- 1 Title: Novel urinary protein panels for NAFLD and fibrosis stages diagnosis
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- 3 Short Title: Urinary NITs for NAFLD diagnosis
- 4 Authors:
- 5 Gong Feng<sup>1, 2\*</sup>, MD; Xiaoxun Zhang<sup>3, 4, 5\*</sup>, MD; Liangjun Zhang<sup>3, 4, 5</sup>, MD; Wen-Yue Liu<sup>6</sup>,
- 6 MD; Shi Geng<sup>7</sup>, MD; Hai-Yang Yuan<sup>8</sup>, MD; Jun-Cheng Sha<sup>9</sup>, MD; Xiao-Dong Wang<sup>10</sup>,
- 7 MD; Dan-Qin Sun<sup>11</sup>, MD; Giovanni Targher<sup>12</sup>, MD; Christopher D. Byrne<sup>13</sup>, MD; Tian-
- 8 Lei Zheng<sup>7,14</sup>, MD; Feng Ye<sup>1</sup>, MD; Ming-Hua Zheng<sup>8,10#</sup>, MD; Jin Chai<sup>3,4,5#</sup>, MD on
- 9 behalf of CHESS-MAFLD consortium
- 10
- <sup>1</sup>The First Affiliated Hospital of Xi'an Jiaotong University, Xi'an 710061, China;
- <sup>2</sup>Xi'an Medical University, Xi'an 710021, China;
- <sup>3</sup>Department of Gastroenterology, Southwest Hospital, Third Military Medical
- 14 University (Army Medical University), Chongqing 400038, China;
- <sup>4</sup>Institute of Digestive Diseases of PLA, Southwest Hospital, Third Military Medical
- 16 University (Army Medical University), Chongqing 400038, China;
- <sup>5</sup>Cholestatic Liver Diseases Center and Center for Metabolic Associated Fatty Liver
- Disease, Southwest Hospital, Third Military Medical University (Army Medical
- 19 University), Chongqing 400038, China;
- <sup>6</sup>Department of Endocrinology, The First Affiliated Hospital of Wenzhou Medical
- 21 University, Wenzhou 325000, China;

- <sup>7</sup>Artificial Intelligence Unit, Department of Medical Equipment Management,
- 23 Affiliated Hospital of Xuzhou Medical University, Xuzhou 221004, China;
- <sup>8</sup>MAFLD Research Center, Department of Hepatology, the First Affiliated Hospital of
- 25 Wenzhou Medical University, Wenzhou 325000, China;
- <sup>9</sup>Interventional radiology, Affiliated Hospital of Xuzhou Medical University, Xuzhou
- 27 221004, China;
- 28 <sup>10</sup>Key Laboratory of Diagnosis and Treatment for The Development of Chronic Liver
- 29 Disease in Zhejiang Province, Wenzhou, Zhejiang 325000, China;
- 30 <sup>11</sup>Department of Nephrology, the Affiliated Wuxi No.2 People's Hospital of Nanjing
- 31 Medical University, Wuxi, Jiangsu Province 214001, China;
- 32 <sup>12</sup>Section of Endocrinology, Diabetes and Metabolism, Department of Medicine,
- 33 University and Azienda Ospedaliera Universitaria Integrata of Verona, Verona, Italy;
- 34 <sup>13</sup>Southampton National Institute for Health and Care Research Biomedical Research
- 35 Centre, University Hospital Southampton, Southampton General Hospital,
- 36 Southampton, UK;
- 37 <sup>14</sup>School of Information and Control Engineering, China University of Mining and
- 38 Technology, Xuzhou 221116, China.

\*These authors contributed equally to this study.

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# Footnote Page 42 43 **Contact Information:** 44 \*Ming-Hua Zheng, M.D., Ph.D., MAFLD Research Center, Department of 45 Hepatology, the First Affiliated Hospital of Wenzhou Medical University, Wenzhou, 46 China. Tel: 86-577-55579611; Fax: 86-577-55578522; E-mail: 47 zhengmh@wmu.edu.cn 48 <sup>#</sup>Jin Chai, M.D., Ph.D., Professor of Gastroenterology and Hepatology in Chongqing 49 50 University School of Medicine and Third Military Medical University. Address: Center for Metabolic Liver Diseases and Center for Cholestatic Liver Diseases, 51 Department of Gastroenterology, The First Affiliated Hospital (Southwest Hospital), 52 Third Military Medical University (Army Medical University) Chongqing, 400038, 53 China; Tel: 86-23-68765331; Fax: 86-23-65410853; E-mail: jin.chai@cldcsw.org 54 55 56 **Total word count:** 4060 words Number of figures/supplementary figures: 4/5 57 58 Number of tables/supplementary tables: 1/7 59 60 **Author contributions to this manuscript:** Ming-Hua Zheng, Jin Chai, and Gong Feng conceived and designed the study; Gong 61

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*OPLS-DA*, orthogonal partial least square discriminant analysis; *UPLC-MS/MS*,

ultra-performance liquid chromatography-mass spectrometry.

