**Exogenous tetracosahexaenoic acid modifies the fatty acid composition of human primary T lymphocytes and Jurkat T cell leukaemia cells contingent on cell type**

Nicola A. Irvine1†, Annette L. West1†, Johanna Von Gerichten2†, Elizabeth A. Miles1, Karen A. Lillycrop3, Philip C. Calder1,4, Barbara A. Fielding2 and Graham C. Burdge1,4\*

1School of Human Development and Health, Faculty of Medicine, University of Southampton, Southampton, Hampshire, United Kingdom,

2Department of Nutritional Sciences, Faculty of Health and Medical Sciences, University of Surrey, Guildford, Surrey, United Kingdom,

3Centre for Biological Sciences, Faculty of Natural and Environmental Sciences, University of Southampton, Southampton, Hampshire, United Kingdom

4National Institute of Health and Care Research Southampton Biomedical Research Centre, University Hospital Southampton National Health Service Foundation Trust and University of Southampton, Southampton, Hampshire, United Kingdom

\*Correspondence to: Professor G.C. Burdge, School of Human Development and Health, Faculty of Medicine, University of Southampton, Southampton, Hampshire, SO16 6YD, United Kingdom [g.c.burdge@soton.ac.uk](mailto:g.c.burdge@soton.ac.uk)

**stract**

Tetracosahexaenoic acid (24:6ω-3) is an intermediate in the conversion of 18:3ω-3 to 22:6ω-3 in mammals. There is limited information about whether cells can assimilate and metabolise exogenous 24:6ω-3. This study compared the effect of incubation with 24:6ω-3 on the fatty acid composition of two related cell types, primary CD3+ T lymphocytes and Jurkat T cell leukaemia, which differ in the integrity of the polyunsaturated fatty acid (PUFA) biosynthesis pathway. 24:6ω-3 was only detected in either cell type when cells were incubated with 24:6ω-3. Incubation with 24:6ω-3 induced similar increments in the amount of 22:6ω-3 in both cell types and modified the homeoviscous adaptations fatty acid composition induced by activation of T lymphocytes. The effect of incubation with 18:3ω-3 compared to 24:6ω-3 on the increment in 22:6ω-3 was tested in Jurkat cells because primary T cells cannot convert 18:3ω-3 to 22:6ω-3. The increment in the 22:6ω-3 content of Jurkat cells incubated with 24:6ω-3 was 19.5-fold greater than that of cells incubated with 18:3ω-3. Acyl-coA oxidase siRNA knockdown decreased the amount of 22:6ω-3 and increased the amount of 24:6ω-3 in Jurkat cells. These findings show exogenous 24:6ω-3 can be incorporated into primary human T lymphocytes and Jurkat cells and induces changes in fatty acid composition consistent with its conversion to 22:6ω-3 via a mechanism involving peroxisomal β-oxidation that is regulated independently from the integrity of the upstream PUFA synthesis pathway. One further implication is that consuming 24:6ω-3 may be an effective alternative means of achieving health benefits attributed to 20:5ω-3 and 22:6ω-3.

**Introduction**

Synthesis of longer-chain ω-3 polyunsaturated fatty acids (PUFAs) from the essential fatty acid α-linolenic acid (18:3ω-3) involves a pathway of mostly alternating desaturation and carbon chain elongation reactions that occur in the endoplasmic reticulum (Sprecher 2000). In rodent (Voss *et al.* 1991) and human (Sibbons *et al.* 2014) hepatocytes, the first, rate-limiting reaction is desaturation at the Δ6 position of 18:3ω-3, which is catalysed by the protein product of the *FADS2* gene, namely Δ6 desaturase, followed by the addition of 2 carbon atoms by elongase-5 activity (Figure 1). Desaturation at the Δ5 position by Δ5 desaturase, yields 20:5ω-3 which is converted by two cycles of chain elongation by elongase-5 then elongase- 2 or 5 activities to form the intermediate 24:5ω-3 which is converted to 24:6ω-3 by Δ6 desaturase. Synthesis of 22:6ω-3 involves translocation of 24:6ω-3 from the endoplasmic reticulum to peroxisomes and removal of 2 carbon atoms by one cycle of β-oxidation (Voss *et al.* 1991; Moore *et al.* 1995) (Figure 1). The reactions downstream of 22:5ω-3 synthesis have been suggested to regulate 22:6ω-3 synthesis independently from the initial desaturation/elongation reactions (Sprecher 1999; Burdge 2004). This view is supported by the findings of whole body tracer studies using stable isotope labelled 18:3ω-3, which showed sexual dimorphism in 22:6ω-3 synthesis (Burdge *et al.* 2002; Burdge & Wootton 2002; Pawlosky *et al.* 2003). Moreover, competition between exogenous 18:3ω-3 and endogenous 24:5ω-3 for Δ6 desaturase activity could modify the synthesis of the terminal product 22:6ω-3 by reducing the desaturation of 24:5ω-3 to 24:6ω-3 (Burdge 2022) which may explain the reduction in blood or tissue 22:6ω-3 content in, at least, some 18:3ω-3 dietary supplementation trials (Gibson *et al.* 2013; Burdge 2022). Alternatively, direct synthesis of 22:6ω-3 by desaturation at the Δ4 position of 22:5ω-3 has been detected in MCF7 breast cancer cells that lack Δ6 desaturase activity (Grammatikos *et al.* 1994; Park *et al.* 2015) and a carnitine-dependent mechanism for this reaction has also been proposed (Infante & Huszagh 2000). One interpretation of these findings is that different cell types differ in their metabolic strategy for 22:6ω-3 synthesis. Although 24:6ω-3 is regarded as a metabolic intermediate that does not accumulate in tissues, rodents can convert dietary (Gotoh *et al.* 2018) or infused (Metherel *et al.* 2019) 24:6ω-3 into 22:6ω-3 *in vivo* and human skin fibroblasts can synthesise 22:6ω-3 from radiolabelled 24:6ω-3 *in vitro* (Moore *et al.* 1995), which suggest that at least some cell types can use exogenous 24:6ω-3 as a substrate Induction of proliferation of T lymphocytes involves homeoviscous adaptations in membrane fatty acid composition (Shires *et al.* 1989; Anel *et al.* 1990; Calder *et al.* 1994; von Gerichten *et al.* 2021) and in the relative proportions of phospholipid classes and individual molecular species (Ferber *et al.* 1975; Lonnberg *et al.* 2013) that are disrupted by incubation with ω-3 PUFAs (Calder *et al.* 1994). Therefore, we investigated the effect of incubating purified quiescent or mitogen-stimulated purified human CD3+ T lymphocytes with 24:6ω-3 on their fatty acid composition as a proxy measure of 24:6ω-3 conversion to 22:6ω-3.

The widely studied Jurkat T lymphocyte model cell line (Abraham & Weiss 2004) expresses *ELOVL2* and can synthesise 22:6ω-3 from 18:3ω-3 (Sibbons *et al.* 2018). We used Jurkat cells to compare the effects of exogenous 18:3ω-3 and 24:6ω-3 on fatty acid composition as a proxy measure to assess the relative effectiveness of 18:3ω-3 and 24:6ω-3 as substrates for 22:6ω-3 synthesis. We also investigated whether peroxisomal and mitochondrial fatty acid β-oxidation are involved in any 24:6ω-3-induced changes in Jurkat fatty scid composition. **Materials and Methods**

*Ethics statement*

The study was reviewed and approved by the East of England - Cambridge Central Research Ethics Committee (approval number 19/EE/0096) and all participants gave written informed consent. The purchase and use of primary T cells from StemCell Technologies UK Ltd. was reviewed and approved by the University of Southampton Faculty of Medicine Ethics Review Committee (submission I.D. 49658 and 58050.A1).

