

1 **Polyunsaturated fatty acids and metabolic health: novel insights**

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17 **Abstract**

18 Purpose of review: This review aims to discuss the potential roles of omega-3 (ω -3) and omega-
19 6 (ω -6) polyunsaturated fatty acids (PUFAs) in the prevention and treatment of metabolic
20 diseases, to provide the latest evidence from epidemiological and clinical studies, and to
21 highlight novel insights into this field.

22 Recent findings: Higher dietary or circulating ω -3 PUFA levels are related to a lower risk of
23 metabolic syndrome. Novel findings in obesity indicate higher proportions of ω -6 and ω -3
24 PUFAs, a modulated oxylipin profile and an altered transcriptome in subcutaneous white
25 adipose tissue, that seem resistant to the effects of ω -3 PUFAs compared with what occurs in
26 normal weight individuals. ω -3 PUFAs may improve the blood lipid profile and glycemic
27 outcomes in patients with type 2 diabetes mellitus and reduce liver fat in non-alcoholic fatty
28 liver disease; the findings of several recent meta-analyses support these effects. Genetic
29 background affects inter-individual variability in the insulin sensitivity response to ω -3 PUFA
30 supplementation. ω -3 PUFAs have prebiotic effects, altering the gut microbiota.

31 Summary: Although evidence for health benefits of ω -3 PUFAs is strong, recent findings
32 suggest a more personalized approach to ω -3 PUFA intake for individuals at high risk for
33 metabolic diseases.

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35 Key words: Metabolic diseases, Risk factors, Omega-3, Omega-6, Diabetes

36 **Introduction**

37 Metabolic syndrome (MetS) is a multifactorial disease that includes various health issues such
38 as abdominal obesity, dyslipidemia, insulin resistance (IR) and hypertension [1*]. Altered lipid
39 metabolism and glycemic control appear to be major risk factors in developing non-alcoholic
40 fatty liver disease (NAFLD) and cardiovascular disease (CVD) [2, 3]. Furthermore, oxidative
41 stress, systemic inflammation, gut dysbiosis, cytokines, adipokines, hepatokines and genetics
42 make a large contribution to complex metabolic disturbances [4]. Together with promoting a
43 healthy lifestyle and a more personalized medication approach, there is growing interest in the
44 use of nutraceuticals and dietary supplements in the treatment or co-treatment of
45 cardiometabolic disease and related features such as dyslipidemia [5*]. The 2019 European
46 Society of Cardiology/European Atherosclerosis Society Guidelines list several nutraceuticals
47 and nutritional supplements including phytosterols (i.e., sterols and stanols), red yeast rice
48 extract, dietary fibers, and omega-3 (ω -3) fatty acids to be considered for the management of
49 dyslipidemias [6]. However, the most relevant and recent American and European guidelines
50 do not encourage any nutraceutical or food supplement approach concerning obesity due to
51 insufficient safety and efficacy data (see [5*] for references). Although several nutraceuticals
52 have been suggested for glycaemic control, alone or as an adjunct to standard medical care (e.g.,
53 berberine, *Morus alba* extract) [7, 8], the updated Standards of Medical Care in Diabetes has
54 not supported the implementation of any nutraceutical or supplement products in the
55 management of type 2 diabetes mellitus (T2DM) [9]. This is similar to a consensus report
56 published in 2018 by the American Diabetes Association and the European Association for the
57 Study of Diabetes [10]. Despite a lack of consensus on the recommendation for patients at high
58 cardiometabolic risk, ω -3 and omega-6 (ω -6) polyunsaturated fatty acids (PUFAs) have been
59 recognized to offer multiple mechanisms of action to counteract a cluster of metabolic

60 disorders. The aim of this article is to discuss the role of PUFAs in the prevention and treatment
61 of metabolic diseases, reviewing the latest evidence and highlighting novel insights in this field.

