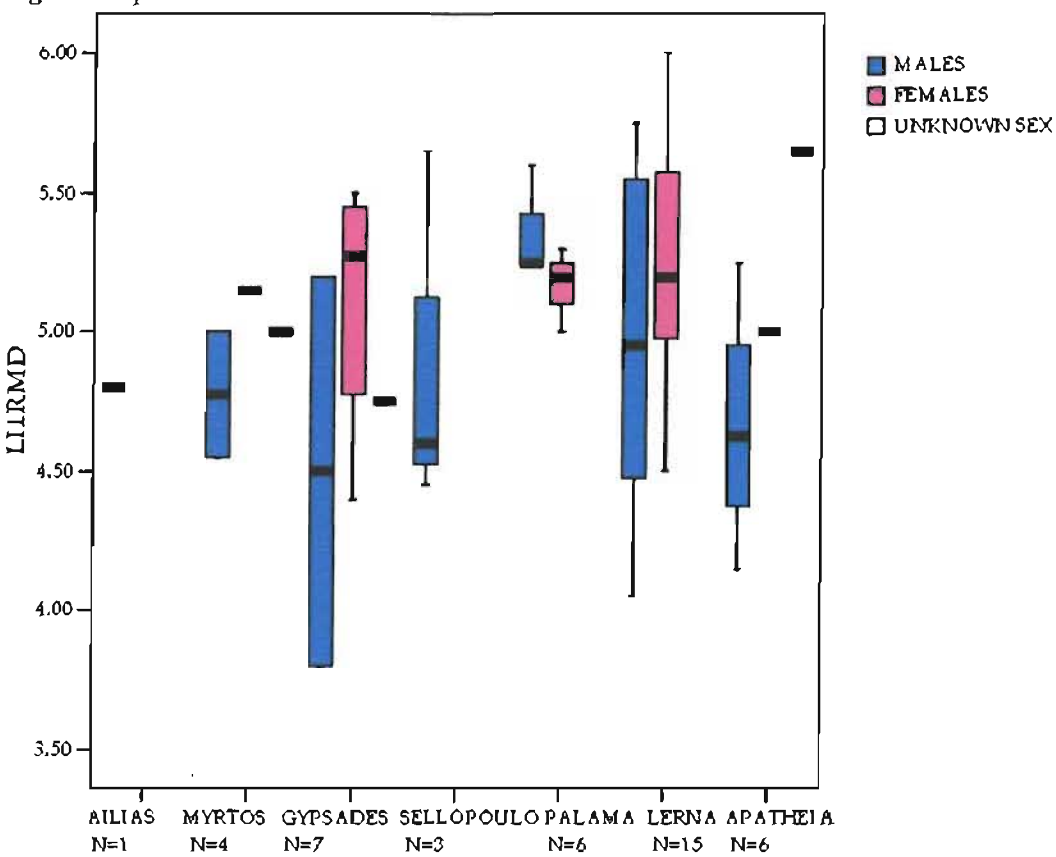


Figure 51 Box and whisker plot of BL crown diameter of 2nd Incisor, Lower, Right side, Cretan and Argolid samples.

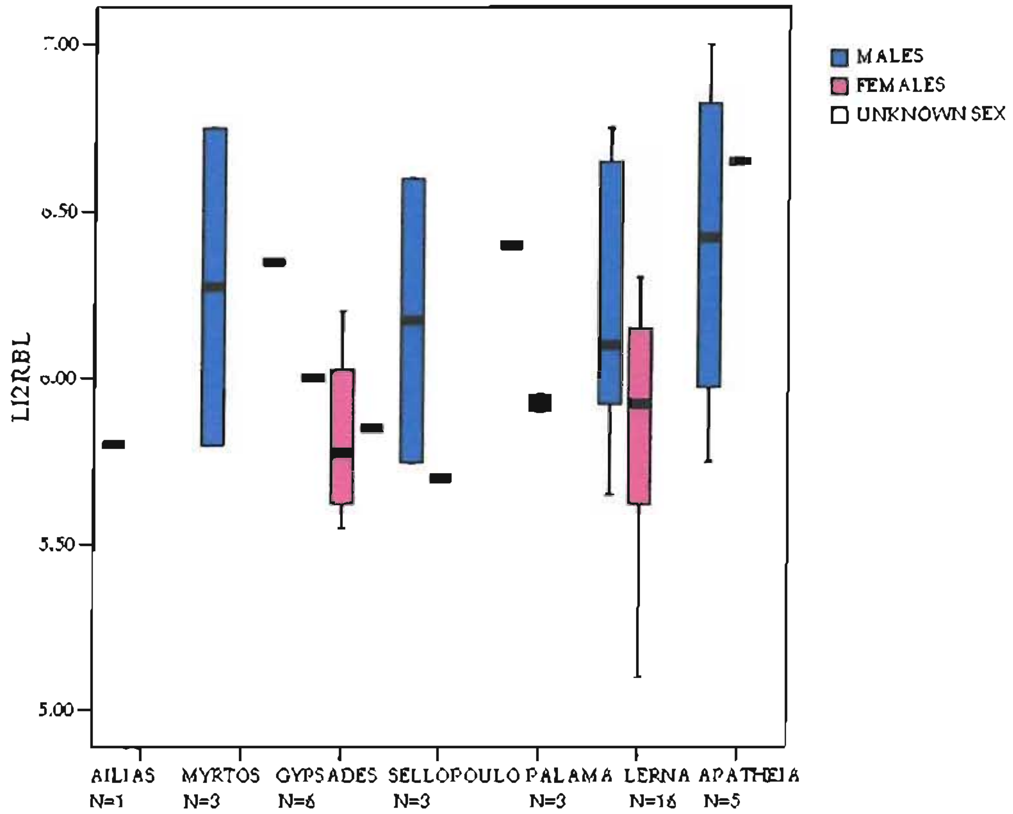


Figure 52 Box and whisker plot of MD crown diameter of 2nd Incisor, Lower, Right side, Cretan and Argolid samples.

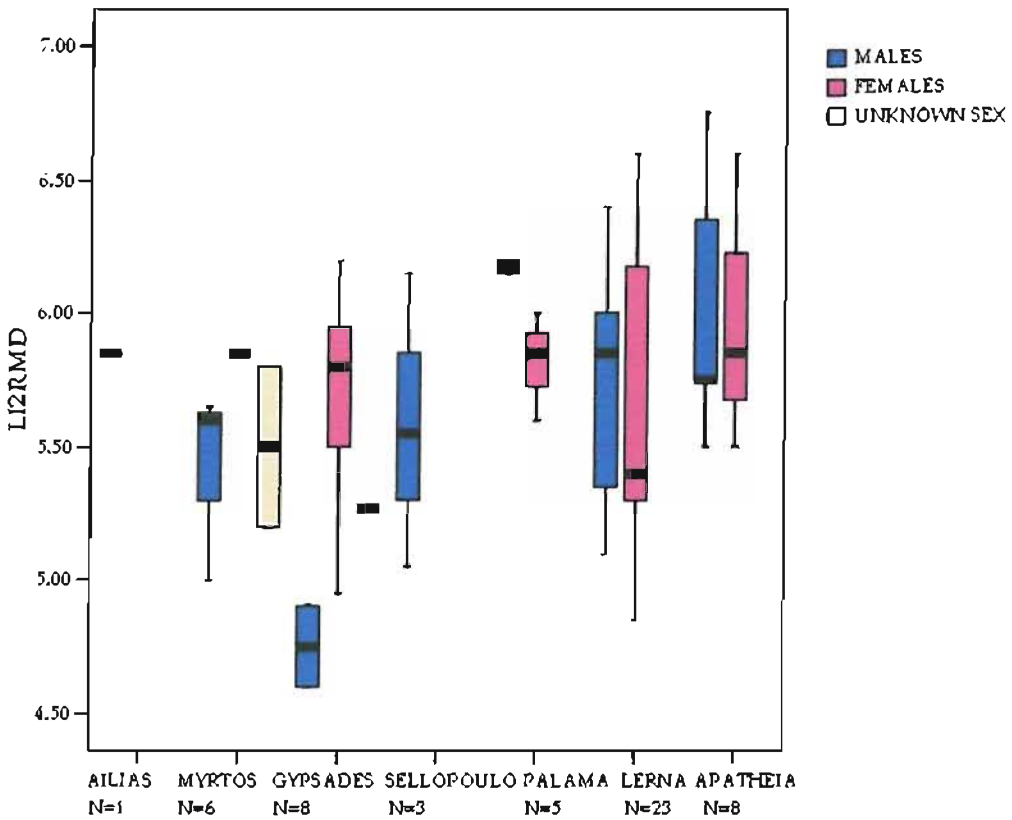


Figure 53 Box and whisker plot of BL crown diameter of Canine, Lower, Right side, Cretan and Argolid samples.

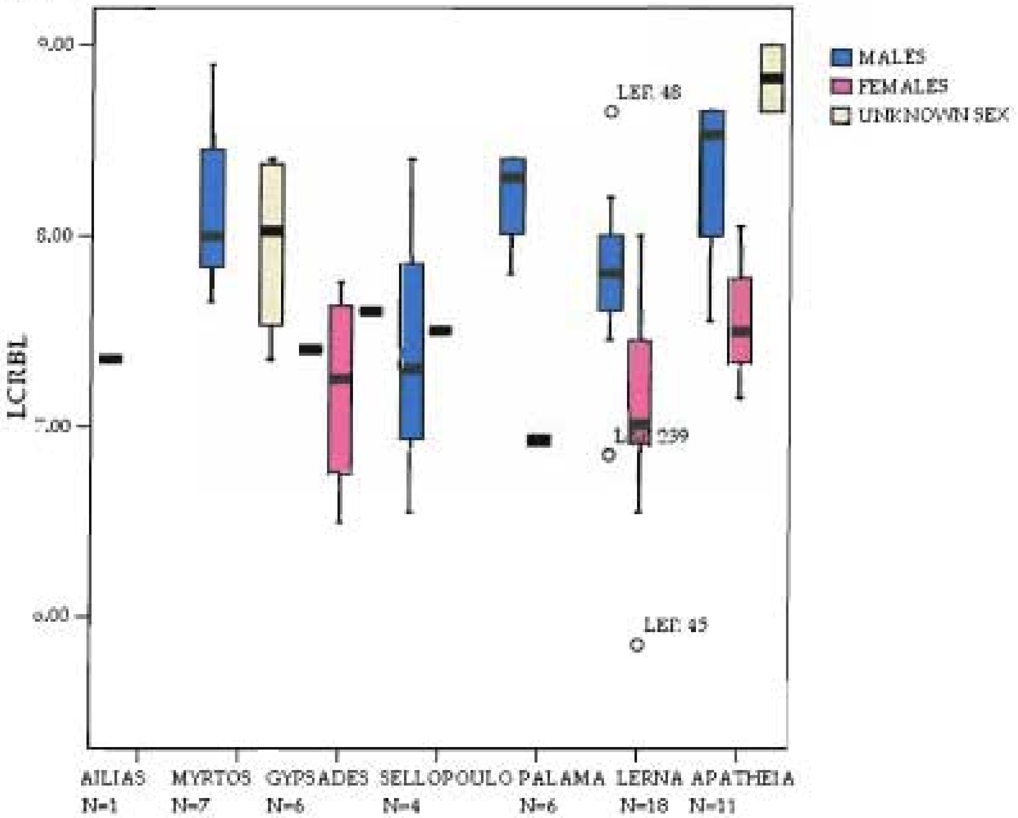


Figure 54 Box and whisker plot of MD crown diameter of Canine, Lower, Right side, Cretan and Argolid samples.

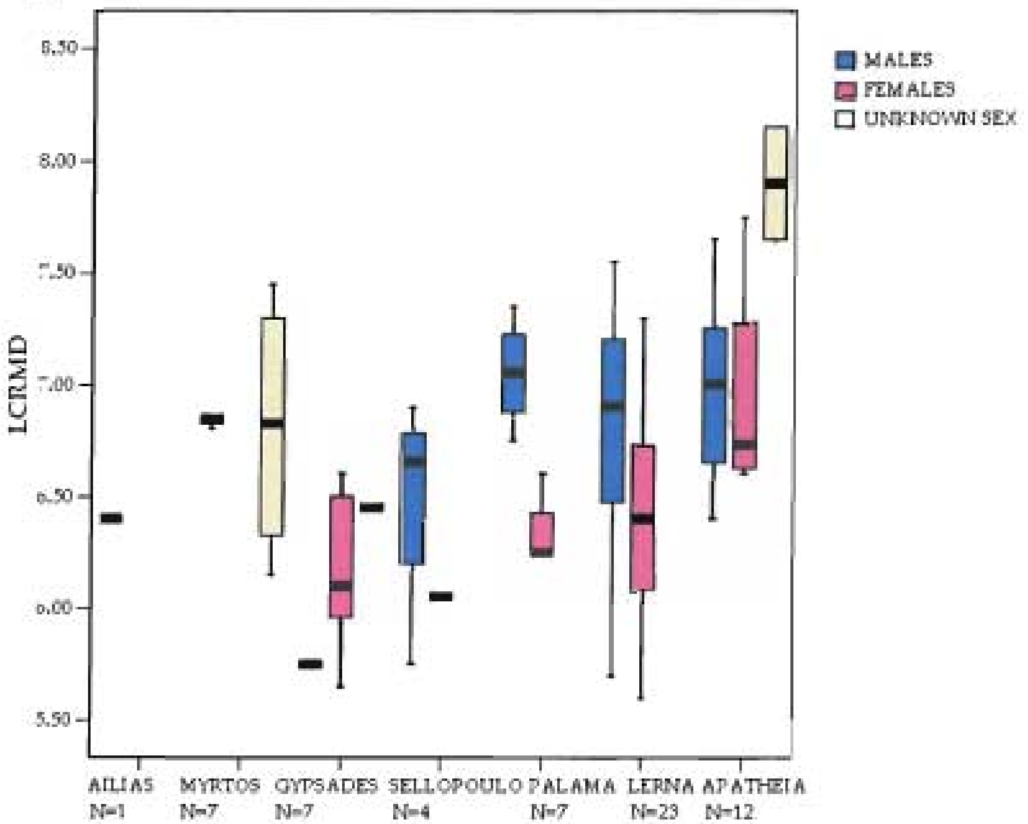


Figure 55 Box and whisker plot of BL crown diameter of 3rd Premolar, Lower, Right side, Cretan and Argolid samples.

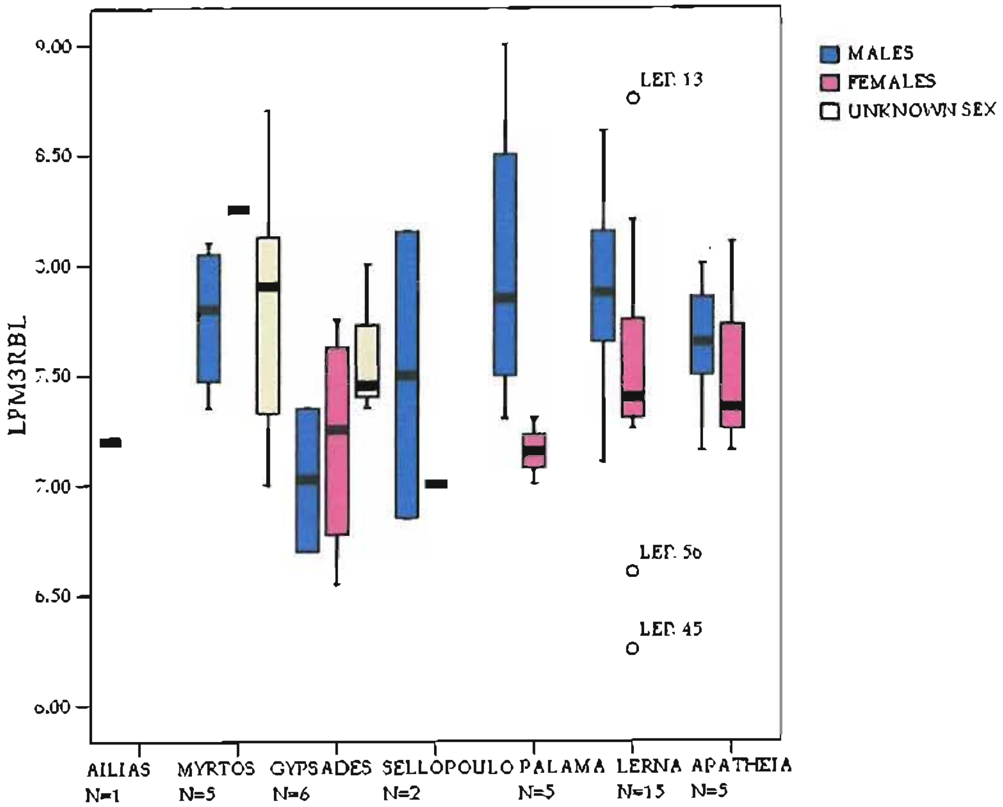


Figure 56 Box and whisker plot of MD crown diameter of 3rd Premolar, Lower, Right side, Cretan and Argolid samples.

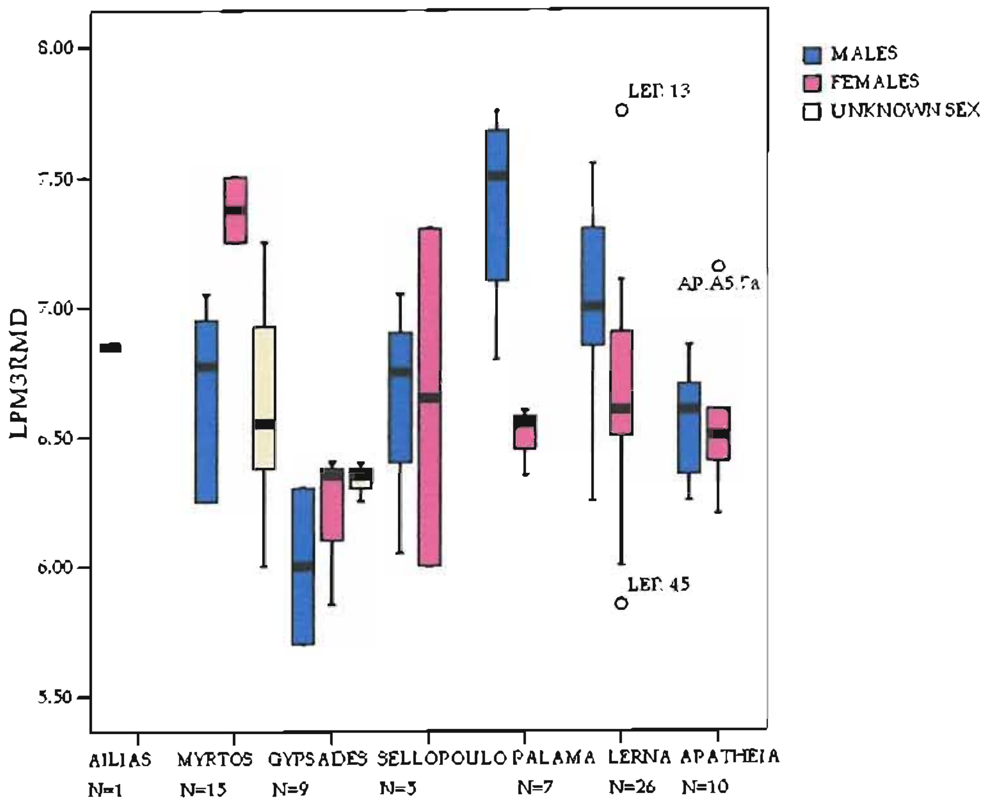


Figure 57 Box and whisker plot of BL crown diameter of 4th Premolar, Lower, Right side, Cretan and Argolid samples.

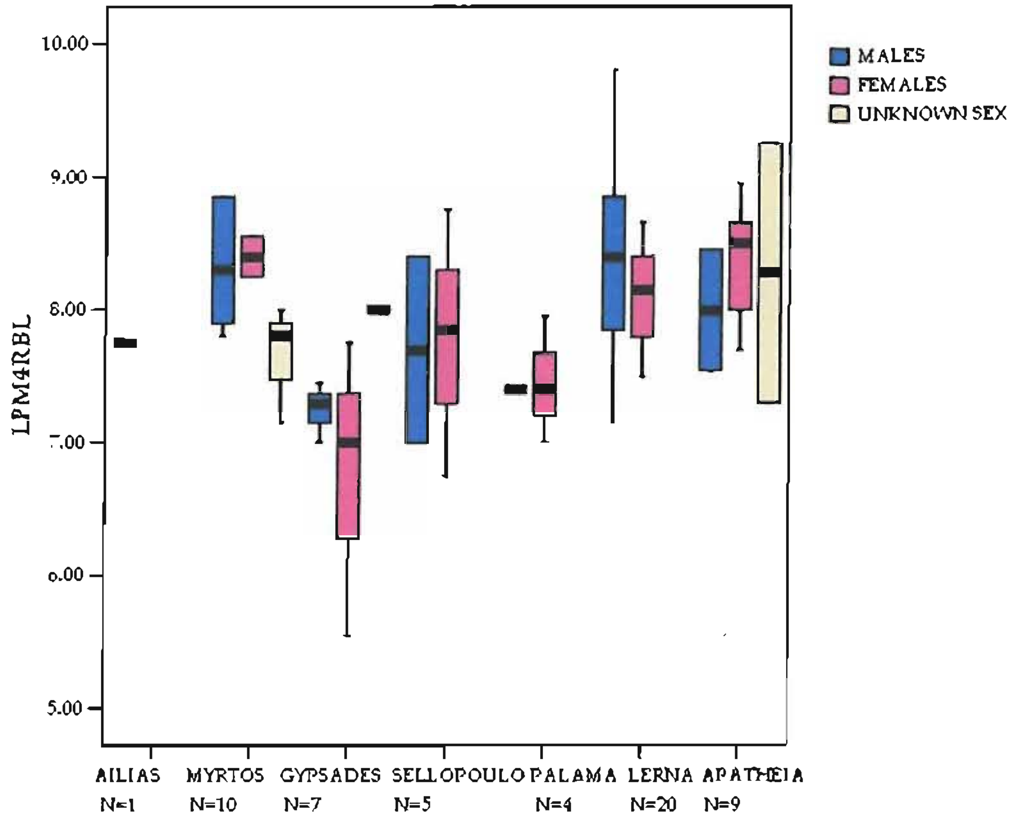


Figure 58 Box and whisker plot of MD crown diameter of 4th Premolar Lower, Right side, Cretan and Argolid samples.

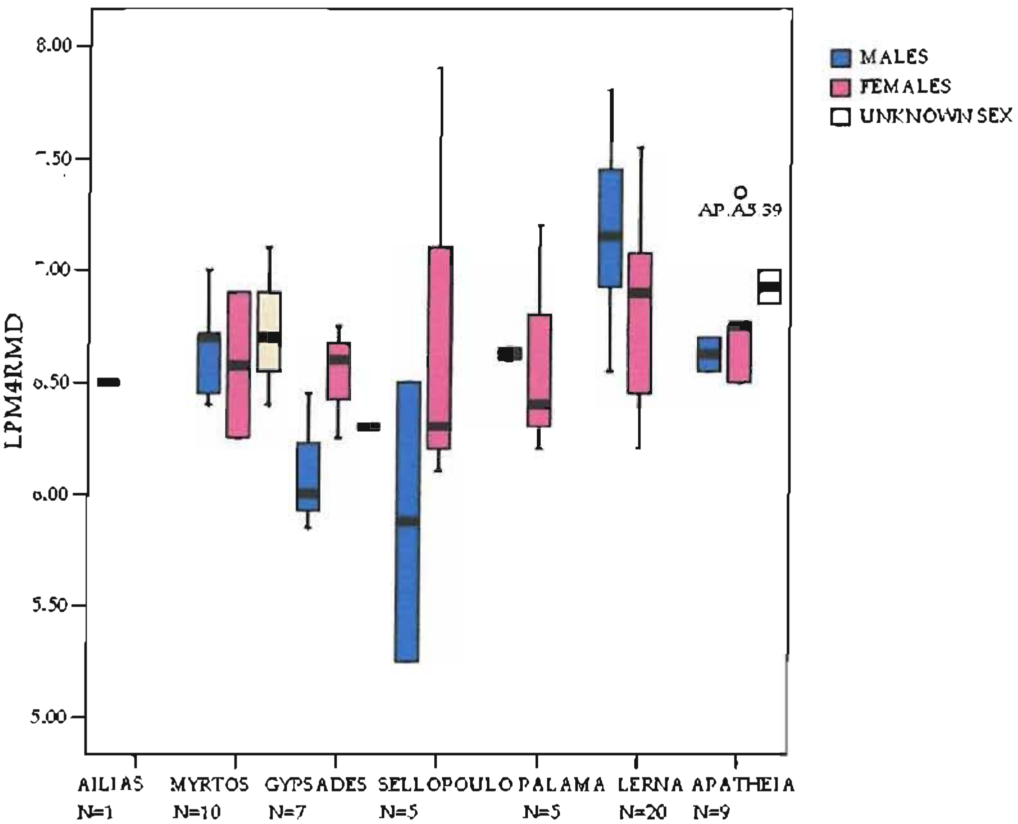


Figure 59 Box and whisker plot of BL crown diameter of 1st Molar, Lower, Right side, Cretan and Argolid samples.

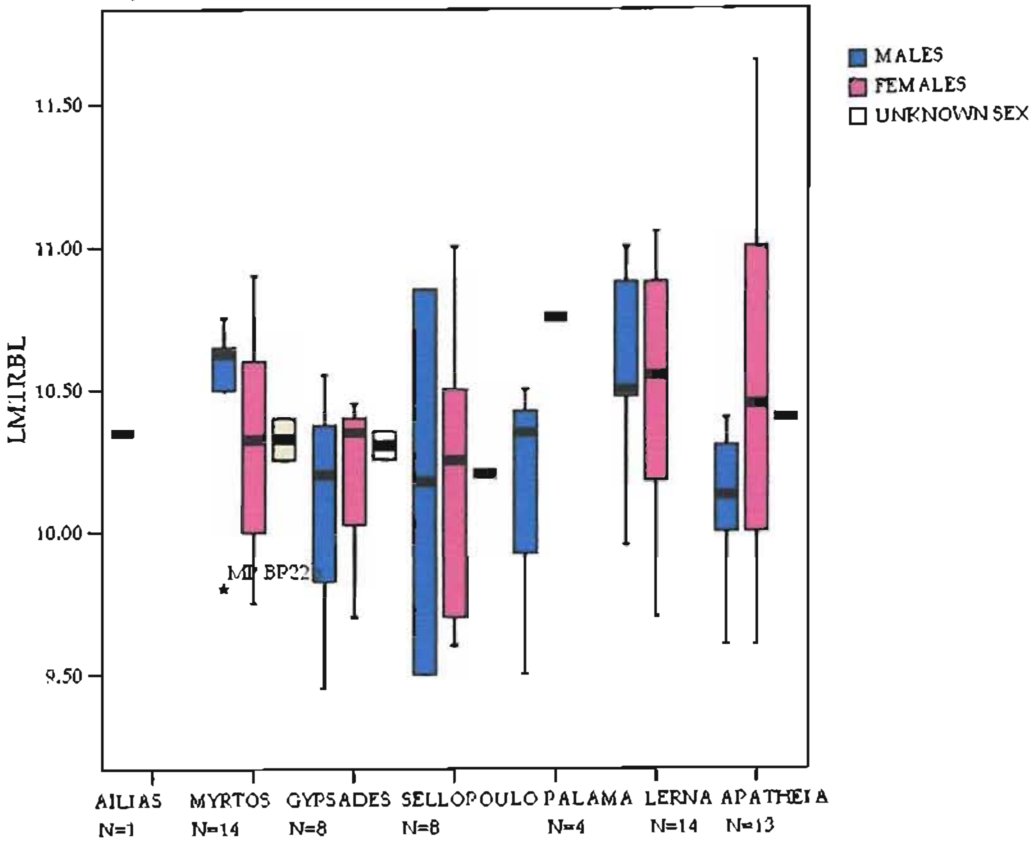


Figure 60 Box and whisker plot of MD crown diameter of 1st Molar, Lower, Right side, Cretan and Argolid samples.

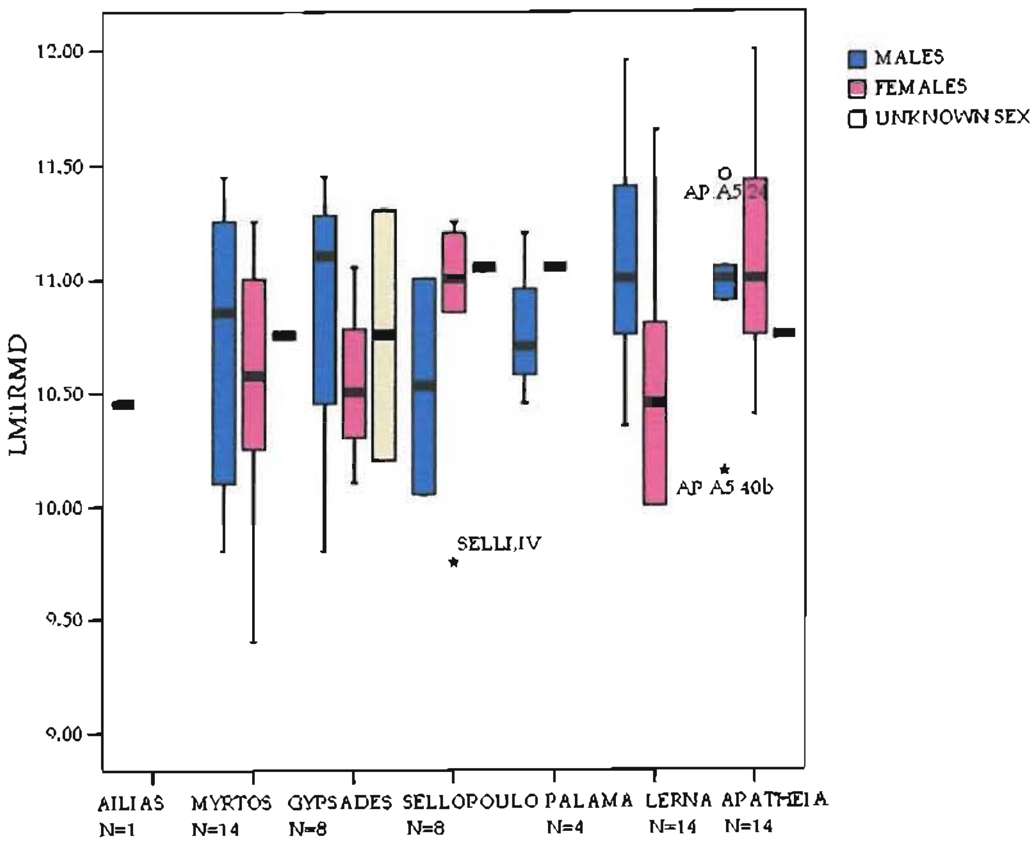


Figure 61 Box and whisker plot of BL crown diameter of 2nd Molar, Lower, Right side, Cretan and Argolid samples.

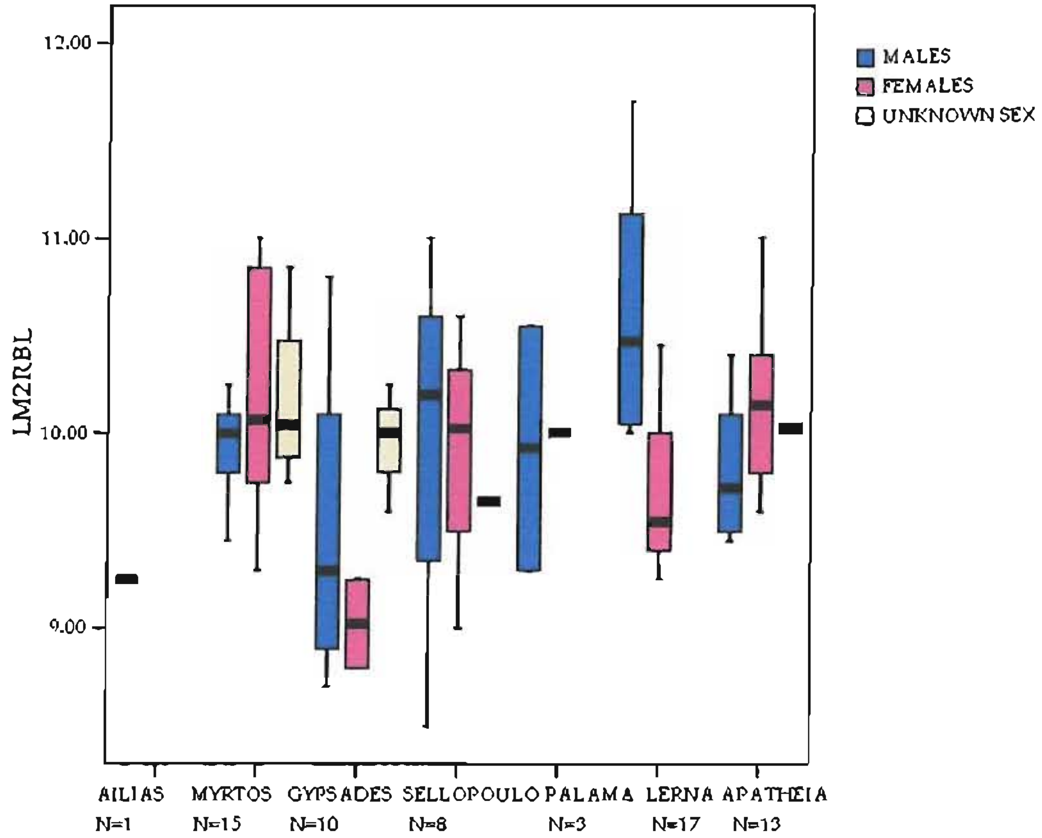


Figure 62 Box and whisker plot of MD crown diameter of 2nd Molar, Lower, Right side, Cretan and Argolid samples.