#### Abstract

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**Background & Aims:** There is an unmet clinical need for simple non-invasive tests 88 to diagnose nonalcoholic fatty liver disease (NAFLD) and fibrosis stages. We aimed 89 to test whether urine samples could be used to diagnose NAFLD, NAFLD with 90 fibrosis (fibrosis stage  $F \ge 1$ ), and NAFLD with significant fibrosis (fibrosis stage  $F \ge 2$ ). 91 **Methods:** We collected urine samples from 100 patients with biopsy-proven NAFLD 92 and 40 healthy volunteers and proteomics and bioinformatics analyses were 93 performed in this derivation cohort. Diagnostic models were developed for detecting 94 95 NAFLD (UP<sub>NAFLD</sub> model), NAFLD with fibrosis (UP<sub>fibrosis</sub> model), or NAFLD with significant fibrosis (UP<sub>significant fibrosis</sub> model). Subsequently, the derivation cohort was 96 divided into training and testing sets to evaluate the diagnostic efficacy of these 97 98 diagnostic models. In a separate independent validation cohort of 100 patients with biopsy-proven NAFLD and 45 healthy controls, urinary enzyme-linked 99 immunosorbent assay (ELISA) analyses were undertaken to validate the accuracy of 100 101 these newly developed diagnostic models. 102 Results: The UP<sub>fibrosis</sub> model and the UP<sub>significant fibrosis</sub> model showed an AUROC of 0.863 (95% CI: 0.725-1.000) and 0.858 (95% CI: 0.712-1.000) in the training set; 103 0.837 (95% CI: 0.711-0.963) and 0.916 (95% CI: 0.825-1.000) in the testing set. The 104 UP<sub>NAFLD</sub> model showed excellent diagnostic performance and the area under the 105 receiver operator characteristic curve (AUROC) exceeded 0.90 in the derivation 106 cohort. In the independent validation cohort, the AUROC for all three of the above 107 diagnostic models exceeded 0.80. 108

Conclusions: Our newly developed models constructed from urine protein

biomarkers have good accuracy for non-invasively diagnosing early liver fibrosis in

NAFLD.

Keywords: Fibrosis, NAFLD, Urinary proteomics, Diagnosis, Liver biopsy

### Introduction

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Nonalcoholic fatty liver disease (NAFLD) has become the most common chronic liver disease, affecting up to ~30% of the world's population. <sup>1,2</sup> NAFLD includes a 117 spectrum of progressive liver conditions ranging from nonalcoholic fatty liver (NAFL) 118 to nonalcoholic steatohepatitis (NASH), cirrhosis and hepatocellular carcinoma.<sup>3-5</sup> 119 Patients with NAFLD may develop varying amounts of liver fibrosis, and previous 120 studies have shown that the severity of liver fibrosis is the strongest histologic risk 121 factor of liver-related complications and mortality.<sup>6,7</sup> Recent studies have also shown 122 that even NAFLD with fibrosis (fibrosis stage  $F \ge 1$ ) or NAFLD with significant 123 fibrosis (fibrosis stage  $F \ge 2$ ) is a predictor of overall and liver-related mortality in 124 patients with NAFLD.<sup>8-11</sup> The assessment of liver fibrosis extends beyond the realm 125 of NAFLD and holds significance in terms of evaluating cardiovascular risk.<sup>12</sup> 126 Noninvasive assessment of hepatic fibrosis may predict cardiovascular events and 127 overall mortality in patients with NAFLD. 12 While the diagnostic performance of 128 traditional non- invasive tests are satisfactory for ruling out advanced fibrosis (fibrosis 129 stage  $F \ge 3$ ), their performance is not adequate for diagnosing fibrosis or significant 130 liver fibrosis. 13,14 Therefore, the detection of NAFLD with fibrosis and NAFLD with 131 significant fibrosis can facilitate a more accurate assessment of the patient's condition, 132 treatment response, and prognostic outcomes. 133 134 To date, liver biopsy remains the 'gold standard' for staging the severity of liver 135

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disease in NAFLD. However, liver biopsy is an invasive method that is expensive,

potentially risky, has possible sampling errors, and patients may not be available to undergoing repeated liver biopsies over time for a benign condition. <sup>13,15,16</sup>

Consequently, it is essential to find pragmatic, less expensive and safer non-invasive methods for diagnosis, staging and monitoring liver fibrosis. Non-invasive diagnosis of each of the stages of NAFLD has been an unmet clinical need. <sup>17,18</sup> Urine tests are simple and also more easily accessible than blood tests; thus, urine has been proposed as a source of potential biomarkers for diagnosis of human diseases in several research fields. <sup>19</sup> Moreover, urine changes may reflect dynamic changes in disease status, which is potentially important for liver disease in NAFLD, especially when clinicians need to know whether the disease is improving or not with a pharmacologic treatment. <sup>20</sup>

Recently, proteomic analyses of liver tissue and blood have been used to discover new biomarkers and therapeutic targets in NAFLD.<sup>21,22</sup> Urine proteomics has also become a focus of research in other chronic diseases, such as diabetic nephropathy, type 1 diabetes, and adult-onset Still's disease.<sup>23,24,25</sup> To date, however, proteomic studies on urine in individuals with NAFLD are scarce, and there is still a lack of any relevant evidence as to whether urine can be used to diagnose NAFLD and the severity of liver fibrosis. Thus, the aim of our study was to explore the urinary protein panels to diagnose NAFLD, NAFLD with fibrosis, and NAFLD with significant fibrosis in a liver biopsy-based derivation cohort of patients with NAFLD.