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63 **Epidemiology of polyunsaturated fatty acids and metabolic health**

64 Much evidence has accumulated from prospective and case-control studies indicating that a
65 higher intake of long-chain PUFAs, especially ω -3 PUFAs, is related to an improved profile of
66 risk factors of MetS. These studies have been summarized in systematic reviews and meta-
67 analyses and are discussed in detail elsewhere recently [11*]. For example, early
68 epidemiological data showed that Greenland Inuit had a much lower prevalence of
69 cardiometabolic disease, including diabetes mellitus and atherosclerosis, considered to be the
70 result of the high ω -3 PUFA content and more optimal ω -6/ ω -3 PUFA ratio of their traditional
71 diet. However, over the last half-century, due to lifestyle changes (e.g., replacing traditional
72 with imported foods, being sedentary) and possibly gene-lifestyle interactions, the Inuit in
73 Greenland have experienced a significant rise in incidence and prevalence of obesity and
74 metabolic disorders [12]. A recent meta-analysis of data from cross-sectional and case-control
75 trials identified that higher blood levels of ω -3 PUFAs are associated with a lower risk of MetS
76 [13]. Another meta-analysis pooled results from prospective cohort studies and confirmed that
77 higher dietary or circulating ω -3 PUFA levels were associated with 26% lower MetS risk than
78 lower dietary or circulating levels (odds ratio (OR)/relative risk (RR) 0.74; 95% confidence
79 interval (CI) 0.62, 0.89) [14*]. Interestingly, docosahexaenoic acid (DHA) appeared to have
80 greater effectiveness (OR/RR 0.66; 95% CI 0.49, 0.88) than other ω -3 PUFAs (i.e., alpha-
81 linolenic acid (ALA), docosapentaenoic acid (DPA) and eicosapentaenoic acid (EPA)), which
82 did not show significant effects. Also, null results were observed concerning the association
83 between circulating or dietary ω -6 PUFAs and MetS. However, a recent meta-analysis of cohort
84 studies evaluated the association between intake of PUFAs and incidence of T2DM, and found

85 that DHA increased risk while the ω -6 linoleic acid (LA) decreased risk of T2DM (RR 1.164;
86 95% CI 1.048 to 1.294 and RR 0.956; 95% CI 0.930 to 0.983, respectively) [15*]. A de novo
87 pooled analysis of 17 prospective cohort studies with 42,466 individuals reported the
88 association between a lower risk for death from cardiovascular disease in patients with the
89 highest versus the lowest quintile of circulating long-chain ω -3 PUFAs [16*].

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91 **Biological actions of PUFAs**

92 Blood triglyceride (TG) levels, blood pressure, fasting blood glucose (FBG), high-density
93 lipoprotein (HDL) cholesterol, and insulin resistance are all improved by ω -3 PUFAs, probably
94 explaining their protective effects on MetS and cardiovascular disease [11*]. Similarly, ω -6
95 PUFAs, instead of saturated fat, can positively impact blood lipid management (especially low-
96 density lipoprotein (LDL) cholesterol) and insulin resistance [11*]. As a structural component
97 of cell membranes, bioactive PUFAs act through various mechanisms influencing the function
98 of membrane proteins, intracellular signalling pathways and gene expression, and altering the
99 production of lipid mediators (i.e., eicosanoids and docosanoids) [17]. Oxylipins are bioactive
100 metabolites generated from PUFAs, including EPA and DHA and the ω -6 PUFA arachidonic
101 acid (AA), through enzymatic and non-enzymatic oxidation [18, 19*, 20, 21*]. Enzymatic
102 oxidation of ω -3 and ω -6 PUFAs shares the same enzymes, i.e., lipoxygenases (LOXs),
103 cyclooxygenases (COXs), and cytochromes P450 (CYP-450) [18, 19*, 20, 21*]. These
104 enzymes are expressed in various cells and tissues. Oxylipins may act as both pro- and anti-
105 inflammatory molecules [20, 21*]. Although oxylipins are triggered in response to
106 inflammatory stimuli, a recent study reported that these same stimuli also programme their
107 removal, for example by upregulation of mitochondrial β -oxidation [22*]. In general, an
108 increased presence of ω -3 PUFAs in cell membranes leads to an increased generation of less-
109 inflammatory and pro-resolving mediators as a result of LOX and COX action on ω -3 rather