*Participants and collection of blood samples*

The inclusion and exclusion criteria used to select participants in the study were described previously (von Gerichten *et al.* 2021). Briefly, donors were healthy men and women with a age of 41 (range 21 – 48) years (n = 10 (4 women)) and median body mass index 25.6 (24.1 – 26.5) kg/m2, blood pressure within age-adjusted normal ranges, non-fasting total cholesterol concentration < 7.5 mmol/L, HbA1c concentration < 42 mmol/mol, and C-reactive protein concentration < 3 mg/L. Participants did not habitually consume fish oil, or dietary oil supplements, smoke tobacco, or report any chronic disease. Volunteers were excluded if they did not meet the inclusion criteria, were pregnant or intending to become pregnant during the study, or were already participating in a clinical study. Non-fasting venous blood samples (100 mL) were collected into tubes containing lithium heparin

*Isolation and culture of CD3+ T cells from whole blood*

Peripheral blood mononuclear cells (PBMCs) were prepared from whole blood using a histopaque density cushion and centrifugation at 845 x g for 15 minutes at room temperature (von Gerichten *et al.* 2021). PBMCs were collected into RPMI1640 medium containing 10% (v/v) heat-inactivated homologous pooled serum (Sigma-Aldrich, Poole, UK) (Complete medium; Table 1). CD3+ T cells were isolated by negative selection using the T cell EasySep kit (StemCell Technologies, UK Ltd., Cambridge, UK) according to the manufacturer’s instructions. Isolated T cells were washed with 10 mL RPMI1640 and collected by centrifugation at 300 x g for 10 minutes at room temperature. CD3+ T lymphocytes were cryopreserved as described (Prescott *et al.* 1999; Noakes *et al.* 2012) and stored in liquid nitrogen. Blood donations by participants were suspended during the United Kingdom national restrictions to mitigate the SARS-CoV-2 pandemic. Consequently, to increase the sample number, purified CD3+ T lymphocytes were purchased from StemCell Technologies UK Ltd (Cambridge, UK) (Catalog number 70024.1); these cells were collected from anonymous donors whose characteristics met the inclusion criteria for the study.

T lymphocyte culture was carried out as described (von Gerichten *et al.* 2021). Cryopreserved cells were thawed, adjusted to a density of 1 × 106 cells/mL and incubated in RPMI1640 containing 2 mM L-glutamine, 100 units/mL penicillin and 100 µg/mL streptomycin and 10% (v/v) heat-inactivated pooled human serum (Sigma-Aldrich) for 96 hours with or without concanavalin A (10 μg/mL) (Con. A; Sigma-Aldrich), and either with or without 6(z), 9(z), 12(z), 15(z), 18(z), 21(z)-24:6ω-3 (30 μM) (Cambridge Bioscience, UK) in a humidified incubator at 37oC in an atmosphere containing 5% (v/v) CO2. Jurkat cells were obtained from local stocks and maintained under the same conditions as T cells, without Con. A (Sibbons *et al.* 2018) for 96 hours either with or without 24:6ω-3(30 µM) (Cambridge bioscience,UK) or 18:3ω-3 (39µM) (Sigma Aldrich). Jurkat and T cells were collected by centrifugation, washed with unsupplemented RPMI1640 as before, and then snap-frozen and stored at -80°C for fatty acid analysis. The purity of both18:3ω-3 and 24:6ω-3 was greater than 95%.

In some experiments, Jurkat cells were treated with the carnitine palmitoyl transferase-1 inhibitor Etomoxir (5 µM) (Sigma-Aldritch) together with 24:6ω-3 (30 µM) for 48 hours. Cells were collected by centrifugation and washed and processed for fatty acid analysis as before.

*siRNA knockdown of acyl-CoA oxidase-1 (ACOX1) in Jurkat cells and RTPCR analysis*

Jurkat cells were suspended in serum-free Accell siRNA delivery media (Horizon Discovery Biosciences Ltd., Cambridge, UK) containing glutamine at the density of 1 × 106 cells /mL and treated with either Accell human *ACOX1* SMARTPool siRNA (1 µM) (Horizon Discovery Biosciences Ltd.) or non-targeted human pool siRNA (1 µM) (Horizon Discovery Biosciences Ltd.) and incubated for 72 hours at 37°C, in an atmosphere containing 5% (v/v) CO2. After 72 hours, the plates were centrifuged at 300 x g for 10 minutes, the supernatant was removed and replaced with RPMI containing 10% human serum and 30 µM of 24:6n-3. The cells were then incubated for a further 96 hours. At the end of the incubation, the cells were collected, washed twice in PBS and pelleted for fatty acid composition analysis and to verify *ACOX1* knockdown. RNA extraction and qRTPCR were carried out essentially as described (von Gerichten *et al.* 2021). Briefly, RNA was extracted using the RNeasy Mini kit (Qiagen) with on-column DNAse activity. RNA was eluted in RNase-free water (30 μl). RNA concentration was measured and purity was assessed using a NanoDrop1000 spectrophotometer. cDNA was synthesised by reverse transcription. The level of the ACOX1 transcript was measured by qRTPCR using QuantiTect assay Hs\_ACOX1\_1\_SG (QT00078960) (Qiagen) with QuantiTect Sybr Green PCR kit (Qiagen). Amplified transcripts were quantified using the standard curve method(Cikos *et al.* 2007) and normalised to the geometric mean of the reference genes 60S ribosomal protein L13-A (RPL13A, Quantitect Primer Assay Hs\_RPL13A Primer design reference gene assay (HK-SY-hu) ) and succinate dehydrogenase complex, subunit A, flavoprotein variant (SDHA), Quantitect Primer Assay Hs\_SDHA\_1\_SG (QT00059486). These reference genes have been shown to be stable in CD3+ T lymphocytes and Jurkat cells (Sibbons *et al.* 2018) by the GeNorm method (Vandesompele *et al.* 2002). The qRTPCR conditions were those specified by the manufacturer.

*Analysis of fatty acid composition by gas chromatography*

CD3+ T lymphocytes and Jurkat cells were thawed and suspended in 0.9% (wt/v) NaCl and the internal standard 17:0 (3 μg) was added. Cell lipids were extracted with chloroform/methanol (2:1, v/v) (Bligh & Dyer 1959), dried, dissolved in toluene and converted to fatty acid methyl esters (FAMEs) by incubation with acidified methanol containing 2% (v/v) sulphuric acid at 50°C for 2 hours (Burdge *et al.* 2000). The reaction mixture was cooled to room temperature and neutralised with KHCO3 (0.25 M) and K2CO3 (0.5 M). FAMEs were collected by hexane extraction (Burdge *et al.* 2000). FAMEs were separated on a BPX-70 fused silica capillary column (30 m × 0.25 mm × 25 μm) (Trajan, Scientific Europe, Milton Keynes, UK) using an Agilent 6890 gas chromatograph (GC) equipped with flame ionisation detection as described (West *et al.* 2016). Chromatograms were integrated manually by a single operator using OpenLAB CDS ChemStation software (version BC.0301.001) (Agilent Technologies, UK). The amounts of individual fatty acids are expressed as mass per million cells at the end of the culture period. Fatty acids were identified by their retention times relative to standards (37 FAMES, Sigma-Aldrich) and confirmed by GC–mass spectrometry (Figure 2) using a 6890 gas chromatograph (Agilent, UK) equipped with an Agilent 5975 mass selective detector set to a mass scan range of m/z 50–550 as described (von Gerichten *et al.* 2021).

*Statistics*

Statistical analyses were carried out using IBM SPSS Statistics for Windows, Version 27.0.

(Armonk, NY: IBM Corp). The data were assessed for normality using the Kolmogorov Smirnov test. Normally distributed data are shown as mean ± SEM, while data that did not follow a normal distribution are reported as median (range). Pairwise comparisons for normally distributed data were made using Student’s t test, or the Mann-Whitney U test for data that were not normally distributed. Statistical testing of the interaction between age and T cell activation status (activation state\*life stage) was by 2-way ANOVA with Tukey’s *post hoc* correction for multiple comparisons. A sample size of n = 6 cultures or participants was calculated to provide 87% power to detect a significant difference in the amount of 22:6ω-3 between treatments of 0.4 nmol/106 cells with α <0.05. Effect sizes are reported either as Cohen’s d, or partial eta squared.