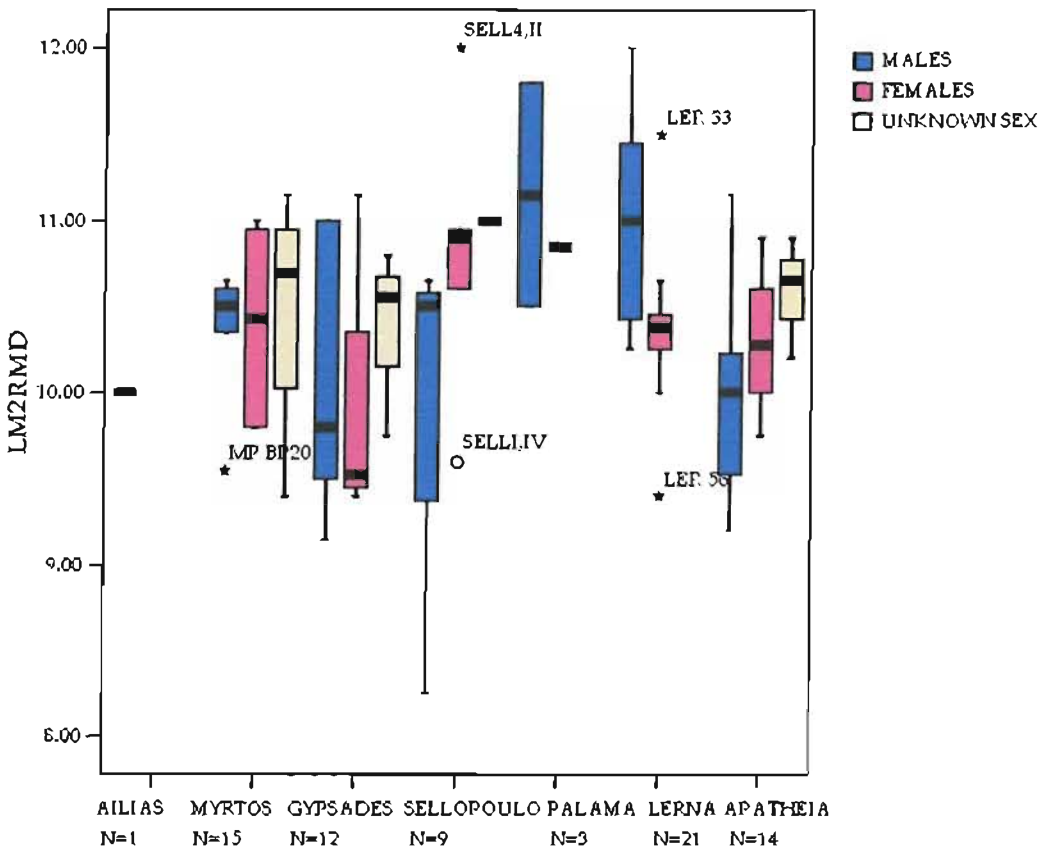


Figure 63 Box and whisker plot of BL crown diameter of 3rd Molar, Lower, Right side, Cretan and Argolid samples.

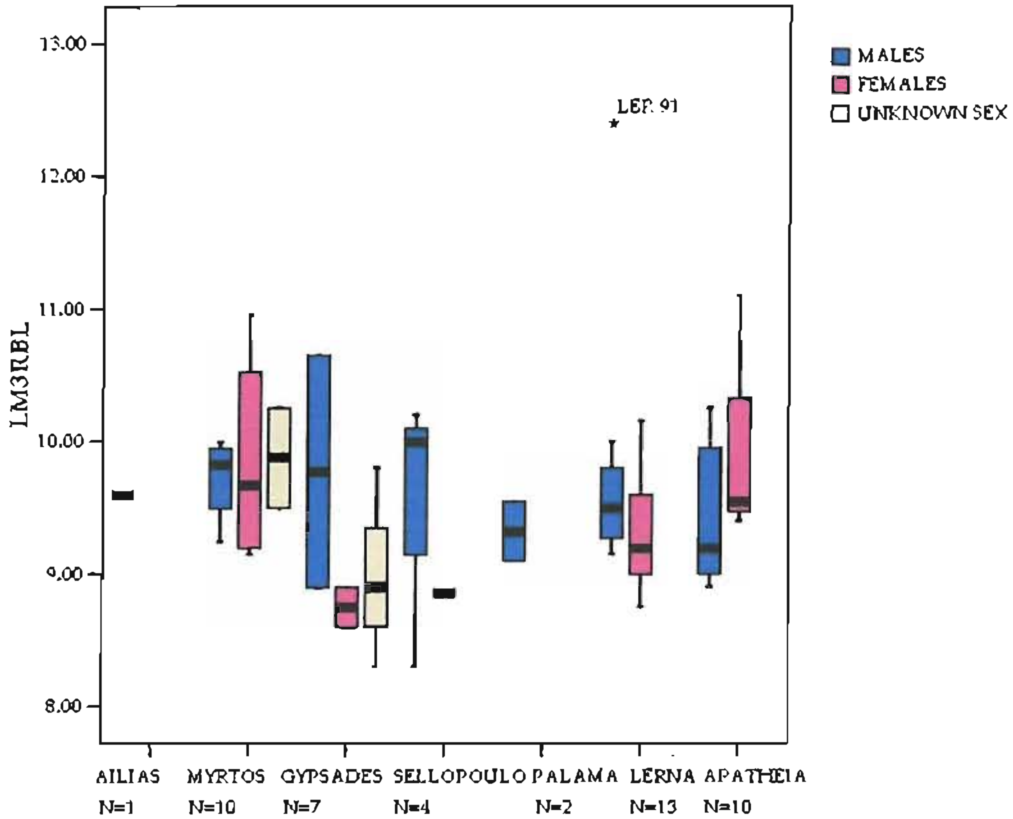
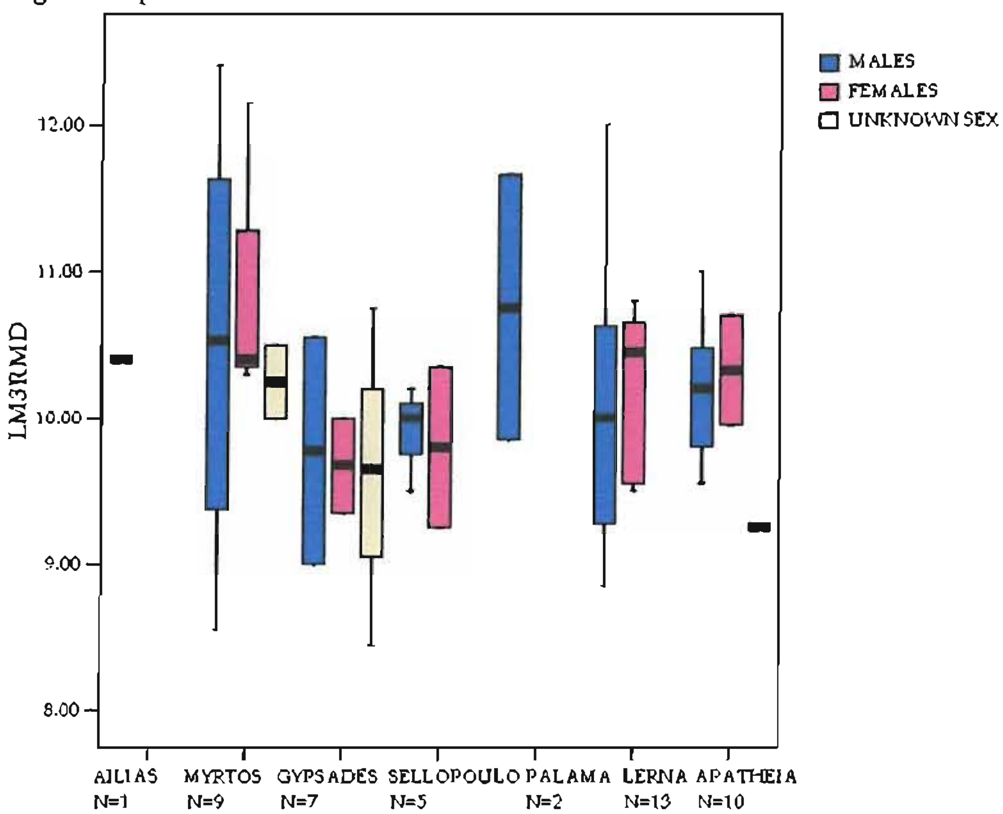


Figure 64 Box and whisker plot of MD crown diameter of 3rd Molar, Lower, Right side, Cretan and Argolid samples.



Maxillary teeth, Right and Left sides Pooled

Figure 65 Box and whisker plot of BL crown diameter of 1st Incisor, Upper, R and L sides pooled, Cretan and Argolid samples.

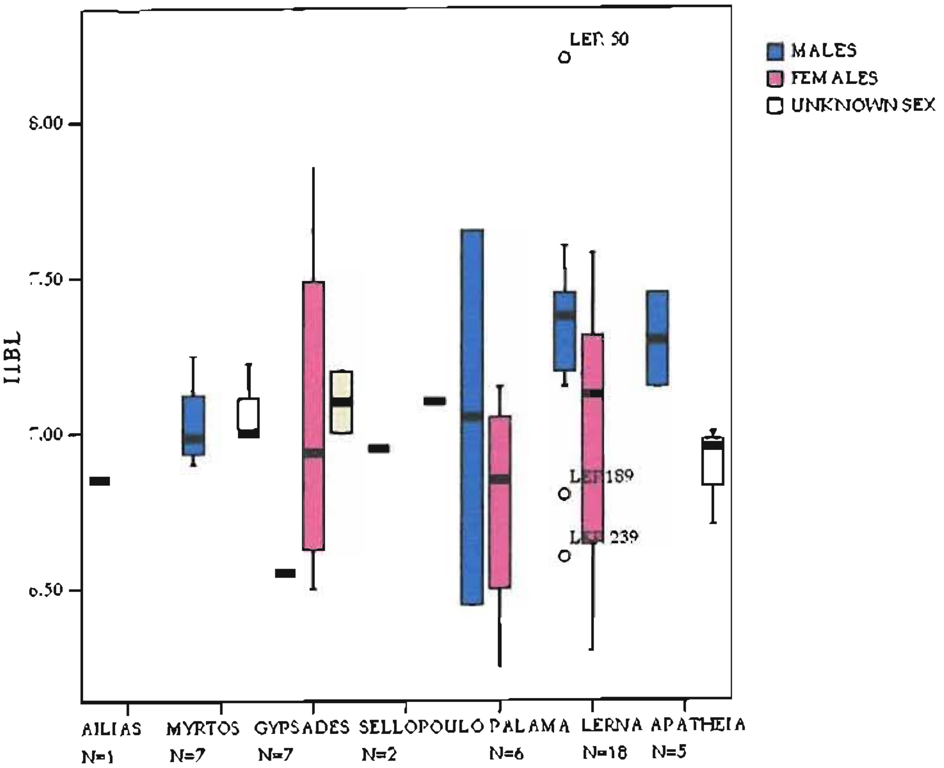


Figure 66 Box and whisker plot of MD crown diameter of 1st Incisor, Upper, R and L sides pooled, Cretan and Argolid samples.

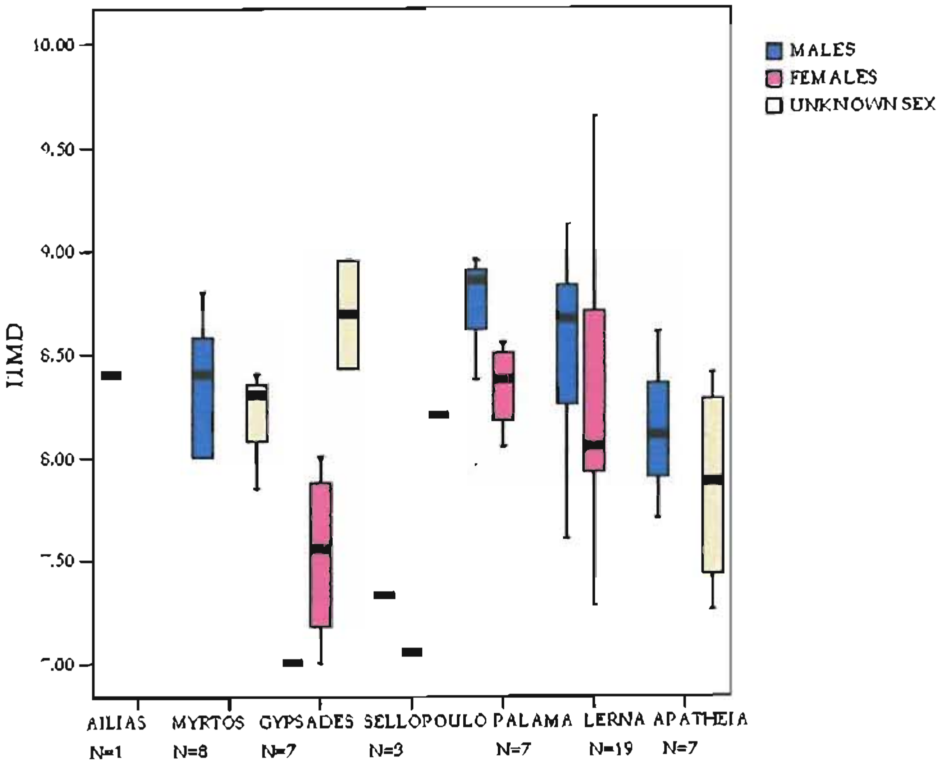


Figure 67 Box and whisker plot of BL crown diameter of 2nd Incisor, Upper, R and L sides pooled, Cretan and Argolid samples.

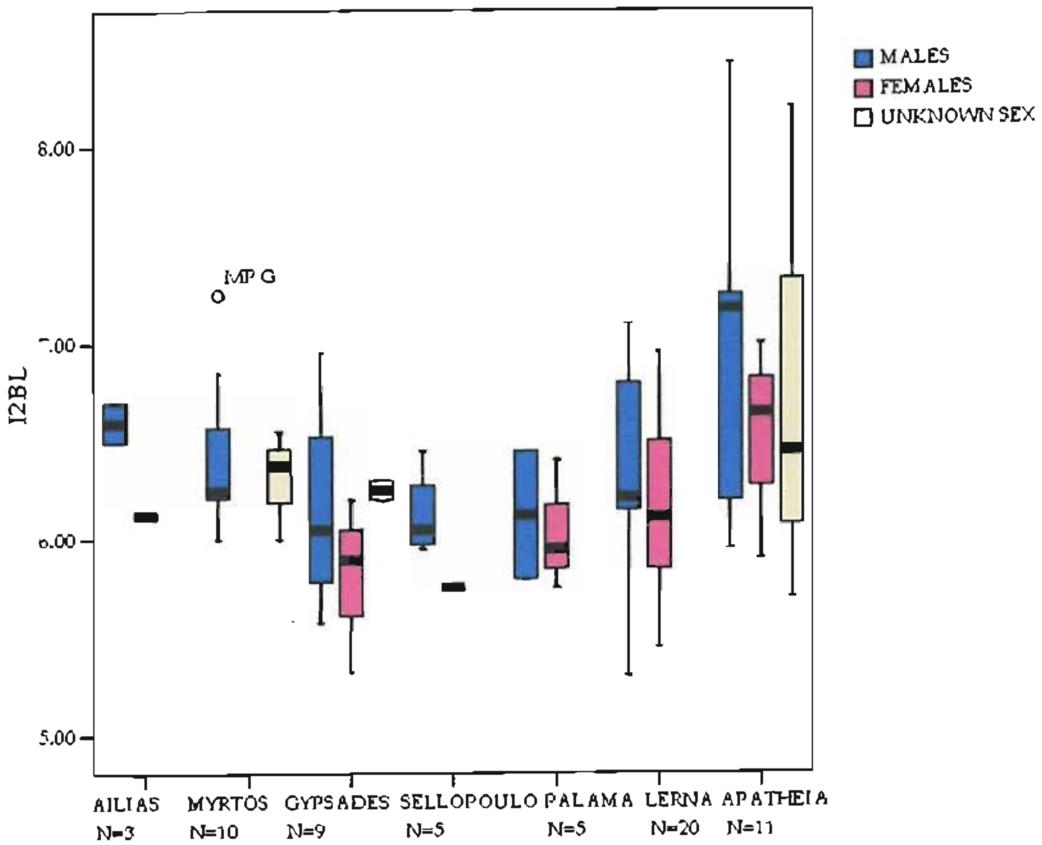


Figure 68 Box and whisker plot of MD crown diameter of 2nd Incisor, Upper, R and L sides pooled, Cretan and Argolid samples.

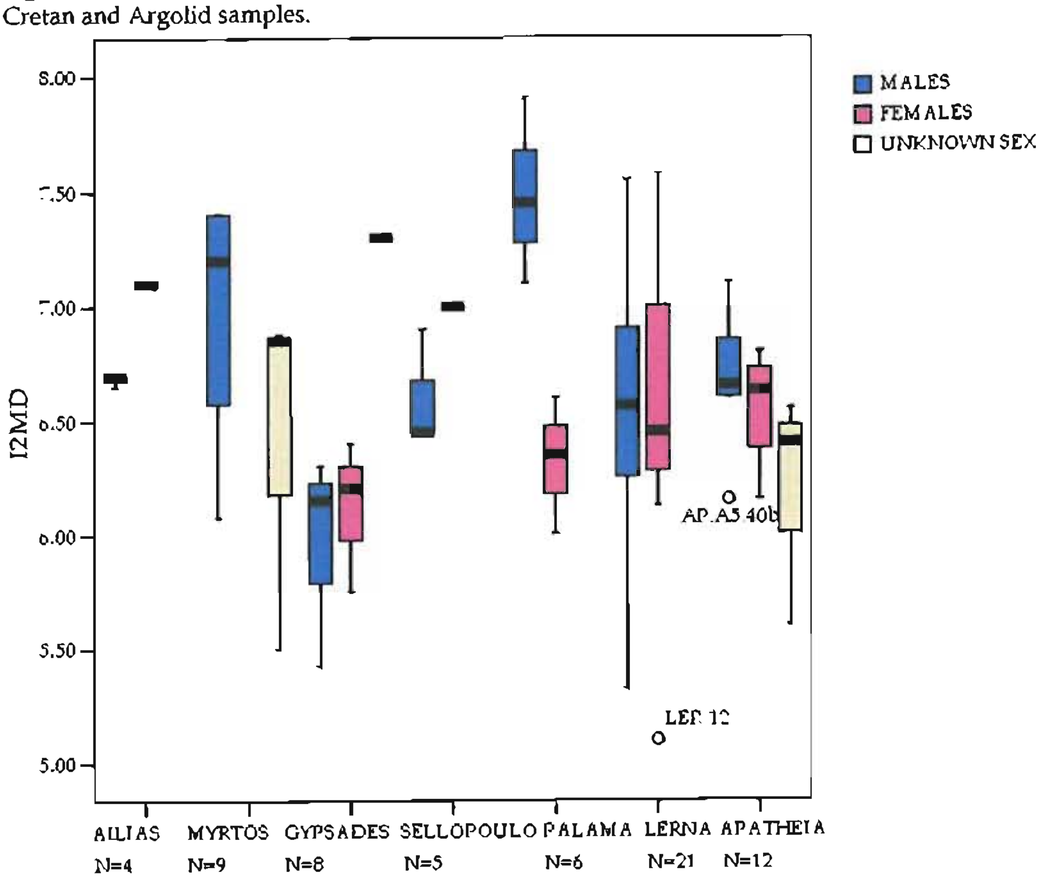


Figure 69 Box and whisker plot of BL crown diameter of Canine, Upper, R and L sides pooled, Cretan and Argolid samples.

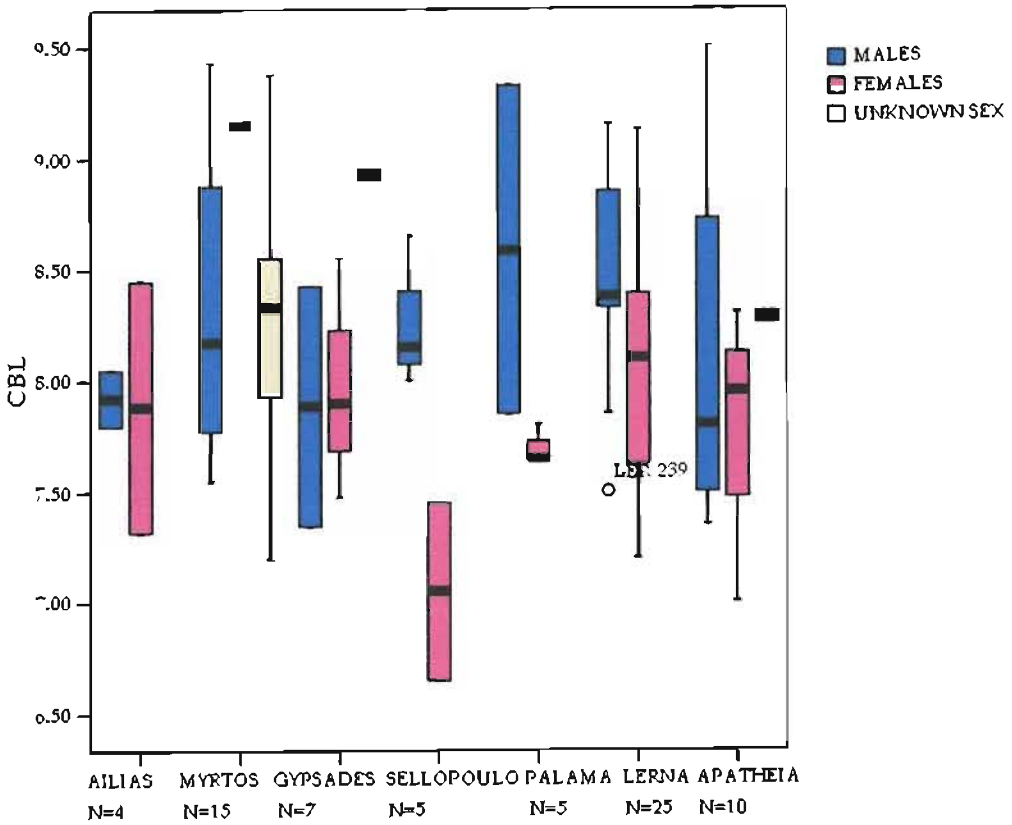


Figure 70 Box and whisker plot of MD crown diameter of Canine, Upper, R and L sides pooled, Cretan and Argolid samples.

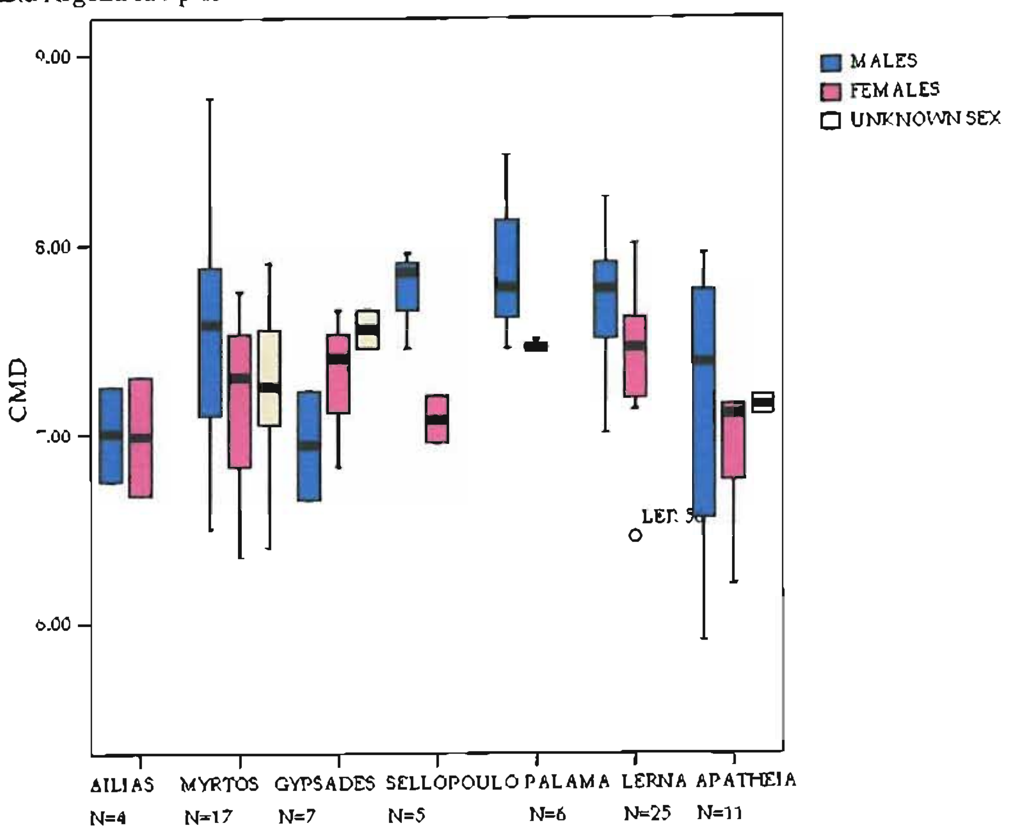


Figure 71 Box and whisker plot of BL crown diameter of 3rd Premolar, Upper, R and L sides pooled, Cretan and Argolid samples.

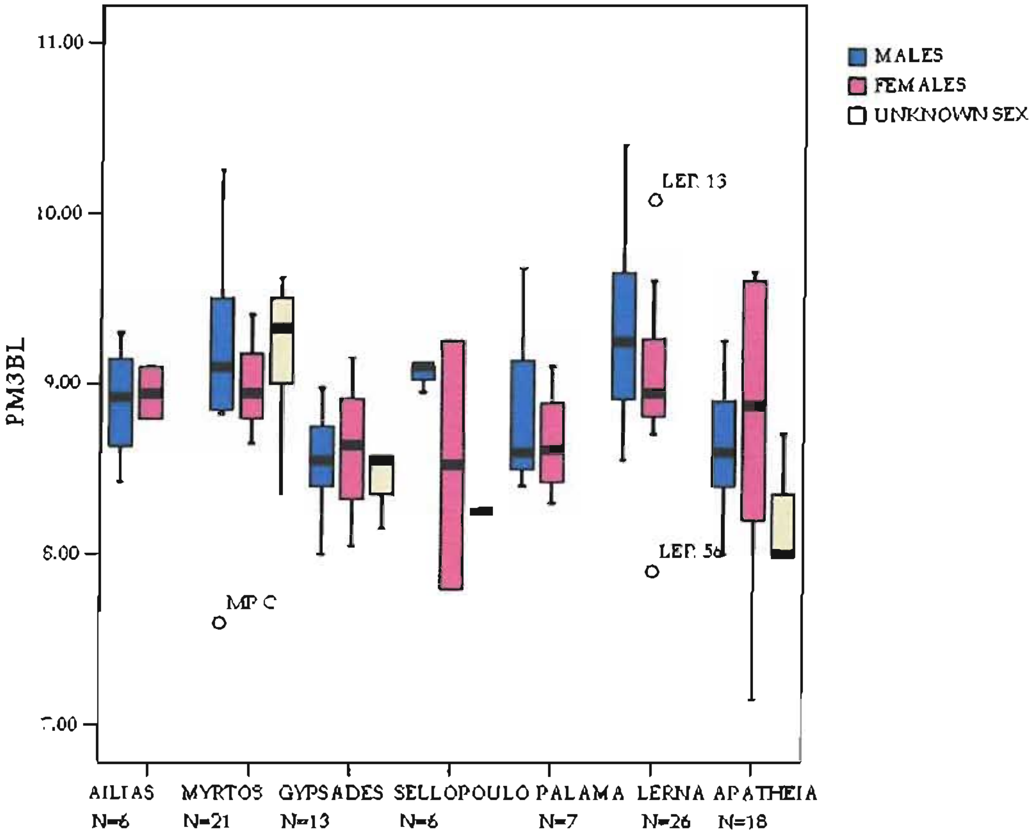


Figure 72 Box and whisker plot of MD crown diameter of 3rd Premolar, Upper, R and L sides pooled, Cretan and Argolid samples.

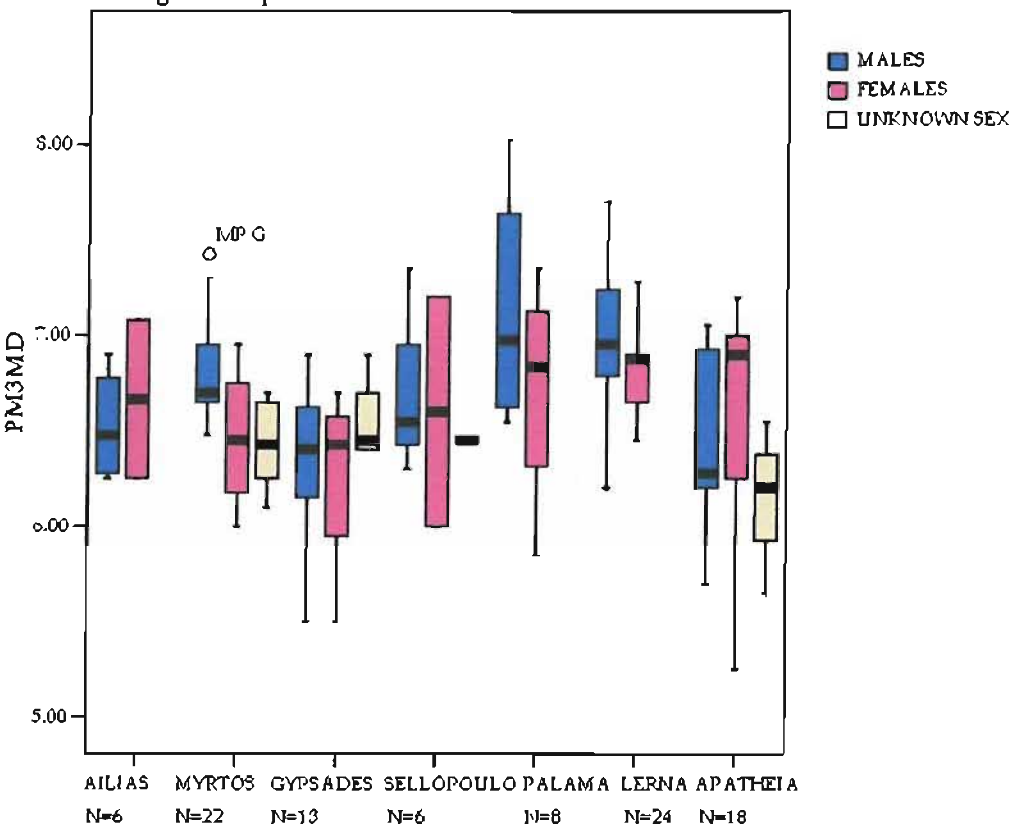


Figure 73 Box and whisker plot of BL crown diameter of 4th Premolar, Upper, R and L sides pooled, Cretan and Argolid samples.

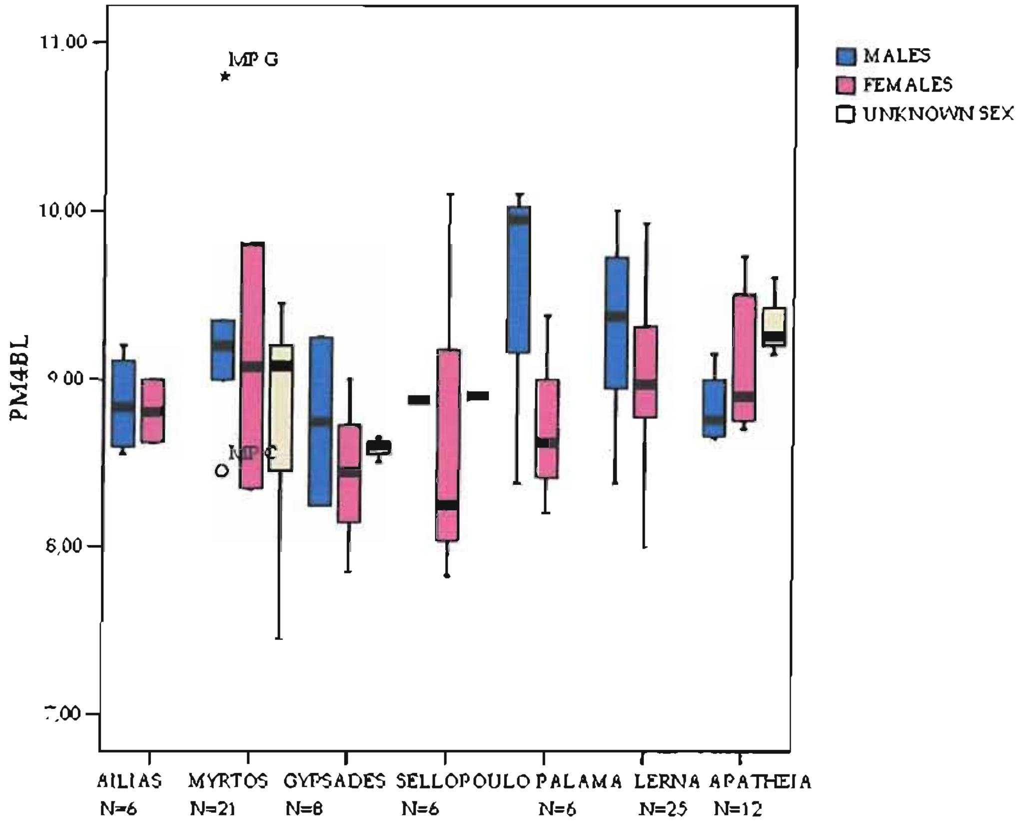


Figure 74 Box and whisker plot of MD crown diameter of 4th Premolar, Upper, R and L sides pooled, Cretan and Argolid samples.