110 than ω -6 PUFAs [20, 23, 24]. Uncontrolled (unresolved) inflammation and continuous release
111 of pro-inflammatory mediators can cause metabolic changes, tissue damage, and loss of
112 function, and oxylipins produced from EPA and DHA are able to both prevent and reverse these
113 effects [25*]. For example, pro-inflammatory activities of AA-derived oxylipins lead to altered
114 lipid metabolism and remodelling and expansion of adipose tissue (see [26*] for references).
115 On the other hand, reduced inflammatory actions and pro-resolving activities of oxylipins
116 derived from EPA and DHA, so-called specialized pro-resolving mediators (SPMs), better
117 regulate the expression of inflammatory cytokines [25*, 27*]. SPMs have been described in
118 human blood and other fluids including breast milk and in human tissues [28*]. The production
119 of SPMs is favoured by the higher EPA and DHA status brought about by increased oral intake
120 of these fatty acids [28*]. SPMs may be responsible for many of the biological actions ascribed
121 to EPA and DHA [28*]. DPA is also a substrate for synthesis of SPMs [29]. Recent findings
122 suggest that genetic/pharmacological targeting of carnitine palmitoyl transferase 1, enhances
123 oxylipin removal via mitochondrial β -oxidation independently of oxidative phosphorylation
124 and energy production [22*]. Upregulation of many genes is observed in a regulatory metabolic
125 checkpoint for oxylipins during inflammation [22*]. Thus, lipidomic profiling targeting
126 oxylipins may contribute to deeper understanding the role of these bioactive metabolites in the
127 development and progression of human diseases. In addition to biological activities, ω -3 PUFAs
128 appear to affect the gut microbiome [30*]. This seems to result in three related effects: altered
129 diversity and abundance of the gut microbial community, modulated levels of pro-inflammatory
130 molecules such as interleukin-17 and lipopolysaccharides, and altered concentrations of short-
131 chain fatty acids and their salts [30*, 31]. Firmicutes (F) and bacteroidetes (B) are the two major
132 bacterial phyla representing about 90% of the human gut microbiota, and an increase in the F/B
133 ratio due to an inappropriate diet including a high ratio of ω -6 to ω -3 PUFAs may lead to
134 overweight, obesity, non-alcoholic fatty liver disease and CVD [32-35].

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Latest clinical trials and meta-analysis of trials related to PUFA supplementation and metabolic disease

A recently published double-blind randomised controlled trial has reported novel insights into fatty acid composition, oxylipin profile and transcriptome in human subcutaneous white adipose tissue (scWAT) in obese individuals [26*]. Fifty healthy normal weight individuals and 50 individuals living with obesity were randomly assigned to receive a supplement providing 1.1 g EPA + 0.8 g DHA or corn oil (as the comparator oil) and were followed up for 12 weeks. Significantly higher proportions of ω -6 and ω -3 PUFAs in scWAT were observed in individuals with obesity compared to normal weight individuals at study entry. The proportion of EPA in scWAT was positively correlated with adipose-IR (ρ 0.248, $P = 0.043$) and the proportion of DPA with homeostatic model assessment of insulin resistance (HOMA2-IR) and adipose-IR (ρ 0.258, $P = 0.038$, and ρ 0.342, $P = 0.005$ respectively). Regarding the oxylipin profile of the whole scWAT, 33 fatty acid metabolites of 111 identified were significantly modified in individuals living with obesity compared to normal weight subjects; typically scWAT from individuals with obesity has lower levels of hydroxy-DHAs and some SPMs. Proportions of several oxylipins were inversely correlated with HOMA2-IR, indicating a link between lower levels of oxylipins with increased insulin resistance [26]. The expression of the genes encoding CYP1B1, ALOX5 (which encodes 5-LOX), and PTGS1 (which encodes COX-1) was upregulated in scWAT from obese individuals. Transcriptional changes, including 622 upregulated and 174 downregulated genes in scWAT in individuals living with obesity, indicated upregulation of inflammatory and immune responses in scWAT in obesity. In addition to dysregulated expression of inflammatory and immune response related genes, nearly 20% of these genes were associated with lipid and carbohydrate metabolism and signalling which may contribute to an upregulation in the T2DM signalling pathway and interruption of whole tissue