**Results**

*The effect of incubation with 24:6ω-3 on the fatty acid composition of T lymphocytes*

There was a significant treatment x activation interaction effect on the amount of 24:0, but not on the amounts of any of the other SFAs or the MUFAs measured in T lymphocytes (Table 2).

Mitogen stimulation significantly increased the amounts of 18:2ω-6 (2.4-fold) and 20:4ω-6 (1.3-fold) and there was a significant treatment x activation interaction effect on the amount of 20:4ω-6, but the amounts of the other ω-6 PUFAs that were measured in T cells were not altered (Table 2). Specifically, the mitogen-induced increment in 20:4ω-6 was greater in T lymphocytes incubated with 24:6ω-3 than in untreated cells. The activation-induced change in the amount of 24:1ω-9 was blunted in T cells incubated with 24:6ω-3 compared to untreated T lymphocytes.

The identities of 24:6ω-3 and 22:6ω-3 were confirmed by comparison of the mass spectra of authentic standards with those of the peaks with the corresponding retention times in T lymphocytes (Figure 2). 24:6ω-3 was not detected in quiescent or activated CD3+ T cells that were maintained in medium lacking this fatty acid (Table 2, Figure 2), but was significantly incorporated into 24:6ω-3-treated T lymphocytes irrespective of activation status (Table 2). Incubation with 24:6ω-3 increased the amount of 22:6ω-3 in quiescent (5-fold) and activated (7-fold) T cells compared to untreated cells (Table 2). Mitogen stimulation increased the amount of 20:3ω-3 in T cells that were not incubated with 24:6ω-3. However, the magnitude of mitogen-induced change in 20:3ω-3 content was less in cells incubated with 24:6ω-3 than in untreated T cells. There was no single factor effect of 24:6ω-3 on the amounts of 20:5ω-3 or 22:5ω-3, but there was a significant treatment x activation effect on the amounts of these ω-3 PUFAs (Table 2).*The effect of incubation with 24:6ω-3 or 18:3ω-3 on the fatty acid composition of Jurkat cells*

Neither 24:6ω-3 nor 18:3ω-3 significantly altered the SFA or MUFA contents of Jurkat cells (Table 23). Incubation with 24:6ω-3 significantly decreased the amounts of 20:3ω-6 (30%) and 20:4ω-6 (51%), but there was no significant effect of incubation with 24:6ω-3 on the 18:2ω-6 and 20:2ω-6 contents of Jurkat cells (Table 3). In contrast, incubation with 24:6ω-3 increased the amounts of 20:5ω-3 (68%), 22:5ω-3 (19%), 24:6ω-3 (0 to 9 nmol / million cells) and 22:6ω-3 (6.9-fold) in Jurkat cells. Incubating Jurkat cells with 18:3ω-3 significantly increased the amounts of 18:3ω-3 (37-fold), 20:3ω-3 (16.6-fold), 20:5ω-3 (13-fold), and 22:5ω-3 (6.5-fold), while the increment in 22:6ω-3 (5%) did not reach statistical significance (Table 3). The increment in the amount of 22:6ω-3 induced in Jurkat cells by incubation with 24:6ω-3 was 19.5-fold greater than the increase due to incubation with 18:3ω-3. 20:4ω-3 was not detected in Jurkat cells after adjustment for the final concentrations of these substrates. *effect of acyl-CoA oxidase-1 siRNA knockdown on the fatty acid composition of Jurkat cells incubated with 24:6ω-3*

Incubation of Jurkat cells with ACOX1 siRNA reduced the median level of the ACOX1 mRNA transcript by 65% (p = 0.0006; Mann Whitney U-test) compared to cells incubated with the non-targeted control siRNA (Figure 3). ACOX1 siRNA knockdown in Jurkat cells incubated with 24:6ω-3 increased the amount of 20:0 by 1.5-fold compared to cells treated with NT siRNA, but there were no statistically significant differences in the amounts of any of the other SFAs measured (Table 4). Jurkat cells treated with ACOX1 siRNA plus 24:6ω-3 contained 68% more 18:1ω-7 than cells treated with NT siRNA, while there were no significant differences between treatments in the amounts of the other MUFAs measured (Table 4). Treatment of Jurkat cells with ACOX1 siRNA plus 24:6ω-3 significantly increased the amount of 20:2ω-6 (43%), 20:3ω-6 and 24:6ω-3 compared to cells treated with NT siRNA plus 24:6ω-3 (Table 4). Jurkat cells incubated with ACOX1 siRNA plus 24:6ω-3 contained 24% less 20:3ω-3, 23% less 22:5ω-3, and 36% less 22:6ω-3, but 2-fold more 20:5ω-3 than cells treated with NT siRNA plus 24:6ω-3.

*The effect of treatment with Etomoxir on the fatty acid composition of Jurkat cells incubated with 24:6ω-3*

There was no significant effect of Etomoxir treatment on the amounts of SFAs, while the amount of 18:1ω-7 was greater (1.4-fold) in Jurkat cells treated with Etomoxir plus 24:6ω-3 than Jurkat cells incubated with 24:6ω-3 alone (Table 34). Treatment of Jurkat cells with Etomoxir plus 24:6ω-3 reduced the amount of 20:4ω-6 by 6%, while there was no significant effect on the amounts of the other ω-6 PUFAs measured. Treatment of Jurkat cells with Etomoxir plus 24:6ω-3 reduced the amounts of 22:6ω-3, 24:6ω-3, and 22:5ω-3 by 46%, 38%, and 45%, respectively. There were no statistically significant effects of treatment with Etomoxir plus 24:6ω-3 on the amounts of the other ω-3 PUFAs measured (Table 34).

**Discussion**

Overall, these findings show that although 24:6ω-3 has been regarded as a poor substrate for phospholipid biosynthesis (Voss et al. 1991), both primary CD3+ T lymphocytes and Jurkat cells accumulated 24:6ω-3 when incubated with exogenous 24:6ω-3, which was greater in activated T cells than unstimulated cells, which is consistent with the general increase in the uptake of exogenous fatty acids by mitogen-stimulated T cells (Rode et al. 1982).

exogenous 24:6ω-3 can be incorporated into primary human T lymphocytes and Jurkat cells The cell type-related changes in fatty acid composition induced by treatment with 24:6 ω-3 and acyl-coA oxidase knockdown are which suggests conversion of 24:6ω-3 to 22:6ω-3 via a mechanism involving peroxisomal β-oxidation that is regulated independently from the upstream reactions of the PUFA synthesis pathway (figure 1). In addition, one possible explanation for increased amounts of saturated or monounsaturated fatty acids is that they represent homeoviscotic adaptations induced by the accumulation of 24:6 ω-3. Hower, because of such homeoviscotic adaptations and uptake of fatty acids from the culture medium, the changes in T cell fatty acid composition induced by incubation with 24:6ω-3 cannot be assumed to reflect metabolic interconversions alone. Nevertheless, the results of previous studies (Moore *et al.* 1995; Metherel & Bazinet 2019; Metherel *et al.* 2019) support the interpretation of the present findings as showing that both CD3+ T lymphocytes and Jurkat cells can utilise 24:6ω-3 as a substrate for 22:6ω-3 synthesis and that such interconversion can occur irrespective of the integrity of the PUFA synthesis pathway. One interpretation of the similarity between Jurkat cells and primary T lymphocytes in the amount of 22:6ω-3 following incubation with 24:6ω-3 is that the post-endoplasmic reticulum reactions of the PUFA synthesis pathway can be regulated independently from the preceding metabolic steps as suggested previously (Sprecher 1999; Burdge 2004).