#### Methods

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Patients and Urine Sample Collection

Patients with NAFLD were diagnosed at the First Affiliated Hospital of Wenzhou Medical University from December 2016 to December 2018, as part of the Prospective Epidemiologic Study of Significantly Characterized Non-Alcoholic Steatohepatitis (PERSONS) cohort study. 14 The inclusion criteria for the study were as follows: (1) individuals aged 18-75 years; (2) individuals with fatty liver diagnosed by imaging and/or elevated serum liver enzymes who were willing to undergo a liver biopsy; (3) individuals with availability of urine samples for protein quantification using label-free quantitative proteomics technology; and (4) individuals who were willing to provide written informed consent. We excluded from the study: (1) individuals with significant alcohol consumption (≥140 g/week in men or ≥70 g/week in women); (2) those who were taking potentially hepatotoxic medications; (3) individuals with viral hepatitis, autoimmune hepatitis, or other known chronic liver diseases; (4) those with pathological liver biopsy suggesting fat content < 5%; and (5) those with incomplete data. Since urine values can vary considerably during a 24-h period, the first-morning urine samples were collected for all participants. Healthy controls were derived from a physical examination population, who was free of fatty liver, as confirmed by ultrasonography. Finally, in the derivation cohort, we obtained 100 patients with biopsy-confirmed NAFLD who had completed urine proteomic abundance measurements from the Wenzhou center and 40 non-steatotic healthy controls from the Southwest Hospital of the Army Medical University, who also

completed urine proteomic abundance measurements. The derivation cohort was then divided into a training set and a testing set (in a 1:1 ratio) to examine the diagnostic efficacy of early patterns diagnosis models for NAFLD. In a separate and independent validation cohort, we included 45 healthy controls from the Southwest Hospital of the Army Medical University and 100 patients with biopsy-confirmed NAFLD from the First Affiliated Hospital of Wenzhou Medical University. The flowchart of the study is summarized in Figure 1. We undertook proteomics and bioinformatics analyses to uncover potential diagnostic biomarkers and develop diagnostic models for identifying NAFLD (UP<sub>NAFLD</sub> model), NAFLD with fibrosis stage F≥1 (UP<sub>fibrosis</sub> model), or NAFLD with significant fibrosis stage F≥2 (UP<sub>significant fibrosis</sub> model), respectively in the derivation cohort. Furthermore, we also validated the diagnostic efficacy of urinary diagnostic panels utilizing enzyme-linked immunosorbent assay (ELISA) in an external validation cohort of patients with biopsy-confirmed NAFLD. The study was conducted in accordance with the ethical guidelines of the Declaration of Helsinki and the International Conference on Harmonization Guidelines for Good Clinical Practice. Written informed consent was obtained from all participants.

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### Liver histology

Ultrasound-guided percutaneous liver biopsy was performed using a 16-gauge

Hepafix needle. Liver biopsy samples were stained with hematoxylin, Masson's

trichrome as well as eosin and subsequently assessed by an experienced liver

pathologist, who was blinded to patients' clinical and laboratory data. Liver biopsy

specimens were required to be >1 cm and the number of portal areas was >6. The histological scoring of NAFLD was assessed by using the NAFLD activity score (NAS), as proposed by the NASH Clinical Research Network.  $^{26}$  The NAS score includes three histologic features including the presence of steatosis, lobular inflammation and hepatocellular ballooning. Liver fibrosis was staged from zero to 4 as follows: 0 = no fibrosis, 1 = perisinusoidal or portal fibrosis; 2 = perisinusoidal and portal/periportal fibrosis; 3 = bridging fibrosis; and 4 = highly suspicious or definite cirrhosis, respectively. The presence of liver fibrosis was defined as fibrosis stage  $\geq F1$ , while significant fibrosis was defined as fibrosis stage  $\geq F2$ , which is clinically relevant for prognostic liver-related outcomes.  $^{27}$ 

## Label-free quantitative proteomics technology

Label-free proteomic quantification was based on the use of the ultra-performance liquid chromatography-mass spectrometry (UPLC-MS/MS). Mass spectrometry can obtain the mass-to-charge ratio and the signal intensity of peptides in a sample, as well as the mass-to-charge ratio and the signal intensity of fragment ions after peptide fragmentation. Usually, the information at the peptide level is referred to as a first-level spectrum and the peptide fragment ion information a second-level spectrum. The information contained in the spectra is very complex, and a database is created to resolve the peptide sequences contained in the spectra. Before searching the database, a theoretical secondary spectrum database is constructed from the protein sequences in the database. Then the secondary spectrum generated by mass spectrometry is

analysed and compared with the theoretical secondary spectrum, and the correct matching theoretical peptide sequences obtained after algorithm scoring and filtering. The protein information contained can be identified by the protein-specific peptides identified. The secondary mass spectrometry data in this experiment was searched using Maxquant (v1.6.15.0).<sup>29</sup> To obtain high-quality data, the search library analysis results required further data filtering. We set the false discovery rate at 1% for the three levels of the spectrum, peptide and protein identification; and the identified proteins had to contain at least one specific (unique) peptide.