160 homeostasis occurring in these early stages of obesity. Following 12 weeks of ω -3 PUFA
161 intervention, an altered response was noticed in individuals with obesity compared with those
162 of normal weight. Although a decrease of AA metabolites in scWAT was observed, modulation
163 of ω -3 PUFA derived oxylipins was impaired in those with obesity compared with what was
164 seen in normal weight individuals suggesting a lack of response in SPM formation and reduced
165 ability to self-resolve inflammation, even when additional ω -3 PUFAs are provided. Despite
166 the incorporation of EPA and DHA resulting in similar levels in scWAT in both groups, this
167 was not sufficient to change ω -6 and ω -3 proportions enough to promote the generation of ω -3
168 PUFA derived oxylipins or have a greater effect on gene expression in obese individuals
169 compared to normal weight individuals. In response to ω -3 PUFA supplementation, 51 and 21
170 genes were differentially expressed in scWAT in normal weight and obese individuals,
171 respectively. The modulation of these genes was linked with the overall downregulation of
172 inflammatory and immune responses as well as upregulation of glucose homeostasis in normal
173 weight individuals, with the absence of these effects in individuals living with obesity.
174 Decreased levels of SPMs and increased expression of genes associated with immune and
175 inflammatory signalling in scWAT, appear to affect whole-body homeostasis.

176 Another study reported the importance of genetic background in interindividual
177 variability in the insulin sensitivity response to ω -3 PUFA supplementation using a genetic
178 score approach [36]. Treatment was with 1.9-2.2 g EPA + 1.1 g DHA per day for six weeks.
179 HOMA-IR was used to classify participants as high or low risk depending on their HOMA-IR
180 change following the ω -3 PUFA supplementation compared to pre-treatment values (some
181 individuals (23.2%) had increased HOMA-IR after ω -3 PUFAs). Of the 210 participants,
182 genome-wide genotyping data were obtained for 138 subjects: eight gene loci had frequency
183 differences between high-risk and low risk participants and a genetic risk score (for increased
184 HOMA-IR with supplemental ω -3 PUFAs) was created. This had a predictive accuracy of 0.85

185 and explained 40% of the variation in HOMA-IR change. These results suggest that genetic
186 background has a role in determining the interindividual variability observed in the insulin
187 sensitivity response following ω -3 PUFA supplementation. The authors suggested that people
188 at risk of insulin sensitivity lowering following ω -3 PUFA supplementation may be able to be
189 identified using genetic-based approaches.

190 In a double-blind clinical trial, the effects of marine-based and plant-based ω -3 PUFAs
191 on glucose and lipids profiles in 150 patients with T2DM were investigated [37]. Patients were
192 randomized in three groups to receive fish oil containing 143 mg EPA and 172 mg
193 DHA/capsule, perilla oil providing 322 mg ALA/capsule, or linseed and fish oil providing
194 105 mg EPA, 60 mg DHA and 140 mg ALA/capsule. All patients were supplemented with 6
195 capsules (3 g of oil) each day for six months. Treatment with perilla oil significantly lowered
196 FBG while fish oil prompted a favorable reduction of serum TG levels compared to baseline
197 values. Additionally, supplementation with ω -3 PUFAs significantly decreased serum total
198 cholesterol, apolipoprotein A1, insulin, C-peptide and IL-6 levels in all the treatment groups
199 compared to initial values. Hence, marine-derived and plant-derived ω -3 PUFAs showed
200 different but overlapping effects on glucose and lipid metabolism.