Fibroblasts from patients with Zellweger’s disease who lack peroxisomes do not synthesise 22:6ω-3 which supports the conclusion that peroxisomal fatty acid β-oxidation is required for 22:6ω-3 synthesis (Moore *et al.* 1995). However, despite this metabolic block, accumulation of 24:6ω-3 has not been reported in tissues or blood from patients who lack peroxisomes (Martinez 1995) or from *Pex-2/Pex-5* dual knockout mice that do not synthesise peroxisomes or express enzymes involved in peroxisomal fatty acid β-peroxidation (Baes *et al.* 1997; Faust & Hatten 1997; Janssen *et al.* 2000). Moreover, *Pex*-2 / *Pex*-5 null mice did not differ in liver 22:6ω-3 content from peroxisome replete mice (Janssen *et al.* 2000). Therefore, the role of peroxisomal fatty acid β-oxidation in 22:6ω-3 biosynthesis remains a matter for debate (Infante & Huszagh 2001). Direct synthesis of 22:5ω-3 by Δ4 desaturation by the protein product of *FADS2* (Park *et al.* 2015) or by a carnitine plus α-tocopherol-dependent mitochondrial pathway (Infante & Huszagh 2000) have been suggested as alternative mechanisms. Jurkat cells were treated with *ACOX1* siRNA iIn order to investigate whether peroxisomal fatty acid β-oxidation was involved in, at least, some of the changes in fatty acid composition induced by incubation with 24:6ω-3, The present findings show that 64% reduction in *ACOX1* mRNA expression by transfection of Jurkat cells with *ACOX1* siRNA was accompanied by lower amounts of 22:6ω-3, 22:5ω-3, 20:5ω-3 and 20:3ω-3, and more 24:6ω-3 when cells were incubated with 24:6ω-3 alone. This finding suggests that peroxisomal fatty acid β-oxidation is involved in the synthesis of other ω-3 PUFAs as well as 22:6ω-3, at least in Jurkat leukaemia cells, although this interpretation requires more rigorous testing by more direct methods. To the best of our knowledge the mechanism by which 24:6ω-3 could be converted to 20:5ω-3 has not been described, although it is possible that this may occur via the recycling of carbon atoms from peroxisomal fatty acid β-oxidation and utilised in the conversion of 18:3ω-3 to 20:3ω-3, although such recycling of carbon atoms from ω-3 PUFAs has only been reported for labelled 18:3ω-3 which can be utilised in cholesterol synthesis in rodent brain (Cunnane *et al.* 1994) and whole body SFA and MUFA synthesis in humans (Burdge & Wootton 2003) and rhesus macaques (Sheaff Greiner *et al.* 1996). The present findings imply that the suggestion that conversion of 24:6ω-3 is a minor source of ω-3 PUFAs (Metherel & Bazinet 2019) may depend on cell type.

Treatment of Jurkat cells with Etomoxir differentially decreased the amounts of

24:6ω-3, 22:5ω-3, 22:6ω-3, 18:1ω-7 and 20:4ω-6 in the cells. One possible interpretation is that the synthesis of these unsaturated fatty acids involves mitochondrial β-oxidation, for example by carbon recycling at least in Jurkat cells as occurs in T lymphocytes incubated with [13C-18:3 ω-3](West *et al*.2022) In the absence of findings from experiments using a 24:6ω-3 tracer, it is not possible to draw robust conclusions about the mechanism of 24:6ω-3 metabolism in T lymphocytes or Jurkat cells, which is an important limitation of the present study.

24:6ω-3 typically accounts for less than 2% of the total fatty acids in marine fish (Tomita 2009), but up to 13% in some species of starfish (Suo *et al.* 2015) and jellyfish (Nichols *et al.* 2003). Jellyfish are consumed frequently in Asian Countries, but rarely in Europe and North America (Bonaccorsi *et al.* 2020). The finding that human T cells can assimilate and metabolise 24:6ω-3, possibly with greater efficiency than 18:3ω-3, suggests that consuming foods that contain 24:6ω-3 could be an alternative to oily fish for achieving the benefits associated with 20:5ω-3 and 22:6ω-3 (Calder 1997, 1998).

**Declarations**

GB has received research funding from Nestle, Abbott Nutrition, and Danone and has served as a member of the Scientific Advisory Board of BASF. PC acts as a consultant to BASF, Smartfish, DSM, Cargill, Danone/Nutricia, and Fresenius-Kabi. KL has received research funding from Nestle, Abbott Nutrition, and Danone.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Funding**

This work was supported by grants from the Biotechnology and Biological Sciences Research Council (BB/S00548X/1 and BB/S005358/1). PC and GB are supported by the National Institute for Health and Care Research through the NIHR Southampton Biomedical Research Centre. Neither funder was involved in the study design, collection, analysis, interpretation of data, the writing of this article, or the decision to submit the manuscript for publication. Publication costs were paid by the University of Southampton.

**Author contributions**

GB, BF, PC, EM, and KL conceived and designed the study. JvG, AW, and NI conducted the experiments and analysed the data with GB. GB wrote the first draft of the manuscript. All authors contributed to drafting the manuscript and approved the submitted version.

**Figure 1.** The ω-3 polyunsaturated fatty acid biosynthesis pathway described in rat liver (Sprecher 1999) plus the modifications found in human primary T lymphocytes (Robichaud *et al.* 2018; von Gerichten *et al.* 2021). [1] The first reaction in T lymphocytes is carbon chain elongation putatively by elongase-5 (Sibbons *et al.* 2018; von Gerichten *et al.* 2021) (Robichaud *et al.* 2018). [2] The first reaction in the hepatic pathway is Δ6 desaturation by the protein product of the *FADS2* gene followed by chain elongation by elongase-5. [3] The protein product of the *FADS2* gene has Δ6 and Δ8 activities (Park *et al.* 2009), which are both expressed in Jurkat cells, while the Δ8 desaturase activity is predominant in T lymphocytes (Sibbons *et al.* 2018). [5] Elongase-2 is not expressed in T lymphocytes (Sibbons *et al.* 2018) (Robichaud *et al.* 2018; von Gerichten *et al.* 2021), therefore, truncating the pathway after synthesis of 22:5ω-3. However, elongase-2 is expressed in Jurkat cells (Sibbons *et al.* 2018). [6] The findings of (Voss *et al.* 1991; Moore *et al.* 1995) summarised by (Sprecher 2000) suggest that the conversion of 24:6ω-3 formed in the endoplasmic reticulum to 22:6ω-3 involves translocation to peroxisomes and carbon chain shortening by one cycle of β-oxidation.



**Figure 2.** Confirmation of 24:6ω-3 and 22:6ω-3 peak identities by gas chromatography (GC)-mass spectrometry (MS). (A) Separation of 37 FAMEs standard mixture by GC-MS, indicating the retention time of 22:6ω-3 methyl ester peak (\*1). (B) The retention time of 24:6ω-3 methyl ester (#2) standard detected by GC-MS. (C) Mass spectrum of the 22:6ω-3 methyl ester standard peak (\*1), (D) mass spectrum of the 24:6ω-3 (THA) methyl ester standard peak (#1), (E) mass spectrum of the 22:6ω-3 methyl ester peak (\*2) from CD3+ T lymphocytes. (F) Mass spectrum of the putative 24:6ω-3 methyl ester peak (#2) from CD3+ T lymphocytes.



Figure 3. The effect of treatment with *ACOX1* siRNA on *ACOX1* mRNA expression in Jurkat cells. Values are median (95% confidence interval, n = 6 culture replicates / treatment) acyl-coA oxidase-1 (ACOX) mRNA levels from individual Jurkat cell cultures treated for 48 hours with either ACOX siRNA or non-targeted siRNA (NT). Statistical comparison was by the Mann-Whitney U-test.