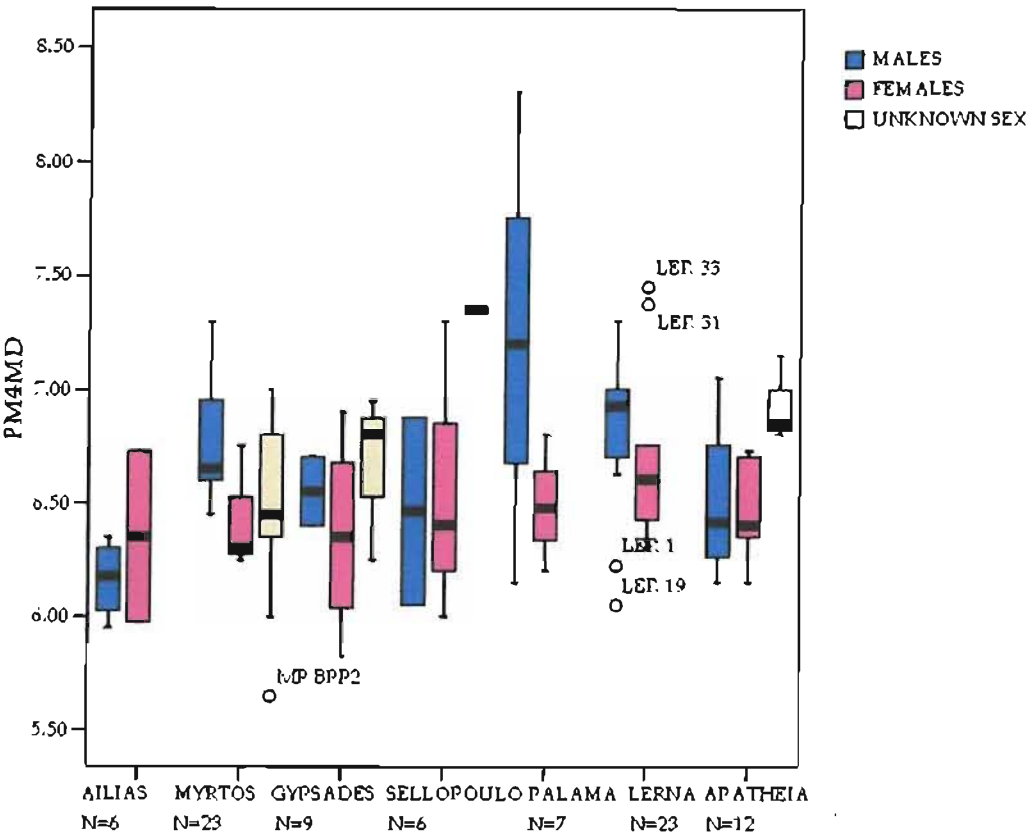


Figure 75 Box and whisker plot of BL crown diameter of 1st Molar, Upper, R and L sides pooled, Cretan and Argolid samples.

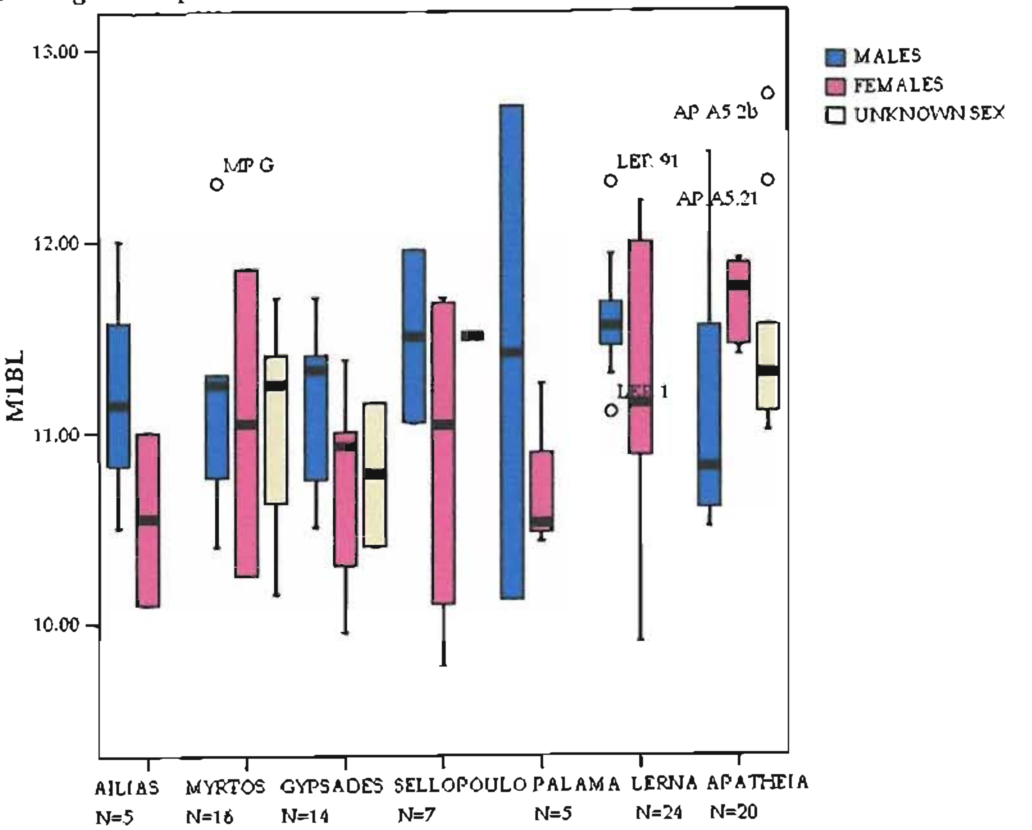


Figure 76 Box and whisker plot of MD crown diameter of 1st Molar, Upper, R and L sides pooled, Cretan and Argolid samples.

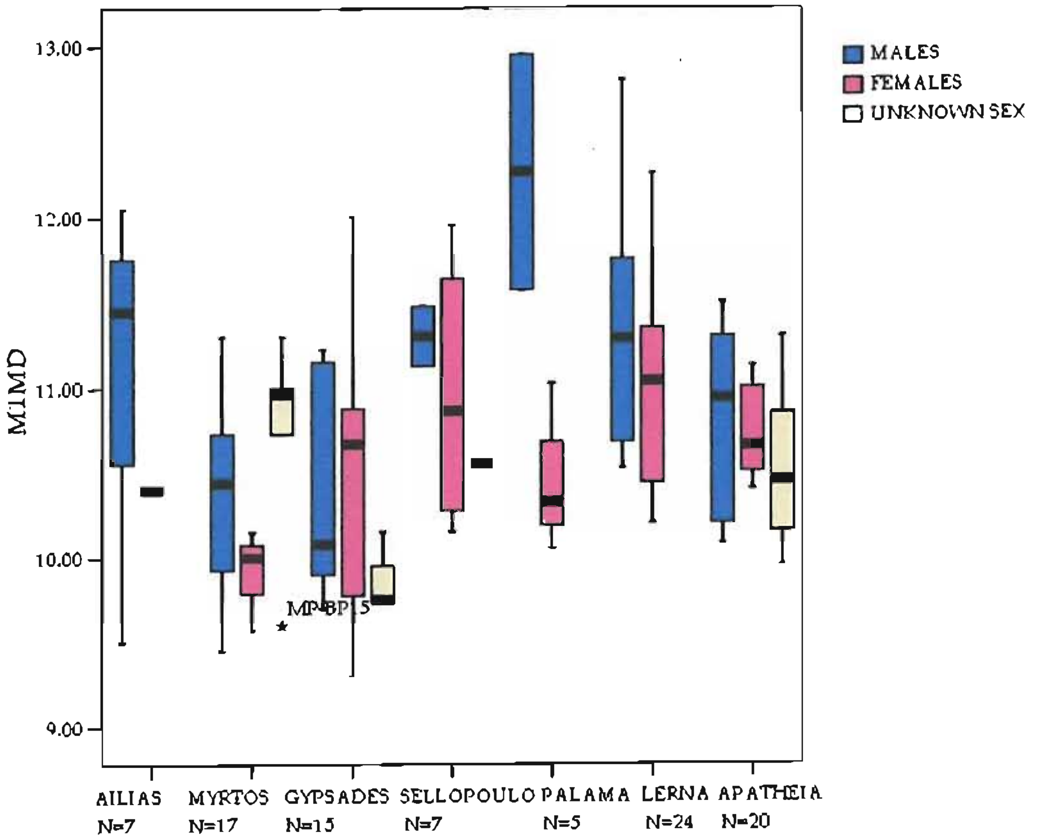


Figure 77 Box and whisker plot of BL crown diameter of 2nd Molar, Upper, R and L sides pooled, Cretan and Argolid samples.

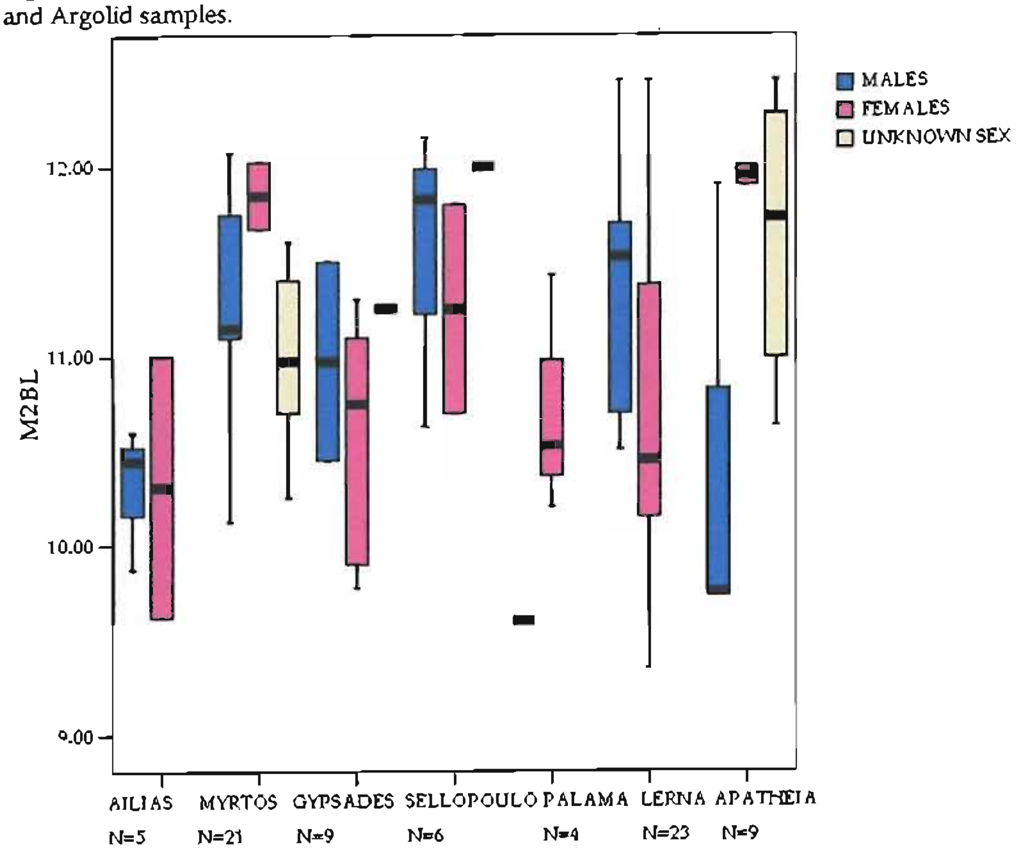


Figure 78 Box and whisker plot of MD crown diameter of 2nd Molar, Upper, R and L sides pooled, Cretan and Argolid samples.

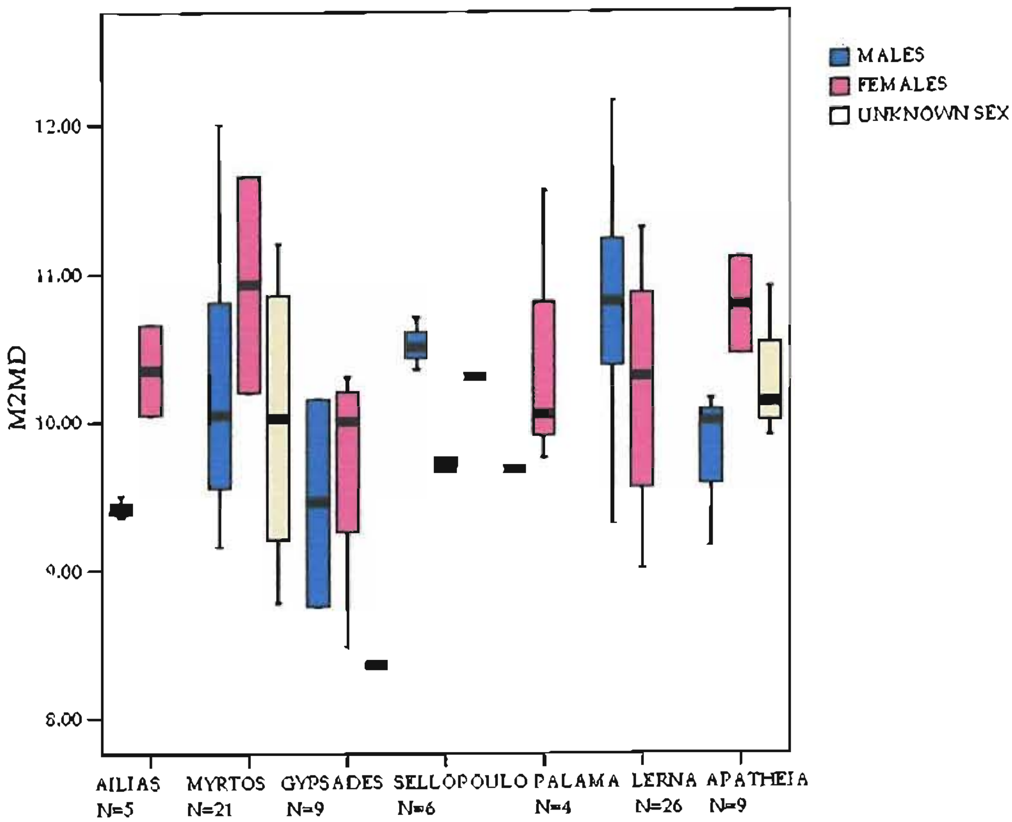


Figure 79 Box and whisker plot of BL crown diameter of 3rd Molar, Upper, R and L sides pooled, Cretan and Argolid samples.

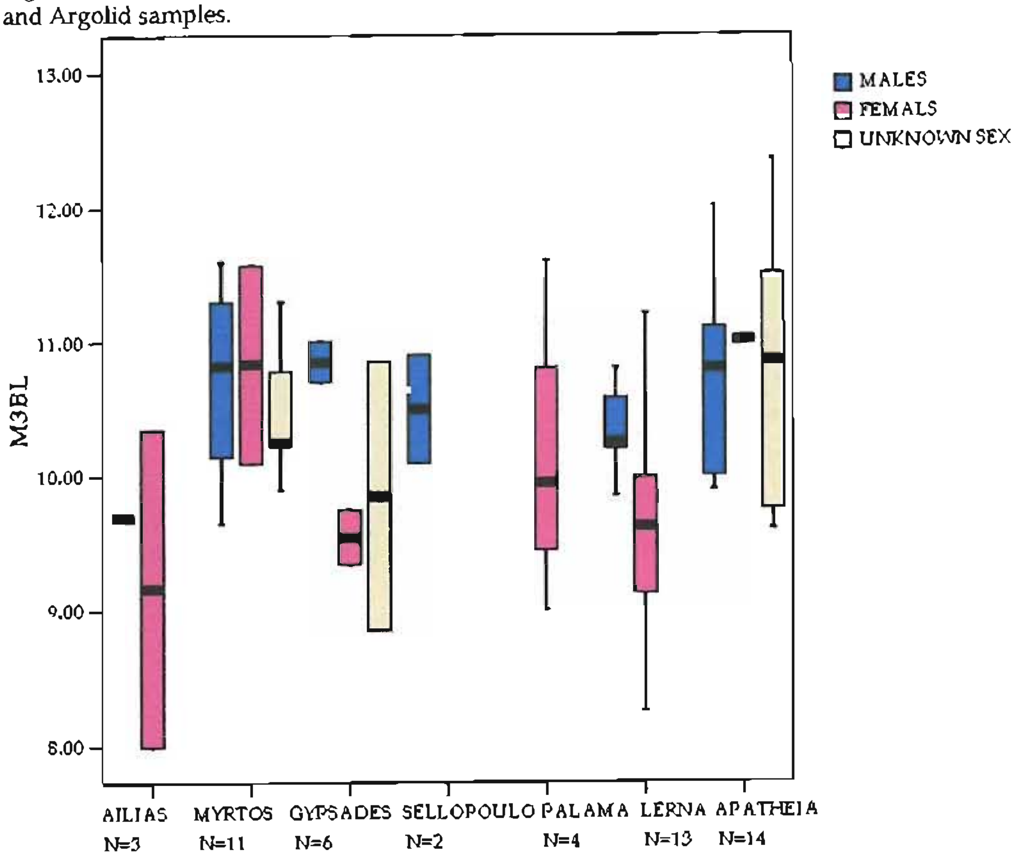
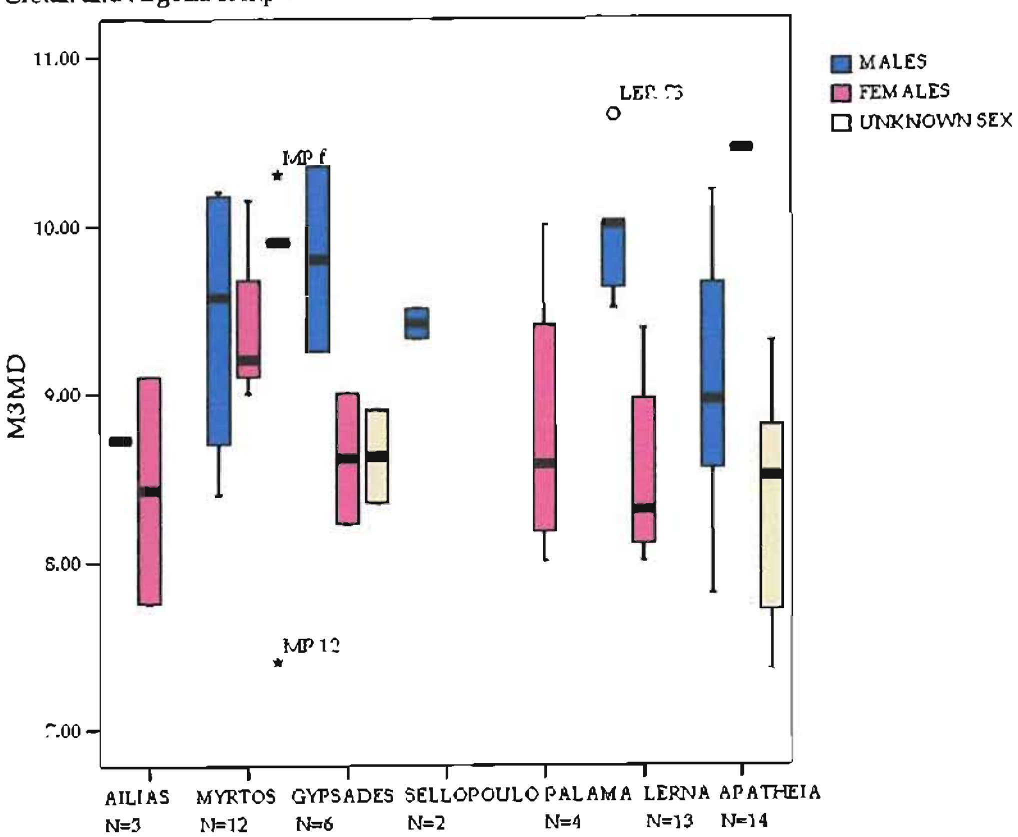


Figure 80 Box and whisker plot of MD crown diameter of 3rd Molar, Upper, R and L sides pooled, Cretan and Argolid samples.



Mandibular teeth, Right and Left sides Pooled

Figure 81 Box and whisker plot of BL crown diameter of 1st Incisor, Lower, R and L sides pooled, Cretan and Argolid samples.

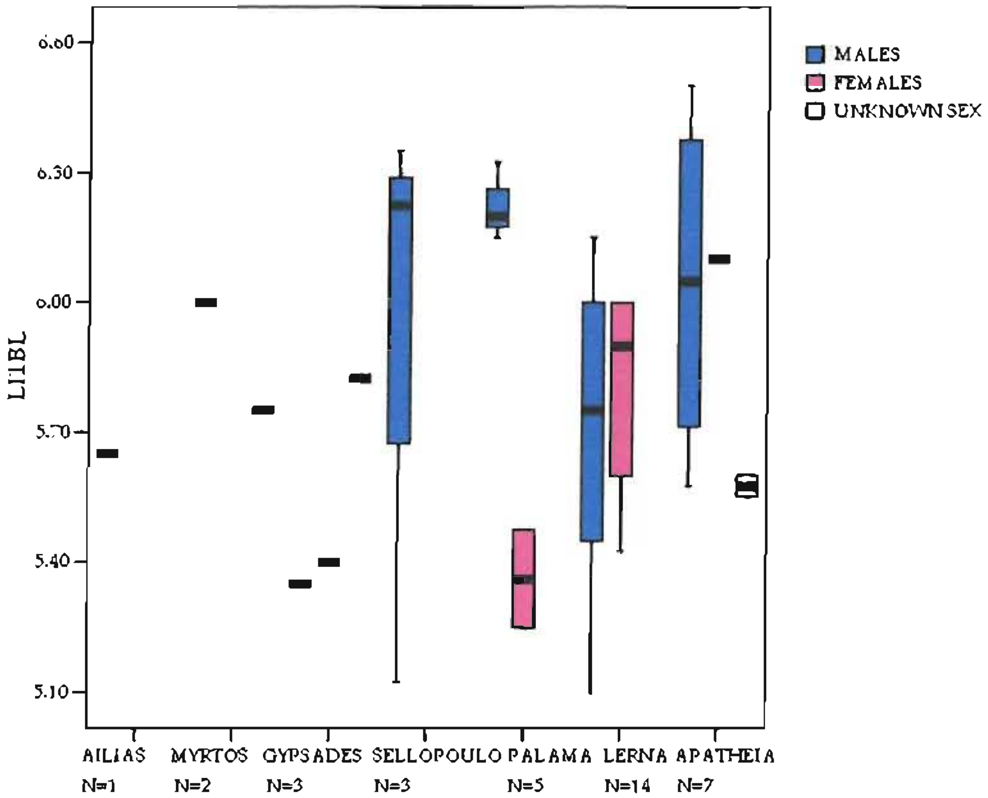


Figure 82 Box and whisker plot of MD crown diameter of 1st Incisor, Lower, R and L sides pooled, Cretan and Argolid samples.

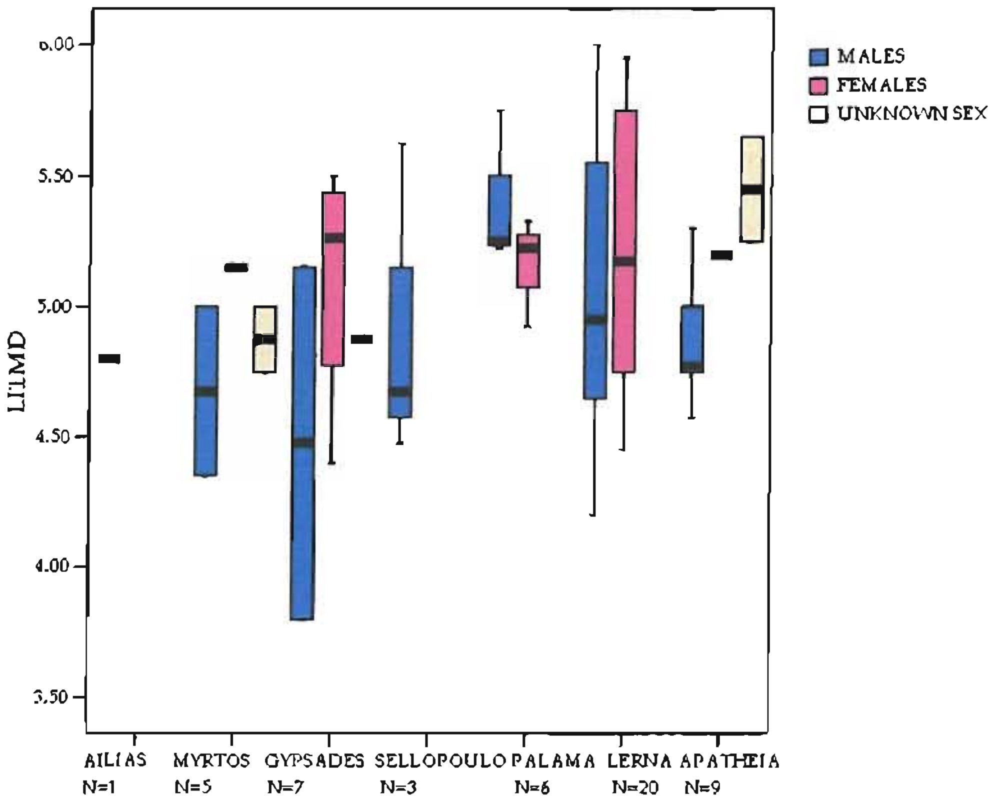


Figure 83 Box and whisker plot of BL crown diameter of 2nd Incisor, Lower, R and L sides pooled, Cretan and Argolid samples.

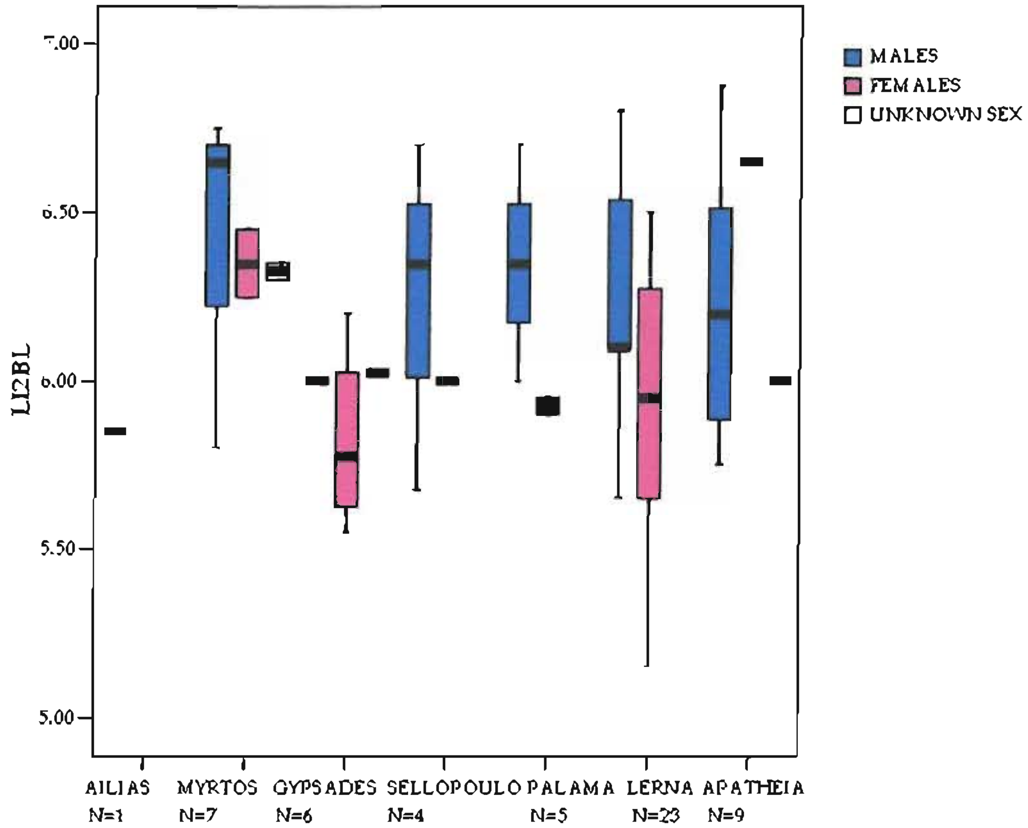


Figure 84 Box and whisker plot of MD crown diameter of 2nd Incisor, Lower, R and L sides pooled, Cretan and Argolid samples.

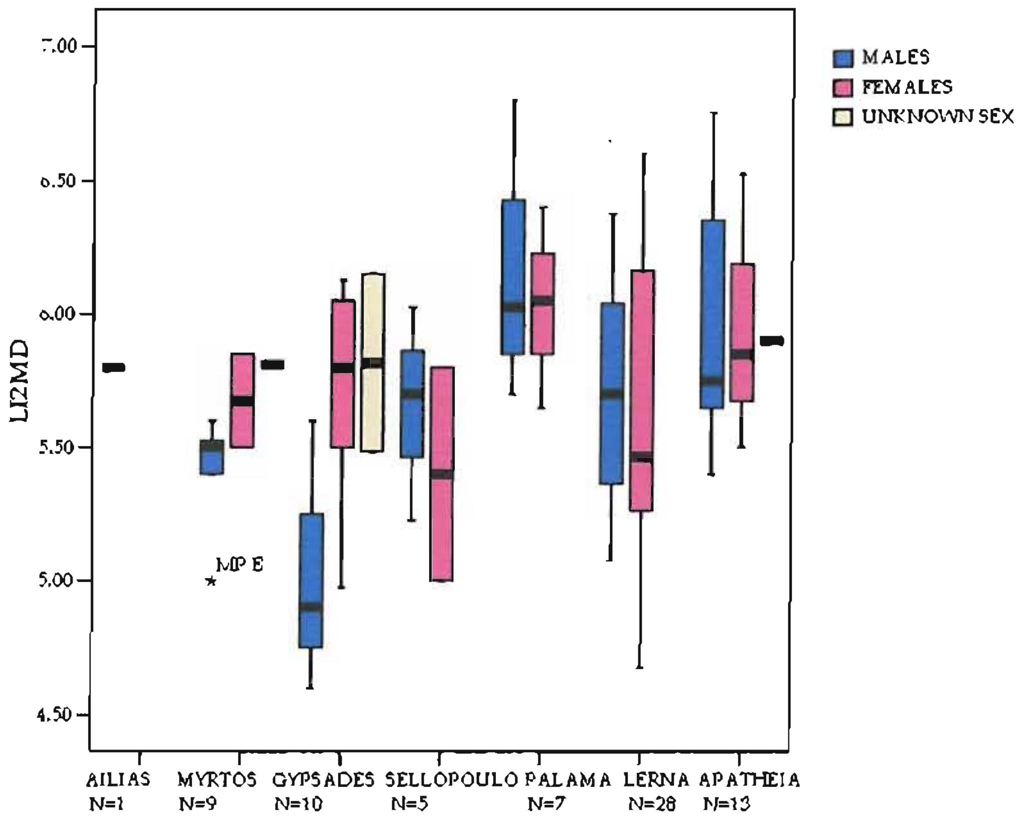


Figure 85 Box and whisker plot of BL crown diameter of Canine, Lower, R and L sides pooled, Cretan and Argolid samples.

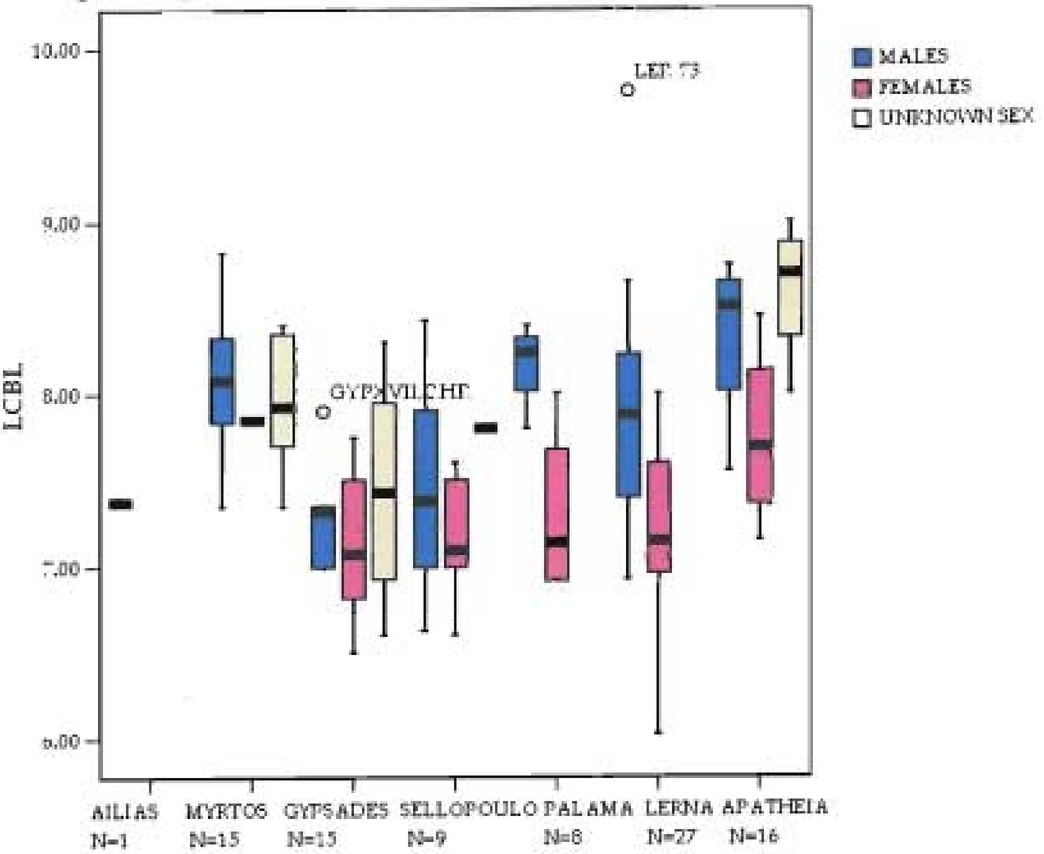


Figure 86 Box and whisker plot of MD crown diameter of Canine, Lower, R and L sides pooled, Cretan and Argolid samples.

