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Convergently evolved placental villi show multiscale structural adaptations to differential placental invasiveness

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Despite having a single evolutionary origin and conserved function, the mammalian placenta exhibits radical structural diversity. The evolutionary drivers and functional consequences of placental structural diversity are poorly understood. Humans and equids both display treelike placental villi, however these villi evolved independently and exhibit starkly different levels of invasiveness into maternal tissue (i.e. the number of maternal tissue layers between placental tissue and maternal blood). The villi in these species therefore serve as a compelling evolutionary case study to explore whether placentas have developed structural adaptations to respond to the challenge of reduced nutrient availability in less invasive placentas. Here, we use three-dimensional X-ray microfocus computed tomography and electron microscopy to quantitatively evaluate key structures involved in exchange in human and equid placental villi. We find that equid villi have a higher surface area to volume ratio and deeper trophoblastic vessel indentation than human villi. Using illustrative computational models, we propose that these structural adaptations have evolved in equids to boost nutrient transfer to compensate for reduced invasiveness into maternal tissue. We discuss these findings in relation to the 'maternal–fetal conflict hypothesis' of placental evolution.

1. Introduction

Placental function underpins fetal development and postnatal health by facilitating physiological exchange between the mother and fetus [1]. The placenta enables the transfer of nutrients and oxygen to the fetus from the mother and removes waste products from the fetal bloodstream, as well as acting as a barrier to toxins and