Table 1. Fatty acid compositions of cell culture media.

|  |  |  |  |
| --- | --- | --- | --- |
|  | Proportions of fatty acids in culture media (moles %) | | |
| Fatty acid | Culture medium | Culture medium plus 24:6ω-3 | Culture medium plus 18:3ω-3 |
| 14:0 | 1.3 | 0.9 | 1.2 |
| 16:0 | 27.6 | 25.4 | 25.8 |
| 16:1ω-7 | 2.2 | 1.9 | 2.0 |
| 18:0 | 8.8 | 8.7 | 8.6 |
| 18:1 ω-9 | 19.9 | 19.1 | 18.7 |
| 18:1 ω-7 | 1.4 | 1.5 | 1.4 |
| 18:2 ω-6 | 30.3 | 28.0 | 28.2 |
| 18:3ω-6 | 0.4 | 0.3 | 0.4 |
| 18:3ω-3 | 0.5 | 0.4 | 6.6 |
| 20:0 | <0.1 | <0.1 | 0.1 |
| 20:1ω-9 | 0.2 | 0.1 | 0.1 |
| 20:2ω-6 | 0.2 | 0.2 | 0.2 |
| 20:3ω-6 | 1.3 | 1.1 | 1.2 |
| 20:4ω-6 | 4.8 | 4.3 | 4.6 |
| 20:5ω-3 | 0.2 | 0.2 | 0.3 |
| 22:5ω-3 | 0.4 | 0.2 | 0.2 |
| 22:6ω-3 | 0.5 | 0.4 | 0.5 |
| 24:6ω-3 | ND | 7.1 | ND |

The total fatty acid composition of culture media was determined by gas chromatography as described in the Material and Methods section. 24:6ω-3 was not detected (ND) in media that were not supplemented with this fatty acid. Culture medium: RPMI1640 medium containing 10% (v/v) heat-inactivated homologous pooled serum.

Table 2 The effect of incubation with 24:6ω-3 on activation-induced changes in the fatty acid composition of CD3+ T lymphocytes

|  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | Amount of fatty acid (nmol/106 cells) | | | | ANOVA | | | | | |
|  | Without 24:6ω-3 | | With 24:6ω-3 | | Tr | ή2p | Ast | p | Tr x Ast | ή2p |
| Fatty acid | Unstimulated | Stimulated | Unstimulated | Stimulated | p |  | p |  | P |  |
| Saturated fatty acids | | | | | | | | | | |
| 14:0 | 0.32 ± 0.049a | 0.39 ± 0.025 | 0.31 ± 0.028 | 0.44 ± 0.051 | 0.82 | n.d. | 0.01 | n.d. | 0.7 | n.d. |
| 16:0 | 0.97 ± 0.092 | 1.72 ± 0.183 | 0.76 ± 0.029 | 2.28 ± 0.139 | 0.14 | n.d. | 1.2e-9 | 0.44 | 0.3 | n.d. |
| 18:0 | 0.73 ± 0.072 | 0.97 ± 0.077 | 0.54 ± 0.028 | 1.10 ± 0.062 | 0.28 | n.d. | 1.2e-14 | 0.85 | 0.06 | n.d. |
| 20:0 | 0.01 ± 0.001 | 0.01 ± 0.000 | 0.00 ± 0.000 | 0.01 ± 0.001 | 0.55 | n.d. | 0.74 | n.d. | 0.4 | n.d. |
| 24:0 | 0.03 ± 0.003a | 0.08 ± 0.008b | 0.02 ± 0.001a | 0.04 ± 0.004a | 0.06 | n.d. | 0.07 | n.d. | <0.001 | 0.39 |
| Monounsaturated fatty acids | | | | | | | | | | |
| 16:1ω-7 | 0.03 ± 0.004 | 0.09 ± 0.007 | 0.02 ± 0.002 | 0.11 ± 0.010 | 0.43 | n.d. | 7.5e-14 | 0.94 | 0.12 | n.d. |
| 18:1ω-9 | 0.42 ± 0.040 | 1.06 ± 0.095 | 0.30 ± 0.013 | 1.23 ± 0.092 | 0.06 | n.d. | 0.02 | 0.23 | 0.36 | n.d. |
| 18:1ω-7 | 0.08 ± 0.008a | 0.14 ± 0.014 | 0.05 ± 0.003a | 0.14 ± 0.008 | 0.07 | n.d. | 9.2e-17 | 0.97 | 0.3 | n.d. |
| 20:1ω-9 | 0.02 ± 0.003a | 0.04 ± 0.006 | 0.02 ± 0.003a | 0.04 ± 0.002 | 0.12 | n.d. | 4.3e-5 | 0.57 | 0.53 | n.d. |
| 24:1ω-9 | 0.03 ± 0.004b | 0.02 ± 0.003 | 0.02 ± 0.001 | 0.02 ± 0.001 | 1.3e-7 | 0.76 | 0.6e-5 | 0.65 | 0.87 | n.d. |
| ω-6 Polyunsaturated fatty acids | | | | | | | | | | |
| 18:2ω-6 | 0.56 ± 0.056a | 1.32 ± 0.108a | 0.42 ± 0.018b | 1.52 ± 0.110b | 0.0003 | 0.49 | 4.6e-7 | 0.73 | 1e-5 | 0.53 |
| 20:2ω-6 | 0.01 ± 0.002 | 0.06 ± 0.010 | 0.01 ± 0.004 | 0.06 ± 0.005 | 0.49 | n.d. | 0.69 | n.d | 0.22 | n.d |
| 20:3ω-6 | 0.05 ± 0.004 | 0.15 ± 0.020 | 0.04 ± 0.002 | 0.12 ± 0.012 | 0.86 | n.d | 1.1e-5 | 0.54 | 0.12 | n.d |
| 20:4ω-6 | 0.44 ± 0.039a | 0.57 ± 0.044a | 0.32 ± 0.020 | 0.54 ± 0.028 | 0.001 | 0.42 | 4.3e-5 | 0.78 | 0.006 | 0.32 |
| ω-3 Polyunsaturated fatty acids | | | | | | | | | | |
| 18:3ω-3 | 0.01± 0.002 | 0.01 ± 0.001 | 0.01 ± 0.004 | 0.02 ± 0.001 | 0.19 | .n.d. | 0.60 | n.d. | 0.40 | n.d. |
| 20:3ω-3 | 0.03 ± 0.003 | 0.10 ± 0.036 | 0.02 ± 0.002 | 0.03 ± 0.010 | 0.001 | 0.43 | 9.3e-8 | 0.47 | 0.33 | n.d. |
| 20:5ω-3 | 0.01 ± 0.002 | 0.01 ± 0.001 | 0.01 ± 0.00a | 0.02 ± 0.00b | 0.85 | n.d. | 0.01 | 0.27 | 0.28 | n.d. |
| 22:5ω-3 | 0.02 ± 0.002a | 0.04 ± 0.004b | 0.01 ± 0.001a | 0.04 ± 0.003b | 0.48 | n.d. | 5.7e-8 | 0.78 | 0.06 | n.d. |
| 22:6ω-3 | 0.02 ± 0.003a | 0.04 ± 0.006b | 0.05 ± 0.004b | 0.28 ± 0.038c | 7.5e-7 | 0.55 | 0.01 | 0.31 | 0.04 | 0.19 |
| 24:6ω-3 | 0.00 ± 0.000 | 0.00 ± 0.000a | 0.00 ± 0.00a | 0.13 ± 0.019b | 3.3e-12 | 0.92 | 0.02 | 0.53 | 0.19 | n.d. |

Values are mean ± SEM amounts of T lymphocyte fatty acids from 6 different participants per treatment. Statistical comparisons were done by 2 way ANOVA with treatment (Tr) (incubation with or without 24:6ω-3) and activation status (Ast;) as fixed factors. Testing *post hoc* was performed by Tukey’s method. p values are reported for single factor and Tr x Ast interaction effects. Effect sizes of means that differed significantly (p < 0.05) are reported asp artial Eta squared ήp but were not determined (n.d) for comparisons that failed to reach statistical Means that do not share superscripted letters differ significantly

Table 3 The effect of incubation with 24:6ω-3 or 18:3ω-3 on the fatty acid composition of Jurkat cells