# Measurement of specific protein levels in urine samples

The levels of ACE2 (CAT#F10272-A), CILP2 (CAT#F111328-A), ENPP7 (CAT#F111336-A), MPST (CAT#F111323-A), OGFOD3 (CAT#F111319-A), P2RX4 (CAT#F111332-A), TMEM256 (CAT#F111315-A), TMEM25 (CAT#F111311-A), or ICAM-1/CD54 (CAT#F0034-A) proteins in the urine samples from patients (n=45 for healthy volunteers and n=100 for biopsy-proven NAFLD patients) were quantified using the corresponding commercial ELISA Kit (Fankew, Shanghai FANKEL Industrial Co., Ltd, Shanghai, China), according to their manufacturer's instructions.

## Statistical analysis

Reporting of clinical data was according to the most frequently obtained data from each participant, with the mean  $\pm$  SD presented for normal continuous variables and the median (interquartile range) for non-normal continuous variables. Otherwise, the

frequency was used for categorical variables. For proteomic abundances, proteins with a missing value ratio higher than 50% in all samples were removed, and the knearest neighbor algorithm was used to fill protein abundances, and log2 transformation was performed for data at the same time. The methods for screening differential proteins include orthogonal partial least square discriminant analysis (OPLS-DA) or linear models for microarray data (limma). OPLS-DA has been widely used in the multi-omics analysis, including proteomic analysis and metabolomics analysis.<sup>30,31</sup> Limma is a differential expression screening method based on generalized linear equations. The R software package limma (version 3.40.6) was used for differential analysis to obtain the differential proteins among different groups. This method has also been used in proteomic analysis. 32,33 Furthermore, we used LASSO regression to pick out the more important proteins.<sup>34</sup> The R package glmnet was used for the LASSO regression analysis and the 10 folds cross-validation method was used to filter the lambda. 35,36 The cross-validation method divided the data into 10 equal parts, first fitting the full data to generate the lambda sequence, then excluding one part of the data at a time and using the remaining 9 parts for validation. The mean and standard deviation of the errors obtained from the 10 validations were calculated. The diagnostic efficacy of each key protein was assessed by the area under the receiver operating characteristics curves (AUROC). The top four ranked proteins of AUROC were considered for inclusion in the pooled analysis of the indicators. The Variance inflation factor (VIF) was calculated to detect the presence of multicollinearity between identified proteins. A cut-off value of 5 was used when

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applying the VIF in the study.<sup>37</sup> We used a stepwise algorithm to find the best combined model including key proteins, which was based on the Akaike information criterion (AIC) principle that involves removing the variables that were not statistically significant.

#### **Results**

Characteristics of NAFLD patients and urine proteomes in the derivation cohort

Patients with NAFLD were divided into two subgroups: patients with liver fibrosis

(defined as stage  $\geq F1$ ; n=81) and those without fibrosis (n=19) or, alternatively,

patients with significant liver fibrosis (defined as stage  $\geq F2$ ; n=16) and those without

significant fibrosis (n=84). **Figure 2** shows the workflow in the derivation cohort.

Baseline characteristics of patients with NAFLD and different fibrosis stages in the

derivation cohort are presented in **Table 1** and **Supplementary Table 1**. Label-free

proteomic analyses identified a total of 4206 proteins among healthy controls and

NAFLD patients with different fibrosis stages.

# Proteomic biomarkers for non-invasive identification of NAFLD in the derivation

cohort

The OPLS-DA method is more sensitive to variables with lower levels of correlation and helps maximize the difference between NAFLD patients and healthy controls based on differential proteins compared with PCA and PLS-DA (**Figure 2e** and **Supplementary Figure 1**). The R2Y represents the interpretation rate of the model to

the Y matrix, and the Q2 represents the prediction ability of the model. The closer these three indexes are to 1, the more stable and reliable the model is. Generally, the model is considered effective with a Q2 value above 0.5. In our modeling results, the parameters of R2Y and O2 were 0.956 and 0.775, respectively, thereby suggesting that the model is reliable and has good prediction ability, and there was also no overfitting in our model (Supplementary Figure 2). The OPLS-DA S-plot clearly shows the distribution of all urinary proteins based on their variable importance in projection (VIP) values. Subsequently, 37 differential proteins were selected by using the following three selection criteria: VIP > 1, p < 0.05 and FC > 2 (or FC < 0.5, Figure 3a). We further used the LASSO regression to screen for important differential proteins and found 15 urinary key proteins (Figure 3b and 3c). The clustering of these 15 key proteins is shown in Supplementary Figure 3a, and their intercorrelations are reported in Supplementary Figure 4a. By enrichment analysis, we found that these key proteins are associated with ferroptosis, oxidoreductase activity, and coenzyme A biosynthesis (Supplementary Figure 3b). Then, using ROC curve analyses, we found that the top four urinary proteins among these 15 proteins were EPHA10, CILP2, TMEM25, and MPST, respectively (Figure 4a). The correlation analyses of these four key proteins showed that EPHA10 correlated with CILP2, TMEM25 and MPST (Supplementary Figure 4b; Supplementary Table 2), but none of these proteins showed evidence of collinearity (VIF < 5). Further, we found by stepwise regression analysis that the combined diagnostic model (UP<sub>NAFLD</sub> model) of the three urinary key proteins (CILP2, TMEM25, and MPST) had an AUROC of

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0.983 (95% CI: 0.961-1.000) in the training set and 0.968 (95% CI: 0.932-1.000) in the testing set from derivation cohort, with excellent diagnostic efficacy (**Figure 4b**). We also found that the UP<sub>NAFLD</sub> model had an AUROC of 0.961 (95% CI: 0.915-1.000) in males and 0.993 (0.979-1.000) in females.