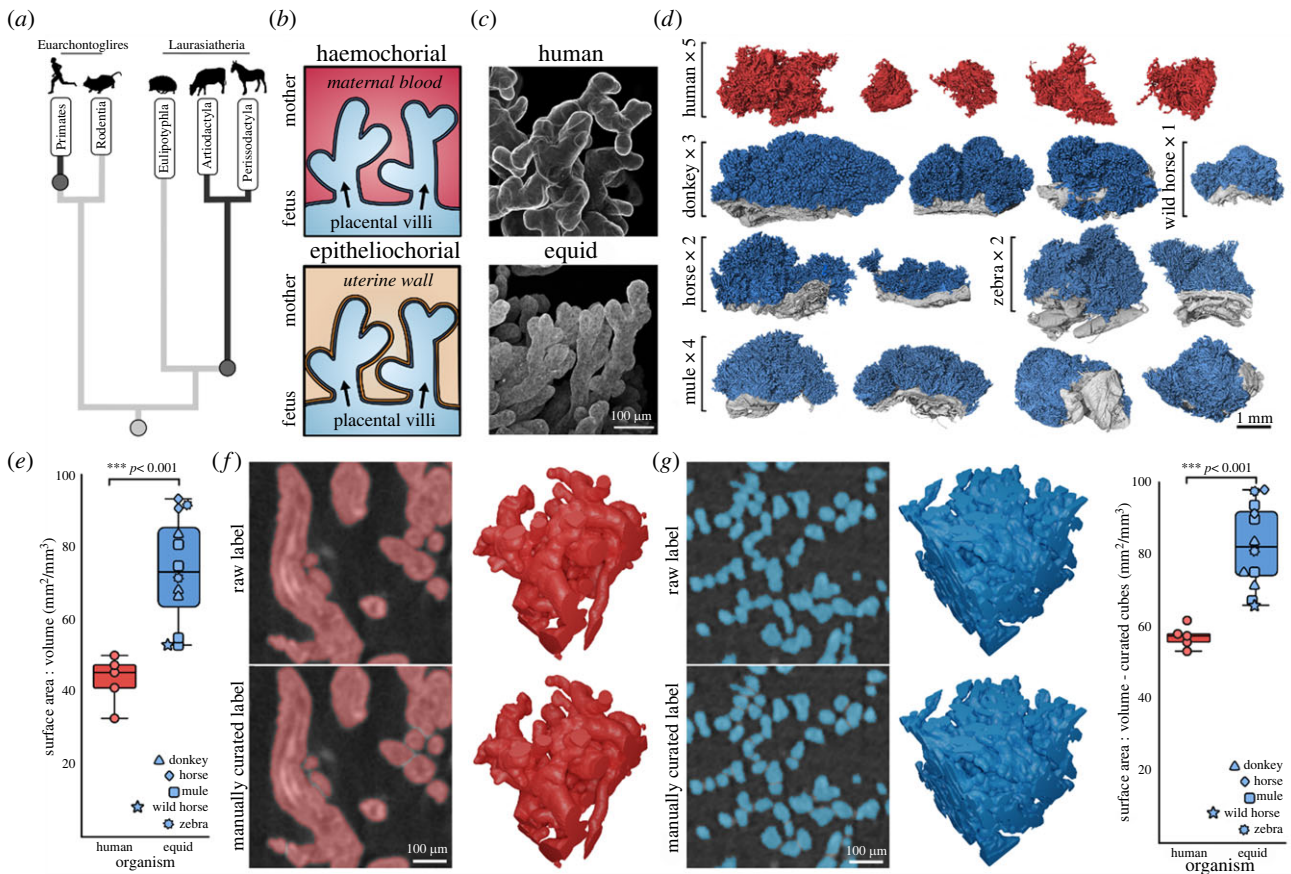


Figure 1. Equid placental villi have a higher surface area to volume ratio (SA : V) than human villi. (a) Placental villi evolved independently in humans and equids (dark lines and circles) from an ancestral labyrinthine placenta (light circle). Simplified tree from the phylogeny outlined in [12]. (b) Human placentas are haemochorial, where villi are in direct contact with maternal blood, whereas equid placentas are epitheliochorial, where villi are only in contact with the maternal uterine epithelium. (c) Fluorescent confocal microscope images of lectin-stained (DSL) human and equid (horse) villi highlighting qualitative differences in morphology. Images are maximum intensity z-projections. (d) Three-dimensional reconstructions from microCT imaging of human (red) and equid (blue) placental villi samples used in this study. Also shown in the equid reconstructions is the non-villous portion of placental samples (grey). (e) MicroCT reconstructions revealed that equid placental villi have a higher SA : V ratio than human placental villi. (f, g) Subvolumes for (f) human and (g) equid (horse) microCT villi were scaled and manually curated to separate connected components. Shown are two-dimensional slices from the volumes (left) and the corresponding three-dimensional reconstruction (right). (h) Equid placental villi were again found to have a higher SA : V than human villi in the curated subvolumes. Animal silhouettes from PhyloPic <https://www.phylopic.org/>.

pathogens [2]. The mammalian placenta is an ephemeral fetal organ which, despite having a single evolutionary origin and conserved functions, exhibits massive morphological and structural diversity across the group [3–5]. The evolutionary pressures governing morphological divergence, and how structure interacts with physiological outcomes, are poorly understood.

Mammalian placentas are grouped into so-called ‘placental types’ [3,4,6–8] according to the way in which maternal and fetal tissues interact (placental interdigitation) and the number of tissue layers separating maternal and fetal circulations (placental invasiveness). Different placental types have been found to correlate with life-history strategies [9], environmental parasite diversity [10] and conflict between maternal and fetus fitness interests [11], however the picture is far from clear. Grouping placental structural variation into qualitative categories may obscure the importance of underlying structural differences across different scales that impact placental function and reproductive success [8].

Human and equid (horses and their kin) placentas are both ‘villous’ placental types, whereby interdigitation with maternal tissue is through treelike branching placental ‘villi’ (figure 1a–c) [7], however their villi evolved independently (figure 1a). Despite showing superficially similar interdigitation, human and equid placentas display very different levels of invasiveness into maternal tissue (figure 1b). In humans, the placenta is ‘haemochorial’ whereby the outer cellular layer of the placental villi (the villous trophoblast) is directly bathed in maternal blood (high invasiveness). By contrast, in equids the placenta is ‘epitheliochorial’ whereby the villous trophoblast is only in contact with the maternal uterine epithelium (low invasiveness). Epitheliochorial placentas are thought to permit less efficient transfer of nutrients from mother to fetus than haemochorial placentas, due to the additional tissue layers between maternal and fetal circulation acting as barriers to transfer [8]. Ancestral trait reconstruction suggests that the ancestral placenta in placental mammals was haemochorial, therefore the epitheliochorial placenta is the derived condition [3,13–16]. Evolutionary pressures proposed to favour the origin of a less invasive placenta include protection of the mother from oxidative stress by fetal metabolism [17], protection of the fetus from vertical transmission of parasites from the mother [10], and the increased ability for the mother to withhold nutrients from the fetus in a phenomenon known as ‘maternal constraint’ [18,19].