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | Amount of fatty acid (nmol/106 cells) | | t test | | Amount of fatty acid (nmol/106 cells | | t test | |
| Fatty acid | Without 24:6ω-3 | With 24:6ω-3 | p | Cohen’s d | Without 18:3ω-3 | With 18:3ω3 | p | Cohen’s d |
| Saturated fatty acids | | | | | | | | |
| 14:0 | 2.15 ± 0.14 | 1.98 ± 0.10 | 0.93 | n.d. | 0.29 ± 0.04 | 0.28 ± 0.04 | 0.7 | n.d. |
| 16:0 | 22.76 ± 0.13 | 21.56 ± 0.33 | 0.62 | n.d. | 2.58 ± 0.33 | 2.67 ± 0.60 | 0.38 | n.d. |
| 18:0 | 13.30 ± 0.09 | 11.81 ± 0.20 | 0.73 | n.d. | 1.24 ± 0.15 | 1.34 ± 0.33 | 0.80 | n.d. |
| 20:0 | 0.19 ± 0.01 | 0.18 ± 0.01 | 0.46 | n.d. | 0.01 ± 0.00 | 0.01 ± 0.00 | 0.31 | n.d. |
| 24:0 | 5.10 ± 0.07 | 3.11 ± 0.18 | 0.13 | n.d. | 0.31 ± 0.03 | 0.33 ± 0.08 | 0.18 | n.d. |
| Monounsaturated fatty acids | | | | | | | | |
| 16:1ω--7 | 1.16 ± 0.01 | 1.13 ± 0.02 | 0.62 | n.d. | 0.15 ± 0.02 | 0.13 ± 0.03 | 0.72 | n.d. |
| 18:1ω-9 | 20.81 ± 0.05 | 17.53 ± 0.08 | 0.76 | n.d. | 1.72 ± 0.21 | 2.00 ± 0.50 | 0.62 | n.d. |
| 18:1ω-7 | 3.93 ± 0.02 | 2.90 ± 0.03 | 0.99 | n.d. | 0.29 ± 0.04 | 0.39 ± 0.10 | 0.38 | n.d. |
| 20:1ω-9 | 1.90 ± 0.03 | 1.88 ± 0.02 | 0.99 | n.d. | 0.16 ± 0.02 | 0.21 ± 0.05 | 0.39 | n.d. |
| 24:1ω-9 | 0.28 ± 0.01 | 0.32 ± 0.01 | 0.82 | n.d. | 0.15 ± 0.02 | 0.13 ± 0.01 | 0.10 | n.d. |
| ω-6 Polyunsaturated fatty acids | | | | | | | |  |
| 18:2ω-6 | 7.98 ± 0.06 | 9.28 ± 0.23 | 0.07 | n.d. | 1.04 ± 0.12 | 0.85 ± 0.21 | 0.45 | n.d. |
| 20:2ω-6 | 1.46 ± 0.02 | 1.52 ± 0.02 | 0.036 | 0.19 | 0.16 ± 0.02 | 0.16 ± 0.04 | 0.95 | n.d. |
| 20:3ω-6 | 6.82 ± 0.04 | 4.74 ± 0.19 | 0.08 | n.d. | 0.50 ± 0.06 | 0.57 ± 0.14 | 0.64 | n.d |
| 20:4ω-6 | 9.36 ± 0.08 | 5.95 ± 0.29 | 0.001 | 1.65 | 0.41 ± 0.05 | 0.62 ± 0.1 | 0.22 | n.d |
| ω-3 Polyunsaturated fatty acids | | | | | | | |  |
| 18:3ω-3 | 0.05 ± 0.01 | 0.17 ± 0.01 | 0.72 | n.d. | 0.01 ± 0.01 | 0.37 ± 0.04 | 1.0e-6 | 4.59 |
| 20-3ω-3 | 0.60 ± 0.04 | 0.49 ± 0.15 | 0.07 | 1.16 | 0.03 ± 0.01 | 0.21 ± 0.03 | 0.005 | 3.77 |
| 20:5ω-3 | 0.16 ± 0.01 | 0.27 ± 0.01 | 2e-5 | 0.56 | 0.01 ± 0.00 | 0.13 ± 0.01 | 5.1e-8 | 4.58 |
| 22:5ω-3 | 1.03 ± 0.01 | 1.23 ± 0.04 | 1.6e-5 | 0.04 | 0.06 ± 0.01 | 0.39 ± 0.05 | 3.3e-9 | 4.07 |
| 22:6ω-3 | 0.77 ± 0.01 | 5.32 ± 0.20 | 2.2e-9 | 0.15 | 0.57 ± 0.00 | 0.60 ± 0.00 | 0.53 | n.d. |
| 24:6ω-3 | 0.00 ± 0.01 | 8.98 ± 0.21 | 3e-5 | 0.23 | 0.00 ± 0.01 | 0.00 ± 0.01 | 0.55 | n.d. |

Values are mean ± SEM amounts of fatty acids (n = 6 culture replicates per treatment). Statistical comparisons were done by Student’s unpaired t test (equal variances were not assumed). Effect sizes of means that differed significantly (p < 0.05) are reported as Cohen’s d, but were not determined (n.d) for comparisons that failed to meet the threshold for statistical significance.

Table 4 The effect of acyl-CoA oxidase-1 SiRNA knockdown and Etomoxir treatments on the fatty acid composition of Jurkat cells incubated with 24:6ω-3

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | Amount of fatty acid (nmol /106 cells) | | t test | | Amount of fatty acid (nmol /106 cells) | | t test | |
| Fatty acid | NT siRNA + 24:6ω-3 | *ACOX1* siRNA + 24:6ω-3 | p | Cohen’s d | Etomoxir control + 24:6ω-3 | Etomoxir + 24:6ω-3 | p | Cohen’s d |
| Saturated fatty acids | | | | | | | | |
| 14:0 | 0.45 ± 0.06 | 0.41 ± 0.11 | 0.93 | n.d. | 1.01 ± 0.12 | 0.25 ± 0.05 | 0.03 | n.d. |
| 16:0 | 17.36 ± 2.87 | 20.82 ± 0.41 | 0.62 | n.d. | 21.60 ± 0.20 | 7.51 ± 0.35 | 0.19 | n.d. |
| 18:0 | 0.66 ± 0.10 | 0.76 ± 0.11 | 0.73 | n.d. | 13.75 ± 0.25 | 4.62 ± 0.15 | 0.06 | n.d. |
| 20:0 | 13.42 ± 2.58 | 20.35 ± 1.10 | 0.046 | 1.8 | 1.85 ± 0.03 | 0.08 ± 0.00 | 0.07 | n.d. |
| Monounsaturated fatty acids | | | | | | | | |
| 16:1ω-7 | 11.62 ± 1.78 | 19.51 ± 0.26 | 0.09 | n.d. | 0.98 ± 0.04 | 0.76 ± 0.11 | 0.44 | n.d. |
| 18:1ω-9 | 2.67 ± 0.43 | 4.85 ± 0.08 | 0.76 | n.d. | 16.93 ± 0.24 | 19.51 ± 0.26 | 0.71 | n.d. |
| 18:1ω-7 | 4.82 ± 0.68 | 8.09 ± 0.27 | 0.019 | 0.17 | 3.44 ± 0.02 | 4.85 ± 0.08 | 0.01 | 0.27 |
| 20:1ω-9 | 0.32 ± 0.08 | 0.31 ± 0.08 | 0.99 | n.d. | 1.59 ± 0.03 | 1.74 ± 0.07 | 0.44 | n.d |
| 24:1ω-9 | 0.18 ± 0.03 | 0.40 ± 0.02 | 0.34 | n.d. | 1.33 ± 0.02 | 0.55 ± 0.02 | 0.04 | 9.21 |
| ω-6 Polyunsaturated fatty acids | | | | | | | | |
| 18:2ω-6 | 1.15 ± 0.14 | 1.74 ± 0.07 | 0.08 | n.d. | 8.02 ± 0.18 | 8.09 ± 0.27 | 0.82 | n.d. |
| 20:2ω-6 | 0.91 ± 0.11 | 1.30 ± 0.05 | 0.04 | 0.39 | 0.17 ± 0.01 | 1.30 ± 0.05 | 0.85 | n.d. |
| 20:3ω-6 | 2.35 ± 0.33 | 2.79 ± 0.09 | 0.01 | 0.89 | 3.97 ± 0.02 | 2.79 ± 0.09 | 0.23 | n.d. |
| 20:4ω-6 | 3.00 ± 0.46 | 4.64 ± 0.10 | 0.001 | 0.65 | 4.95 ± 0.03 | 4.64 ± 0.10 | 0.01 | 4.42 |
| ω-3 Polyunsaturated fatty acids | | | | | | | | |
| 18:3ω-3 | 0.14 ± 0.02 | 0.33 ± 0.01 | 0.54 | n.d. | 0.22 ± 0.01 | 0.41 ± 0.08 | 0.16 | 4.6 |
| 20:3ω-3 | 0.33 ± 0.05 | 0.25 ± 0.02 | 0.07 | n.d. | 0.46 ± 0.01 | 0.33 ± 0.01 | 1.94 | n.d. |
| 20:5ω-3 | 0.28 ± 0.04 | 0.55 ± 0.02 | 0.14 | n.d. | 2.05 ± 0.03 | 2.13 ± 0.07 | 0.29 | n.d. |
| 22:5ω-3 | 0.95 ± 0.14 | 0.73 ± 0.02 | 0.56 | n.d. | 1.33 ± 0.02 | 0.73 ± 0.02 | 0.04 | 0.34 |
| 22:6ω-3 | 4.91 ± 0.42 | 3.14 ± 0.17 | 0.01 | 0.23 | 10.22 ± 0.08 | 5.49 ± 0.17 | 0.01 | 0.76 |
| 24:6ω-3 | 2.69 ± 0.25 | 3.27 ± 0.13 | 0.002 | 0.15 | 6.85 ± 0.34 | 4.27 ± 0.13 | 0.02 | 0.211 |
|  |  |  |  |  |  |  |  |  |