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# Proteomic biomarkers for non-invasive identification of fibrosis stage ≥F1 in the

### derivation cohort

The R software package limma was used to obtain urinary differential proteins between patients with liver fibrosis ≥F1 and those without fibrosis. A total of 56 differential urinary proteins were subsequently selected by using the criteria of p < 0.05 and FC > 1.5 (or FC < 0.7, **Figure 3d).** In addition, 22 key proteins were identified using the LASSO regression (Figures 3e and 3f). The clustering results of these 22 key proteins are shown in **Supplementary Figure 3c**, and the correlations among them are shown in **Supplementary Figure 4c**. Our enrichment analysis revealed that these key proteins were associated with the renin-angiotensin system, cornified envelope, humoral immune response, and regulation of inflammatory response (Supplementary Figure 3d). After analyzing these 22 key proteins using ROC curves, we found that OGFOD3, ACE2, ENPP7, and P2RX4 were the top four key proteins (Figure 4c). Correlation analysis of these four key proteins revealed that ACE2 correlated with OGFOD3, ENPP7, and P2RX4 (Supplementary Figure 4d; Supplementary Table 3), but none of these variables showed evidence of collinearity (VIF < 5). Moreover, we discovered that the combined diagnostic model (UP<sub>fibrosis</sub>

model) of the 3 key proteins (OGFOD3, ENPP7, and P2RX4) had an AUROC of 0.863 (95% CI: 0.725-1.000) in the training set and 0.858 (95% CI: 0.712-1.000) in the testing set using stepwise regression analysis (**Figure 4d**). We also discovered that UP<sub>fibrosis</sub> model had an AUROC of 0.877 (95% CI: 0.765-0.989) in males and 0.857 (95% CI: 0.669-1.000) in females.

Proteomic biomarkers for non-invasive identification of fibrosis stage  $\geq$ F2 in the derivation cohort

By limma analysis, we identified 44 differential urinary proteins with the selection criteria of p < 0.05 and FC > 1.5 (or FC < 0.7, Figure 3g). We also identified 12 key proteins using LASSO regression (Figure 3h and 3i). Supplementary Figure 3e shows the clustering results of these 12 key proteins, and the correlations among them are reported in Supplementary Figure 4e. Using enrichment analysis, we identified these key proteins as being associated with dense platelet granule lumens and collagen-containing extracellular matrix (Supplementary Figure 3f). Using the AUROC analyses, we found PECAM1, TMEM256, MSRA, and ICAM1 to be the top four key proteins (Figure 4e). TMEM256 had the highest AUROC of 0.772 (95% CI: 0.658-0.886), with a sensitivity of 0.750 and specificity of 0.786 for significant fibrosis in NAFLD. PECAM1 was correlated with TMEM256, MSRA, and ICAM1 (Supplementary Figure 4f; Supplementary Table 4), but none of these four key proteins showed evidence of collinearity (VIF < 5). Furthermore, using a stepwise regression analysis, we found that in the combined diagnostic model (UPsignificant fibrosis

model) two key proteins (TMEM256 and ICAM1) had an AUROC of 0.837 (95% CI: 0.711-0.963) in the training set and 0.916 (95% CI: 0.825-1.000) in the testing set, respectively (**Figure 4f**). We also found UP<sub>significant fibrosis</sub> model had an AUROC of 0.829 (95% CI: 0.698-0.960) in men and 0.833 (95% CI: 0.690-0.977) in women.

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# Validation by ELISA analyses

ELISA analyses were performed to validate the combined diagnostic models of the key proteins in above urine samples. In the independent validation cohort, we included 45 healthy controls and 100 patients with biopsy-confirmed NAFLD. The baseline characteristics of this validation cohort are presented in **Supplementary Table 5.** Of these 100 patients with NAFLD, 57 had fibrosis stage ≥F1 and 13 had significant fibrosis (stage ≥F2). Baseline characteristics of NAFLD patients with different fibrosis stages belonging to the validation cohort are presented in Supplementary Table 6 and Supplementary Table 7. It was found that the model consisting of three urinary proteins, CILP2, TMEM25 and MPST, achieved an AUROC of 0.850 (95% CI: 0.784-0.915) for differentiating NAFLD patients from healthy controls (Supplementary Figure 5a). The combined model consisting of three urinary proteins, OGFOD3, ENPP7 and P2RX4, showed an AUROC of 0.804 (95% CI: 0.718-0.889) for differentiating patients with fibrosis (stage F  $\geq$ 1) from those without fibrosis (Supplementary Figure 5b). The combined model comprised of two urinary proteins, TMEM256 and ICAM-1, was found to have an AUROC of 0.807 (95% CI: 0.715-0.899) for differentiating between patients with significant

fibrosis (stage  $F \ge 2$ ) and those without significant fibrosis (F0 + F1 stages)

(Supplementary Figure 5c).