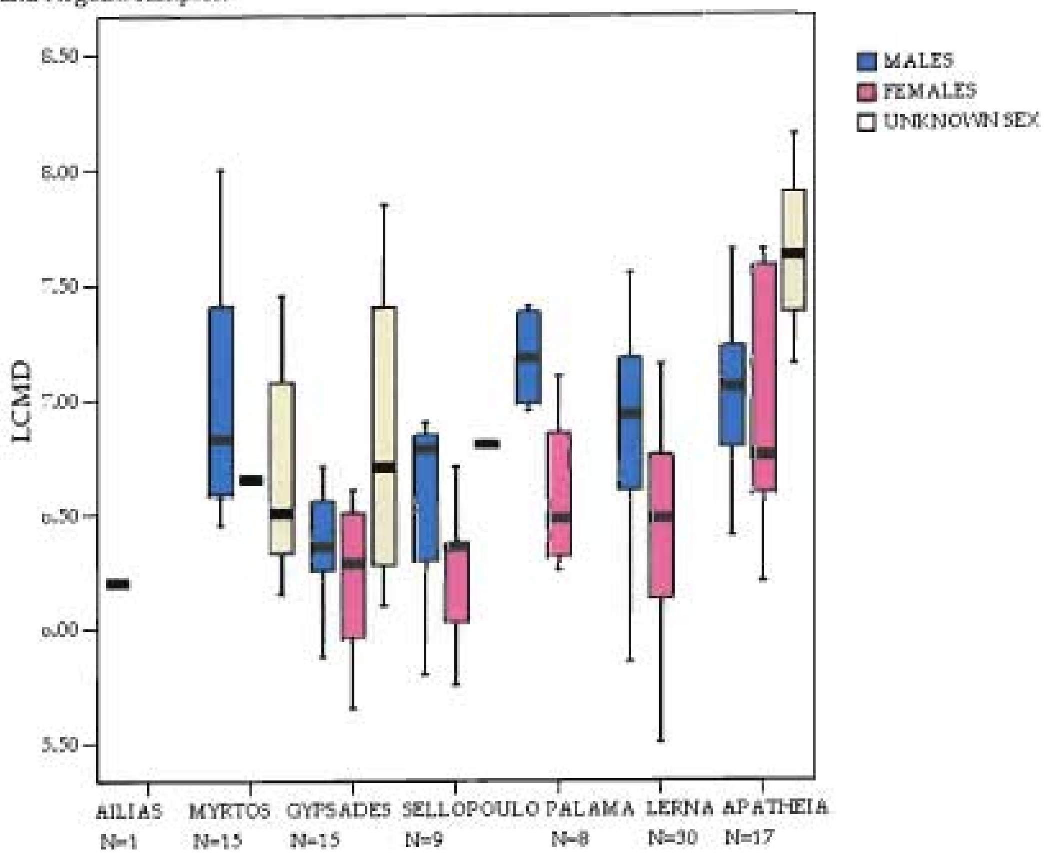


Figure 87 Box and whisker plot of BL crown diameter of 3rd Premolar, Lower, R and L sides pooled, Cretan and Argolid samples.

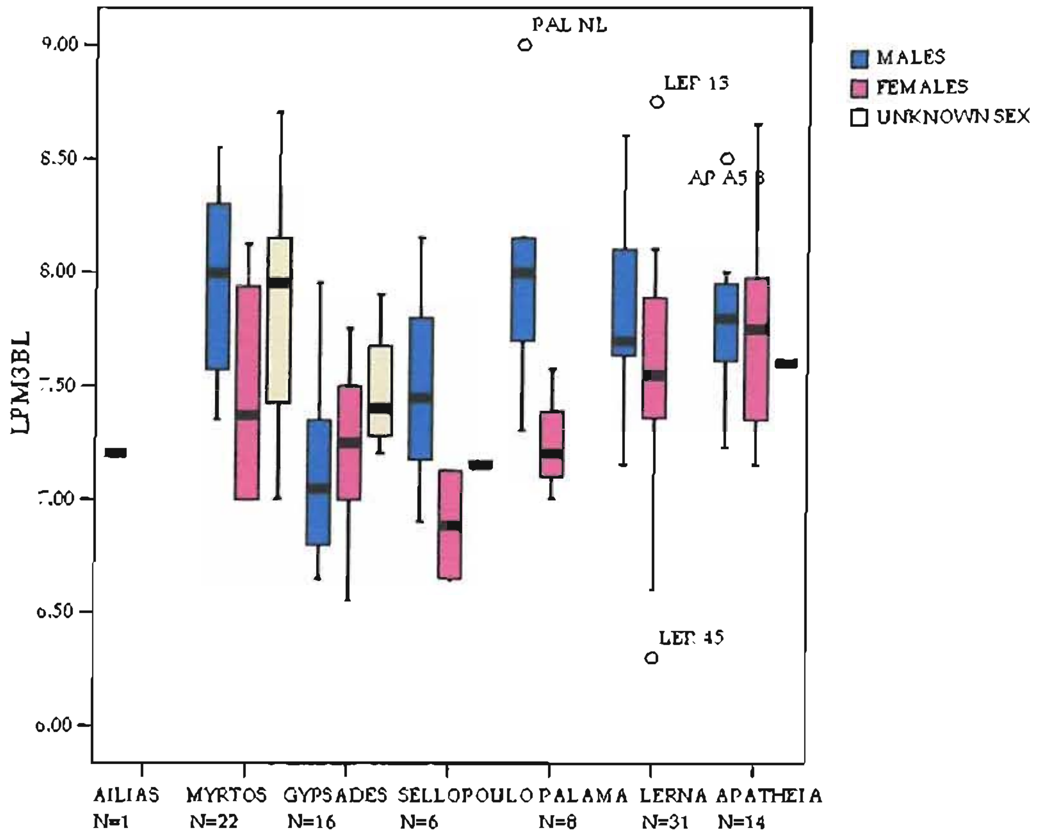


Figure 88 Box and whisker plot of MD crown diameter of 3rd Premolar, Lower, R and L sides pooled, Cretan and Argolid samples.

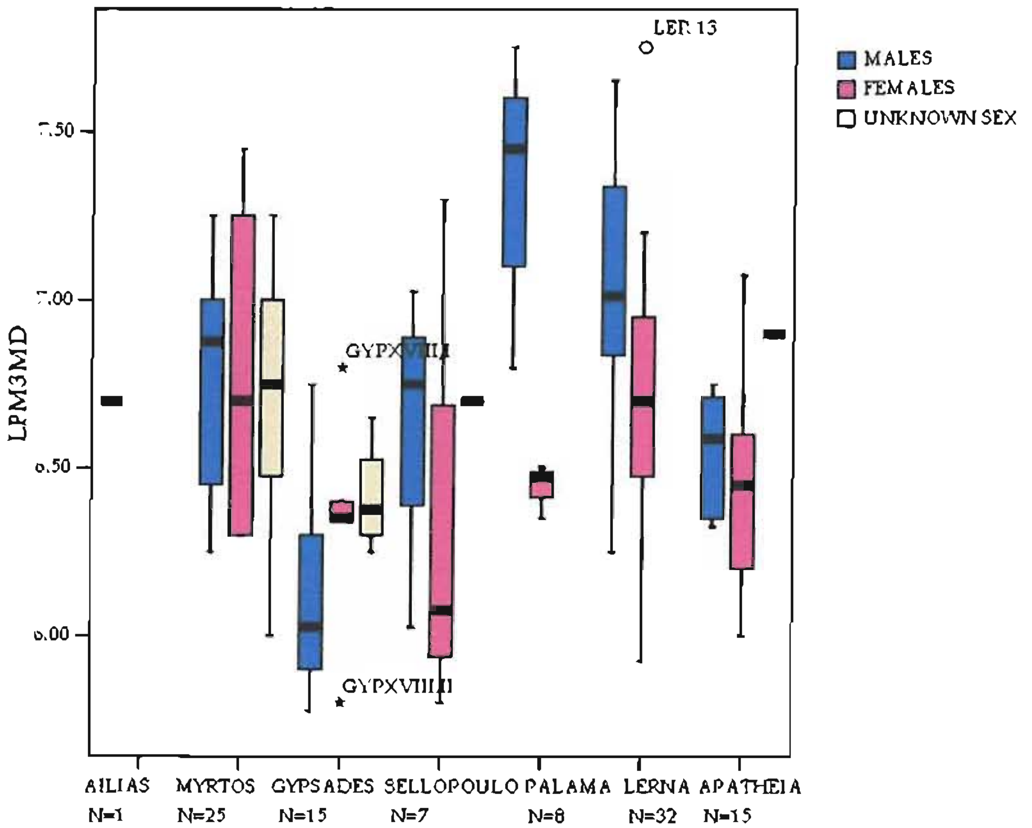


Figure 89 Box and whisker plot of BL crown diameter of 4th Premolar, Lower, R and L sides pooled, Cretan and Argolid samples.

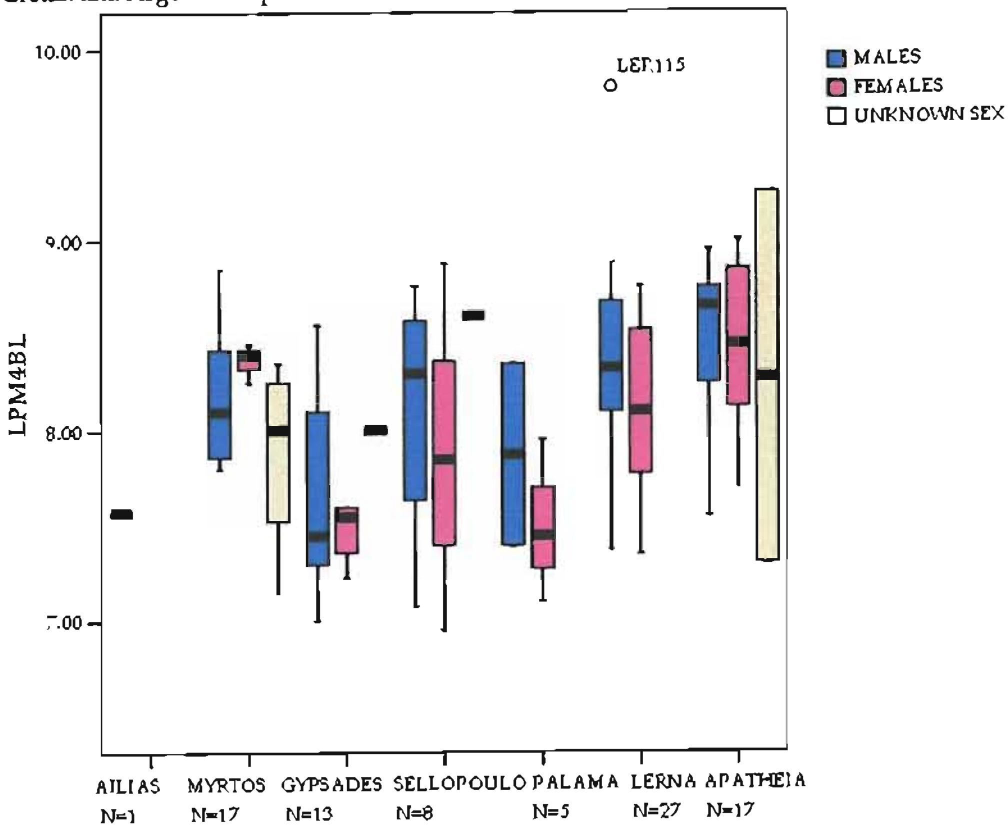


Figure 90 Box and whisker plot of MD crown diameter of 4th Premolar, Lower, R and L sides pooled, Cretan and Argolid samples.

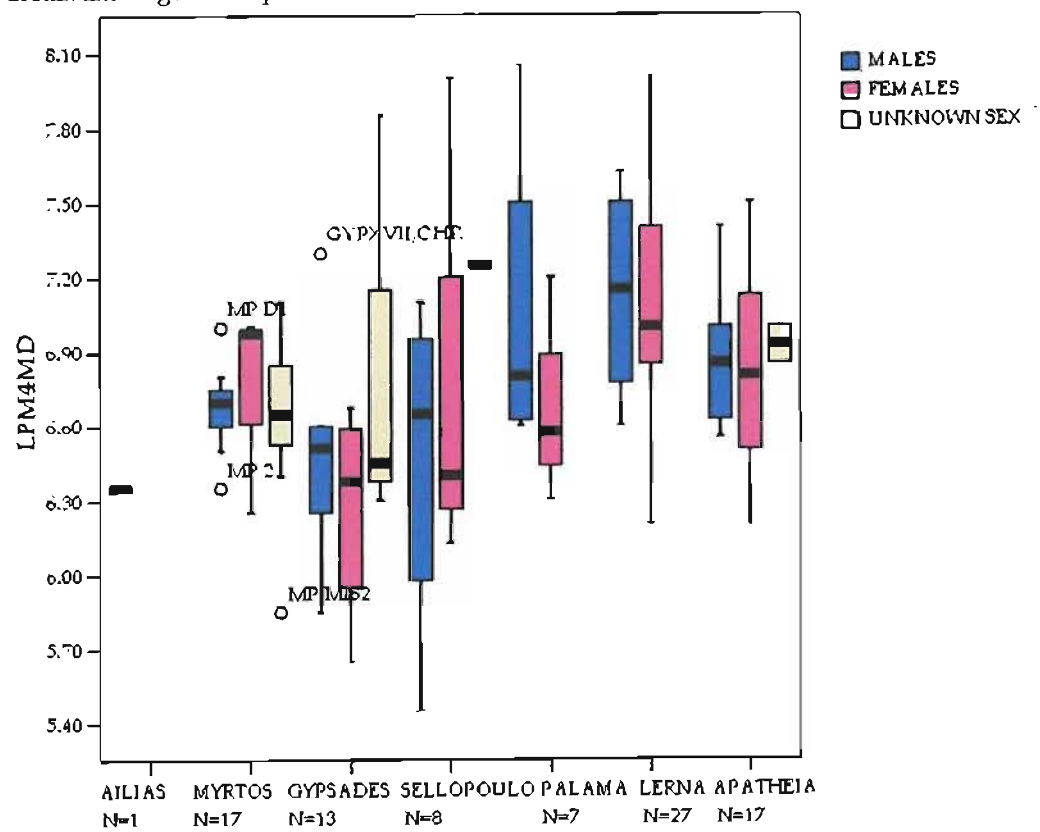


Figure 91 Box and whisker plot of BL crown diameter of 1st Molar, Lower, R and L sides pooled, Cretan and Argolid samples.

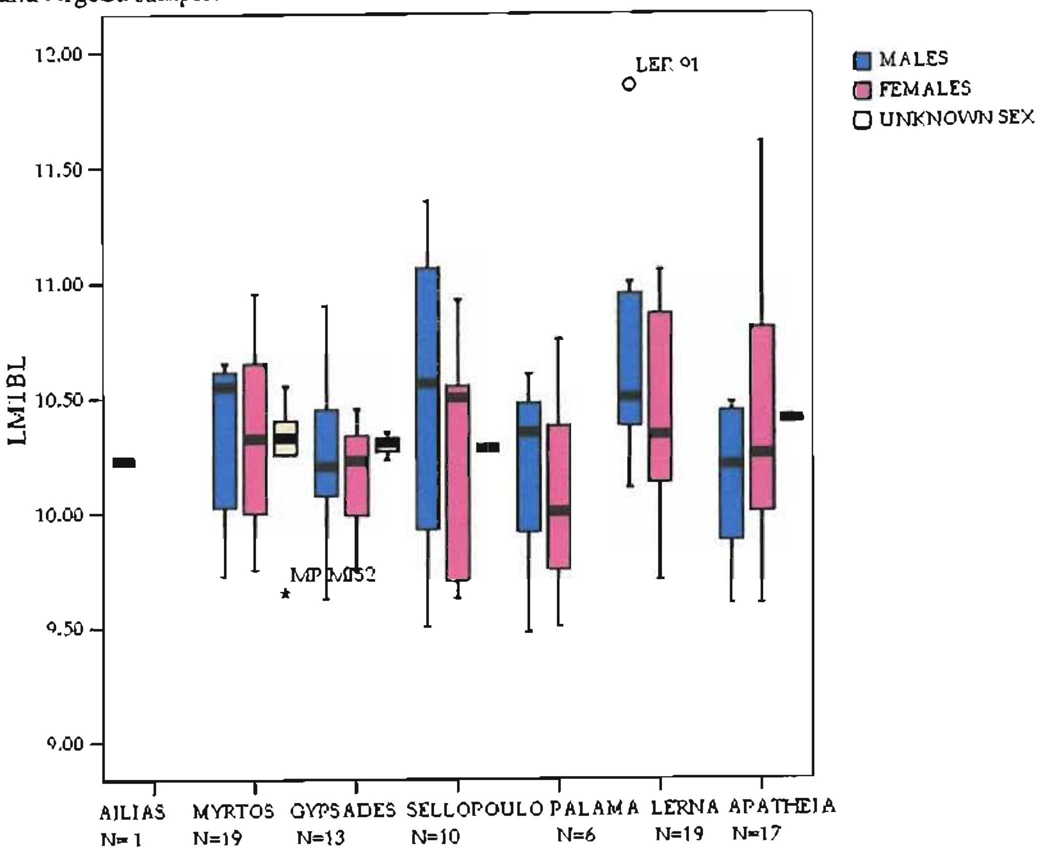


Figure 92 Box and whisker plot of MD crown diameter of 1st Molar, Lower, R and L sides pooled, Cretan and Argolid samples.

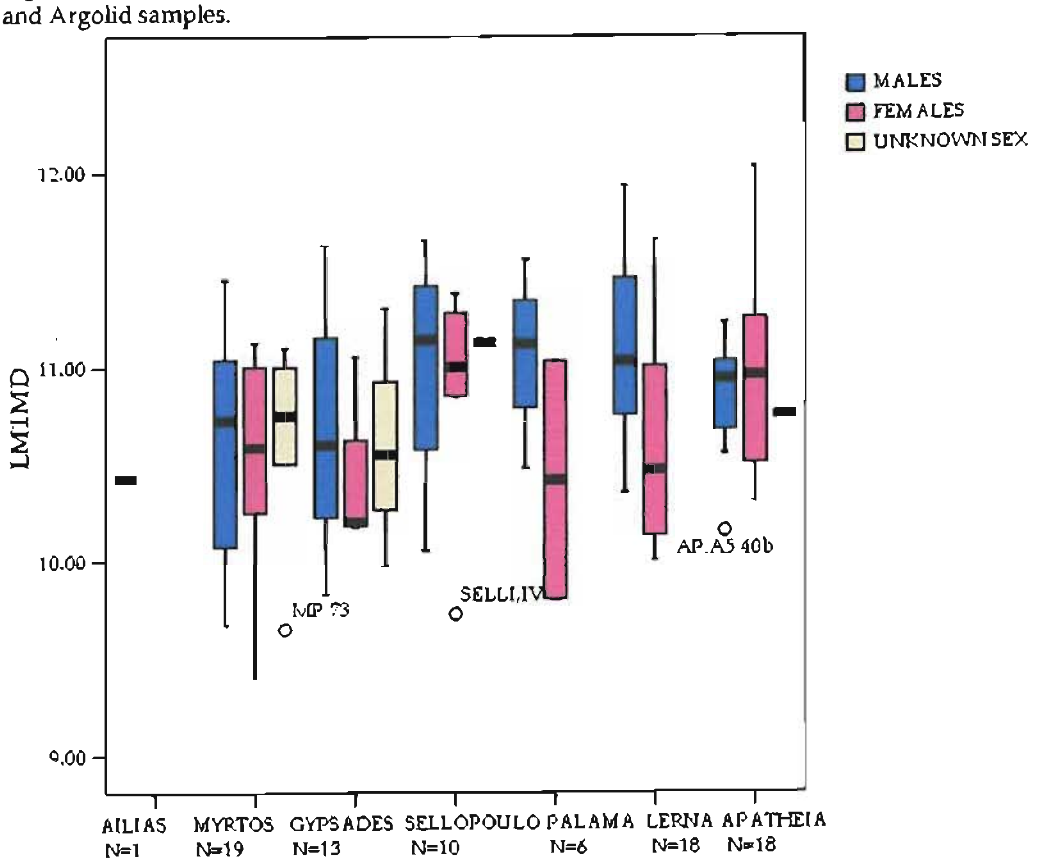


Figure 93 Box and whisker plot of BL crown diameter of 2nd Molar, Lower, R and L sides pooled, Cretan and Argolid samples.

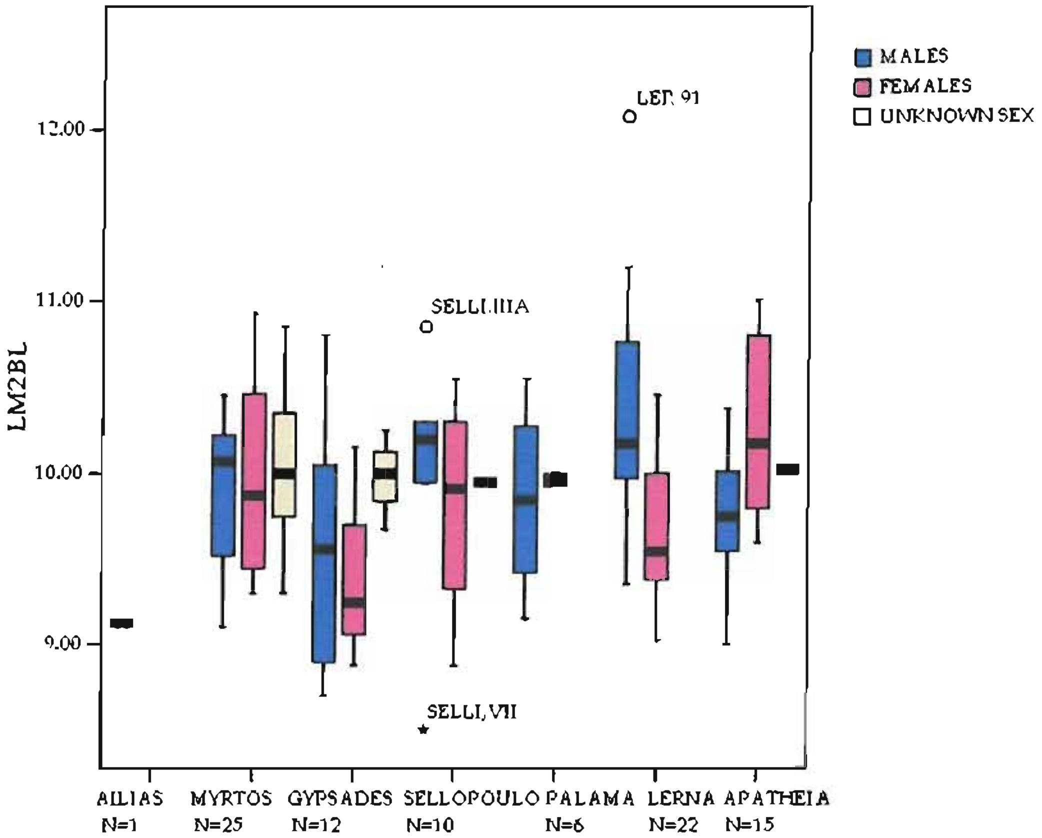


Figure 94 Box and whisker plot of MD crown diameter of 2nd Molar, Lower, R and L sides pooled, Cretan and Argolid samples.

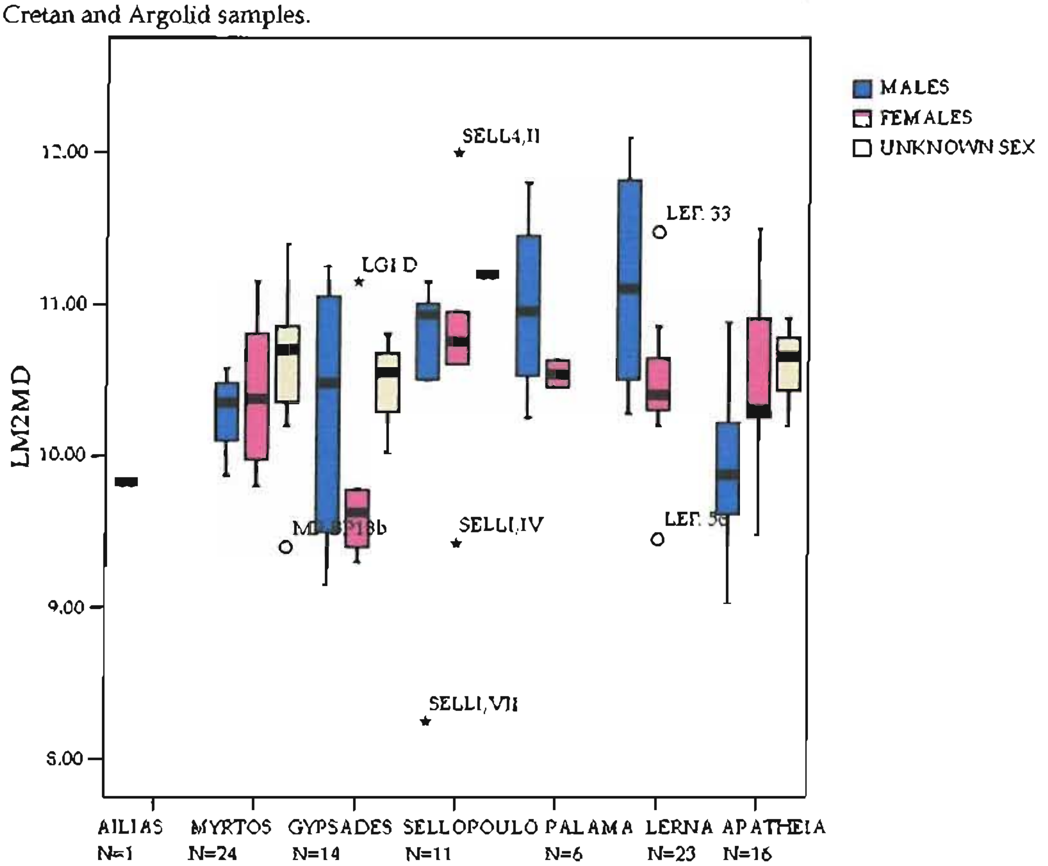


Figure 95 Box and whisker plot of BL crown diameter of 3rd Molar, Lower, R and L sides pooled, Cretan and Argolid samples.

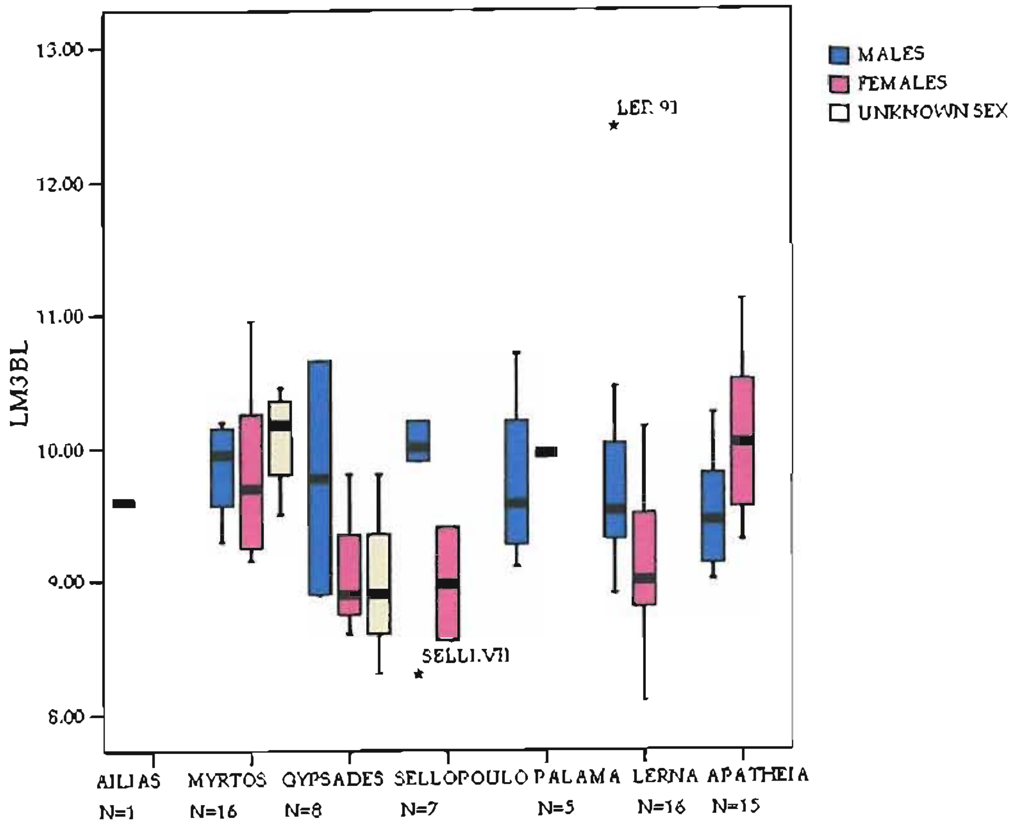
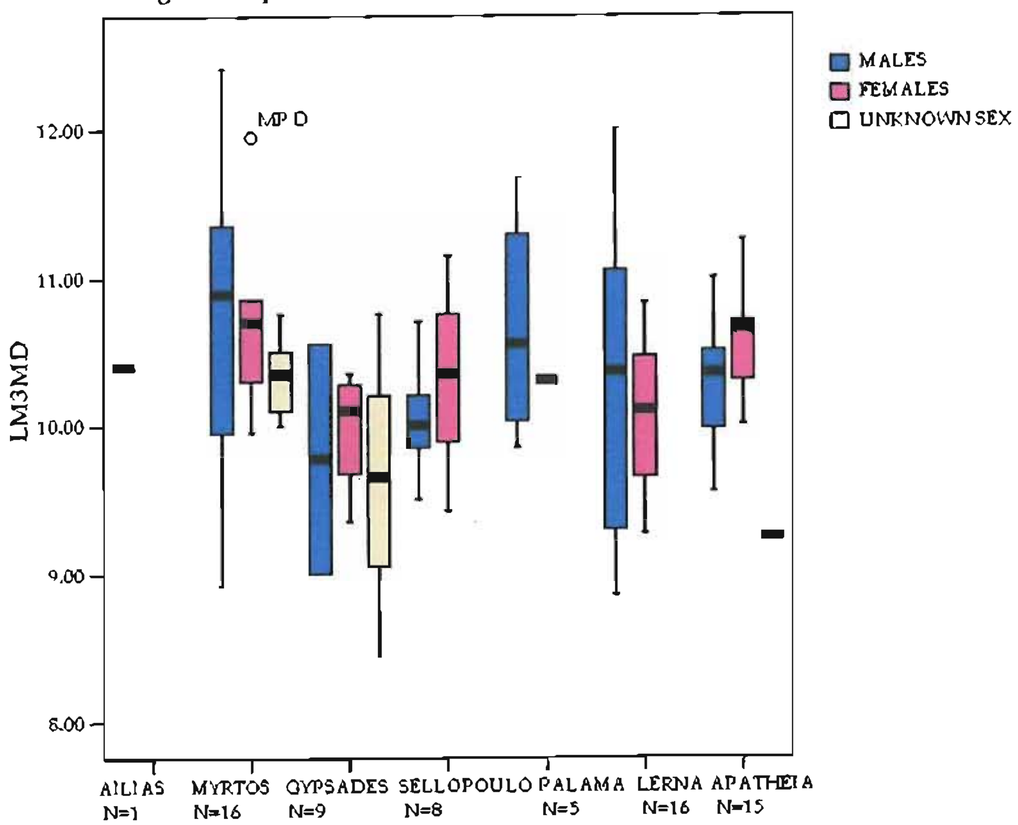


Figure 96 Box and whisker plot of MD crown diameter of 3rd Molar, Lower, R and L sides pooled, Cretan and Argolid samples.



APPENDIX E
NON-METRIC TRAITS:
FREQUENCIES (%) AND NUMBER (N) OF INDIVIDUALS FOR ALL TESTED SAMPLES

Table 6.1 Percentages (%) of cranial non-metric traits and number of individuals (N) scored for Lerna and Apatheia samples.