Values are mean ± SEM (n = 6 culture replicates per treatment). All cultures contained 24:6ω-3 (25 µM). Statistical comparisons were done by Student’s unpaired t test (equal variances were not assumed). Effect sizes of means that differed significantly (p < 0.05) are reported as Cohen’s d, but were not determined (n.d) for comparisons which failed to meet the threshold for statistical significance. ACOX1, acyl-CoA oxidase; NT non-targeted siRNA control.

`References

Abraham, R.T., Weiss, A., 2004. Jurkat T cells and development of the T-cell receptor signalling paradigm. Nat Rev Immunol 4, 301-8

Anel, A., Naval, J., Gonzalez, B., Torres, J.M., Mishal, Z., Uriel, J., Pineiro, A., 1990. Fatty acid metabolism in human lymphocytes. I. Time-course changes in fatty acid composition and membrane fluidity during blastic transformation of peripheral blood lymphocytes. Biochim.Biophys Acta 1044, 323-331

Baes, M., Gressens, P., Baumgart, E., Carmeliet, P., Casteels, M., Fransen, M., Evrard, P., Fahimi, D., Declercq, P.E., Collen, D., van Veldhoven, P.P., Mannaerts, G.P., 1997. A mouse model for Zellweger syndrome. Nat Genet 17, 49-57

Bligh, E.G., Dyer, W.J., 1959. A rapid method of total lipid extraction and purification. Can J Biochem Physiol 37, 911-7

Bonaccorsi, G., Garamella, G., Cavallo, G., Lorini, C., 2020. A Systematic Review of Risk Assessment Associated with Jellyfish Consumption as a Potential Novel Food. Foods 9

Burdge, G., 2004. Alpha-linolenic acid metabolism in men and women: nutritional and biological implications. Curr.Opin.Clin.Nutr Metab Care 7, 137-144

Burdge, G.C., 2022. alpha-linolenic acid interconversion is sufficient as a source of longer chain omega-3 polyunsaturated fatty acids in humans: An opinion. Lipids

Burdge, G.C., Jones, A.E., Wootton, S.A., 2002. Eicosapentaenoic and docosapentaenoic acids are the principal products of alpha-linolenic acid metabolism in young men\*. Br J Nutr 88, 355-63

Burdge, G.C., Wootton, S.A., 2002. Conversion of alpha-linolenic acid to eicosapentaenoic, docosapentaenoic and docosahexaenoic acids in young women. Br.J Nutr 88, 411-420

Burdge, G.C., Wootton, S.A., 2003. Conversion of alpha-linolenic acid to palmitic, palmitoleic, stearic and oleic acids in men and women. Prostaglandins Leukot.Essent.Fatty Acids 69, 283-290

Burdge, G.C., Wright, P., Jones, A.E., Wootton, S.A., 2000. A method for separation of phosphatidylcholine, triacylglycerol, non-esterified fatty acids and cholesterol esters from plasma by solid-phase extraction. Br.J Nutr 84, 781-787

Calder, P.C., 1993. The effects of fatty acids on lymphocyte functions. Braz.J Med.Biol.Res. 26, 901-917

Calder, P.C., 1995. Fatty acids, dietary lipids and lymphocyte functions. Biochem.Soc.Trans. 23, 302-309

Calder, P.C., 1997. N-3 polyunsaturated fatty acids and immune cell function. Adv.Enzyme Regul. 37, 197-237

Calder, P.C., 1998. Dietary fatty acids and lymphocyte functions. Proc.Nutr Soc. 57, 487-502

Calder, P.C., 2015. Marine omega-3 fatty acids and inflammatory processes: Effects, mechanisms and clinical relevance. Biochim Biophys Acta 1851, 469-484

Calder, P.C., Yaqoob, P., Harvey, D.J., Watts, A., Newsholme, E.A., 1994. Incorporation of fatty acids by concanavalin A-stimulated lymphocytes and the effect on fatty acid composition and membrane fluidity. Biochem J 300 ( Pt 2), 509-18

Cikos, S., Bukovska, A., Koppel, J., 2007. Relative quantification of mRNA: comparison of methods currently used for real-time PCR data analysis. BMC.Mol.Biol. 8, 113

Cunnane, S.C., Williams, S.C., Bell, J.D., Brookes, S., Craig, K., Iles, R.A., Crawford, M.A., 1994. Utilization of uniformly labeled 13C-polyunsaturated fatty acids in the synthesis of long-chain fatty acids and cholesterol accumulating in the neonatal rat brain. J Neurochem 62, 2429-36

Faust, P.L., Hatten, M.E., 1997. Targeted deletion of the PEX2 peroxisome assembly gene in mice provides a model for Zellweger syndrome, a human neuronal migration disorder. J Cell Biol 139, 1293-305

Ferber, E., De Pasquale, G.G., Resch, K., 1975. Phospholipid metabolism of stimulated lymphocytes. Composition of phospholipid fatty acids. Biochim Biophys Acta 398, 364-76

Gibson, R.A., Neumann, M.A., Lien, E.L., Boyd, K.A., Tu, W.C., 2013. Docosahexaenoic acid synthesis from alpha-linolenic acid is inhibited by diets high in polyunsaturated fatty acids. Prostaglandins Leukot Essent Fatty Acids 88, 139-46

Gotoh, N., Nagao, K., Ishida, H., Nakamitsu, K., Yoshinaga, K., Nagai, T., Beppu, F., Yoshinaga-Kiriake, A., Watanabe, H., Yanagita, T., 2018. Metabolism of Natural Highly Unsaturated Fatty Acid, Tetracosahexaenoic Acid (24:6n-3), in C57BL/KsJ-db/db Mice. J Oleo Sci 67, 1597-1607

Grammatikos, S.I., Subbaiah, P.V., Victor, T.A., Miller, W.M., 1994. n-3 and n-6 fatty acid processing and growth effects in neoplastic and non-cancerous human mammary epithelial cell lines. Br J Cancer 70, 219-27

Infante, J.P., Huszagh, V.A., 2000. Secondary carnitine deficiency and impaired docosahexaenoic (22:6n-3) acid synthesis: a common denominator in the pathophysiology of diseases of oxidative phosphorylation and beta-oxidation. FEBS Lett 468, 1-5