### **Discussion**

Our urine proteomics profiling showed for the first time that there is a pattern of urinary proteins in patients with biopsy-proven NAFLD and liver fibrosis in a Han Chinese population. We discovered for the first time that TMEM256 had an AUROC of 0.772 (95% CI: 0.658-0.886), with a sensitivity of 0.750 and specificity of 0.786 for significant fibrosis in NAFLD. The diagnostic model consisting of CILP2, TMEM25, and MPST was useful to distinguish NAFLD patients from healthy controls. The diagnostic models composed of OGFOD3, ENPP7, and P2RX4 for NAFLD with fibrosis, and composed of TMEM256 and ICAM-1 for NAFLD with significant fibrosis provide promising results for future clinical investigation in other ethnic groups. Although Liu et al. identified some potential urinary biomarkers for NAFLD diagnosis, these authors did not use liver biopsy (i.e., the gold standard) for verifying each of the stages of liver disease in NAFLD. In addition, the sample size of the Liu study was small in both the discovery and validation cohorts (neither exceeded 30 cases).

The utilization of urinary protein panels in personalized medicine for the management of NAFLD and metabolic disorders has the potential to greatly impact current approaches to patient care. Urinary protein panels can help identify individuals who

are at risk of developing NAFLD, NAFLD with fibrosis, or NAFLD with significant fibrosis, even before the onset of symptoms. This early detection can aid in the prompt initiation of lifestyle changes and therapeutic interventions that can prevent or slow the progression of the disease. Urinary protein panels can also help monitor the effectiveness of therapeutic interventions and track the progression of the disease over time by changes in model scores. Meanwhile, NAFLD is closely associated with metabolic disorders and may exacerbate the conditions of metabolic disorders. Therefore, utilizing these models to manage NAFLD can help patients improve their metabolic status and reduce the risk of metabolic disorder-related diseases such as cardiovascular disease, stroke, and kidney disease, on the basis of effectively managing NAFLD.

We found that the model composed of MPST, CILP2 and TMEM25 was a reliable non-invasive diagnostic tool for identifying NAFLD. Li et al. suggested that free fatty acids increased hepatic MPST expression and inhibited the CSE/H<sub>2</sub>S pathway, thus leading to NAFLD. MPST might be a potential therapeutic target for NAFLD. Genome-wide association studies have reported multiple loci associated with NAFLD, including CILP2. Meanwhile, we discovered that the diagnostic model composed of OGFOD3, ENPP7 and P2RX4 was useful for identifying NAFLD with fibrosis. Xie et al. showed that disrupting HIF-2 $\alpha$  in the intestine specifically reduced liver steatosis, along with diminished HIF-2 $\alpha$  signaling in the small intestine, several mRNAs encoded by ceramide-synthesis-related genes, were significantly downregulated,

including ENPP7.<sup>42</sup>

We also found that TMEM256 and ICAM-1 could be used as reliable diagnostic biomarkers for significant fibrosis (stage  $F \ge 2$ ) in NAFLD in the future. TMEM256, which localized within the mitochondrial membrane, belongs to mitochondrial proteins and is involved in oxidative phosphorylation.<sup>43</sup> More than 40% of the mitochondrial proteome is associated with human diseases, including NAFLD.<sup>44</sup> Mitochondrial dysfunction and endoplasmic reticulum stress are strongly implicated in the development and progression of NAFLD. As an important member of the immunoglobulin superfamily, the increased expression of ICAM-1 on the cell surface is associated with development of several diseases, and can be used as a predictor of liver fibrosis.<sup>45</sup> The results of animal experiments showed that in liver fibrosis tissues, ICAM1 is expressed in hepatocytes, mostly in the portal zone, inflammatory zone, and focal necrotic zone, and the intensity of its expression increases progressively with the severity of liver inflammation and fibrosis.<sup>46</sup>

Urine is a readily available and highly accepted test analyte by patients and has become a specimen for monitoring and detection of many diseases or physiological characteristics. <sup>47-49</sup> In our study, GO and KEGG analyses showed that urine proteins in NAFLD patients were mainly associated with ferroptosis, oxidoreductase activity, and coenzyme A biosynthesis. As a newly discovered form of iron-dependent programmed cell death, ferroptosis is involved in the development and progression of

NAFLD.<sup>50</sup> A typical feature of NAFLD is development of oxidative stress and redox imbalance.<sup>51</sup> In recent years, an increasing number of studies have found that coenzyme A is closely related to NAFLD from different perspectives. Zhou et al. revealed that inhibition of stearoyl-coenzyme A desaturases-1 ameliorated hepatic steatosis. 52 Huang et al. suggested that enoyl coenzyme A hydratase-1 protected against high-fat-diet-induced hepatic steatosis and insulin resistance.<sup>53</sup> In our study, urine proteins in NAFLD with fibrosis were associated with the renin-angiotensin system, cornified envelope, humoral immune system, and regulation of inflammatory response. A prior report has suggested that ACE2/Ang-(1-7)/Mas may also contribute to NAFLD development.<sup>54</sup> NAFLD is characterized by a dysregulated immune response.<sup>55</sup> It is believed that inflammation and upregulation of inflammatory mediators play a secondary role in the pathogenesis of NAFLD.<sup>56</sup> In our study, urine proteins in NAFLD with significant fibrosis were associated with dense platelet granule lumens and collagen-conjugate lumens. Yang et al. discovered that counting platelets may help to determine the severity of liver injury and liver fibrosis. <sup>57</sup> The above enrichment analysis suggests that urine proteomic data may reflect NAFLD disease characteristics.