201 The potential prebiotic effects of ω -3 fatty acids were investigated in 69 participants
202 who were randomised to take ω -3 capsules containing 165 mg EPA and 110 mg DHA daily or
203 20 g of inulin fiber for a period of 6 weeks [38*]. ω -3 PUFA supplementation resulted in marked
204 increases in *Bacteroides* spp and *Coprococcus* spp and significant decreases in the fatty-liver
205 related *Collinsella* spp. On the other hand, similar to the inulin fiber arm which resulted in
206 significant increases in butyrate, iso-butyrate and iso-valerate, ω -3 PUFA supplementation
207 showed favorable increases in iso-butyrate and isovalerate and an almost significant increase in
208 butyrate. *Coprococcus*, which was significantly higher after the treatment with ω -3 PUFAs, was
209 found to be positively correlated with isobutyric acid and negatively correlated with serum

210 lipids such as VLDL-TG after adjusting for confounders. Thus, ω -3 PUFA supplementation
211 altered gut microbiota composition and some microbiota-mediated metabolic effects, indicating
212 that ω -3 PUFAs may be a helpful prebiotic nutrient.

213 Ten RCTs were summarized in a meta-analysis to evaluate the effects of supplemental
214 ω -3 PUFAs on proteinuria, estimated glomerular filtration rate (eGFR) and metabolic
215 biomarkers among patients with T2DM and type 1 diabetes mellitus (T1DM) [39]. Although
216 ω -3 PUFAs reduced the rate of proteinuria among diabetic patients, this was significant only in
217 patients with T2DM (SMD = -0.29; 95% CI: -0.54, -0.03; p = 0.03). Additionally, patients who
218 were supplemented for at least 2 years with EPA or EPA+DHA showed significant lower
219 proteinuria compared to controls (SMD = -0.30; 95% CI: -0.58, -0.02; p = 0.04). Regarding
220 eGFR, there was an increasing trend in the ω -3 PUFA group, but the effect was not statistically
221 significant (WMD = 1.56 mL/min/1.73 m²; 95% CI: -1.53, 4.65; p = 0.32). A pilot RCT included
222 27 subjects with T1DM who were assigned to receive 3.3 g/day of encapsulated ω -3 PUFAs
223 (i.e., 2.8 EPA + 0.8 DHA g/day) or encapsulated corn oil placebo for 6 months [40]. No
224 significant differences were found between ω -3 PUFA and placebo groups in metabolic,
225 glycemic or vascular outcomes. It is important to note the low sample size of this trial.

226 A meta-analysis of 30 RCTs published in 2021 reported the effects of ω -3 PUFA
227 supplementation on metabolic and inflammatory biomarkers, weight, and body mass index
228 (BMI) in patients with T2DM [41*]. Glycemic factors including FBG, glycated hemoglobin
229 (HbA1c), and HOMA-IR were significantly reduced in ω -3 PUFA supplemented groups [-0.36
230 (-0.71 to -0.01), -0.74 (-1.13 to -0.35), -0.58 (-1.13 to -0.03), respectively]. ω -3 PUFAs were
231 associated with statistically significant reductions in concentrations of total cholesterol (-0.60
232 (-0.88 to -0.32)), LDL cholesterol (-0.54 (-0.85 to -0.23)), HDL cholesterol (0.60 (0.23 to 0.96))
233 and TG (-0.27 (-0.37 to -0.18)). Inflammatory biomarkers such as tumor necrosis factor-alpha
234 (TNF- α) and C-reactive protein (CRP) were not significantly decreased. Furthermore, there was