Infante, J.P., Huszagh, V.A., 2001. Zellweger syndrome knockout mouse models challenge putative peroxisomal beta-oxidation involvement in docosahexaenoic acid (22:6n-3) biosynthesis. Mol Genet Metab 72, 1-7

Ishihara, K., Murata, M., Kaneniwa, M., Saito, H., Shinohara, K., Maeda-Yamamoto, M., Kawasaki, K., Ooizumi, T., 1998. Effect of tetracosahexaenoic acid on the content and release of histamine, and eicosanoid production in MC/9 mouse mast cell. Lipids 33, 1107-14

Janssen, A., Baes, M., Gressens, P., Mannaerts, G.P., Declercq, P., Van Veldhoven, P.P., 2000. Docosahexaenoic acid deficit is not a major pathogenic factor in peroxisome-deficient mice. Lab Invest 80, 31-5

Lonnberg, T., Yetukuri, L., Seppanen-Laakso, T., Lahesmaa, R., Oresic, M., 2013. T-cell activation induces selective changes of cellular lipidome. Front Biosci (Elite Ed) 5, 558-73

Martinez, M., 1995. Polyunsaturated fatty acids in the developing human brain, erythrocytes and plasma in peroxisomal disease: therapeutic implications. J Inherit Metab Dis 18 Suppl 1, 61-75

Metherel, A.H., Bazinet, R.P., 2019. Updates to the n-3 polyunsaturated fatty acid biosynthesis pathway: DHA synthesis rates, tetracosahexaenoic acid and (minimal) retroconversion. Prog Lipid Res 76, 101008

Metherel, A.H., Lacombe, R.J.S., Chouinard-Watkins, R., Bazinet, R.P., 2019. Docosahexaenoic acid is both a product of and a precursor to tetracosahexaenoic acid in the rat. J Lipid Res 60, 412-420

Moore, S.A., Hurt, E., Yoder, E., Sprecher, H., Spector, A.A., 1995. Docosahexaenoic acid synthesis in human skin fibroblasts involves peroxisomal retroconversion of tetracosahexaenoic acid. J Lipid Res 36, 2433-43

Nichols, P.D., Danaher, K.T., Koslow, J.A., 2003. Occurrence of high levels of tetracosahexaenoic acid in the jellyfish Aurelia sp. Lipids 38, 1207-10

Noakes, P.S., Vlachava, M., Kremmyda, L.S., Diaper, N.D., Miles, E.A., Erlewyn-Lajeunesse, M., Williams, A.P., Godfrey, K.M., Calder, P.C., 2012. Increased intake of oily fish in pregnancy: effects on neonatal immune responses and on clinical outcomes in infants at 6 mo. Am J Clin Nutr 95, 395-404

Park, H.G., Park, W.J., Kothapalli, K.S., Brenna, J.T., 2015. The fatty acid desaturase 2 (FADS2) gene product catalyzes Delta4 desaturation to yield n-3 docosahexaenoic acid and n-6 docosapentaenoic acid in human cells. FASEB J 29, 3911-9

Park, W.J., Kothapalli, K.S., Lawrence, P., Tyburczy, C., Brenna, J.T., 2009. An alternate pathway to long-chain polyunsaturates: the FADS2 gene product Delta8-desaturates 20:2n-6 and 20:3n-3. J Lipid Res 50, 1195-202

Pawlosky, R., Hibbeln, J., Lin, Y., Salem, N., Jr., 2003. n-3 fatty acid metabolism in women. Br.J.Nutr. 90, 993-994

Prescott, S.L., Macaubas, C., Smallacombe, T., Holt, B.J., Sly, P.D., Holt, P.G., 1999. Development of allergen-specific T-cell memory in atopic and normal children. Lancet 353, 196-200

Robichaud, P.P., Munganyiki, J.E., Boilard, E., Surette, M.E., 2018. Polyunsaturated fatty acid elongation and desaturation in activated human T-cells: ELOVL5 is the key elongase. J Lipid Res 59, 2383-2396

Rode, H.N., Szamel, M., Schneider, S., Resch, K., 1982. Phospholipid metabolism of stimulated lymphocytes. Preferential incorporation of polyunsaturated fatty acids into plasma membrane phospholipid upon stimulation with concanavalin A. Biochim Biophys Acta 688, 66-74

Sheaff Greiner, R.C., Zhang, Q., Goodman, K.J., Giussani, D.A., Nathanielsz, P.W., Brenna, J.T., 1996. Linoleate, alpha-linolenate, and docosahexaenoate recycling into saturated and monounsaturated fatty acids is a major pathway in pregnant or lactating adults and fetal or infant rhesus monkeys. J Lipid Res 37, 2675-86

Shires, S.E., Kelleher, J., Trejdosiewicz, L.K., 1989. Effects of linoleic acid and mitogenic stimulation on the fatty acid composition of human lymphocytes. Biochim Biophys Acta 1002, 74-8

Sibbons, C.M., Brenna, J.T., Lawrence, P., Hoile, S.P., Clarke-Harris, R., Lillycrop, K.A., Burdge, G.C., 2014. Effect of sex hormones on n-3 polyunsaturated fatty acid biosynthesis in HepG2 cells and in human primary hepatocytes. Prostaglandins Leukot Essent Fatty Acids 90, 47-54

Sibbons, C.M., Irvine, N.A., Pérez-Mojica, J.E., Calder, P.C., Lillycrop, K.A., Fielding, B.A., Burdge, G.C., 2018. Polyunsaturated Fatty Acid Biosynthesis Involving Δ8 Desaturation and Differential DNA Methylation of FADS2 Regulates Proliferation of Human Peripheral Blood Mononuclear Cells. Frontiers in Immunology 9

Sprecher, H., 1999. An update on the pathways of polyunsaturated fatty acid metabolism. Curr.Opin.Clin.Nutr.Metab Care 2, 135-138

Sprecher, H., 2000. Metabolism of highly unsaturated n-3 and n-6 fatty acids. Biochim Biophys Acta 1486, 219-31

Suo, R., Li, H., Yoshinaga, K., Nagai, T., Mizobe, H., Kojima, K., Nagao, K., Beppu, F., Gotoh, N., 2015. Generation of Tetracosahexaenoic Acid in Benthic Marine Organisms. J Oleo Sci 64, 721-7

Tomita, Y., Ando, Y., 2009. Reinvestigation of positional distribution of tetracosahexaenoic acid in triacyl-sn-glycerols of flathead flounder flesh. FIsheries Science 75, 75:445–451

Vandesompele, J., De Preter, K., Pattyn, F., Poppe, B., Van Roy, N., De Paepe, A., Speleman, F., 2002. Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. Genome Biol 3, RESEARCH0034

von Gerichten, J., West, A.L., Irvine, N.A., Miles, E.A., Calder, P.C., Lillycrop, K.A., Fielding, B.A., Burdge, G.C., 2021. The Partitioning of Newly Assimilated Linoleic and alpha-Linolenic Acids Between Synthesis of Longer-Chain Polyunsaturated Fatty Acids and Hydroxyoctadecaenoic Acids Is a Putative Branch Point in T-Cell Essential Fatty Acid Metabolism. Front Immunol 12, 740749

Voss, A., Reinhart, M., Sankarappa, S., Sprecher, H., 1991. The metabolism of 7,10,13,16,19-docosapentaenoic acid to 4,7,10,13,16,19-docosahexaenoic acid in rat liver is independent of a 4-desaturase. J Biol Chem. 266, 19995-20000

West, A.L., Burdge, G.C., Calder, P.C., 2016. Lipid structure does not modify incorporation of EPA and DHA into blood lipids in healthy adults: a randomised-controlled trial. Br J Nutr 116, 788-97

[1] A.L. West, J. von Gerichten, N.A. Irvine, E.A. Miles, K.A. Lillycrop, P.C. Calder, B.A. Fielding, G.C. Burdge, 2022 Fatty acid composition and metabolic partitioning of alpha-linolenic acid are contingent on life stage in human CD3(+) T lymphocytes. Front Immunol 13 1079642.