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In the field of proteomics research, UPLC-MS/MS based techniques for the detection and relative quantification of thousands of proteins in biological samples have been widely used, and two mainstream methods have been developed for labeled quantification and unlabeled quantification (label-free), both of which are based on

data-dependent acquisition. 48 These methods are based on the data-dependent acquisition (DDA) model to acquire protein spectral data. The label-free proteomic quantification was used in this study. Label-free proteomic quantification is a new protein quantification technique that does not rely on isotope labeling and analyzes enzymatic peptides by liquid-liquid mass spectrometry. 58 This technique simply analyzes the mass spectrometry data generated during the large-scale identification of proteins and compares the signal intensities of the corresponding peptides in different samples for the relative quantification of the corresponding proteins. It facilitates the pursuit of more precise qualitative and quantitative results and avoids experimental errors introduced by labeling and liquid-phase grouping. 59 In this study, a series of frontier technologies such as protein extraction, enzymatic digestion, liquid chromatography-mass spectrometry tandem analysis, and bioinformatics analysis were combined to finally achieve quantitative proteomic data from the samples.

There were some important limitations in the present study that should be mentioned.

First, the number of patients with NAFLD included in our study was relatively small.

Second, this study only explored the diagnostic biomarkers of NAFLD and associated

fibrosis from the perspective of high-throughput omics, but the underlying

mechanisms between these biomarkers and the disease were not addressed in depth.

In conclusion, the diagnostic model consisting of CILP2, TMEM25, and MPST was useful for distinguishing NAFLD patients from healthy controls. The diagnostic

- 489 models comprising OGFOD3, ENPP7 and P2RX4 (for NAFLD with fibrosis), and
- 490 comprising TMEM256 and ICAM-1 (for NAFLD with significant fibrosis) now need
- 491 to be further tested in other ethnic groups. Urine-based biomarkers are more
- accessible and less invasive than blood tests; and may circumvent the need to invasive
- liver biopsy to refine the staging of liver disease severity in people with NAFLD.

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# Table legend 648 Table 1. Baseline characteristics of patients with biopsy-proven NAFLD in the 649 650 derivation cohort grouped by the severity of fibrosis (fibrosis stages < 1 vs. fibrosis stages $\geq 1$ ). 651 652 Figure legends 653 Figure 1. Flow-chart of the study design, including the derivation cohort 654 (subsequently divided into the training set and testing set) and the validation cohort. 655 656 *Note:* Diagnostic models were developed for detecting NAFLD (UP<sub>NAFLD</sub> model), NAFLD with fibrosis stage F≥1 (UP<sub>fibrosis</sub> model), or NAFLD with significant fibrosis 657 stage F\ge 2 (UP\_significant fibrosis model), respectively. 658 659 (a) Derivation cohort. (b) Validation cohort. 660 Figure 2. The diagram illustration of discovering biomarkers of NAFLD and related 661 fibrosis from the perspective of urine proteomics. 662 (a) Grouping of patients with liver biopsy-proven NAFLD in the derivation cohort. (b) 663 Label-free quantitative proteomics technology data analysis process. (c) Proteomics 664 identification results. The x-axis represents the identified substances and the y-axis 665 represents the number of substances. (d) Molecular weight of the identified proteins. 666 The X-axis represents the size of the molecular weight and the Y-axis represents the 667 number of molecular weights in this range. (e) OPLS-DA plots of urine proteomics 668 from NAFLD and healthy individuals (R2Y, 0.956; Q2, 0.775). 669

Figure 3. Volcano plots and LASSO regression of differential proteins.

(a) Volcano map of differential proteins between NAFLD patients and healthy controls. (b, c) LASSO regression screening for important differential key proteins between NAFLD patients and healthy controls. (d) Volcano map of differential key proteins between NAFLD patients with fibrosis stage F≥1 and those without fibrosis. (e, f) LASSO regression screening for important differential proteins between NAFLD with fibrosis and NAFLD without fibrosis. (g) Volcano plot of differential key proteins between NAFLD patients with significant fibrosis stage F≥2 and those without significant fibrosis. (h, i) LASSO regression screening for important differential proteins between NAFLD patients with significant fibrosis and those

without significant fibrosis.