CRANIAL NON-METRIC TRAITS	LERNA		APATHEIA	
	Lside	Rside	Lside	Rside
Highest Nuchal Line present	% 65.3 N 26	% 66.6 N 27	% 92.8 N 14	% 82.3 N 17
Ossicle at Lambda	% 11.5 N 26		% 21.7 N 23	
Lambdoid ossicle present	% 21.7 N 23	%20.8 N 24	% 38.8 N 18	% 46.15 N 13
Parietal foramen present	% 42.3 N 26	% 28 N 25	% 50 N 20	% 40 N 20
Bregmatic bone present	% 0 N 32		% 0 N 24	
Metopic suture	% 14.7 N 34		% 8 N 25	
Coronal ossicle present	% 0 N 23	% 0 N 28	% 0 N 14	% 0 N 11
Epipteric bone present	% 0 N 26	% 0 N 26	% 0 N 11	% 0 N 13
Fronto-temporal articulation	% 4.5 N 22	% 5 N 20	% 0 N 13	% 0 N 12
Parietal notch bone present	% 0 N 20	% 0 N 21	% 0 N 17	% 0 N 11
Ossicle at asterion	% 5.2 N 19	% 4.3 N 23	% 16.6 N 18	% 0 N 12
Auditory torus present	% 3.7 N 27	% 6 N 33	% 0 N 16	% 0 N 19
Foramen of Huschke present	% 86.9 N 23	% 87.5 N 24	% 92.8 N 14	% 93.7 N 16
Mastoid foramen exsutural	% 76 N 25	% 64.2 N 28	% 77.7 N 18	% 78.9 N 19
Mastoid foramen absent	% 72.2 N 18	% 82.3 N 17	% 41.6 N 12	% 22.2 N 9
Posterior condylar canal patent	% 0 N 4	% 0 N 1	% 0 N 1	% 50 N 2
Condylar facet double	% 0 N 16	% 0 N 14	% 20 N 5	% 0 N 7
Pharyngeal tubercle present	% 61.5 N 13		% N 0	
Precondylar tubercle present	% N 0	% N 0	% N 0	% N 0
Anterior condylar canal double	% 31.2 N 16	% 2 N 10	% 50 N 8	% 37.5 N 8
Foramen ovale incomplete	% 0 N 14	% 11.1 N 9	% 0 N 8	% 0 N 8
Foramen spinosum open	% 9 N 11	%0 N 9	% 0 N 6	% 0 N 6
Accessory lesser palatine foramen present	% N 0	% 84.6 N 13	% N 0	% 100 N 3
Palatine torus present	% N 0	% N 0	% N 0	% N 0
Maxillary torus present	% 3.8 N 26	% 8.6 N 23	% 0 N 4	% 0 N 4

CRANIAL NON-METRIC TRAITS	LERNA		APATHEIA	
	Lside	Rside	Lside	Rside
Maxillary bridge present	% N 0	% N 0	% N 0	% N 0
Zygomatic – facial foramen absent	% 10.7 N 28	% 23.3 N 30	% 25 N 12	% 7.6 N 13
Supraorbital foramen complete	% 33.3 N 27	% 26.6 N 30	% 9.5 N 21	% 20 N 20
Frontal notch / foramen present	% 77.7 N 27	% 75.8 N 29	% 80 N 20	% 84 N 19
Anterior ethmoid foramen exsutural	% 80 N 5	% 100 N 1	% 100 N 1	% 33.3 N 3
Posterior ethmoid foramen absent	% N 0	% N 0	% N 0	% N 0
Accessory infraorbital foramen present	% 100 N 24	% 100 N 25	% 50 N 2	% 100 N 3
Inca bone	% 9.3 N 32		% 0 N 22	
Infraorbital suture	% 15.3 N 13	% 31.5 N 19	% 0 N 2	% 20 N 5
Nasal foramen	% 100 N 17	% 100 N 13	% 100 N 4	% 100 N 6
Trochlear spur	% N 0	% N 0	% N 0	% N 0
Trochlear fossa	% N 0	% N 0	% N 0	% N 0
Squamo- parietal ossicles	% 0 N 22	% 0 N 21	% 0 N 13	% 0 N 13
Processus marginalis	% N 0	% N 0	% N 0	% 0 N 0
Zygomatico-temporal foramen	% 7.1 N 14	% 0 N 10	% 0 N 6	% 0 N 8
Occipito mastoid ossicle	% 0 N 22	% 0 N 19	% 0 N 12	% 0 N 10
Intermediate condylar canal	% N 0	% N 0	% 33.3 N 3	% 0 N 1
Post condylar tubercle	% 53.3 N 15	% 57.1 N 7	% 75 N 4	% 50 N 4
Jugular foramen bridge	% 0 N 21	% 3.8 N 26	% 0 N 6	% 0 N 6
Foramen of vesalius	% N 0	% N 0	% N 0	% N 0
Pterygo-basal bridge	% 38.8 N 18	% 41.6 N 12	% 0 N 8	% 0 N 5
Pterygo-spinous bridge	% 0 N 15	% 10 N 10	% 33.3 N 9	% 33.3 N 6
Spino-basal bridge	% 11.7 N 17	% 18.1 N 11	% 0 N 5	% 0 N 3
Foramen ovale spine	% N 0	% N 0	% N 0	% N 0
Accessory foramen spinosum	% 0 N 14	% 0 N 7	% 14.2 N 7	% 0 N 5
Lateral pterygoid perforated	% N 0	% N 0	% N 0	% N 0
Pterygoid spurs	% N 0	% N 0	% N 0	% N 0
Palatine bridge	% N 0	% N 0	% N 0	% N 0
Zygomatico- facial foramen multiple	% 73.9 N 23	% 44.4 N 27	% 58.3 N 12	% 81.81 N 11

Table 6.2 Percentages of cranial non-metric traits and number of individuals scored (n) for Moni, Palaikastro, Ailias and Myrtos Pyrgos samples.

CRANIAL NON-METRIC TRAITS	MONI		PALAIKASTRO		AILIAS		MYRTOS PYRGOS	
	Lside	Lside	Lside	Lside	Lside	Lside	Rside	Lside
Highest Nuchal Line present	% 58.8 N 17	% 64.7 N 17	% 100 N 22	% 100 N 22	% 58.6 N 58	% 59.01 N 61	% 66.6 N 21	% 69.5 N 23
Ossicle at Lambda	% 15.7 N 19		% 16.6 N 24		% 14.7 N 61		% 3.8 N 26	
Lambdoid ossicle present	% 7.1 N 14	% 35.7 N 14	% 10.5 N 19	% 63.6 N 22	% 30.6 N 49	% 31.25 N 48	% 33 N 21	% 10.5 N 19
Parietal foramen present	% 60 N 20	% 50 N 20	% 63.6 N 22	% 61.9 N 21	% 33.8 N 65	% 41.9 N 62	% 68 N 25	% 60.8 N 23
Bregmatic bone present	% 0 N 19		% 4 N 25		% 0 N 68		% 0 N 26	
Metopic suture	% 0 N 18		% 8.3 N 24		% 2.8 N 70		% 9 N 33	
Coronal ossicle present	% 0 N 18	% 0 N 17	% 0 N 21	% 0 N 23	% 1.6 N 59	% 3.4 N 58	% 0 N 19	% 0 N 16
Epipteric bone present	% 0 N 17	% 0 N 15	% 0 N 19	% 0 N 23	% 0 N 56	% 0 N 52	% 0 N 19	% 0 N 16
Fronto-temporal articulation	% 0 N 8	% 0 N 12	% 0 N 14	% 0 N 13	% 5.1 N 39	% 6.6 N 30	% 0 N 9	% 11.1 N 9
Parietal notch bone present	% 11.1 N 9	% 11.1 N 9	% 29.4 N 17	% 6 N 15	% 5.4 N 37	% 6.4 N 31	% 0 N 13	% 0 N 10
Ossicle at asterion	% 8.3 N 12	% 15.3 N 13	% 5.2 N 19	% 5.2 N 19	% 2.3 N 43	% 4.4 N 45	% 0 N 18	% 0 N 14
Auditory torus present	% 0 N 14	% 0 N 14	% 0 N 17	% 0 N 22	% 2.5 N 39	% 0 N 38	% 18.1 N 22	% 12.5 N 24
Foramen of Huschke present	% 93.3 N 15	% 93.3 N 15	% 100 N 14	% 100 N 20	% 94.7 N 38	% 91.8 N 37	% 100 N 20	% 100 N 22
Mastoid foramen exsutural	% 87.5 N 16	% 87.5 N 16	% 58.8 N 17	% 57.8 N 19	% 75 N 36	% 69.4 N 36	% 71.4 N 21	% 84.21 N 19
Mastoid foramen absent	% 77.7 N 9	% 63.6 N 11	% 33.3 N 9	% 35.7 N 14	% 64.7 N 17	% 68.1 N 22	% 37.5 N 8	% 57.14 N 7

CRANIAL NON-METRIC TRAITS	MONI		PALAIKASTRO		AILIAS		MYRTOS PYRGOS	
	Lside	Lside	Lside	Lside	Lside	Lside	Rside	Lside
Posterior condylar canal patent	% N 0	% N 0	% N 0	% N 0	% 60 N 5	% 40 N 5	% 66.6 N 3	% 75 N 4
Condylar facet double	% 0 N 4	% 0 N 1	% 0 N 2	% 0 N 2	% 0 N 8	% 0 N 8	% 0 N 14	% 0 N 13
Pharyngeal tubercle present	% 100 N 9		% 100 N 2		% 81.8 N 11		% 91.6 N 12	
Precondylar tubercle present	% N -	% 20 N 5	% N 0	% 0 N 1	% 33.3 N 9	% 11.1 N 9	% 23 N 13	% 33.3 N 15
Anterior condylar canal double	% 33.3 N 6	% 14.2 N 7	% 0 N 2	% 0 N 2	% 20 N 10	% 33.3 N 9	% 40 N 20	% 35.2 N 17
Foramen ovale incomplete	% 0 N 5	% 0 N 6	% 0 N 5	% 0 N 6	% 0 N 9	% 0 N 12	% 0 N 11	% 0 N 8
Foramen spinosum open	% 0 N 4	% 33.3 N 6	% 20 N 5	% 0 N 7	% 33.3 N 9	% 42.8 N 7	% 0 N 7	% 12.5 N 8
Accessory lesser palatine foramen present	% 50 N 2	% 50 N 2	% 100 N 1	% 0 N 1	% 100 N 6	% 100 N 3	% 66.6 N 3	% 50 N 4
Palatine torus present	% 20 N 5	% 20 N 5	% 0 N 2	% 0 N 1	% 11.1 N 9	% 11.1 N 9	% 18.1 N 11	% 18.1 N 11
Maxillary torus present	% 0 N 3	% 0 N 3	% 0 N 1	% 0 N 2	% 0 N 12	% 0 N 9	% 14.2 N 7	% 25 N 8
Maxillary bridge present	% 66.6 N 3	% 75 N 4	% 100 N 2	% 0 N 1	% 60 N 10	% 50 N 8	% 87.5 N 8	% 100 N 8
Zygomatic – facial foramen absent	% 0 N 8	% 0 N 8	% 0 N 2	% 0 N 4	% 0 N 13	% 0 N 9	% 0 N 23	% 0 N 22
Supraorbital foramen complete	% 20 N 15	% 41.6 N 12	% 11.7 N 17	% 20 N 20	% 33.3 N 51	% 19.1 N 47	% 33.3 N 27	% 22.7 N 22
Frontal notch / foramen present	% 93.3 N 15	% 92.3 N 13	% 95 N 20	% 89.4 N 19	% 86 N 50	% 85.7 N 49	% 79.3 N 29	% 79.16 N 24
Anterior ethmoid foramen exsutural	% 50 N 2	% 0 N 3	% 66.6 N 3	% 0 N 1	% 100 N 4	% 100 N 2	% 100 N 1	% 100 N 1
Posterior ethmoid foramen absent	% 0 N 2	% 0 N 3	% 0 N 2	% 0 N 2	% 0 N 5	% 0 N 3	% N 0	% 100 N 1

CRANIAL NON-METRIC TRAITS	MONI		PALAIKASTRO		AILIAS		MYRTOS PYRGOS	
	Lside	Lside	Lside	Lside	Lside	Lside	Rside	Lside
Accessory infraorbital foramen present	% 25 N 4	% 100 N 2	% 75 N 4	% 75 N 4	% 100 N 7	% 100 N 4	% 88.8 N 9	% 85.8 N 7
Inca bone	% 0 N 19		% 4.3 N 23		% 1.5 N 64		% 0 N 30	
Infraorbital suture	% 100 N 2	% 100 N 3	% 100 N 1	% 33.3 N 3	% 22.2 N 9	% 60 N 5	% 60 N 5	% 100 N 3
Nasal foramen	% 100 N 1	% 100 N 2	% 100 N 4	% 66.6 N 3	% 50 N 2	% 75 N 4	% 100 N 3	% 100 N 3
Trochlear spur	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0
Trochlear fossa	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0
Squamo- parietal ossicles	% 0 N 13	% 0 N 10	% 0 N 17	% 0 N 18	% 0 N 47	% 0 N 35	% 5.5 N 18	% 6.6 N 15
Processus marginalis	% 77.7 N 9	% 80 N 5	% 50 N 2	% 66.6 N 3	% 69.2 N 13	% 75 N 8	% 23.8 N 21	% 27.2 N 18
Zygomatico-temporal foramen	% 0 N 7	% N 0	% 0 N 1	% N 0	% 0 N 6	% 20 N 5	% 0 N 10	% 0 N 6
Occipito mastoid ossicle	% 7.6 N 13	% 7.1 N 14	% 0 N 12	% 0 N 15	% 0 N 32	% 0 N 27	% 12.5 N 8	% 8.3 N 12
Intermediate condylar canal	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0
Post condylar tubercle	% 20 N 5	% 33.3 N 3	% 100 N 3	% 25 N 4	% 71.4 N 7	% 66.6 N 6	% 50 N 2	% 100 N 1
Jugular foramen bridge	% 25 N 8	% 0 N 6	% 0 N 1	% 0 N 3	% 0 N 12	% 12.5 N 8	% 0 N 10	% 0 N 8
Foramen of vesalius	% 100 N 5	% 60 N 5	% 100 N 3	% 100 N 6	% 90 N 10	% 60 N 10	% 45.4 N 11	% 55.5 N 9
Pterygo-basal bridge	% 33.3 N 6	% 33.3 N 9	% 44.4 N 9	% 40 N 10	% 28.5 N 14	% 28.5 N 14	% 6.6 N 15	% 18.1 N 11
Pterygo-spinous bridge	% 0 N 4	% 0 N 5	% 0 N 8	% 0 N 8	% 0 N 12	% 0 N 11	% 10 N 10	% 14.2 N 7

CRANIAL NON-METRIC TRAITS	MONI		PALAIKASTRO		AILIAS		MYRTOS PYRGOS	
	Lside	Lside	Lside	Lside	Lside	Lside	Rside	Lside
Spino-basal bridge	% 40 N 5	% 33.3 N 9	% 28.5 N 7	% 36.3 N 11	% 61.53 N 13	% 41.66 N 12	% 40 N 15	% 41.6 N 12
Foramen ovale spine	% 0 N 5	% 0 N 7	% 12.5 N 8	% 0 N 6	% 10 N 10	% 0 N 12	% 14.2 N 14	% 14.2 N 7
Accessory foramen spinosum	% 0 N 5	% 0 N 4	% 0 N 8	% 0 N 9	% 0 N 12	% 0 N 10	% 0 N 9	% 0 N 9
Lateral pterygoid perforated	% N 0	% N 0	% N 0	% N 0	% N 0	% 0 N 1	% 50 N 2	% 0 N 2
Pterygoid spurs	% N 0	% N 0	% N 0	% N 0	% N 0	% 0 N 1	% 0 N 1	% 0 N 1
Palatine bridge	% 0 N 3	% 0 N 3	% 0 N 1	% 0 N 1	% 0 N 1	% 0 N 1	% 0 N 2	% 0 N 2
Zygomatico- facial foramen multiple	% 75 N 8	% 66 N 6	% 100 N 1	% 100 N 4	% 83.3 N 12	% 88.8 N 9	% 57.1 N 21	% 52.9 N 17

Table 6.3 Percentages of Cranial non-metric traits and number of individuals scored (n) for Gypsades, Sellopoulo and Mavrospelio samples.

CRANIAL NON-METRIC TRAITS	GYPSADES		SELLOPOULO		MAVROSPELIO		PALAMA	
	Lside	Rside	Lside	R side	Lside	Lside	Rside	Lside
Highest Nuchal Line present	% 88.8 N 9	% 88.8 N 9	% 80 N 5	% 75 N 4	% 66.6 N 6	% 57.1 N 7	% 75 N 8	% 77.7 N 9
Ossicle at Lambda	% 20 N 10		% 25 N 4		% 0 N 7		% 0 N 10	
Lambdoid ossicle present	% 0 N 4	% 75 N 4	% 50 N 4	% 0 N 4	% 33.3 N 6	% 33.3 N 6	% 11.1 N 9	% 33.3 N 9
Parietal foramen present	% 40 N 15	% 64.2 N 14	% 0 N 4	% 40 N 5	% 14.2 N 7	% 28.5 N 7	% 45.4 N 11	% 54.5 N 11
Bregmatic bone present	% 0 N 14		% 0 N 6		% 0 N 7		% 0 N 13	
Metopic suture	% 5 N 20		% 0 N 6		% 14.28 N 7		% 7.6 N 13	
Coronal ossicle present	% 0 N 8	% 0 N 11	% 0 N 5	% 0 N 5	% 0 N 7	% 0 N 7	% 0 N 9	% 10 N 10
Epipteric bone present	% 0 N 8	% 0 N 7	% 0 N 5	% 0 N 6	% 0 N 7	% 0 N 6	% 0 N 10	% 0 N 8
Fronto-temporal articulation	% 0 N 4	% 0 N 0	% 0 N 5	% 0 N 3	% 0 N 7	% 0 N 5	% 0 N 8	% 0 N 5
Parietal notch bone present	% 0 N 3	% 0 N 3	% 0 N 3	% 0 N 3	% 0 N 4	% 0 N 5	% 12.5 N 8	% 0 N 5
Ossicle at asterion	% 0 N 4	% 0 N 3	% 0 N 5	% 0 N 3	% 16.6 N 6	% 20 N 5	% 0 N 10	% 0 N 5
Auditory torus present	% 0 N 8	% 0 N 10	% 80 N 5	% 20 N 5	% 0 N 4	% 20 N 5	% 0 N 7	% 16.6 N 6
Foramen of Huschke present	% 10 N 4	% 80 N 5	% 100 N 3	% 66.6 N 3	% 100 N 4	% 100 N 5	% 88.8 N 9	% 90 N 10
Mastoid foramen exsutural	% 37.5 N 8	% 76.9 N 13	% 25 N 4	% 75 N 4	% 75 N 4	% 75 N 4	% 63.6 N 11	% 37.5 N 8
Mastoid foramen absent	% 50 N 2	% 10 N 2	% 100 N 1	% 33.3 N 3	% 66.6 N 3	% 33.3 N 3	% 50 N 8	% 80 N 5
Posterior condylar canal patent	% 0 N 1	% N 0	% 50 N 2	% 33.3 N 3	% 100 N 1	% 0 N 1	% 0 N 1	% 0 N 1

CRANIAL NON-METRIC TRAITS	GYPSADES		SELLOPOULO		MAVROSPELIO		PALAMA	
	Lside	Rside	Lside	R side	Lside	Lside	Rside	Lside
Condylar facet double	% N 0	% N 0	% 0 N 3	% 0 N 2	% 0 N 1	% 0 N 1	% 0 N 2	% 0 N 2
Pharyngeal tubercle present	% N 0		% 100 N 3		% 100 N 2		% 40 N 5	
Precondylar tubercle present	% N 0	% 0 N 1	% 33.3 N 3	% 33.3 N 3	% 100 N 1	% 0 N 1	% 0 N 3	% 0 N 2
Anterior condylar canal double	% N 0	% 0 N 1	% 0 N 4	% 25 N 4	% 0 N 2	% 50 N 2	% 28.7 N 7	% 20 N 5
Foramen ovale incomplete	% 0 N 1	% 0 N 1	% 0 N 2	% 0 N 3	% 0 N 3	% 0 N 4	% 0 N 5	% 0 N 4
Foramen spinosum open	% 100 N 1	% 0 N 1	% 0 N 1	% 66.6 N 3	% 0 N 1	% 0 N 3	% 0 N 3	% 25 N 4
Accessory lesser palatine foramen present	% 0 N 2	% 0 N 2	% 0 N 1	% 0 N 1	% 0 N 1	% 100 N 1	% 0 N 1	% N 0
Palatine torus present	% 0 N 1	% 0 N 2	% 66.6 N 3	% 66.6 N 3	% 0 N 2	% 0 N 2	% 0 N 6	% 0 N 5
Maxillary torus present	% 100 N 1	% 100 N 2	% 25 N 4	% 25 N 4	% 0 N 2	% 0 N 2	% 0 N 2	% 0 N 3
Maxillary bridge present	% 0 N 7	% 0 N 9	% 100 N 2	% 100 N 2	% 50 N 2	% 50 N 2	% 33.3 N 3	% 66.6 N 3
Zygomatic - facial foramen absent	% 10 N 10	% 18.1 N 11	% 20 N 5	% 0 N 4	% 0 N 1	% 0 N 1	% 28.7 N 7	% 0 N 5
Supraorbital foramen complete	% 72.7 N 11	% 72.7 N 11	% 20 N 5	% 25 N 4	% 40 N 5	% 14.2 N 7	% 30 N 10	% 57.1 N 7
Frontal notch / foramen present	% 100 N 1	% N 0	% 75 N 4	% 75 N 4	% 80 N 5	% 85.7 N 7	% 70 N 10	% 42.8 N 7
Anterior ethmoid foramen exsutural	% N 0	% N 0	% 100 N 1	% 0 N 1	% 33.3 N 3	% 50 N 4	% 100 N 1	% 100 N 2
Posterior ethmoid foramen absent	% N 0	% N 0	% 0 N 1	% 0 N 1	% 0 N 4	% 25 N 4	% 0 N 1	% 0 N 2
Accessory infraorbital foramen present	% N 0	% N 0	% 50 N 2	% 50 N 2	% 100 N 2	% 100 N 2	% 100 N 3	% 100 N 4

CRANIAL NON-METRIC TRAITS	GYPSADES		SELLOPOULO		MAVROSPELIO		PALAMA	
	Lside	Rside	Lside	R side	Lside	Lside	Rside	Lside
Inca bone	% 6 N 15		% 0 N 6		% 0 N 7		% 0 N 12	
Infraorbital suture	% N 0	% N 0	% 0 N 1	% 100 N 1	% 100 N 1	% 100 N 2	% 66.6 N 3	% 25 N 4
Nasal foramen	% N 0	% N 0	% 0 N 1	% 50 N 2	% 100 N 2	% 100 N 3	% 83.3 N 6	% 80 N 5
Trochlear spur	% N 0	% N 0	% 100 N 2	% 100 N 1	% N 0	% N 0	% N 0	% N 0
Trochlear fossa	% N 0	% N 0	% 0 N 2	% 0 N 1	% N 0	% N 0	% N 0	% N 0
Squamo- parietal ossicles	% 0 N 2	% 0 N 1	% 0 N 4	% 0 N 3	% 0 N 6	% N 0	% 0 N 7	% 0 N 7
Processus marginalis	% 25 N 8	% 50 N 6	% N 0	% 0 N 1	% 100 N 1	% 50 N 2	% 57.1 N 7	% 80 N 5
Zygomatico-temporal foramen	% 0 N 1	% 0 N 3	% 0 N 2	% 0 N 2	% N 0	% N 0	% 0 N 2	% 0 N 1
Occipito mastoid ossicle	% N 0	% 0 N 1	% 0 N 3	% 0 N 2	% 0 N 4	% 0 N 5	% 0 N 9	% 0 N 8
Intermediate condylar canal	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0
Post condylar tubercle	% N 0	% N 0	% N 0	% N 0	% 100 N 1	% 100 N 1	% 100 N 3	% 100 N 4
Jugular foramen bridge	% N 0	% N 0	% 0 N 1	% 0 N 1	% 0 N 1	% 0 N 1	% 0 N 4	% 0 N 3
Foramen of vesalius	% 50 N 2	% N 0	% 100 N 1	% 100 N 1	% 100 N 3	% 75 N 4	% 0 N 6	% 0 N 4
Pterygo-basal bridge	% 0 N 2	% 0 N 1	% 0 N 3	% 50 N 4	% 0 N 3	% 33.3 N 3	% 33.3 N 6	% 20 N 5
Pterygo-spinous bridge	% 0 N 2	% N 0	% 50 N 2	% 100 N 1	% 0 N 3	% 0 N 4	% 0 N 6	% 0 N 5
Spino-basal bridge	% 100 N 2	% 100 N 1	% 33.3 N 3	% 0 N 2	% 66.6 N 3	% 100 N 3	% 0 N 6	% 25 N 4

CRANIAL NON-METRIC TRAITS	GYPSADES		SELLOPOULO		MAVROSPELIO		PALAMA	
	Lside	Rside	Lside	R side	Lside	Lside	Rside	Lside
Foramen ovale spine	% 0 N 2	% 0 N 1	% N 0	% N 0	% 0 N 3	% 0 N 4	% 16.6 N 6	% 25 N 4
Accessory foramen spinosum	% 0 N 2	% 0 N 1	% 0 N 2	% 0 N 2	% 0 N 3	% 0 N 3	% 0 N 6	% 0 N 4
Lateral pterygoid perforated	% N 0	% N 0	% N 0	% 0 N 1	% 0 N 1	% 0 N 1	% 0 N 1	% 0 N 1
Pterygoid spurs	% N 0	% N 0	% N 0	% N 0	% N 0	% N 0	% 0 N 1	% N 0
Palatine bridge	% N 0	% N 0	% N 0	% N 0	% 0 N 2	% 0 N 2	% 0 N 1	%0 N 1
Zygomatico- facial foramen multiple	% 57.14 N 7	% 37.1 N 8	% 40 N 5	% 40 N 5	% 100 N 1	% 100 N 1	% 33.3 N 6	% 80 N 5

Table 6. 4 Percentages (%) of dental non-metric traits, left side maxillary, and number of individuals (N) scored for Cretan and Argolid samples.

LEFT SIDE MAXILLARY DENTAL NON-METRICS	GYPSADES	SELLOPOULO	MYRTOS PYRGOS	PALAMA	APATHEIA	LERNA
Incisor 1st Shoveling	% 0 N 2	% 100 N 2	% 0 N 6	% 0 N 6	% 0 N 4	% 6.2 N 16
Incisor 2nd Shoveling	% 0 N 3	% 66.6 N 3	% 28.5 N 7	% 28.5 N 7	% 0 N 5	% 70 N 20
Canine Mesial ridge	% 0 N 6	% 0 N 3	% 0 N 12	% 0 N 5	% 0 N 4	% 4.7 N 21
Canine Distal Accessory ridge	% 20 N 5	% 33.3 N 3	% 0 N 7	% 0 N 5	% 25 N 4	% 0 N 17
Molar 1st Metaconule (cusp 5)	% 9 N 11	% 20 N 5	% 7.1 N 14	% 20 N 5	% 15.3 N 13	% 10.5 N 19
Molar 1st Carabelli's trait	% 20 N 5	% 40 N 5	% 0 N 11	% 40 N 5	% 50 N 10	% 37.5 N 16
Molar 1st Enamel extension	% 0 N 11	% 0 N 5	% 0 N 14	% 0 N 5	% 0 N 13	% 5 N 20
Molar 1st Enamel Pearl	% 0 N 10	% 0 N 5	% 0 N 14	% 0 N 4	% 0 N 13	% 5 N 20
Molar 2nd Hypocone (cusp 4)	% 50 N 6	% 75 N 4	% 62.5 N 16	% 100 N 3	% 100 N 3	% 77.7 N 18
Molar 3rd Metacone (cusp 3)	% 100 N 6	% 100 N 1	% 1 N 6	% 100 N 1	% 100 N 9	% 100 N 11
Molar 3rd Parastyle	% 0 N 5	% 0 N 1	% 0 N 7	% 50 N 2	% 12.5 N 8	% 0 N 11
Molar 3rd Congenital Absence	% 0 N 6	% 0 N 2	% 0 N 11	% 0 N 3	% 0 N 9	% 26.3 N 19

Table 6. 5 Percentages (%) of dental non-metric traits, right side maxillary, and number of individuals (N) scored for Cretan and Argolid samples.

RIGHT SIDE MAXILLARY DENTAL NON-METRICS	GYPSADES	SELLOPOULO	MYRTOS PYRGOS	PALAMA	APATHEIA	LERNA
Incisor 1st Shoveling	% 0 N 2	% 50 N 2	% 0 N 6	% 0 N 4	% 0 N 2	% 0 N 14
Incisor 2nd Shoveling	% 28.5 N 7	% 50 N 2	% 55.5 N 9	% 0 N 2	% 12.5 N 8	% 58.8 N 17
Canine Mesial ridge	% 0 N 4	% 0 N 3	% 0 N 13	% 0 N 3	% 0 N 4	% 5.5 N 18
Canine Distal Accessory ridge	% 33.3 N 3	% 0 N 3	% 0 N 12	% 0 N 3	% 0 N 2	% 0 N 17
Molar 1st Metaconule (cusp 5)	% 0 N 9	% 25 N 4	% 30 N 10	% 16.6 N 6	% 8.3 N 12	% 27.7 N 18
Molar 1st Carabelli's trait	% 25 N 8	% 50 N 4	% 20 N 10	% 40 N 5	% 57.1 N 7	% 33.3 N 15
Molar 1st Enamel extension	% 0 N 11	% 0 N 4	% 0 N 13	% 0 N 4	% 0 N 13	% 4.7 N 21
Molar 1st Enamel Pearl	% 0 N 10	% 0 N 4	% 0 N 13	% 0 N 4	% 0 N 13	% 4.7 N 21
Molar 2nd Hypocone (cusp 4)	% 50 N 8	% 60 N 5	% 62.9 N 13	% 100 N 4	% 100 N 5	% 69.5 N 23
Molar 3rd Metacone (cusp 3)	% 66.6 N 3	% 100 N 2	% 100 N 7	% 100 N 3	% 100 N 8	% 100 N 12
Molar 3rd Parastyle	% 0 N 3	% 0 N 2	% 0 N 8	% 0 N 4	% 50 N 8	% 0 N 11
Molar 3rd Congenital Absence	% 0 N 3	% 0 N 2	% 0 N 11	% 0 N 5	% 0 N 9	% 23.8 N 21

Table 6. 6 Percentages (%) of dental non-metric traits, left side mandibular, and number of individuals (N) scored for Cretan and Argolid samples.

LEFT SIDE MANDIBULAR DENTAL NON-METRICS	GYPSADES	SELLOPOULO	MYRTOS PYRGOS	PALAMA	APATHEIA	LERNA
Incisor 1st Shoveling	% 0 N 4	% 0 N 3	% 0 N 2	% 0 N 6	% 0 N 6	% 0 N 13
Incisor 2nd Shoveling	% 0 N 7	% 0 N 5	% 0 N 6	% 0 N 7	% 0 N 7	% 0 N 27
Canine Mesial Ridge	% 0 N 8	% 12.5 N 8	% 0 N 11	% 0 N 5	% 0 N 9	% 8.3 N 24
Canine Distal Accessory Ridge	% 0 N 7	% 14.2 N 7	% 0 N 10	% 0 N 5	% 0 N 4	% 0 N 27
Premolar 1st Number of Cusps (›1)	% 57.14 N 7	% 50 N 6	% 35.2 N 17	% 16.6 N 6	% 5 N 10	% 59.2 N 27
Premolar 2nd Number of Cusps (›1)	% 55.5 N 9	% 71.4 N 7	% 54.5 N 11	% 50 N 4	% 100 N 8	% 70 N 20
Molar 1st Number of Cusps (›4)	% 100 N 9	% 100 N 7	% 92.8 N 14	% 62.5 N 8	% 100 N 10	% 90 N 10
Molar 2nd Number of Cusps (›3)	% 85.7 N 7	% 100 N 8	% 94.7 N 19	% 100 N 5	% 100 N 9	% 90.0 N 11
Molar 3rd Protostylid	% 0 N 1	% 0 N 4	% 0 N 11	% 25 N 4	% 0 N 11	% 0 N 7
Molar 3rd Congenital Absence	% 0 N 3	% 0 N 6	% 0 N 14	% 0 N 6	% 0 N 14	% 0 N 10

Table 6. 7 Percentages (%) of dental non-metric traits, right side mandibular, and number of individuals (N) scored for Cretan and Argolid samples.

RIGHT SIDE MANDIBULAR DENTAL NON-METRICS	GYPSADES	SELLOPOULO	MYRTOS PYRGOS	PALAMA	APATHEIA	LERNA
Incisor 1st Shoveling	% 0 N 5	% 0 N 3	% 0 N 3	% 0 N 5	% 0 N 4	% 0 N 3
Incisor 2nd Shoveling	% 0 N 9	% 0 N 3	% 0 N 7	% 0 N 5	% 0 N 6	% 0 N 5
Canine Mesial Ridge	% 0 N 4	% 0 N 4	% 0 N 10	% 0 N 6	% 0 N 10	% 0 N 7
Canine Distal Accessory Ridge	% 0 N 5	% 0 N 4	% 0 N 5	% 0 N 6	% 16.6 N 6	% 0 N 4
Premolar 1st Number of Cusps (›1)	% 37.5 N 8	% 66.6 N 6	% 50 N 14	% 50 N 6	% 60 N 5	% 60 N 10
Premolar 2nd Number of Cusps (›1)	% 20 N 5	% 20 N 5	% 50 N 10	% 25 N 4	% 42.8 N 7	% 42.8 N 7
Molar 1st Number of Cusps (›4)	% 100 N 8	% 62.5 N 8	% 91.6 N 12	% 100 N 3	% 100 N 12	% 83.3 N 6
Molar 1st Protostylid	% 16.6 N 6	% 0 N 7	% 0 N 13	% 25 N 4	% 0 N 12	% 0 N 6
Molar 2nd Number of Cusps(›3)	% 100 N 11	% 100 N 7	% 93.7 N 16	% 100 N 3	% 100 N 15	% 87.5 N 8
Molar 3rd Protostylid	% 0 N 7	% 0 N 4	% 0 N 11	% 0 N 2	% 0 N 8	% 0 N 6
Molar 3rd Congenital Absence	% 0 N 8	% 0 N 7	% 8.3 N 12	% 25 N 4	% 7.1 N 14	% 14.2 N 7

APPENDIX F
VALUES FOR STRONTIUM ISOTOPE RATIO ANALYSIS

Table 7.1 Strontium Isotope Ratio analysis values, all samples analysed.