235 no significant reduction in weight and BMI. Sub-group analysis of supplemental ω -3 PUFAs
236 according to prior defined doses (i.e., <1, 1 to 2, and >2 g/d) and duration \leq 8 week/ $>$ 8 week
237 showed that supplementation with 1 to 2 g/d for more than 8 weeks significantly affected FBG
238 level and HOMA-IR, while a significant reduction was found for HbA1c at all the 3 dose sub-
239 groups and both \leq 8 week/ $>$ 8 weeks. All 3 dose sub-groups significantly lowered TG and total-
240 cholesterol levels during both \leq 8 week/ $>$ 8 weeks. The findings revealed that ω -3 PUFA doses
241 >1 g/d significantly changed LDL and HDL levels. However, the statistically significant
242 reduction in LDL concentration occurred only when ω -3 PUFAs were consumed for more than
243 8 weeks, while improvement in HDL level was noticed during all analyzed periods (41*). A
244 previous meta-analysis of 45 RCTs, involving 2647 patients with T2DM, showed an association
245 between ω -3 PUFA supplementation and favorable improvement in lipid profile, inflammatory
246 markers and HbA1c level [42].

247 Lee et al. [43*] included 22 RCTs with 1366 participants in a meta-analysis of the effect
248 of ω -3 PUFAs in treating NAFLD. The dosage of ω -3 PUFAs used in the included trials was in
249 range of 0.25 to 5 g/day and duration was 3 to 18 months. Treatment with ω -3 PUFA
250 supplements significantly reduced liver fat compared to placebo (pooled RR 1.52; 95% CI:
251 1.09, 2.13). ω -3 PUFA supplementation also improved the levels of TG, total cholesterol and
252 HDL cholesterol, and BMI, with a pooled mean difference and 95% CI being -28.57 (-40.81 to
253 -16.33), -7.82 (-14.86 to -0.79), 3.55 (1.38 to 5.73), and -0.46 (-0.84 to -0.08), respectively.
254 These effects were obtained mainly with the treatment course of at least six months. Liver
255 enzymes, LDL cholesterol, HOMA-IR, or FBG did not show a remarkable improvement in
256 NAFLD patients taking supplemental ω 3 PUFAs.

257

258 **Summary, discussion & conclusion**

259 Current epidemiological studies and intervention trials suggest that higher ω -3 PUFA intake
260 may effectively lower the prevalence of metabolic diseases and mechanistic studies suggest that
261 this is through multiple actions of bioactive fatty acids (EPA and DHA) and their bioactive
262 metabolites. DHA has attracted more attention with its greater effectiveness in reducing MetS
263 risk and incidence of T2DM than other ω -3 PUFAs. The main biological activities of ω -3
264 PUFAs are improvement in blood lipids, fasting blood glucose and insulin resistance, and
265 promoting anti-inflammatory and pro-resolving actions. Additionally, prebiotic activities of ω -
266 3 PUFAs have recently been recognized: these fatty acids appear to modulate gut microbiota
267 composition with promising effects on metabolic disease risk.

268 Over the last years, there has been increasing evidence from clinical trials suggesting
269 that dysregulated expression of inflammatory and immune response-related genes and
270 processes in obesity may be a reason for the lack of response to ω -3 PUFA interventions in
271 obese individuals [26*, 44]. Altered adipose tissue fatty acid composition, modified oxylipin
272 profile, and dysregulation of endocannabinoid concentrations and gene expression profiles in
273 the early stages of obesity seem to be closely related. On the other hand, the genetic background
274 affects inter-individual variability in the insulin sensitivity response to ω -3 PUFA
275 supplementation [36]. Several studies and meta-analyses confirmed favorable improvement in
276 lipid profile and glycemic outcomes in patients with T2DM taking supplemental ω -3 PUFAs.
277 However, similar findings were not observed in patients with T1DM. Evidence from
278 metabolomic studies implicates that a sub-optimal fatty acid profile in early life may signal the
279 risk of pancreatic islet autoimmunity [45-47]. The possible effect of ω -3 PUFAs regarding the
280 prevention of T1DM is unclear. A meta-analysis of patients with NAFLD revealed that ω -3
281 PUFA supplementation considerably improved liver fat and blood lipids except for LDL
282 cholesterol, while liver enzymes and glycemic parameters remained unchanged [43*].