- **Figure 4.** ROC curve analyses of key proteins and combined models in the derivation cohort.
- (a) ROC curves of EPHA10, CILP2, TMEM25, and MPST. (b) The combined diagnostic model (UP<sub>NAFLD</sub> model) of three key proteins (CILP2, TMEM25, and MPST) for NAFLD patients. (c) ROC curves of ACE2, OGFOD3, ENPP7, and P2RX4. (d) The combined diagnostic model (UP<sub>fibrosis</sub> model) of three key proteins (OGFOD3, ENPP7, and P2RX4) for NAFLD patients with fibrosis. (e) ROC curves of PECAM1, TMEM256, MSRA, and ICAM1. (f) The combined diagnostic model (UP<sub>significant fibrosis</sub>) of the two key proteins (TMEM256, and ICAM1) for NAFLD

- patients with significant fibrosis.
- D set, training set; V set: testing set.
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| 696 | ONLINE-ONLY SUPPLEMENTARY MATERIAL   |
| 697 | Supplementary Table 1. Baseline characteristics of patients with biopsy-proven         |
| 698 | NAFLD in the derivation cohort grouped by stages of fibrosis (fibrosis stage $F \le 2$ |
| 699 | and fibrosis stage $F \ge 2$ ).  |
| 700 | Supplementary Table 2. Correlation analyses of four key proteins (MPST, EPHA10,        |
| 701 | TMEM25 and CILP2) between NAFLD patients and healthy controls.                         |
| 702 | Supplementary Table 3. Correlation analyses of four key proteins (OGFOD3,              |
| 703 | ENPP7, ACE2 and P2RX4) between NAFLD patients with fibrosis and those without          |
| 704 | fibrosis.  |
| 705 | Supplementary Table 4. Correlation analyses of four key proteins (PECAM1,              |
| 706 | TMEM256, MSRA and ICAM1) between NAFLD patients with significant fibrosis              |
| 707 | and those without significant fibrosis.  |
| 708 | Supplementary Table 5. Baseline characteristics of healthy controls and NAFLD          |
| 709 | patients in the validation cohort.   |
| 710 | Supplementary Table 6. Baseline characteristics of patients with biopsy-proven         |
| 711 | NAFLD in the validation cohort grouped by stages of fibrosis (fibrosis stage $F \le 1$ |
| 712 | and fibrosis stage $F \ge 1$ )   |
| 713 | Supplementary Table 7. Baseline characteristics of patients with biopsy-proven         |
| 714 | NAFLD in the validation cohort grouped by stages of fibrosis (fibrosis stage $F \le 2$ |
| 715 | and fibrosis stage $F \ge 2$ )   |
| 716 | Supplementary Figure 1. Plots of PCA (a) and PLS-DA (b).                               |
| 717 | Supplementary Figure 2. Permutation test of OPLS-DA.                                   |

- 718 (a) The parameters of R2Y, and Q2 in the permutation test of OPLS-DA. (b)
- 719 Permutation analysis to verify whether the model has overfitting.
- 720 **Supplementary Figure 3.** Heatmaps and enrichment analysis.
- 721 (a) Heatmap of hierarchical cluster analysis of differential key proteins in the NAFLD
- patient group (A) and in the healthy control group (B). (b) GO and KEGG enrichment
- analysis of 15 key proteins in NAFLD patients. (c) Heatmap of hierarchical cluster
- analysis of differential key proteins in NAFLD patients with fibrosis stage  $F \ge 1$  (C)
- and in those without fibrosis (D). (d) Enrichment analysis of 22 key proteins in
- NAFLD patients with fibrosis. (e) Heatmap of hierarchical cluster analysis of
- 727 differential key proteins in NAFLD patients with significant fibrosis stage  $F \ge 2$  (E)
- and those without significant fibrosis (F) groups. (f) Enrichment analysis of 12 key
- 729 proteins in NAFLD patients with significant fibrosis.
- 730 **Supplementary Figure 4.** Correlation analyses between urinary key proteins.
- 731 (a) Correlation analyses among 15 differential proteins selected to distinguish
- NAFLD patients from healthy controls. (b) Correlation analyses of four key proteins
- 733 (EPHA10, CILP2, TMEM25, and MPST). (c) Correlation analyses among 22
- differential proteins selected to distinguish NAFLD patients with fibrosis from those
- without fibrosis. (d) Correlation analyses of four key proteins (OGFOD3, ACE2,
- 736 ENPP7, and P2RX4). (e) Correlation analyses among 12 differential proteins selected
- to distinguish NAFLD patients with significant fibrosis from those without significant
- fibrosis. (f) Correlation analyses of four key proteins (PECAM1, TMEM256, MSRA,
- 739 and ICAM1).

| 740 | Supplementary Figure 5. ROC curves of combined models in the independent                  |
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| 741 | validation cohort.  |
| 742 | (a) The model consisting of three indicators, CILP2, TMEM25, and MPST, for                |
| 743 | differentiating NAFLD patients from healthy controls (UP <sub>NAFLD</sub> model). (b) The |
| 744 | combined model consisting of three indicators, OGFOD3, ENPP7, and P2RX4, for              |
| 745 | differentiating NAFLD patients with fibrosis from those without fibrosis (UPfibrosis      |
| 746 | model). (c) The combined model comprised of two indicators, TMEM256 and ICAM-             |
| 747 | 1, for differentiating NAFLD patients with significant fibrosis from those without        |
| 748 | significant fibrosis (UP <sub>significant fibrosis</sub> model).                          |
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