GEOGRAPHICAL REGION	SKELETAL COLLECTION	INDIVIDUAL	ELEMENT	Sr ISOTOPE RATIO ($^{87}Sr/^{86}Sr$) (values)
Central Crete	Sellopoulo	SELI, VII	Tooth (M1)	0.708933
	Sellopoulo	SELI, VII	Bone (Femur)	0.709000
	Sellopoulo	SEL4, 3	Tooth (M1)	0.708943
	Sellopoulo	SELI, IV	Tooth (M1)	0.708604
	Sellopoulo	SELI, IV	Tooth (M1)	0.708984
	Sellopoulo	SELI, II	Tooth (M1)	0.709062
	Sellopoulo	SELI, III NE	Tooth (M1)	0.708934
	Sellopoulo	SELII, N.BOTHROS	Tooth (M1)	0.709016
	Sellopoulo	SELI, III	Tooth (M1)	0.708967
	Sellopoulo	SELI, III	Bone (Tibia)	0.709044
	Sellopoulo	SELI A (S/G)	Tooth	0.708679
	Knossos	KN A (PIG)	Tooth	0.708985
	Upper Gypsades	GYPXVIII, III	Tooth (M1)	0.709053
	Upper Gypsades	GYPXVIII, II	Tooth (M1)	0.709021
	Upper Gypsades	GYPXVIII, III	Tooth (M1)	0.709029
	Upper Gypsades	GYPXVIII,VII	Tooth (M1)	0.709025
	Upper Gypsades	GYPXVIII,VI	Tooth (M1)	0.709058
	Lower Gypsades	LGI, Larnax	Tooth (M1)	0.709033
	Lower Gypsades	LGI, Larnax	Bone (Femur)	0.709022
	Lower Gypsades	LGI, F	Tooth (M2)	0.708997
	Lower Gypsades	LGI, F5	Tooth (PM4)	0.709035
	Lower Gypsades	LGI, E6	Tooth (M1)	0.709085
	Lower Gypsades	LGI, Balk A-B (S/G)	Tooth	0.708989
	Ailias	AIL 102	Tooth (M1)	0.708999
	Ailias	AIL 2	Tooth (M1)	0.708970
	Ailias	AIL 6	Tooth (M1)	0.709000
	Ailias	AIL 90	Tooth (M1)	0.709014
	Ailias	AIL 15	Tooth (M1)	0.709042
	Ailias	AIL 1	Tooth (M1)	0.709212

GEOGRAPHICAL REGION	SKELETAL COLLECTION	INDIVIDUAL	ELEMENT	Sr ISOTOPE RATIO ($^{87}Sr/^{86}Sr$) (values)
Central Crete	Ailias	AIL 103	Tooth (M1)	0.709050
	Ailias	AIL 98	Tooth (M1)	0.709227
	Knossos Makry Teichos	KN1 SN	Shell	0.708991
	Knossos Makry Teichos	KN2 SN	Shell	0.709026
	Knossos Stratigraphical Museum	KN3 SN	Shell	0.708974
	Knossos Stratigraphical Museum	KN4 SN	Shell	0.708952
	Ano Asites	AAS (R)	Tooth (I1)	0.708957
	Maroulas	MLS2, 1	Tooth (M1)	0.709097
	Maroulas	MLS2, 1	Bone (femur)	0.709151
	EPISKOPI	EPIS 1	Tooth (M1)	0.709072
	EPISKOPI	EPIS 2	Tooth (M1)	0.708467
	EPISKOPI	EPIS 3	Tooth (M1)	0.708482
Eastern Crete	Myrtos Pyrgos	MP 25B	Tooth (M1)	0.708792
	Myrtos Pyrgos	MP A	Tooth (M1)	0.708545
	Myrtos Pyrgos	MP H	Tooth (M1)	0.708781
	Myrtos Pyrgos	MP 25A	Tooth (M1)	0.708773
	Myrtos Pyrgos	MP 5	Tooth (M1)	0.708931
	Myrtos Pyrgos	MP D (2nd individual)	Tooth (M1)	0.708675
	Myrtos Pyrgos	MP E	Bone (Femur)	0.708856
	Myrtos Pyrgos	MP A	Bone (Femur)	0.708860
	PALAIKASTRO D131	PK D131	Tooth (M1)	0.709112
Western Crete	Kastelos	KAS 2	Tooth (M1)	0.709042
	Kastelos	KAS P	Tooth (M1)	0.709013
	Kastelos	KAS P	Bone (Femur)	0.709054
	Margarites	MRG 1	Tooth (M1)	0.708986

GEOGRAPHICAL REGION	SKELETAL COLLECTION	INDIVIDUAL	ELEMENT	Sr ISOTOPE RATIO ($^{87}Sr/^{86}Sr$) (values)
Western Crete	Margarites	MRG 2	Tooth (M1)	0.708975
	Margarites	MRG 3	Tooth (M1)	0.709037
	Margarites	MRG 4	Bone (Femur)	0.709080
	Palama	PAL15 A	Tooth (M1)	0.709585
	Palama	PAL15 C	Tooth (M1)	0.709086
	Palama	PAL16, 1	Tooth (M1)	0.709062
	Palama	PAL16, 2	Tooth (M1)	0.708916
	Palama	PAL16, 2	Bone (Femur)	0.709022
	Palama	PAL NI	Tooth (M1)	0.709085
	Palama	PAL8, 2	Tooth (M1)	0.709052
	Palama	PAL10, 2	Tooth (M1)	0.709044
	Palama	PAL3	Tooth (M1)	0.709051
	Palama	PAL3	Bone (Femur)	0.709127
	Palama	PAL16, 2	Bone (Femur)	0.709022
Naxos	Tsikniades	TSIK 114	Tooth (M1)	0.709412
	Tsikniades	TSIK 114	Bone (Femur)	0.709487
	Tsikniades	TSIK 120	Tooth (M1)	0.709024
	Tsikniades	TSIK 121	Tooth (M1)	0.709514
	Tsikniades	TSIK1 SN	Shell	0.709279
	Tsikniades	TSIK2 SN	Shell	0.709261
	Aplomata	APL K	Tooth (M1)	0.709406
	Aplomata	APL XXIII	Tooth (M1)	0.709452
	Aplomata	APLI C	Tooth (M1)	0.709372
	Aplomata	APLB	Tooth M1	0.709440
	Vasalakis	VAS 3A	Tooth (M1)	0.709038
	Vasalakis	VAS 3A	Bone (Femur)	0.709680
	Vasalakis	VAS 0282	Bone (Femur)	0.709427
	Vasalakis	VAS 0282	Tooth (M1)	0.710474
	Vasalakis	VAS C (COW)	Tooth (M3)	0.709495
	Vasalakis	VAS P (PIG)	Tooth (M1)	0.710039
	Kamini	KAM 3	Tooth (M2)	0.709457
	Kamini	KAM 3B	Tooth (M1)	0.709467
	Kamini	KAM K3	Tooth (M1)	0.709431

GEOGRAPHICAL REGION	SKELETAL COLLECTION	INDIVIDUAL	ELEMENT	Sr ISOTOPE RATIO ($^{87}Sr/^{86}Sr$) (values)
Naxos	Kamini	KAM A1	Bone (Tibia)	0.709543
Argolid - Mainland	Mycenae 2km from Acropolis	MYC1 SN	Shell	0.708262
	Mycenae Acropolis	MYC2 SN	Shell	0.708254
	Mycenae Acropolis	MYC3 SN	Shell	0.708226
	Mycenae Acropolis	MYC4 SN	Shell	0.708328

Key: M1=1st Molar, M2= 2nd molar, S/G=Sheep/Goat, SN=Snail, R=Rabbit.

REFERENCES

Acsaki, G. & Nemeskeri, J. 1970. History of Human Life Span and Mortality. Budapest: Akademiaj Kiado.

Agelarakis, A. 1995. An Anthology of Hellenes involved with the field of Physical Anthropology. *International Journal of Anthropology*, 10: 149-62.

Ager, D. V. 1986. The geology of Europe. Swansea: University College.

Alberti, L. 2004a. Oi nekropoleis tis Knosoy tin Ysterominoiki II-IIIA1 periodo. Athens: Unpublished PhD thesis, University of Athens.

Alberti, L. 2004b. The Late Minoan II-IIIA1 warrior graves at Knossos: the burial assemblages. In G. Cadogan, E. Hatzaki, & A. Vasilakis (eds), *Knossos: Palace, City, State. Proceedings of the Conference in Herakleion organized by the British School at Athens and the 23rd Ephoreia of Prehistoric and Classical Antiquities of Herakleion, in November 2000, for the Centenary of Sir Arthur Evans's Excavations at Knossos, BSA Studies 12*, pp.127-36. London.

van Alden, T.H., Runnels, C.N., & Pope, K. 1986. Five Thousand Years of Land Use and Abuse in the Southern Argolid, Greece. *Hesperia*, 55: 103-128.

van Alden, T.H., & Runnels, C.N. 1987. The Evolution of the Settlement in the Argolid, Greece: An Economic Explanation. *Hesperia*, 56: 303- 34.

van Andel, T.J & Runnels, C.N. 1988. An essay 'on the emergence of civilization' in the Aegean world. *Antiquity*, 62: 234-47.

Anderson, J.E. 1968. Skeletal "anomalies" as genetic indicators. In D.R. Brothwell (ed), *The skeletal biology of earlier populations*, pp.135-48. Oxford: Pergamon Press.

Andreou, S. 1978. Pottery Groups of the Old Palace Period in Crete. PhD thesis, University of Cincinnati.

Angel, J.L. 1951. Population size and microevolution in Greece. *Cold Spring Harbor Symposium on Quantitative Biology, 15*: 343-351.

Angel, J.L. 1959. Early Helladic skulls from Agios Kosmas. In G.E. Mylonas (ed), *Agios Kosmas*, pp. 167-169. Princeton: University Press.

Angel, J.L. 1971. The People of Lerna: Analysis of a Prehistoric Aegean Population. Washington: Smithsonian Institution Press.

Angel, J. L. 1973a. Human Skeletons from Grave Circles at Mycenae. In G.E. Mylonas (ed), *Tafikos Kyklos B ton Mykinon*, pp. 379-397. Athens: Library of the Archaeological Society at Athens, 73.

Angel, J.L.1973b. Neolithic human remains (Franchti Cave). *Hesperia*, 42: 277-282.

Angel. J. L. 1977. Human Skeletons. In J. E.Coleman (ed), *Kephala. A late Neolithic settlement and cemetery, pp. 133-156.* Princeton: American School of Classical Studies at Athens.

Angel, J.L. 1982a. A new measure of growth efficiency: skull base height. *American Journal of Physical Anthropology*, 58: 297-305.

Angel, J.L. 1982b. Ancient skeletons from Asine. In S. Dietz (ed), *Asine II. Results of the excavations east of the Acropolis 1970-1974*, pp. 105-138. Stockholm: Paul Åströms Förlag.

Anthony, D. W. 1990. Migration in Archaeology: The Baby and the Bathwater. *American Anthropologist*, 92: 895-914.

Ascadi, G., & Nemeskéri, J. 1970. History of Human Life Span and Mortality. Budapest: Akademiai Kiado.

Åström, P. 1994. The Mycenaean Citadel of Midea. *Journal of Prehistoric Religion*, 8: 5-6.

Barber, R.L.N. 1978. The Cyclades in the Middle and Late Bronze Age. PhD dissertation, University of St Andrews.

Barber, R.L.N. & MacGillivray, J.A. 1980. The Early Cycladic Period: Matters of Definition and Terminology. *American Journal of Archeology*, 84, (2): 141-157.

Barber, R.L.N. 1983. The Definition of the Middle Cycladic Period. *American Journal of Archaeology*, 87: 76-81.

Barber, R. 1984. The status of Phylakopi in Creto-Cycladic Relations. In R. Hägg, & N. Marinatos (eds), *The Minoan Thalassocracy: Myth and Reality. Proceedings of the Third International Symposium at the Swedish Institute in Athens, 31 May-5 June, 1982. Skrifter Utgivna av Svenska Institutet i Athen, 4º, XXXII*, pp. 179-81. Stockholm: Paul Åströms Förlag.

Barber, R.L.N. 1987. The Cyclades in the Bronze Age. London: Gerald Duckworth & Co. Ltd.

Barber, R., & Hadjianastasiou, O. 1989. Mikre Vigla: A Bronze Age Settlement on Naxos. *Annual of the British School at Athens*, 84: 63-162.

Barbujani, G., Magagni, A., Minch, E., & Cavalli-Sforza, L.L. 1997. An apportionment of human DNA diversity. *Proceedings of National Academy of Sciences of the United States of America*, 94: 4516-19.

Bass, W.M. 1995. Human Osteology: A Laboratory and Field Manual. Columbia, Missouri: Missouri Archaeological Society.

Betancourt, P.P. 1976. The end of the Greek Bronze Age. *Antiquity*, 50: 40-7.

Beals, K.L., Smith, C.L., & Dodd, S.M. 1984. Climate and the Evolution of Brachycephalization. *American Journal of Physical Anthropology*, 62: 1-13.

Bennet, J. 1999. La Crète mycénienne. Actes de la Table Ronde Internationale organisée par l'Ecole Française d'Athènes, 26-28 Mars 1991. *American Journal of Archaeology*, 103: 554-55.

Berry, A.C., & Searly, A.G. 1963. Epigenetic Polymorphism of the Rodent Skeleton. *Proceedings Zoological Society of London*, 140: 557-615.

Berry, A. C., & Berry R.J. 1967. Epigenetic variation in the human cranium. *Journal of Anatomy*, 101: 361-379.

Berry, A.C. 1968. The biology of non-metrical variation in mice and men. In D.R. Brothwell (ed), *The skeletal biology of earlier human populations*, pp. 103-133. Oxford: Pergamon Press.

Berry, A. C. 1978. Anthropological and family studies of minor variants of the dental crown. In P.M Rutter, & K. A. Joysey (eds), *Development, function and evolution of teeth*, pp. 81-98. London: Academic Press.

Betancourt, P.P. 1976. The end of the Greek Bronze Age. *Antiquity*, 50: 40-7.

Betancourt, P.P. 2000. The Aegean and the Origin of the Sea Peoples. In E.D. Oren (ed), *The Sea Peoples and Their World: A reassessment. University Museum Monograph 108, University Museum Symposium Series 11*, pp. 297-303. Philadelphia: The University Museum, University of Pennsylvania.

Binford, L. R. 1965. Archaeological systematics and the study of culture process. *American Antiquity*, 31: 203-10.

Bintliff, J.L. 1977. Natural environment and Human Settlement in Prehistoric Greece based on original fieldwork. British Archaeological Reports Supplementary Series 28. Oxford: British Archaeological Reports.

Bintliff, J. 1994. Time, Structure, and Agency: The Annales, Emergent Complexity, and Archaeology. In J. Bintliff, *A Companion to Archaeology*, pp.174-94. Oxford: Blackwell.

Bisel, S. C. 1982. The human Skeletal Remains. In W.A. McDonald & Wilkie (eds), *Excavations at Nichoria in Southwest Greece*, Vol. II. Bronze Age Occupation, pp. 345-358.

Bisel, S.C., & Angel, J.L. 1985. Health and Nutrition in Mycenaean Greece: A study in human skeletal remains. In N.C. Wilkie, & W.D.E. Coulson (eds), *Contributions to Aegean Archaeology: Studies in honor of W.A. McDonald.* University of Minnesota.

Blegen, C. W. 1928. Zygouries: A Prehistoric Settlement in the Valley of Cleonae. Cambridge.

Blom, D.E., Hallgrimsson, B., Keng,L., Lozada, M.C., & Buikstra, J.E. 1998. Tiwanaku 'Colonization': Bioarchaeological Implications for Migrations in the Moquegua Valley, Peru. *World Archaeology*, 30:238-61.

Boardman, J. 1964. The End of the Mycenaean Age. *The Classical Review*, 14: 311-13.

Bodmer, W.F., & Cavalli-Sforza, L.L. 1976. Genetics, evolution and man. San Francisco: W.H. Freeman.

Boulle, E-L. 2001. Evolution of Two Human Skeletal Markers of the Squatting Position: A Diachronic Study From Antiquity to the Modern Age. *American Journal of Physical Anthropology*, 115: 50-56.

Bourdieu, P. 1990. The logic of practice. Stanford, California: Stanford University Press.

Bowyer, D. 1972. Chromatography analyses for lipids in clay. In P. M. Warren (ed), *Myrtos: An Early Bronze Age Settlement in Crete*, pp.330-1. London: Thames &

Hudson. The British School of Archaeology at Athens, Supplementary Volume No. 7.

Boyd, D.C. 1988. A functional model for masticatory-related mandibular, dental and craniofacial microevolutionary change derived from a selected southeastern Indian skeletal temporal series. Unpublished PhD dissertation. University of Tennessee, Knoxville.

Brace, L.C 1964. A non-racial approach towards the understanding of human diversity. In A. Montagu (ed), *The Concept of Race*, pp. 103-152. New York: The Free Press of Glencoe.

Brace, L.C. 1980. Australian Tooth-Size Clines and the Death of a Stereotype. *Current Anthropology*, 21: 141-64.

Brace, L.C. 1982. The roots of the race concept in American physical anthropology. In F. Spencer (ed), *A History of American Physical Anthropology 1930-1980*, pp. 11-29. New York: Academic Press.

Brace, L.C., & Hunt, K.D. 1990. A Non-racial Craniofacial Perspective on Human Variation: A(ustralia) to Z(uni). *American Journal of Physical Anthropology*, 82: 341-360.

Brace, C.L, Rosenberg, K.R., & Hunt, K.D. 1991. What big teeth you had grandma! Human tooth size, past and present. In M.A. Kelly, & C.S. Larsen (eds), *Advances in dental anthropology*, pp. 33-57. New York: Wiley-Liss.

Branigan, K. 1967. Further Light on Prehistoric Relations between Crete and Byblos. *American Journal of Archaeology*, 71: 117-21.

Branigan, K. 1968. A Transitional Phase in Minoan Metallurgy. *Annual of the British School at Athens*, 63: 185-203.

Branigan, K. 1989. Minoan Foreign Relations in Transition. *Aegaeum*, 3: 65-71.

Branigan, K. 1993. Dancing with Death: Life and Death in Southern Crete c. 3000-2000 BC. Amsterdam: Adolf M. Hakkert.

Braudel, F. 1972. The Mediterranean and the Mediterranean World in the Age of Philip II (translation of the 2nd edition). London: Collins.

Bridges, P., Blitz, J., & Solano, J. 2000. Changes in long bone diaphyseal strength with horticultural intensification in west- central Illinois. *American Journal of Physical Anthropology*, 112(2): 217-238.

Bridges, P.S. 1989. Changes in activities with the shift to agriculture in the Southeastern United States. *Current Anthropology*, 75: 113-27.

Broodbank, C. 1989. The longboat and society in the Cyclades in the Keros-Syros culture. *American Journal of Archaeology*, 93: 319-37.

Broodbank, C., & Strasser, T. F. 1991. Migrant farmers and the Neolithic colonization of Crete. *Antiquity*, 65: 233-245.

Broodbank, C. 1993. Ulysses without Sails: Trade, Distance, Knowledge and Power in the Early Cyclades. *World Archaeology*, 24: 315-331.

Broodbank, C. 2000. An Island Archaeology of the Early Cyclades. Cambridge: Cambridge University Press.

Brooks, S., & Suchey, J.M. 1990. Skeletal age determination based on the os pubis: A comparison of the Acsadi-Nemeskeri and Suchey – Brooks methods. *Human Evolution*, 5: 227-238.

Brothwell, D.R. 1981. *Digging Up Bones*. Ithaka. New York: Cornell University Press.

Buikstra J.E., Frankenberg, S.R., & Konigsberg, L.W. 1990. Skeletal Biological Distance Studies in American Physical Anthropology: Recent Trends.

Buikstra, J. E., & Ubelaker, D.H. 1994. Standards for Data Collection from Human Skeletal Remains. Fayetteville, Arkansas: Arkansas Archaeological Survey Report Number 44.

Butler, P.M. 1939. Studies of the mammalian dentition – and of differentiation of the postcanine dentition. *Proceedings of the Zoological Society*, 107 (B): 103-132. London.

Buxton, L.H.D. 1920. The Inhabitants of the Eastern Mediterranean. *Biometrika*, 13: 97-112.

Buxton, L.H.D. 1938. Platymeria and Platychemia. *Journal of Anatomy*, 73: 31-6.

Cadogan, G. 1977-78. Pyrgos, Crete, 1970-7. *Archaeological Reports for 1977-78*: 70-84. British School at Athens.

Cadogan, G. 1983. Early Minoan and Middle Minoan Chronology. *American Journal of Archaeology*: 507–18.

Cadogan, G. 2000. Domestic Life at Minoan Myrtos-Pyrgos. *Pepragmena 8:Proceedings of the 8th International Cretological Congress*, pp. 169-74.

Capasso, L., Kennedy, K.A.R., & Wilczak, C. A. 1999. *Atlas of Occupational Markers on Human Remains*. Journal of Paleontology, Monograph Publication 3. Teramo: Edigrafital S.p.A.

Carlson, A.K. 1996. Lead isotope analysis of human bone for addressing cultural affinity: a case study from Rocky Mountain House, Alberta. *Journal of Archaeological Science*, 23 (4): 557-68.

Garn, S.M., Guzman, M.A., & Wagner, B. 1969. Subperiosteal gain and endosteal loss in protein-calorie malnutrition. *American Journal of Physical Anthropology*, 30: 153-156.

Carr, H. G. 1960. Some dental characteristics of the Middle Minoans. *Man*, 157: 119-22.

Caskey, J.L. 1954. Excavations at Lerna, 1952-53. *Hesperia*, 23: 3-30.

Caskey, J.L. 1955. Excavations at Lerna, 1954. *Hesperia*, 24: 25-49.

Caskey, J.L. 1956. Excavations at Lerna, 1955. *Hesperia*, 25: 147-73.

Caskey, J.L. 1957. Excavations at Lerna, 1956. *Hesperia*, 26: 142-62.

Caskey, J.L. 1958. Excavations at Lerna, 1957. *Hesperia*, 27: 125-44.

Caskey, J.L. 1959. Activities at Lerna, 1958-59. *Hesperia*, 28: 202-7.

Caskey, J.L. 1960. The Early Helladic Period in the Argolid. *Hesperia*, 29: 285-303.

Caskey, J.L. 1969. Crises in the Minoan-Mycenaean World. *Proceedings of the Philosophical Society*, 113: 433-49.

Casti, J.L. 1994. Complexification: Explaining a paradoxical world through the science of surprise. London: Abacus.

Castrigiano, D. P. L., & Hayes, S.A. 1993. Catastrophe Theory. Wokingham: Addion - Wesley.

Catling, H. 1957. Archaeology in Greece, 1957. *Archaeological Reports*, 22-3.

Catling, H. 1981. Archaeology in Greece, 1980-81. *Archaeological Reports*, 36.

Cavanagh, W., & Mee, C. 1998. A Private Place: Death in Prehistoric Greece. Studies in Mediterranean Archaeology, Vol. CXXV. Sweden: Paul Åströms Förlag.

Cavalli-Sforza, L.L., Piazza, A., Menozzi, P., & Mountain, J. 1988. Reconstruction of human evolution: bringing together genetic, archaeological, and linguistic data. *Proceedings of the National Academy of Sciences of the United States of America*, 85(16): 6002-6.

Chambellan, M. 1883. Étude anatomique et anthropologique sur le os wormiens. Paris.

Charles, P. 1965. Anthropologie Archéologique de le Crète par Rober. Etudes Crétoises, XIV. Ecole Française D'Athènes.

Chatfield, C., & Collins, A. J. 1980. Introduction to multivariate analysis. London: Chapman and Hall.

Cherry, J. F. 1984. The emergence of state in prehistoric Aegean. *Proceedings of Cambridge Archaeological Society*, 30:18-48.

Cheverud, J.M, & Buikstra, J.E.1981. Quantitative genetics of skeletal nonmetric traits in the rhesus macaques on Cayo Santiago.I. Single trait heritabilities. *American Journal of Physical Anthropology*, 54: 43-49.

Cheverud, J.M. 1988. A Comparison of Genetic and Phenotypic Correlations. *Evolution*, 42: 958-68.

Chippindale, C. 2000. Capta and Data: On the True Nature of Archaeological Information. *American Antiquity*, 65: 605- 12.

Coleman, J. 1979. Chronological and Cultural Divisions of the Early Cycladic Period: a critical appraisal. In J.L. Davis, & J.F. Cherry (eds), *Papers in Cycladic Prehistory*, pp. 48- 50. Institute of Archaeology, Monograph 14. Los Angeles: University of California.

Coleman, J. 1992. Greece, the Aegean and Cyprus.In R.W. Ehrich (ed), *Chronologies in Old World Archaeology*, pp.255-64. Chicago: University of Chicago Press.

Colledge, S., Conolly, J., & Shennan, S. 2004. Archaeobotanical evidence for the spread of farming in the Eastern Mediterranean. *Current Anthropology*, 45 (Supplement): 35-58.

Cook, J. M. 1951. Archaeology in Greece. Journal of Hellenic Studies, LXXII: 108.

Cook, J. M. & Boardman, J. 1954. Archaeology in Greece, 1953. *Archaeological Reports*, pp. 142-169.

Coon, C.S. 1962. The origin of Races. New York: Alfred A. Knopf.

Coppa, A. Cucina, A. , Mancinelli, D., Vargiu, R., & Calcagno, J.M. 1998. Dental Anthropology of Central-Southern, Iron Age Italy: The Evidence of Metric Versus Nonmetric Traits. *American Journal of Physical Anthropology*, 107: 371-86.

Corrucini, R.S. 1974. An examination of the meaning of cranial discrete traits for human skeletal biological studies. *American Journal of Physical Anthropology*, 40: 425-45.

Crow, J.F. & Kimura, M. 1970. An Introduction to Population Genetics Theory. New York: Harper and Row.

Cullen T. 2001. Aegean Prehistory: a review. Boston: Archaeological Institute of America

Czarnetzki, A. 1971. Epigenetische skelettmerkmale im populationvergleich. I. Rechts-links-unterschiede bilateral angelegter merkmale. *Zeitschrift für morphologie und Anthropologie*, 63: 238-54.

Dahlberg, A.A. 1945. The changing dentition of man. *Journal of American Dentist Association*, 32: 676-90.

Darwin, C. 1998. The Origin of Species, (first published in 1859, London: John Murray). Oxford: Oxford University Press.

Davaras, K. 1971. Πρωτομινωικόν νεκροταφείον Αγίας Φωτιάς Σητείας. *Archaeologika Analekta Athinon*, 4: 392-97.

Davies, D. M. 1972. The Influence of Teeth, Diet, and Habits on the Human Face. London: W. Heinemann.

Davis, J.L. 1982. The earliest Minoans in the south-east Aegean: a reconsideration of the evidence. *Anatolian Studies*, 32: 33-41.

Davis, J.L., & Cherry, J.F. 1984. Phylakopi in Late Cycladic I: a pottery seriation study. In J.A. MacGillivray, & R.L.N. Barber (eds), *The Prehistoric Cyclades . Contributions to a workshop on Cycladic chronology*, pp. 148-161. Edinburgh: Department of Classical Archaeology, University of Edinburgh.

Davis, J.L. 1987. Perspectives on the Prehistoric Cyclades: an archaeological introduction. In P. Getz-Preziosi (ed), *Early Cycladic Art in North American Collections*, pp. 4-45. Richmond: Virginia Museum of Fine Arts.

Davis, J.L. 2001. The Islands of the Aegean. In T. Cullen (ed), *Aegean Prehistory: a review*, pp. 19-76. Boston: Archaeological Institute of America.

Day, P.M, Wilson, D.E., & Kiriatzi, E. 1998. Pots, Labels, and People: Burying Ethnicity in the Cemetery at Aghia Photia, Siteias. In K. Branigan (ed), *Cemetery and Society in the Aegean Bronze Age. Sheffield Studies in Aegean Archaeology 1*, pp. 133-49. Sheffield, Sheffield Academic Press.

Debets, C.F. 1961. A nekotorykh napravleniax izmenenii v stroenii cheloveka sovremennogo vida. *Sovetskaya etnografiya*, 2: 9-23.

Demakopoulou, K. 1995. Mycenaean Citadels: Recent Excavations on the Acropolis of Midea in the Argolid. *Bulletin of the Institute of Classical Studies of the University of London*, pp. 151-61.

Desborough, V.R.d'A. 1964. The last Mycenaeans and Their Successors: An Archaeological Survey c. 1200-c.1000BC. Oxford.

Devor, E.J. 1987. Transmission of Human craniofacial dimensions. *Journal of Craniofacial Genetics Developmental Biology*, 7: 95-106.

Dickinson, O.T.P.K. 1977. The origins of Mycenaean civilization. *Studies in Mediterranean Archaeology, 49*. Gothenburg: Paul Åströms Förlag.

Dickinson, O.T.P.K. 1986. Early Mycenaean Greece and the Mediterranean. In M. Marazzi, S. Tusa, & L. Vagnetti (eds), *Traffici Micinei del Mediterraneo. Problemi storici e documentazione archeologica. Atti del convegno di Palermo (11-12 Maggio e 3-6 Dicembre 1984)*, pp.271-76. Taranto.

Dickinson, O.T.P.K. 1994. The Aegean Bronze Age. Cambridge: Cambridge University Press.

Dietz, S. 1982. Asine II. Results of the excavations east of the Acropolis 1970-1974, Stockholm: Paul Åströms Förlag.

Donlon, D.A. 2000. The Value of Infracranial Nonmetric Variation in Studies of Modern Homo sapiens: An Australian Focus. *American Journal of Physical Anthropology*, 113: 349-68.

Dorsey, G.A. 1897. Wormian bones in artificially deformed Kwakiutl cranial. *American Anthropology*, 10: 169-173.

Doumas, C. 1977a. An historical survey of Early Cycladic research. In J. Thimme (ed). *Art and Culture of the Cyclades*, pp. 185-191. Chicago: The University of Chicago Press.

Doumas, C. 1977b. Early Bronze Age Burial Habits in the Cyclades. *Studies in Mediterranean Archaeology, 48*. Göteborg: Paul Åströms Förlag.

Doumas, C. 1978. The Thera and the Aegean World I. London: Thera and the Aegean World.

Doxey, D. 1987. Causes and effects of the fall of Knossos in 1375 B.C. *Oxford Journal of Archaeology*, 6: 301-317.

Drews, R. 1993. The end of the Bronze Age: Changes in Warfare and the Catastrophe ca. 1200BC. New Jersey: Princeton.

Driessen, J. & MacDonald, C. 1984. Some Military Aspects of the Aegean in the Late Fifteenth and Early Fourteenth Centuries B.C. *Annual of the British School at Athens*, 79: 49-74.

Driessen, J. 1990. An Early Destruction in the Mycenaean Palace at Knossos: A New Interpretation of the Excavation Field-Notes of the South-East Area of the West Wing. *Acta Archaeologica Lovaniensia Monographiae 2.* Leuven, Belgium.

Driessen, J. & MacDonald, C. F. 1997. The Troubled Island. Liège: Austin.

Duckworth, W.L.H. 1903. Excavations at Palaikastro II, Ossuaries at Roussolakkos. *Annual of the British School at Athens*, 9: 350-55.

Eller, E. 1999. Population Substructure and Isolation by Distance in Three Continental Regions. *American Journal of Physical Anthropology*, 108: 147-59.

El-Najjar, M.Y., & Dawson, G.L. 1977. The effect of artificial cranial deformation on the incidence of wormian bones in the lambdoidal suture. *American Journal of Physical Anthropology*, 46: 155-60.

Enlow, D.H. 1990. Facial growth. Philadelphia: Saunders.

Evans, A. 1921. The palace of Minos at Knossos,Vol. I. London: Macmillan and Co.

Evans, A. 1928. The palace of Minos at Knossos,Vol. II. London: Macmillan and Co.

Evans, A. 1930. The palace of Minos at Knossos,Vol. III. London: Macmillan and Co.

Evans, A. 1935. The palace of Minos at Knossos,Vol. IV. London: Macmillan and Co.

Evershed, R., Vaughan, S.I., Dubb, S.N., & Soles, J. 1997. Fuel for Thought? Beeswax in Lamps and conical cups from Late Minoan Crete. *Antiquity*, 71: 979-85.

Ezzo, J.A., Johnson, C.M., & Price, T.D. 1997. Analytical perspectives on prehistoric migration: a case study from east-central Arizona. *Journal of Archaeological Science*, 24: 447-66.

Falk, D., & Corruccini, R. 1982. Efficacy of cranial versus dental measurements for separating human populations. *American Journal of Physical Anthropology*, 57: 123-28.

Faure, G. & Powel, T. 1972. Strontium isotope geology. Springer-Verlag: New York.

