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Method development to extract microplastic particles from fishmeal



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Background

- There is increasing concern of microplastic pollution entering human food chain, e.g. commercially available bivalves contain 2.2 particles (mean items per g of tissue)⁽¹⁻⁴⁾. Harmful chemicals adsorbed onto microplastics may become bioavailable in the digestive system. Pollutants in fishmeal may enter the human food chain indirectly through feed for poultry, pigs and aquaculture^(5,6).
- Assessments of gastrointestinal (GI) tracts show that marine fish contain 1.0 to 4.9 items (mean items per individual)⁽⁷⁻¹¹⁾, other body parts have not been assessed. Fish and their by-products, including GI tracts, are used for fishmeal production^(5,6).
- Microplastic extractions from fish GI tracts are often conducted with potassium hydroxide (KOH)⁽¹²⁻¹⁴⁾. KOH has also been shown to be suitable for digestions of muscle and skin tissue of fish⁽¹⁵⁾.
- Initial trials found that fishmeal digests with 10 % KOH could not be filtered, possible due to very fine bone fragments warranting a new protocol for extracting particles from fishmeal. The suitability of density separation using brine solutions is assessed.

Methods and results

Comments

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Workflow and results of method development to extract microplastics from fishmeal samples: **Fishmeal Treatment 1** Filtration <u>NOT</u> possible (pore size 0.45 to 20 µm) samples 10 % KOH (3 x volume of sample) Digestion at room Filtration <u>NOT</u> possible temperature for 2 weeks overnight at 60° C (pore size 0.45 to $20 \mu m$) Treatment 2 Density Filtration **NOT** possible 10 % KOH (3 x separation with (pore size 20 µm) volume of sample) Sodium chloride [sample turned into overnight at 60° C (NaCI) jelly] Treatment 3 Filtration of supernatant Density separation (pore size 20 µm) with NaCl [pellets did not dissolve completely]

Workflow and results of recovery rate of microplastic particles from spiked fishmeal samples:

Transparent PP - Polypropylene (c. 0.85 g cm⁻³)

• Green PET - Polyethylene terephthalate (c. 1.37 g cm⁻³)



Addition of microfragments (respective relative densities in brackets)

• White PS – Polystyrene (c. 1.06 g cm⁻³)

Treatment 3

- Add 50 ml NaCl solution, soak overnight
- Add further 150 ml and decant top 50 ml & repeat this step once
 - Filter supernatant

Microfragments 0 8 0 0 0 			Spiked	Table - Recovery of particles according to density (sample mean)		
6 gu			Recovered		Spiked	% recovered
4 tra				Fragments		
Q 2				(relative density <1.2)	20	17.8
= 0				Fragments		
	PP	PS	PET	(relative density >1.2)	10	2.2
Fig. 1 – M						
per polyn	ner and	sample				



Fig. 2 – PS fragments (red circles) remaining on sample surface

Discussion and conclusion

Density separation with a brine solution appears to be a suitable method for extracting microplastics from fishmeal. Brine solutions are commonly used to extract microplastics from sediments^(1,16-20). Using a brine solution of NaCl of approximate density 1.21 g cm⁻³, it was not expected to recover particles of a relative density above this value (table). Using Zinc chloride (density >1.7 g cm⁻³) instead of NaCl should result in improved recovery of microplastics of relative densities above 1.21 g cm^{-3 (20)}.

Interestingly, 27.8 % of polystyrene fragments were recovered, but only 7.8 % of polypropylene fragments (Fig. 1). An underestimation of polypropylene fragments may have occurred due to the similarity of clear polypropylene and bone fragments when using a low magnification microscope. However, general low recovery rates of polypropylene and polystyrene were likely due to the lack of transfer of microfragments when decanting the supernatant. A large number of white polystyrene fragments were still visible on the solution surface of the sample after the removal of the supernatant (Fig. 2). Removing the top layer in a more controlled manner using a separating funnel, pipette or an overflow method is suggested.

References

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