283 The evidence base for the relationship between ω -3 PUFA (i.e. EPA and DHA) intakes
284 and blood levels on one hand and “health”, including biomarkers, risk factors and clinical
285 outcomes, on the other hand includes both observational studies and intervention studies. It is
286 important to consider the intakes and blood levels that these different study types might reflect,
287 particularly in the context of intake recommendations. Different organisations make different
288 recommendations for the combined intake of EPA and DHA that is thought necessary to support
289 good health. For example, the Food and Agricultural Organisation of the United Nations
290 recommends a minimum of 250 mg EPA+DHA per day for adult males and non-pregnant or
291 non-lactating adult females [48] and the European Food Safety Authority states the adequate
292 intake as 250 mg/day for adult males and non-pregnant adult females [49]. The French Agency
293 for Food, Environmental and Occupational Health Safety sets a target of 400 to 500 mg/day for
294 adults in the general population [50], while the United Kingdom recommendation based upon
295 fish consumption is a minimum of 450 mg/day [51]. The Australian National Health and
296 Medical Research Council recommends a target of 430 to 610 mg EPA+DHA per day for adults
297 in the general population [52]. In comparison to these recommendations most adults consume
298 less than 200 mg EPA+DHA per day [53,54]. Higher intakes can be achieved by eating fatty
299 fish regularly or by using supplements that contain EPA and DHA. A standard one g fish oil
300 capsule will provide about 300 mg EPA+DHA [55]. Observational studies that associate ω -3
301 PUFA intakes to health-related outcomes report intakes of EPA and DHA across the range of
302 tens to hundreds of mg/day. Intervention studies using ω -3 PUFA supplements typically
303 provide EPA and DHA intakes in excess of recommendations, often over 1000 mg/day, as
304 described earlier. Blood levels of EPA and DHA strongly relate to intakes of these fatty acids;
305 this is clearly demonstrated in intervention studies that report linear associations between intake
306 and levels of EPA and DHA in blood lipids and blood cells [56-59]. Most intervention studies
307 with ω -3 PUFAs have a duration of 4 weeks to 3 months, although there are studies of longer

308 duration. With increased daily intake of ω -3 PUFAs from supplements, net incorporation of the
309 fatty acids into blood lipids and some blood cells becomes detectable within a few days [60],
310 although a new steady state is not reached for blood lipids until after a few weeks [56-59].
311 Incorporation into blood cells is slower, because cells turn over at a slower rate than blood
312 lipids; incorporation into erythrocytes is slower than into platelets and leukocytes because
313 erythrocytes turn over more slowly. Platelets and leukocytes reach a new steady state after about
314 one to two months and for erythrocytes this is not reached until about six months [56,59].

315 Recent evidence from epidemiological and intervention trials summarized here supports
316 a role for ω -3 PUFAs (EPA and DHA) in prevention of cardiometabolic disease and in control
317 of several recognized risk factors. However, even though ω -3 PUFAs are recommended as
318 effective therapeutic agents in managing dyslipidemias [61], further investigations are needed
319 to clarify the dose-dependent effects of EPA and DHA, separately and together, on metabolic
320 disease risk factors and related clinical outcomes.

321

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324

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327

328 **Conflicts of interest**

329 ID has no conflict of interest to declare. PCC acts as an advisor/consultant to DSM, BASF AS,
330 Cargill, Smartfish, Fresenius-Kabi, Bayer Consumer Care and GSK Consumer Healthcare.

331

332 **Key points**

- 333 • Higher dietary and circulating ω -3 PUFAs may decrease the prevalence of metabolic
334 diseases through multiple biological actions
- 335 • DHA has a higher potential to reduce metabolic disease risks than other ω -3 PUFAs
- 336 • ω -3 PUFAs modulate gut microbiota and may act as a prebiotic agents
- 337 • Dysregulated expression of inflammatory and immune response-related genes in obesity
338 may be a reason for the lack of response to ω -3 PUFA interventions in obese individuals
- 339 • ω -3 PUFAs improve lipid profile and glycemic outcomes in patients with type 2
340 diabetes mellitus and decrease liver fat in patients with non-alcoholic fatty liver disease

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