Faure, G. 1986. Principles of Isotope Geology. New York: John Wile & Sons.

Finnegan, M. 1974. Discrete non-metric variation of the post-cranial skeleton in man. *American Journal of Physical Anthropology*, 40: 135-6.

Finnegan, M. 1978. Non-metric variation of the infracranial skeleton. *Journal of Anatomy*, 125: 23-37.

Flannery, K. V. 1968. Archaeological systems theory and early Mesoamerica. In B.J. Meggers (ed), *Anthropological archaeology in the Americas*, pp. 67-87. Washington: Anthropological Society of Washington.

Forsen, J. 1992. The Twilight of the Early Helladics: A study of the Disturbances in East-Central and Southern Greece towards the End of the Early Bronze Age. Studies in Mediterranean Archaeology, Pocket-book 116. Jonsered: Paul Åströms Förlag.

Forsdyke, E.J. 1926-7. The Mavro Spelio Cemetery at Knossos. *Annual of the British School at Athens*, 28: 243-96.

Fortuin, A.R. 1977. Stratigraphy and Sedimentary History of the Neogene Deposits in the Ierapetra Region, Eastern Crete. Geologische Uitgaven. Amsterdam Papers of Geology, Series I, 8.

Frayer. D.W., & Wolpoff, M. H. 1985. Sexual Dimorphism. *Annual Reviews in Anthropology*, 14: 429-73.

French, E.B., & Wardle, K.A. 1988. Problems in Greek Prehistory. *Papers Presented at the Centenary Conference of the British School of Archaeology at Athens, Manchester, April 1986.* Bristol: Bristol Classical Press.

French, E. B. 1998. The Ups and Downs of Mycenae: 1250-1150 BCE. In S. Gitin, A. Mazar, & E. Stern (eds), *Mediterranean Peoples in Transition, Thirteenth to Early Tenth Centuries BC*, pp. 2-5. Jerusalem.

Frodin, O. & Persson, A. 1938. Asine: Results of the Swedish Excavations 1922-1930.

Frost, H.M. 1997. Indirect Way to Estimate Peak Joint Loads in Life and in Skeletal Remains (Insights from a New Paradigm). *The Anatomical Record*, 248: 475-483.

Futuyma, D.J. 1995. Evolutionary Biology, 2nd Edition (Translated in Greek). Herakleio: Panepistimiakes Ekdoseis Kritis.

Gale, N.H., & Stos-Gale, Z.A. 1981. Lead and silver in the ancient Aegean. *Scientific American*, 244 (6): 142-52.

Gale, N.H. 1990. The Provenance of Metals for Early Bronze Age Crete – Local or Cycladic? In V. Niniou-Kindel (ed), *Pepragmena toy ST' Diethnoys Kritologikoy Synedrioy (Chania, 24-30 Augoustou 1986)*, Vol. A, pp. 299-316. Chania: Filologikos Syllogos «O Chrysostomos».

Garn, S.M., Guzman, M.A., & Wagner, B. 1969. Subperiosteal gain and endosteal loss in protein-calorie malnutrition. *American Journal of Physical Anthropology*, 30: 153-156.

Giddens, A. 1984. The constitution of society: outline of the theory of structuration. Cambridge: Polity Press.

Gilbert, R., & Gill, G.W. 1990. Metric technique for identifying American Indian femora. In G. W. Gill, & S. Rhine (eds), *Skeletal Attribution of Race: Methods for Forensic Anthropology*, pp. 97-99. Albuquerque, NM: Maxwell Museum of Anthropology.

Gimbutas, M. 1974. The destruction of Aegean and East Mediterranean urban civilization around 2300B.C. In (eds) R. A. Crossland, & A. Birchall, *Bronze Age Migration in the Aegean*, pp. 129-139. Great Britain.

Gödecken, K.B. 1988. A contribution to the early history of Miletus. In E.B. French, & K.A. Wardle (eds), *Problems in Aegean Prehistory. Papers presented at the centenary conference of the British School of Archaeology at Athens, Manchester April 1986*, pp. 307-15. Bristol: Bristol Classical Press.

González-Jose, R., Dahinten, S. L., Luis, M.A., & Hernández, M. 2001. Craniometric Variation and the Settlement of the Americas: Testing Hypotheses by Means of R-Matrix and Matrix Correlation Analyses. *American Journal of Physical Anthropology*, 116: 154-65.

Gould, S. J. 2003. I Have Landed. Splashes and Reflections in Natural History. London: Vintage.

Graustein, W.C. 1989. $^{87}Sr/^{86}Sr$ ratios measure the sources and flow of strontium in terrestrial ecosystems. In P.W. Rundel, J.R. Ehleringer, & K.A. Nagy (eds), *Stable isotopes in ecological research*, pp. 491-512. New York: Springer-Verlag.

Green, R.F., and Myers S.J., 1976. The Use of Inverse Sine Transformations in the Analysis of Non-Metric Cranial Data, *American Journal of Physical Anthropology*, 45: 61-68.

Grewal, M.S. 1962. The rate of genetic divergence of sublines in the C57BL strain of mice. *Genetical Research*, 3: 226-37.

Grüneberg, H. 1952. Genetical studies on the skeleton of the mouse. Quasicontinuous variations. *Journal of Genetics*, 51: 95-114.

Grüneberg, H. 1963. The pathology of development. Oxford: Blackwell.

Grupe, G., Price, T.D, Schorter, P., Sollner, F., Johnson, C. & Beard, B. 1997. Mobility of Bell Beaker people revealed by stable strontium isotope ratios of teeth and bones. A study of southern Bavarian skeletal remains. *Applied Geochemistry*, 12: 517-25.

Gulson, B.L., Jameson, C.W., & Gillings, B.R. 1997. Stable lead isotopes in teeth as indicators of past domicile-a potential new tool in forensic science? *Journal of Forensic Science*, 42: 787-91.

Hägg, R. & Marinatos, N. 1984. The Minoan Thalassocracy: Myth and Reality. *Proceedings of the Third International Symposium at the Swedish Institute in Athens, 31 May-5 June, 1982. Skrifter Utgivna av Svenska Institutet i Athen, 4^{o}, XXXII.* Stockholm: Paul Åströms Förlag.

Hallager, E. 1988. Final palatial Crete. An essay in Minoan Chronology. In A. Damsgaard-Madsen, E. Christiansen, & E. Hallager, (eds). *Studies in Ancient History and Numismatics presented to Rudi Thomsen*, pp. 11-21. Aarhus: Aarhus University Press.

Hallager, B.P., & McGeorge, T. 1992. Late Minoan III Burials at Khania. The tombs, finds and deceased in Odos Palama. Studies in Mediterranean Archaeology, Vol. XCIII. Göteborg: Paul Åströms Förlag.

Hallgrímsson, B., Ó Donnbhain, B., Walters, G.B., Cooper, D.M.L., D. Guobjartsson, & Stefánsson, K. 2004. Composition of the Founding population of Iceland: Biological Distance and Morphological Variation in Early Historic Atlantic Europe. *American Journal of Physical Anthropology*, 124: 257-74.

Halstead, P. 1977. Demography in the study of past populations economy and society. *Annual of the British School at Athens*, 72: 107-111.

Halstead, P. 1981. From determinism to Uncertainty: Social Storage and the Rise of the Minoan Palace. In A. Sheridan, & G. Bailey (eds), *Economic Archaeology*, pp. 187-213. British Archaeological Reports, 96.

Halstead, P. 1996. The development of agriculture and pastoralism in Greece: when, how, who and what? In D.R. Harris (ed), *The Origins and Spread of Agriculture and Pastoralism in Eurasia*, pp. 296-309. London: UCL Press.

Hamilakis, Y. 2002a. What Future For the 'Minoan' Past? Rethinking Minoan Archaeology. In (ed) Y. Hamilakis, Y. *Labyrinth Revisited. Rethinking 'Minoan' Archaeology*. Oxford: Oxfow Books.

Hamilakis, Y. 2002b. Too many Chiefs? Factional Competition in Neopalatial Crete. In J. Driessen, I. Schoep, & R. Laffineur (eds), *Monuments of Minos. Rethinking the Minoan Palaces. Proceedings of the International Workshop "Crete of the hundred Palaces?" held at the Universite Catholique de Louvain, Louvain-la-Neuve, 14-15 December 2001*, pp. 179-99.

Hanihara, T. 1993. Population Prehistory of East Asia and th ePacific as Viewed from Craniofacial Morphology: The Basic Populations of East Asia, VII. *American Journal of Physical Anthropology*, 91: 173-187.

Hanihara, T. 1994. Craniofacial continuity and disconuity of Far Easterns in the late Pleistocen and Holocene. *Journal of Human Evolution*, 27: 417-441.

Hanihara, T. 1996. Comparison of craniofacial features of major human groups. *American Journal of Physical Anthropology*, 99: 389-412.

Hanihara, T. 1997. Craniofacial affinities of Marian Islanders and circum-Pacific peoples. *American Journal of Physical Anthropology*, 104: 411-25.

Hanihara, T. & Ishida, H. 2001a. Frequency variations of discrete traits in major human populations: I. Supernumerary ossicle variations. *Journal of Anatomy*, 198: 689-706.

Hanihara, T. & Ishida, H. 2001b. Frequency variations of discrete traits in major human populations: II. Hypostotic variations. *Journal of Anatomy*, 198: 707-25.

Hanihara, T. & Ishida, H. 2001c. Frequency variations of discrete traits in major human populations: III. Hyperostotic variations. *Journal of Anatomy*, 199: 251-72.

Hanihara, T. & Ishida, H. 2001d. Frequency variations of discrete traits in major human populations: IV. Vessel and nerve related variations. *Journal of Anatomy*, 199: 273-87.

Hanihara, T. & Ishida, H. 2005. Metric dental variation of major human populations. *American Journal of Physical Anthropology*, 128: 287-98.

Hankey,V. 1967. Mycenaean pottery in the Middle East: notes on finds since 1951. *Annual of the British School at Athens*, 62: 107-47.

Hankey, V. 1973. Late Minoan finds in the South-east Mediterranean. *Pepragmena toy G' Dietnoys Kritologikoy Synedrioy (Rethymno 18-23 Septemvrioy 1971)*, Vol. 1, pp. 104-10. Athens.

Hansen, J. M. 1988. Agriculture in the prehistoric Aegean: data versus speculation. *American Journal of Archaeology*, 92: 39-52.

Harpending, H.C., & Ward, R.H. 1982. Chemical systematics and human populations. In M. Nitecki (ed), *Biochemical Aspects of Evolutionary Biology*, pp. 213-56. Chicago: University of Chicago.

Harris, E.F. & Rathburn, T.A. 1991. Ethnic differences in the apportionment of tooth sizes. In M.A. Kelley, & C.S. Larsen (eds), *Advances in Dental Anthropology*, pp. 121-42. New York: Wiley-Liss.

Hartenberger, B., & Runnels, C. 2001. The Organization of Flaked Stone Production at Bronze Age Lerna. *Hesperia*, 70(3): 255-83.

Hauser, G., & De Stefano, G.F. 1989. Epigenetic Variants of the Human Skull. Stuttgart: E. Schweizerbart'sche Verlagsbuchhandlung.

Hadjianastasiou, O. 1996. A Mycenaean Pictorial Vase from Naxos. *Atti e memorie d'el secondo congresso internazionale di Micenologia, Roma-Napoli, 1991*, pp. 1433-41. Roma: Gruppo Editoriale Internazionale.

Hemphill, B.E. 1999. Biological affinities and adaptations of Bronze Age Bactrians: IV. A craniometric investigation of Bactrian origins. *American Journal of Physical Anthropology*, 108: 173-92.

Hertzog, K. 1968. Associations between discontinuous cranial traits. *American Journal of Physical Anthropology*, 29: 397-404.

Hides, D.S. 1995. Consuming Identities. *Sociology Review*, 5(2): 30-3.

Hiernaux, J. 1972. The analysis of multivariate biological distances between human populations: Principles, and applications to sub- Saharan Africa. In J.S. Weiner, & J. Huizinga (eds), *The assessment of populations affinities in man*, pp. 96-114. Oxford: Clarendon Press.

Higa, T., Hanihara, T., Sunakawa, H., & Ishida, H. 2003. Dental Variation of Ryukyu Islanders: A Comparative Study Among Ryukyu, Ainu, and Other Asian Populations. *American Journal of Human Biology*, 15: 127-43.

Higgins, M.D. & Higgins, R. 1996. A Geological Companion to Greece and the Aegean. London: Duckworth.

Hill, P.A. 1998. Bone remodeling. British Journal of Orthopaedics, 25: 101-7.

Hiller, E.S. 1977. Das minoische Kreta nach den Augrabungen des letzten Jahrzehntes. Vienna.

Hillson, S. 2002. Dental anthropology (3rd Edition). Cambridge: Cambridge University Press.

Hillson, S., FitzGerald, C., & Flinn, H. 2005. Alternative Dental Measurements: Proposals and relationships with other measurements. *American Journal of Physical Anthrpology*, 126: 413-26.

Himes, J.W., Matorell, R., Habicht, J-P., Yarbrough, C., Malina, R.M., & Klein, R.E. 1975. Patterns of cortical bone growth in moderately malnourished preschool children. *Human Biology*. 47: 337-350.

Hiorns, R.W., Harrison, G.A., Boyce, A.J. & Kuchemann, C.F. 1969. A mathematical analysis of the effects of movement on the relatedness between populations. *Annual of Human Genetics*, 32: 237-51.

Hofreiter, M., Serre, David, Poinar, H.N., Kuch, M., & Paabo, S. 2001. Ancient DNA. Nature Reviews, *Genetics*, 2: 353-8.

Hood, M.S.F., & Boardman, J. 1955. Archaeology in Greece, 1955. *Archaeological Reports*, 2: 3-38.

Hood, S., Huxley, G., Sandars N.1958-59. A Minoan Cemetery on Upper Gypsades (Knossos Survey 156). *Annual of the British School at Athens*, 53-54: 194-262.

Hood, M.S.F. 1961. Archaeology in Greece, 1960-61. *Archaeological Reports*, 7: 26-27.

Hood, M.S.F. 1962. Archaeology in Greece, 1961-62. *Archaeological Reports*, 8: 25-27.

Hood, M.S.F. 1973. The Eruption of Thera and its Effects in Crete in Late Minoan I. *Ta Pepragmena tou G' Diethnous Kritologikou Synedriou (Rethymno 18-23 Septemvrioy 1971)*, Vol.1, pp. 111-118, Athens.

Hood, M.S.F., & Smyth, D. 1981. Archaeological Survey of the Knossos area. *British School at Athens Supplementary Volume 14*. London: Thames and Hudson.

Hood, S., & Taylor, W. 1981. The Bronze Age Palace at Knossos. London: The British School at Athens. London: Thames and Hudson.

Hood, M.S.F. 1985. Warlike Destruction in Crete c.1450 B.C. *Ta Pepragmena tou E' Diethnoys Kritologikoy Synedrioy (Agios Nikolaos, 25 Septemvrioy – 1 Oktovrioy 1981)*, Vol. 1, pp. 170-78.Irakleio: Etairia Kritikon Istorikon Meleton.

Hood, S. 1986. Evidence for invasions. In G. Cadogan (ed), *The End of the Early Bronze Age in the Aegean*, pp. 31-68. Leiden: E.J. Brill.

Hope Simpson, R., & Dickinson, O.T.P.K. 1979. A Gazetteer of Aegean Civilization in the Bronze Age 1: The Mainland and the Islands. *Studies in Mediterranean Archaeology*, 52. Göteborg: Paul Åströms Förlag.

Howell, R.J. 1992. Middle Helladic Settlement: Stratigraphy and Architecture. In W.A. McDonald, & N.C. Wilkie (eds), *Excavations at Nichoria in Southwest Greece II: The Bronze Age Occupation*, pp. 15-42. Minneapolis.

Howells, W.W.1973. Cranial Variation in Man: A Study by Multivariate Analysis of Patterns of Difference among Recent Human Populations. Harvard University, *Papers of the Peabody Museum of Archaeology and Ethnology*, 67.

Howells, W.W. 1986. Physical anthropology of the prehistoric Japanese. In R. J. Pearson, G.L.Barnes, & K.L.Hutterer (eds), *Windows on the Japanese Past*, pp. 85-90. Ann Arbor:Center for Janpanese Studies , University of Michigan.

Howells, W.W. 1989. Skull Shapes and the map: Craniometric analysis in the dispersion of modern Homo. *Papers of the Peabody Museum of Archaeology and Ethnology*, 79. Cambridge: Harvard University Press.

Iakovides, S. 1983. Late Helladic Citadels on Mainland Greece. Leiden.

Iakovides, S. 1986. Destruction Horizons at Late Bronze Age Mycenae. In *Filia Epe es G.E. Mylonan I.*, pp. 233-60. En Athinais Archaeologike Etaireia, Vivliothike, 103. (Φίλια 'Επη εις Γ.Ε. Μυλωνάν Ι. Εν Αθήναις Αρχαιολογική Εταιρεία, Βιβλιοθήκη, 103).

Immerwahr, S. A. 1960. Mycenaean trade and colonization. *Archaeology*, 13 (1): 4-13.

Irish, J. D. 2005. Population Continuity vs. Discontinuity Revisited: Dental Affinities Among Late Paleolithic Through Christian-Era Nubians. *American Journal of Physical Anthropology*, 128: 520-35.

Irish, J.D. 2006. Who were the ancient Egyptians? Dental affinities among Neolithic through Postdynastic peoples. *American Journal of Physical Anthropology*, 129: 529-43.

Ishida, H. & Kondo, O. 1998. Human Cranial Variations Based on Different Sets of Measurement Variables. *Archaeological Science*, 106: 41-60.

Israel, H. 1977. The dichotomous pattern of craniofacial expansion during aging. *American Journal of Physical Anthropology*, 47: 47-52.

J. M. Hansen. 1988. Agriculture in the Prehistoric Aegean: Data versus speculation. *American Journal of Physical Anthropology*, 92:39-52.

Jacobs, K. 1993. Human Postcranial Variation in the Ukrainian Mesolithic-Neolithic. *Current Anthropology*, 34 (3): 311-24.

Jameson, M.H. 1976. The Southern Argolid: The setting for Historical and Cultural Studies. In M. Dimen, & E. Freidl (eds), *Regional Variations in Modern Greece and Cyprus: Towards a Perspective on the Ethnography of Greece*. New York: Academy of Sciences Annals 268.

Jantz, R.L., Hunt, D.R., Falsetti, A.B, & Key, P.J. 1992. Variation among northern Amerindians: Analysis of Boas's anthropometric data. *Human Biology*: 64: 435-61.

Johnson, A.L., & Lovell, N.C. 1994. Biological differentiation at Predynastic Naqada, Egypt: An analysis of dental morphological traits. *American Journal of Physical Anthropology*, 93: 427-33.

Jones, S. 1997. The archaeology of ethnicity constructing identities in the past and present London: Routledge.

Jones, S. 2000. The language of the genes. Great Britain: Clays Ltd, St Ives plc.

Jorde, L.B., Bamshad, M.J., Watkins, W.S., Zenger, R., Fraley, A.E., Krakowiak, P.A., & Carpenter, K.D. 1995. Origins and affinities of modern humans: a comparison of mitochondrial and nuclear genetic data. *American Journal of Human Genetics*, 57: 523-38.

Jorgensen, N.O., Morthorst, J., & Holm, P.M. 1999. Strontium isotope studies of "brown water" (organic-rich groundwater) from Denmark. *Hydrogeology Journal*, 7: 533-9.

Kanta, A. 1980. The Late Minoan III Period on Crete: A Survey of Sites, Pottery and their Distribution. Studies in Mediterranean Archaeology, 58. Göteborg: Paul Åströms Förlag.

Karagiorgis, B. 1992. Kypros. To Stavrodromi tis Anatolikis Mesogeioy, 1600-500π.X (Κύπρος. Το Σταυροδρόμι της Ανατολικής Μεσογείου, 1600-500π.X). Milan: Mondadori Electa Spa.

Kardara, K. 1977. Keneta euvrimata tafon A and B (Κινητά ευρήματα τάφων Α και Β). Athena. Etnike Archaeologike Etaireia.

Kasai, K., Kanazawa, E., Aboshi, H., Tuisuva, J., Takahashi, M., & Matsuno, M. 1988, Comparative Study of Craniofacial Morphology and Bite Force in Fijians and Japanese, *American Journal of Human Biology*, Vol. 10, 33-72.

Kasai, K. Richards, L.C., Brown, T. 1993. Comparative study in Japanese and Australian Aboriginal populations. *Human Biology*, 65: 821-834.

Keita, S. O. Y. 1988, An Analysis of Crania from Tell-Duweir Using Multiple Discriminant Functions, *American Journal of Physical Anthropology*, 75: 375-390.

Kelley, M.A., & Larsen, C. S. 1991. Advances in Dental Anthropology. New York: Wiley-Liss.

Key, P.J. 1983. Craniometric Relationships among Plains Indians. University of Tennessee. Department of Anthropology, *Reports of Investigations*, 34.

Kieser, J.A. 1990. Human adult odontometrics. Cambridge: Cambridge University Press.

Kilian-Dirlmeier, I. 1985. Noch einmal zu den 'Kriegergräbern' von Knossos. *Jahrbuch des Römisch-Germanischen Zentralmuseums Mainz*, 32: 196-214.

Kilian, K. 1988. Mycenaeans Up to Date, Trends, and Changes in Recent Research. In E.B. French & K.A. Wardle (eds), *Problems in Greek Prehistory*, pp. 115-52. Bristol: Bristol Classical Press.

Killen, J.T. 1985. The Linear B tablets and the Mycenaean economy. In A.D. Morpurgo & Y. Duhoux (eds). *Linear B: a 1984 survey*, pp. 241-305. Biblioteque des Cahiers de l'Institut de Linguistique de Louvain 26. Louvain-la-Neuve : Cabay and Publications Linguistiques de Louvain.

Knapp, A.B. 1993. Thalassocracies in Bronze Age Eastern Mediterranean Trade: Making and Breaking a Myth. *World Archaeology*, 24: 332-47.

Knappett, C. 1999. Assessing a Polity in Protopalatial Crete: The Malia-Lasithi State. *American Journal of Archaeology*, 103:615-39.

Koch, P.L., Behrensmeyer, A.K., Tuross, N., & Fogel, M.L. 1990. Isotopic fidelity during bone weathering and burial. Annual Report of the Geophysical Lab. Washington, DC: Carnegie Institute.

Koch, P.L., Tuross, N., & Fogel, M.L. 1997. The effects of sample treatment and diagenesis on the isotopic integrity of carbonate in biogenic hydroxylapatite. *Journal of Archaeological Science*, 24: 417-29.

Kohn, L.A.P. 1991. The role of genetics in craniofacial morphology and growth. *Annual Review of Anthropology*: 20: 261-78.

Kohn, M.J., Schoninger, M.J.,& Barker, W.W. 1999. Altered states: effects of diagenesis on fossil tooth chemistry. *Geochimica et Cosmochimica Acta*, 63: 2737-47.

Kolata, G.B. 1977. Catastrophe theory: The emperor has no clothes. Science,

Kondoleon, N.M. 1958. Ανασκαφαί Νάξου. *Praktika tis en Athinais Archaeologikis Etaireias*: 228-9.

Kondoleon. N.M. 1959. Ανασκαφαί Νάξου. *Praktika tis en Athinais Archaeologikis Etaireias*: 180-7.

Kondoleon, N. M. 1961. Μυκηναική Νάξος. *Praktika tis en Athinais Archaeologikis Etaireias:* 600-8.

Konigsberg, L.W.1987. Population genetic models for interpreting prehistoric intra-cemetery biological variation. PhD Thesis. Illinois.

Konigsberg, L.W., & Ousley, S.D. 1993. Multivariate quantitative genetics of anthropometrics from the Boas data. *American Journal of Physical Anthropology Supplement*, 16: 127-28.

Konigsberg, L.W., & Ousley, S.D.1995. Multivariate quantitative genetics of anthropometric traits from the Boas data. *Human Biology*, 67: 481-498.

Konsolaki- Yannopoulou, E. 2001. New evidence for the practice of libations in the Aegean Bronze Age. In R. Laffineur, & R. Hagg (eds), *Potnia. Deities and Religion in the Aegean Bronze Age. Proceedings of the 8th International Aegean Conference. Aegaeum*, 22: 213-219.

Konsolaki- Yannopoulou, E. 2003. Η Μαγούλα στον Γαλατά της Τροιζηνίας. Ένα νέο ΜΕ-ΥΕ Κέντρο στον Σαρωνικό. In E. Konsolaki- Yannopoulou (ed), *Argosaronikos. Praktika 1st Diethoys Synedrioy Istorias kai Arxaiologias toy Saronikou*, Vol. A, pp.159-181. Athens.

Kossinna, G. 1911. Dei Herkunft der Germanen. Leipzig: Kabitzsch.

Kotsakis, K. 1991. The powerful past: theoretical trends in Greek archaeology. In I. Hodder, ed. Archaeological Theory in Europe: the last three decades, pp. 65-90. London: Routledge.

Koumaris, J. 1948. On the Morphological Variety of Modern Greeks. *Man*, 48: 126-7.

Krause, W. 1880. Anatomische Varietaten. Hanover: Hahn'sche Hofbuchhandlung.

Krogman, W.M, & İşcan, M.Y. 1986. The Human Skeleton in Forensic Medicine. Springfield, Illinois: C.C. Thomas.

Kruts, S.I. 1984. Paleoantropologicheskie issledovaniya stepnogo pridneprovya. Kiev: Naukova Dumka.

Kuniholm, P.I.1990. Archaeological evidence and non-evidence for climate change. Philosophical Transactions of the Royal Society of London, Series A, 330: 645-55.

Lahr, M.M. 1995. Patterns of modern human diversification: implications for Amerindian origins. *Yearbook of Physical Anthropology*, 38: 163-98.

Lahr, M.M. 1996. The evolution of modern human diversity. Cambridge: Cambridge University Press.

Larsen, C. S. 1999. Bioarchaeology. Interpreting behavior from the human skeleton. Cambridge: Cambridge University Press.

Latter, B.D.H. 1980. Genetic differences within and between populations of the major human subgroups. *American Naturalist*, 116: 220.

Lewontin, R.C. 1972. The apportionment of human diversity. In T. Dobzhansky, M.K. Hecht, W.C. Steere (eds), *Evolutionary Biology*, 6, pp. 381-398. New York: Appleton-Century-Crofts.

Lieberman, D.E. 1996. How and why humans grow thin skulls: experimental evidence for systemic cortical robusticity. *American Journal of Physical Anthropology*, 101: 217-36.

Lloyd-Jones, J. 1997. Calculating bio-distance using dental morphology. In S. Anderson, & K. Boyle (eds), *Computing and Statistics in Osteoarchaeology. Proceedings of the second meeting of the Osteoarchaeological Research Group held in London on 8th April 1995*, pp. 23-34.

Lovejoy, C.O., Meindl, R.S, Pryzbeck, T.R, & Mensforth, R.P. 1985. Chronological metamorphosis of the auricular surface of the ilium: A new method for the determination of adult skeletal age at death. *American Journal of Physical Anthropology*, 68: 1-14.

Lukacs, J.R. 1985. Tooth size variation in Prehistoric India. *American Anthropologist*, 87: 811-25.

Macbeth, H. 1993. Ethnicity and human biology. In M. Chapman (ed), *Social and Biological Aspects of Ethnicity*, pp. 47-91. New York: Oxford University Press,

Macdonald, C.F. 2002. The Neopalatial Palaces of Knossos. In J. Driessen, I. Schoep, & R. Laffineur (eds), Monuments of Minos. *Rethinking the Minoan Palaces. Proceedings of the International Workshop "Crete of the hundred Palaces?" held at* the *Universite Catholique de Louvain, Louvain-la-Neuve, 14-15 December 2001*, pp. 35-54.

MacGillivray, J.A., & Sackett, L.H. 1994. Report on the 1994 Excavations at Palaikastro, British School at Athens. Annual Report of the Managing Committee for the Season 1993-94, pp. 20-21.

MacKenzie, D. 1904. The successive settlements at Phylakopi in their Aegeo-Cretan relations. In T.D. Atkinson, R.C. Bosanquet, R.C. Edgar, A.J. Evans, D.G. Hogarth, D. MacKenzie, C. Smith, & F.B. Welch (eds). *Excavations at Phylakopi in Melos conducted by the British School at Athens*, pp. 238-72. London: Macmillan.

Manning, S.W. 1986. The Military Function in Late Minoan I Crete: A Note. *World Archaeology*, 18: 284-88.

Manning, S.W., & Weninger, B. 1992. A light in the dark: archaeological wiggle matching and the absolute chronology of the close of the Aegean Late Bronze Age. *Antiquity*, 66: 636-63.

Manning, S.W. 1994. The emergence of the divergence: development and Decline on the Bronze Age Crete and the Cyclades. In C. Mathers, & S. Stodart (eds), *Development and Decline In the Mediterranean Bronze Age*, pp. 221-270. Sheffield Archaeological Monographs 8.University of Sheffield: J.R. Collis.

Manning, S.W. 1995. The Absolute Chronology of the Aegean Early Bronze Age. Sheffield: Sheffield University Press.

Manolis, S. 1991. Anthropological research on the composition of Bronze Age populations from Southern Greece. Athens: Unpublished PhD thesis.

Manzi, Giorgio and Vienna, Alessandro, 1997, Cranial non-metric traits as indicators of hypostosis or hyperotosis, *Rivista di Antropologia* (Roma), Vol. 75, 41-61.

Marinatos, S. 1939. The volcanic destruction of Minoan Crete. *Antiquity*, 13: 425-39.

Marinatos, N. 1993. Minoan Religion. Ritual, Image and Symbol. Columbia: University of South Carolina Press.

Mariolopoulos, H. 1953. Episkopisis toy Klimatos tis Ellados (Επισκόπησις του Κλίματος της Ελλάδος). Athens.

Mascie-Taylor. C.G.N, Madsen. H., & Mohamed. G.H. 1995. Data Handling and Biostatistics: Use of SPSS for Windows. Cambridge: Department of Biological Anthropology.

Matsumura, H., & Hudson, M.J. 2005. Dental Perspectives on the Population History of Southeast Asia. *American Journal of Physical Anthropology*, 127: 182-209.

Matthäus, H. 1983. Minoische Kriegergräber. In O, Krzyszkowska, & L. Nixon, (eds). *Minoan Society. Proceedings of the Cambridge Colloquium 1981*. Bristol: Bristol Classical Press, pp. 203-15.

Mays, S. 1998. The Archaeology of Human Bones. London: Routledge/English Heritage.

McDonald, W.A., & Rapp, G.R. 1972. The Minnesota Messenia Expedition: Reconstructing a Bronze Age Regional Environment. Minneapolis: The University of Minnesota Press.

McDonald, W.A. & Wilkie, N. 1992. Excavations at Nichoria II: The Bronze Age Occupation. Minneapolis: The University of Minnesota Press.

McGeorge, T. 1983. The Minoans: physical variation and affinities. London: Unpublished Phd thesis, University College of London.

McGeorge, T. 1988. Health and diet in Minoan times. In R.E. Jones, & H.W. Catling (eds), *New aspects of Archaeological science in Greece. Proceedings of a Meeting held at the British School at Athens. January 1987*, pp. 47-54. Athens: Occasional Paper 3 of the Fitch Laboratory.

McKern, T., & Stewart, T.D. 1957. Skeletal Age Changes in Young American Males, Analyzed from the Standpoint of Identification. Technical Report EP-45. Massachusetts: Headquarters, Quartemaster Research and Development Command.

Mee, C. 1982. Rhodes in the Bronze Age. Warminster Wits.

Mee, C. 1988. A Mycenaean thalassocracy in the eastern Aegean? In E.B. French & K.A. Wardle (eds). *Problems in Greek Prehistory. Papers Presented at the Centenary Conference of the British School of Archaeology at Athens, Manchester April 1986*,pp. 301-6. Bristol: Bristol Classical Press.

Melas, E. 1985. The Islands of Karpathos, Saros and Kasos in the Neolithic and Bronze Age. Göteborg: Paul Åströms Förlag.

Mellaart, J. 1971. Prehistory of Anatolia and its relations with the Balkans. *Studia Balcanica. L'ethnogenese des peoples balkaniques*, pp. 119-37. Sofia: Academie Bulgare des Sciences.

Mellaart, J. 1975. The Neolithic of the Near East. London: Thames & Hudson.

Mellersh, H.E.L. 1970. The Destruction of Knossos. London.

Mellink, M. 1986. The Early Bronze Age in Western Anatolia. In G.Cadogan (ed), *The End of the Early Bronze Age*, pp. 139-52. Leiden.

Molnar, S. 1971. Human Tooth Wear, Tooth Function and Cultural Variability. *American Journal of Physical Anthropology*, 34: 175-90.

Molto, J.E. 1983. Biological relationships of southern Ontario Woodland peoples: the evidence of discontinuous cranial morphology. Ottawa: National Museum of Man.

Momigliano, N. 1991. MMIA pottery from Evans' excavations at Knossos: a reassessment. *Annual of the British School at Athens*, 86: 149-271.

Montelius, O. 1903. Die Typologische Methode: Die älteren kulturperioden im Orient und in Europa, Vol.I. Stockholm: Selbstverlag.

Moody, J., Rackham, O., & Rapp, G.J. 1996. Environmental Archaeology of Prehistoric NW Crete. *Journal of Field Archaeology*, 23(3): 273-297.

Moorrees, C.F.A. 1957. The Aleut dentition. Cambridge, Massachusetts: Harvard University Press.

Musgrave, J.H. 1980. By strangers honor'd: a statistical study of ancient crania from Crete, Mainland Greece, Cyprus, Israel and Egypt. *Journal of Mediterranean Anthropology and Archaeology*, 1: 50-107.

Musgrave, J.H. 1985. The skull of Philip II of Macedon. In S.J.W. Linsey & B. Mathews (eds), *Current Topics in Oral Biology*, pp. 1-16. Bristol: Press of the University of Bristol.

Musgrave, J.H. 1990. The cremated remains from Tombs II and III at Nea Michaniona and Tomb Beta at Derveni. *Annual of the British School at Athens*, 85: 301-25.

Musgrave, J.H., Neave, R.A.H, Prag, A.J.N.W., Sakellarakis, E., & Sakelarakis, J.A. 1994. The priest and priestess from Archanes-Anemospilia:reconstructing Minoan faces. *Annual of the British School at Athens*, 89: 89-100.

Musgrave, J. H., Neave, R. A. H., & Prag, A. J. N. W. 1995. Seven faces from Grave Circle B at Mycenae. *Annual of the British School at Athens*, 90: 107-136.

Najem, W.J. 1997. The correlation of craniofacial form between siblings. Unpublished MS thesis, Department of Orthodontics, Case Western Reserve University.

Nevros, K.I., & Zvorykin, I.A. 1939. Zur Kenntnis der Boeden der Insel Kreta. *Soil Research*, 6: 242–307.

Niemeier, W.D. 1982. Mycenaean Knossos and the age of Linear B. *Studi miceni ed anatolici*, 23: 219-287.

Niemeier, W.D. 1983. The character of the Knossian palace society in the second half of the fifteenth century B.C.: Mycenaean or Minoan? In O, Krzyszkowska, & L. Nixon (eds), *Minoan Society. Proceedings of the Cambridge Colloquium 1981*, pp. 217-36. Bristol: Bristol Classical Press.

Niemeier, W.D. 1985. The archaeological dating of the Knossian Linear B tablets reconsidered. A Summary. In Th. Detorakis (ed), *Pepragmena tou E' Diethnous Kritologikou Synedriou (Agios Nikolaos, 25 Septemvriou - 1 Oktovriou 1981)*, pp. 259-263. Irakleio, Kritis: Etairia Kritikon Istorikon Meleton..

Osborne, R.H., Horowitz, S.L., & de George, F.V. 1958. Genetic variation of tooth dimensions: a twin study of the permanent anterior teeth. *American Journal of Human Genetics*, 10: 350-56.

Ossenberg, N.S. 1969. Discontinuous morphological variation of the human cranium. Unpublished PhD dissertation. University of Toronto.

Ossenberg, N.S. 1970. The influence of artificial cranial deformation on discontinuous morphological traits. *American Journal of Physical Anthropology*, 33: 357-71.

Ossenberg, N.S. 1976. Within and between race distances in population studies based on discrete traits of human skull. *American Journal of Physical Anthropology*, 45: 701-16.

Owsley, D. W., & Jantz, R. L. 1994. Skeletal Biology in the Great Plains. Migration, Warfare, Health, and Subsistence. United States of America.

Papadopoulou, E. 1997. Μαργαρίτες. *Archaeologikon Deltion*, 52 (Chronika, B3): 1040-41.

Papathanasiou, A. 2001. A bioarchaeological analysis of Neolithic Alepotrypa cave, Greece. British Archaeological Reports International Series 961.

Parfitt, A.M. 1983. The physiologic and clinical significance of bone data. In R. R. Recker (ed), *Bone histomorphometry: techniques and interpretation*, pp. 143-223. CRC Press, Boca Raton, FL.

Pearson, O.M. 2000. Postcranial Remains and the Origins of Modern Humans. *Evolutionary Anthropology*, 9: 229-47.

Pendlebury, J.D.S. 1939. The Archaeology of Crete. London: Methuen.

Pe-Piper & Piper, G. 2002. The Igneous Rocks of Greece. The anatomy of an orogen. *Beiträge zür Regionalen Geologie der Erde. Band 30.* Berlin.

Perizonius, W.R.K. 1984. Closing and Non- Closing Sutures in 256 Crania of Known Age and Sex from Amsterdam (A.D. 1883-1909). *Journal of Human Evolution*, 13: 201-206.

Perlés, C. 1987. Les industries lithiques taillées de Franchthi (Argolide, Grèce) I : Présentation générale et industries Paléolithiques. Excavations at Franchthi Cave, Greece, Fascicle 3.Bloomington: Indiana University Press.

Perzigian, A.J. 1984. Human odontometric variation: an evolutionary and taxonomic assessment. *Anthropologie*, 22; 193-98.

Philaniotou, O. in press. Naxos, Tsikniades: An Early Cycladic Cemetery.

Pietrusewsky, M. 1990. Craniofacial Variation in Australasian and Pacific Populations. *American Journal of Physical Anthropology*, 82: 319-40.

Pietrusewsky, M., & Douglas, M.T. 1993. The skeletal biology of an historic Hawaiian cemetery: Familial relationships. *Homo*, 43: 245-62.

Platon, N. 1952. Ανασκαφή ΥΜΙΙΙ λαξευτών τάφων εις την περιοχή Επισκοπής και Σταμνίων Πεδιάδος Ηρακλείου. *Praktika tis en Athinais Archaeologikis Etaireias:* 619-648.

Popham, M., Catling, E.A., & Catling, H.W. 1974. Sellopoulo Tombs 3 and 4. Two late Minoan graves near Knossos. *Annual of the British School at Athens*, 69: 195-257.

Popham, M. 1975. Late Minoan II Crete: A Note. *American Journal of Archaeology*, 79: 372-74.

Popham, M.R. 1976. Mycenaean-Minoan Relations between 1450 and 1400 BC. *Bulletin of the Institute of Classical Studies of the University of London*, 23: 119-21.

Popham. M.R. 1994. Late Minoan II to the End of the Bronze Age. In D. Evely, H. Hughes-Brock, & N. Momigliano (eds), *Knossos: A Labyrinth of History. Papers Presented in Honour of Sinclair Hood, British School at Athens*, pp. 89-103. Oxford: Oxbow Books.

Potter, R.H.Y., Yu, P.L., Dahlberg, A.A., Merritt, A.D., & Connelly, P.M. 1968. Genetic studies of tooth size factors in Pima Indian families. *American Journal of Human Genetics*, 20: 89-100.

Potter, R.Y. 1972. Univariate versus multivariate differences in tooth size according to sex. *Journal of Dental Research*, 51: 716-22.

Potter, R.H., Alcazaren, A.B., Herbosa, F.M., &, Tomaneng, J. 1981. Dimensional Characteristics of the Filipino Dentition. *American Journal of Physical Anthropology*, 55 (1): 33-42.

Powell, J.R. 1989. Metric versus Non-Metric Skeletal Traits. Which is the more reliable indicator of Genetic Distance ? With special reference to crania from ancient Greece and Egypt. Unpublished PhD Thesis, Department of Anatomy, Bristol Medical School.

Powell, J. F., & Neves, W.A. 1999. Craniofacial Morphology of the First Americans: Pattern and Process in the Peopling of the New World. *Yearbook of Physical Anthropology*, 42: 153-88.

Pretty, G.L., Henneberg, M., & Lambert, K.M. 1998. Trends in stature in the South Australian Aboriginals Murraylands. *American Journal of Physical Anthropology*, 106: 505-14.

Price, T.D., Johnson, C.M., Ezzo, J.A., Burton, J.H., & Ericson, J.A. 1994. Residential mobility in the late prehistoric Southwest: A preliminary study using strontium isotope ratios. *Journal of Archaeological Science*, 24: 315-330.

Price, T.D., Grupe, G., & Schröter, P. 1998. Migration and mobility in the Bell Beaker period in Central Europe. *Antiquity*, 72: 405-11.

Price, T.D., Manzanilla, L., & Middleton, W.H. 2000. Residential mobility at Teotihuacan: a preliminary study using strontium isotopes. *Journal of Archaeological Science*, 27: 903-13.

Price, T.D., Burton, J.H., & Bentley, R.A. 2002. The characterization of biologically available strontium isotope ratios for the study of prehistoric migration. *Archaeometry*, 44: 117-35.

Prowse, T.L., & Lovell, N.C. 1996. Concordance of Cranial and Dental Morphological Traits and Evidence for Endogamy in Ancient Egypt. *American Journal of Physical Anthropology*, 101: 237-46.

Pullen, D.J. 1987. Asine, Berbati, and the Chronology of Early Bronze Age Greece. *American Journal of Archaeology*, 91: 533-44.

Rackham, O., & Moody, J. 1996. The making of the Cretan landscape. Manchester: Manchester University Press.

RehaK, P., & Younger, J.G. 2001. In T.Cullen (ed), *Aegean Prehistory: a review*, pp. 383-465. Boston: Archaeological Institute of America.

Relethford, J.H, & Lees, F.C. 1982. The Use of Quantitative Traits in the Study of Human Population Structure. *Yearbook of Physical Anthropology*, 25: 113-132.

Relethford, J.H., & Blangero, J. 1990. Detection of differential gene flow from patterns of quantitative variation. *Human Biology*, 62: 5-25.

Relethford, J.H. & Harpending, H.C. 1994. Craniometric Variation, Genetic Theory, and Modern Human Origins. *American Journal of Physical Anthropology*, 95: 249-70.

Relethford, J.H, Crawford, M.H, & Blangero, J. 1997. Genetic Drift and Gene Flow in Post-Famine Ireland. *Human Biology*, 69: 443-65.

Renfrew, C. 1972. The emergence of civilization: The Cyclades and the Aegean in the Third Millennium BC. London: Methuen.

Renfrew, C. 1977. Seeds from Area K. In J.E. Coleman (ed), *Keos I: Kephala*, pp. 127-28. Princeton.

Renfrew, C. 1978. Trajectory, discontinuity and morphogenesis. The implications of Catastrophe Theory for archaeology. *American Antiquity*, 43: 203-44.

Renfrew, C., & Cooke, K.L. 1979. Transformations: Mathematical Approaches to Culture Change. London: Academic Press.

Renfrew, C. 1979. Systems collapse as social transformation: catastrophe and anastrophe in early state society. In C. Renfrew & K.L. Cooke (eds), *Transformations: Mathematical Approaches to Culture Change*, pp. 481-506. London: Academic Press.

Renfrew, C. 1980. The great tradition versus the great divide: archaeology as anthropology? *American Journal of Archaeology*, 84: 287-98.

Renfrew, C. 1983. Palaeoethnobotany: the prehistoric food plants of the Near East and Europe. London: Methuen.

Renfrew, C. 1991. The Cycladic spirit: masterpieces from the Nicholas P. Goulandris Collection. London: Thames and Hudson.

Renfrew, C. 1996. Who were the Minoans? Towards a population history of Crete. In W.F. Bakker, R.F. Willets, & C. Davaras (eds), *Cretan Studies*, Vol.5, pp. 1-29. Amsterdam: Adolf M. Hakkert.

Rethemiotakis, G. 2002. Evidence on Social and Economic Changes at Galatas and Pediada in the New-Palace Period. In J. Driessen, I. Schoep, & R. Laffineur (eds), *Monuments of Minos. Rethinking the Minoan Palaces. Proceedings of the International Workshop "Crete of the hundred Palaces?" held at the Universite Catholique de Louvain, Louvain-la-Neuve, 14-15 December 2001*, pp. 55-69.

Richman, E. A., Michel, M.E., Schulter-Ellis, F.P., Corrucini, R.S. 1979. Determination of sex by discriminant function analysis of postcranial skeletal measurements. *Journal of Forensic Science*, 24: 159-67.

Richman, J.M. 1995. Craniofacial genetics makes headway. *Current Biologist*, 5: 435-48.

Richtsmeier, J.T., & McGrath, J.W. 1986. Quantitative genetics of cranial nonmetric traits in random bred mice: heritability and etiology. *American Journal of Physical Anthropology*, 69: 51-58.

Rightmire, G.P. 1970. Bushman, Hottentot and South African Negro Crania Studied by Distance and Discrimination. *American Journal of Physical Anthropology*, 33: 147-68.

Rightmire, G.P. 1972. Cranial measurements and discrete traits compared in distance studies of African Negro Skulls. *Human Biology*, 44: 263-76.

Rightmire, G.P. 1976. Metric versus discrete traits in African skulls. In E.Giles & J.S. Friedlaender (eds), *The measure of man*, pp.383-407. Cambridge: Peabody Museum Press.

Risnes, S. 1989. Ectopic tooth enamel. An SEM study of the structure of enamel in enamel pearls. *Advances in Dental Research*, 3: 258-64.

Roberts, D.F. 1973. Climate and human variability. Reading: Addison-Wesley.

Rodden, J. 1981. The development of the Three Age System: archaeology's first paradigm. In G. Daniel (ed), *Towards a History of Archaeology*, pp. 51-68. London: Thames and Hudson.

Rogers, G., & Hawkesworth, C.J. 1989. A geochemical traverse across the North Chilean Andes: evidence for crust generation from the mantle wedge. *Earth and Planetary Science letters*, 91: 271-85.

Rosing, F.W. 1982. Discrete des Menschlichen Skeletts-ein Kritischer Uberblick. *Homo*, 33: 100-25.

Ross, A.H. 2004. Regional Isolation in the Balkan Region: An Analysis of Craniofacial Variation. American Journal of Physical Anthropology, 124: 73-80.

Ruff, C. 1980. Age differences in craniofacial dimensions among adults from Indian Knoll, Kentucky. *American Journal of Physical Anthropology*, 1: 108-138.

Ruff, C.B., & Hayes, W.C. 1983. Cross-sectional geometry of Pecos Pueblo femora and tibiae: A biomechanical investigation: II, Sex, age and side differences. *American Journal of Physical Anthropology*, 60: 383-400.

Ruff, C.B., Larsen, C.S., & Hayes, W.C. 1984. Structural changes in femur with the transition to agriculture on the Georgia coast. *American Journal of Physical Anthropology*, 64: 125-36.

Ruff, C.B. 1988. Hind limb articular surface allometry in Hominid and Macaca, with comparisons to diaphyseal scaling. *Journal of Human Evolution*, 17: 687-714.

Ruff, C.B., Scott, W.M., & Liu, A.Y.C. 1991. Articular and diaphyseal remodeling of the proximal femur with changes in body mass in adults. *American Journal of Physical Anthropology*, 86: 397-413.

Runnels, C.N. & van Andel, T.H. 1987. The evolution in the Southern Argolid, Greece: An Economic Explanation. *Hesperia*, Vol. 56, No.3: 303-34.

Rutter, J.R. 1979. Ceramic Change in the Aegean Early Bronze Age. Los Angeles: University of California.

Rutter, J. 1983. Some Observations on the Cyclades in the Later Third and Early Second Millenia. *American Journal of Archaeology*, 87: 69-76.

Rutter, J.R., & Zerner, C.W. 1984. Early Hellado-Minoan contacts. In R. Hagg, & N. Marinatos (eds), *The Minoan Thalassocracy: Myth and Reality. Proceedings of the Third International Symposium at the Swedish Institute in Athens*, pp. 75-83. Stockholm: Paul Åströms Förlag.

Rutter, J. R. 2001. The Prepalatial Bronze Age of the Southern and Central Greek Mainland. T.Cullen (eds), *Aegean Prehistory: a review*, pp. 95-147. Boston: Archaeological Institute of America.

Ryman, N., Chakraborty, R., & Nei, M. 1983. Differences in the relative distribution of human gene diversity between electrophoretic and red and white cell antigen loci. *Human Heredity*, 33: 93-102.

Sakellarakis, J. 1972. A Mycenaean burial enclosure in Crete. Archaeologika Analekta ex Athinon (Αρχαιολογικά Ανάλεκτα εξ Αθηνών), pp. 399-419.

Sandars, N. 1978. The Sea People: Warriors of the Ancient Mediterranean 1250 – 1150 BC. London: Thames and Hudson.

Sardi, M.L., Ramirez Rozzi, F., González-Jose, R., & Pucciarelli, H.M. 2005. South Amerindian craniofacial morphology: Diversity and implication for Amerindian Evolution. *American Journal of Physical Anthropology*, 128: 747-56.

Saunders, S.R. 1978. The development and distribution of discontinuous morphological variation of the human infracranial skeleton. Archaeological Survey of Canada, Paper No 81. Ottawa: National Museum of Canada.

Saunders, S. R. 1989. Nonmetric skeletal variation. In M.Y. Iscan & K.A.R. Kennedy (eds), *Reconstruction of Life from the Skeleton*, pp. 95-108. New York: Alan R. Liss.

Schaeffer, C.F. 1939. Ugaritica 1. Mission de Ras Shamra 3. Bibliothèque Archéologique et Historique 31. Paris : Geuthner.

Schallin, A.L. 1993. Islands under Influence. The Cyclades in the Late Bronze Age and the nature of the Mycenaean presence. Studies in Mediterranean Archaeology, Vol. CXI. Jonsered: Paul Åstroms Förlag.

Schelmerdine, C. W. 2001. In T. Cullen (ed), *Aegean Prehistory: a review*, pp. 329-377. Boston: Archaeological Institute of America.

Schillaci, M.A., & Stojanowski, C.M. 2005. Craniometric Variation and Population History of the Prehistoric Tewa. *American Journal of Physical Anthropology*, 126: 404-12.

Schliemann, H. 1886. Tiryns. The Prehistoric Palace of the Kings of Tiryns. London: John Murray.

Schoep, I. 1999. Tablets and Territories? Reconstructing Late Minoan IB Political Geography through Undeciphered Documents. *American Journal of Archaeology*, 103: 201-21.

Schofield, G. 1959. Metric and Morphological Features of the Femur of the New Zealand Maori. *The Journal of the Royal Anthropological Institute of Great Britain and Ireland*, 89: 89-105.

Scott, G.R., & Turner, C.G. 1988. Dental Anthropology. *Annual Reviews in Anthropology*, 17: 99-126.

Sealy, J.C. 1989. Reconstruction of Later Stone Age diets in the South-western Cape, South Africa: evaluation and application of five isotopic and trace element techniques. PhD thesis, University of Cape Town.

Self, S.G., & Leamy, L. 1978. Heritability of quasi-continuous skeletal traits in a random bred population of house mouse. *Genetics*, 88: 109-20.

Shear, I. M. 1987. The Panagia Houses at Mycenae. Philadelphia: University Museum Monograph 68.

Shennan, S. 1997. Quantifying Archeology. Edinburgh: Edinburgh University Press.

Sherratt, E.S., & Crouwel, J.H. 1987. Mycenaean Pottery from Cilicia in Oxford. *Oxford Journal of Archaeology, 6:* 325-352.

Sherratt, A., & Sherratt, S. 1991. From luxuries to commodities: the nature of Mediterranean Bronze Age trading systems. In N.H. Gale (ed), *Bronze Age Trade in Mediterranean. Papers Presented at the Conference held at Rewley House, Oxford, in December 1989*, pp. 351-86. Studies in Mediterranean Archaeology 90. Jonsered: Paul Åströms Förlag.

Sillen, A., Hall, G, & Armstrong, R. 1998. $^{87}Sr/^{86}Sr$ ratios in modern and fossil food-webs of the Sterkfontein Valley: implications for early hominid habitat preference. *Geochimica et Cosmochimica Acta*, 62: 2463-78.

Sjøvold, T. 1984. A Report on the Heritability of Some Cranial Measurements and Non-metric Traits. In G.N. Van Vark & W.W. Howells (eds), *Multivariate Statistics in Physical Anthropology*, pp. 223-246. Dordrecht: D. Reidel.

Smith, C.L., & Beals, K.L. 1990. Cultural correlates with Cranial Capacity. *American Anthropologist*, 92: 193-200.

Snyder, L.M., & Klippel, W.E. 2003. From Lerna to Kastro: Further thoughts of dogs as food in ancient Greece; perceptions, prejudices and reinvestigations. In E. Kotjabopoulou, Y. Hamilakis, P. Halstead, C. Gamble, & P. Elefanti (eds), *Zooarchaeology in Greece: Recent Advances*, pp. 221-231. The British School at Athens, Studies 9. London.

Sofaer, J.A. 1969. The genetics and expression of a dental morphological variant in the mouse. *Archives of Oral Biology*, 14: 1213-23.

Sofaer, J.A., Smith, P., & Kaye, E. 1986. Affinities between contemporary and skeletal Jewish and non-Jewish groups based on tooth morphology. *American Journal of Physical Anthropology*, 70: 265-75.

Sokal, R.R. 1966 Numerical Taxonomy. *Scientific American*, 215: 106-16.

Spielman, R.S., & Smouse, P.E. 1976. Multivariate classification of human populations I. Allocation of Yanomama Indians to villages. *American Journal of Human Genetics*, 28: 317-31.

van Spronsen, P.H, Weijs,W.A., Prahal-Andersen, B., Valk, J., & Van Ginkel, F. 1992. A comparison of jaw muscle cross sections of long-face and normal adults. *Journal of Dental Research*, 71: 1279-1285.

Steadman, D.W. 1998. The population shuffle in the central Illinois Valley: a diachronic model of Mississipian biocultural interactions. *World Archaeology*, 30: 306-26.

Steele, D.G., & Bramblett, C.A. 1988. The Anatomy and Biology of the Human Skeleton. College Station, TX: Texas University.

Steele, D.G., & Mays, S. 1995. Handedness and Directional Asymmetry in the Long Bones in the Human Upper Limb. *International Journal of Osteoarchaeology*, 5: 39-49.

Stefan, V.H., & Chapman, P.M. 2003. Cranial Variation in the Marquesas Islands. *American Journal of Physical Anthropology*, 121: 319-31.

Stock, J., & Pfeiffer, S. 2001. Linking Structural Variability in Long Bone Diaphyses to Habitual Behaviors: Foragers From the Southern African Later Stone Age and the Andamen Islands. *American Journal of Physical Anthropology*, 115: 337-348.

Stojanowski, C.M. 2003. Differential phenotypic variability among the Apalachee Mission populations of La Florida: A diachronic perspective. *American Journal of Physical Anthropology*, 120: 352-63.

Stos-Gale, Z. 1989. Cycladic copper metallurgy. In E.P. Hauptmann & G.A. Wagner (eds). *Old World Archaeometallurgy*. Der Anschnitt Beiheft 7. Bochum:

Vereinigung der Freunde von Kunst und Kultur in Bergbau e. V. Bochum, pp. 279- 92.

Stos-Gale, Z., & MacDonald, C. 1991. Sources of Metals and Trade in the Bronze Age Aegean. In N. Gale (ed), *Bronze Age Trade in the Mediterranean*, pp. 249-88. Jorsered: Studies in Mediterranean Archaeology, 90.

Strachan, T., & Read, A.P. 1996. Human molecular genetics. Oxford: BIOS Scientific Publishers.

Tainter, J. A. 1988. The Collapse of Complex Societies. Cambridge: Cambridge University Press.

Templeton, A.R., Routman, E., & Phillips, C.A. 1995. Separating population structure from population history: A cladistic analysis of the geographical distribution of mitochondrial DNA haplotypes distribution in the tiger salamander, Ambystoma tigrinum. *Genetics*, 140: 767-82.

Templeton, A.R. 1998. Nested clade analyses of phylogeographic data: testing hypotheses about gene flow and population history. *Molecular Ecology*, 7: 632-50.

Tomlinson, R.A. 1995. Archaeology in Greece, 1994-95. *Archaeological Reports* 45: 69-76.

Torgersen, J.1951. Hereditary factors in the sutural pattern of the skull. *Acta Radiologica*, 36: 374-82.

Torrence, R. 1982. The obsidian quarries and their use. In C. Renfrew & M. Wagstaff (eds). *An Island Polity. The Archaeology of Exploration in Melos*, pp. 193-221. Cambridge: Cambridge University Press.

Torrence, R. 1986. Production and Exchange of Stone Tools: Prehistoric Obsidian in the Aegean. Cambridge: Cambridge University Press.

Treuil, R., Darcque, P., Poursat, J., & Touschais, G.I. 1996. Oi Politismoi toy Aegaeoy kata tin Epochi toy Chalcoy (Οι Πολιτισμοί του Αιγαίου κατά τη Νεολιθική και την Εποχή του Χαλκού). Αθήνα : Μ. Καρδαμίτσα.

Triantaphyllou, S. 2001. A bioarchaeological Approach to Prehistoric Cemetery Populations from Central and Western Greek Macedonia. British Archaeological Reports International Series 976.

Trigger, B.G. 1989. A History of Archaeological Thought. Cambridge: Cambridge University Press.

Tsountas, Ch. & Manatt, J.I. 1897. The Mycenaean age. London.

Tsountas, K. 1898. Κυκλαδικά. *Archaeologike Ephemeris*, 137- 212.

Tsountas, K. 1899. Κυκλαδικά II. *Archaeologike Ephemeris*, 73-134.

Turner, C., Nichol, C., & Scott, G. 1991. Scoring procedures for key morphological traits for the permanent dentition: The Arizona State University Dental Anthropology System. In M. Kelley & C. Larsen (eds), *Advances in Dental Anthropology*, pp. 13-31. New York: Wiley Liss.

Tyrell, A. J., & Chamberlain, A. T., 1988, Non-metric Trait Evidence for Modern Human Affinities and the distinctiveness of Neanderthals, *Journal of Human Evolution*, 34: 549-554.

Ubelaker, D.H. 1989. Human Skeletal Remains. Washington: Taraxacum Press.

Utermohle, C.J., & Zegura, S.L. 1982. Intra and Interobserver Error in Craniometry: A cautionary Tale. *American Journal of Physical Anthropology*, 57: 303-310.

Varela, H. H., & Cocilovo, J. A. 2002. Genetic Drift and Gene Flow in a Prehistoric Population of the Azapa Valley and Coast, Chile. *American Journal of Physical Anthropology*, 118: 259-67.

Vasilakis, A. 1989-90. Προϊστορικές θέσεις στη Μονή Οδηγήτριας. *Kritike Estia*, 3: 64-5.

Vecci, F., Coppa, A., & Priori, R. 1987. Morphological distance between four ethnic groups in southern Mexico. *Rivista di Antropologia*, 65: 223-234.

Vermeule, E. 1963. The Fall of Knossos and the Palace Style. *American Journal of Archaeology*, 67: 195-99.

Voutsaki, S. 1998. Mortuary Evidence, Symbolic Meanings and Social Change: A Comparison between Messenia and the Argolid in the Mycenaean Period. In K. Branigan (ed), *Cemetery and Society in the Aegean Bronze Age*, pp. 41-58. Sheffield Studies in Aegean Archaeology, 1. Sheffield: Sheffield Academic Press Ltd.

Wace, A.J.B. 1921-23. Excavations at Mycenae IX: the Tholos tombs. *Annual of the British School at Athens*, 25: 283-402.

Wace, A.J.B. 1950. Excavations at Mycenae, 1939. *Annual of the British School at Athens*, 45: 203-228.

Wace, A.J.B. 1953a. Mycenae 1939-1952, part I: preliminary report on the excavations of 1952. *Annual of the British School at Athens*, 48: 3- 18.

Wace, A.J.B. 1953b. Mycenae 1939-1952, part IV: the Epano Phournos tholos tomb. *Annual of the British School at Athens*, 48: 69-83.

Wace, A.J.B. 1954. Mycenae 1939- 1953, part I: preliminary report on the excavations of 1953. *Annual of the British School at Athens*, 49: 231-243.

Wace, A.J.B. 1955. Mycenae 1939-1954. *Annual of the British School at Athens*, 50: 175-250.

Wace, A.J.B. 1956. Mycenae 1939- 1955, part I: preliminary report on the excavations of 1955. *Annual of the British School at Athens*, 51: 103- 122.

Wall, S. M., Musgrave, J.H., & Warren, P.M. 1986. Human bones from a Late Minoan IB House at Knossos. *Annual of the British School at Athens*, 81: 334-388.

Walberg, G. 1983. Middle Minoan Provincial Pottery. Mainz-am-Rhein: Philipp von Zabern.

Warren, P.M. 1969. Minoan Stone Vases. Cambridge: University Printing House.

Warren, P.M. 1973. Crete, 3000-1400BC: Immigration and the archaeological evidence. In R.A Crossland, & A. Birchall (eds), *Bronze Age Migrations in the Aegean: Archaeological and Linguistic Problems in Greek Prehistory*, pp. 41-7. London: Gerald Duckworth & Ltd.

Warren, P. 1980. Problems in Crete and the Aegean in the 3rd and earlier 2nd millenium B.C.

Warren, P.M. 1985. Minoan Palaces. *Scientific American*, 253 (1): 205-211.

Warren, P.M., & Hankey, V. 1989. Aegean Bronze Age Chronology. Bristol: Bristol Classical Press.

Warren, P.M. 1991. The Minoan Civilization of Crete and the Volcano of Thera. *Journal of the Ancient Chronology Forum*, 4: 29-39.

Waterhouse, H. & Hope-Simpson, R. 1960. Prehistoric Laconia: Part I. *Annual of the British School at Athens*, 65.

Watrous, L.V. 1985. Late Bronze Age Kommos: imported pottery as evidence for foreign contact. In J.W. Shaw, & M.C. Shaw (eds), *A Great Minoan Triangle in South-central Crete: Kommos, Hagia Triadha, Phaistos*, pp. 7- 11. Scripta Mediterranea 6. Toronto: Society for Mediterranean Studies.

Watrous, L.V. 2001. Crete from Earliest Prehistory through the Protopalatial Period. In T. Cullen (ed), *Aegean Prehistory: a review*, pp. 157-215. Boston: Archaeological Institute of America.

Weinstein, K.J. 2005. Body Proportions in Ancient Andeans from high and low altitudes. *American Journal of Physical Anthropology*, 128: 569-85.

Wells, P. 1992. Crisis Years? The 12th Century B.C. in Central and Southern Europe. In W. A. Ward & M. Joukowsky (eds), *The Crisis Years: The 12th Century B.C.*, pp.31-39. Dubuque: Kendall/Hunt.

Wheeler, T. 1974. Early Bronze Age Burial Customs in Western Anatolia. *American Journal of Archaeology*, 78: 415-25.

White, C.D., Spence, M.W., Stuart-Williams, H.L.Q., & Schwarcz, H.P. 1998. Oxygen isotopes and the identification of geographical origins: the Valley of Oaxaca versus the Valley of Teotihuacan. *Journal of Archaeological Science*, 25: 643-55.

White, C.D., Spence, M.W., & Law, K.R. 2000. Testing the nature of Teotihuacan imperialism at Kaminaljuju using phosphate oxygen-isotope ratios. *Journal of Archaeological Research*, 56: 535.

Wiencke, M.H. 1986. Building BG at Lerna. In R. Hagg, & D. Konsola (eds), *Early Helladic Architecture and Urbanization. Proceedings of a seminar held at the Swedish Institute in Athens (June 8, 1985)*, pp. 41-45. Studies in Mediterranean Archaeology, 76. Jonsered: Paul Åstroms Förlag

Wiencke, M.H. 1989. Change in the Early Helladic II. *American Journal of Archaeology*, 93: 495-509.

Wiencke, M.H. 1998. Mycenaean Lerna. *Hesperia*, 67 : 125-214.

Wilkinson, R.G., & Norelli, R. J. 1981. A Biocultural Analysis of Social Organization at Monte Alban. *American Antiquity*, 46: 743-58.

Williams, P.L, & Warwick, R. 1980. Gray's anatomy. Edinburgh: Churchill Livingstone.

Williams-Blangero, S. & Blangero, J. 1989. Anthropometric variation and the genetic structure of the Jirels of Nepal. *Human Biology*, 61:1-12.

Winder, S.C. 1981. Infracranial non-metric variation: an assessment of its value for biological distance studies. Unpublished PhD thesis, University of Indiana.

Wood-Jones, F. 1931. The non-metrical morphological characters of the skull as criteria for racial diagnosis. Part I. General discussion of the morphological characters employed in racial diagnosis. *Journal of Anatomy*, 65: 179-95.

Wood, J.W., Milner G.R., Harpending H.C., & Weiss, K.M. 1992.The Osteological Paradox: Problems of Inferring Health from Skeletal Samples. *Current Anthropology*, 33: 343-370.

Wright, H.E. 1972. Vegetation History. In W.A. McDonald & G.R. Rapp (eds). *The Minnesota Messenia Expedition: Reconstructing a Bronze Age Regional Environment*, pp. 188-99. Minneapolis.

Wright, J.C. 1984. Changes in the Form and Function of the Palace at Pylos. In C.W. Shelmerdine, & T.G. Palaima (eds), *Pylos Comes Alive: Industry and Administration in a Mycenaean Palace*, pp.19-29. New York.

Wright, J.C., Cherry, J. Davis, J.L., E. Mantzourani, Sutton, S.B, & Sutton, R.F. 1990. The Nemea valley Archaeological Project: A preliminary Report. *Hesperia*, 59: 579- 659.

Yoffee, N. & Cowgill, G.L. 1988. The Collapse of Ancient States and Civilizations. Tucson: University of Arizona Press.

Zafeiropoylos, N. 1960. Ανασκαφαί Νάξου. Praktika tis en Athinais Archaeologikis Etaireias.

Zahler, R.S., & Sussmann, H.J. 1977. Claims and Accomplishments of Applied Catastrophe Theory. *Nature*, 269: 759-63.

Zakrzewski, S.R. 2001. Continuity and Change: A Biological History of Ancient Egypt. Unpublished PhD thesis, University of Cambridge.

Zakrzewski, S.R. 2003. Variation in Ancient Egyptian Stature and Body Proportions. *American Journal of Physical Anthropology*, 121: 219-29.

Zohary, D., & Hopf, M. Domestication in the Old World. *Science*, 182: 887-894.