Maternal constraint is particularly important when understood in relation to the ‘maternal–fetal conflict hypothesis’ in placental evolution. According to this hypothesis, the long-term fitness interests of the mother and fetus can become misaligned and the

resulting conflict has been proposed to present a key driver in placental evolution [11,20–22]. For example, according to the conflict hypothesis, fetuses with highly invasive placentas could extract more nutrients from the mother than it is optimal for her to give, benefitting fetal fitness in terms of increased growth of the current fetus at the expense of the mother's future reproduction. In species with large precocial offspring which demand high levels of energy investment, such as horses, additional maternal tissue layers may have evolved to allow the mother to constrain the transfer of nutrients to any given offspring to benefit her overall lifetime fitness [19,23]. The pressure for constraint would not be as strong in species with more altricial young, where energy investment is relatively lower. The extent to which the placenta would structurally respond to a reduction in invasiveness and nutrient availability is not known.

Do placentas show structural adaptations to compensate for reduced invasiveness into maternal tissue? This question gains importance when considering that human and equid placentas, despite displaying superficially similar villous interdigitation, differ significantly in placental invasiveness. Their comparison represents a powerful evolutionary case study to explore this idea. Here, we used 3D X-ray microfocus computed tomography (microCT) and electron microscopy (EM) to structurally quantify placental villi from humans and equids across scales. We hypothesized that, given the epitheliochorial organization of the equid placenta, equid villi would display structural adaptations to boost nutrient transfer in response to reduced placental invasiveness relative to humans at different scales.

2. Methods

(a) Sample collection and ethics

Human (*Homo sapiens*) placentas ($n = 5$) were collected following delivery from uncomplicated pregnancies with written informed consent and ethical approval from the Southampton and Southwest Hampshire Local Ethics Committee (11/SC/0529). Equid placentas ($n = 12$) were collected non-invasively after foaling at spontaneous placental delivery from healthy full-term pregnancies as part of routine placental checks. Saddlebred horse (*Equus caballus*) placentas ($n = 2$) were collected at the experimental farm of Institut Français du Cheval et de l'Équitation (IFCE), France. Grévy's zebra (*E. grevyi*) placentas ($n = 2$) and a Przewalski's wild horse (*E. ferus*) placenta ($n = 1$) were collected at Marwell Wildlife, UK. Donkey (*E. asinus*) placentas ($n = 3$) and mule (*E. asinus* × *E. caballus*) placentas ($n = 4$) were collected at the Department of Veterinary Sciences at Pisa University, Italy. Sample analysis in Southampton received local approval from the University of Southampton (47770.A1/46381.A2).

(b) Sample processing and imaging

Small pieces of villous placental tissue (approximately 0.5×0.5 cm) were immediately fixed in 3% glutaraldehyde 0.1 M cacodylate buffer (pH 7.4) upon collection and stored at 4°C until sample processing. In equids, all samples were collected from the area <10 cm from the umbilical cord insertion, in the 'body' of the placenta, in order to standardize sampling. Samples were processed, imaged, and segmented according to the protocol detailed in [24]. Briefly, pieces of fixed placental tissue (approx. 3–5 mm) were dissected, heavy-metal stained (with osmium tetroxide, uranyl acetate and lead nitrate), dehydrated and embedded in Spurr's resin. The resin blocks were then scanned using a custom Nikon XTH 225 ST (Nikon X-Tek Systems Ltd) microCT scanner at the μ -VIS X-ray Imaging Centre, three-dimensional X-ray Histology Facility, University of Southampton (www.muvis.org) [25], generating image volumes of 4.8 μ m isotropic voxels. Scanned blocks were then trimmed by ultramicrotomy using an EM UC7 ultramicrotome (Leica, Germany). Villous tissue subvolumes of interest identified from microCT datasets were excised from the resin blocks and mounted onto aluminium pins (TABB—G312/1) with conductive glue (CircuitWorks Conductive Epoxy CW2400; ITW Chemtronics). Pins were imaged by block face scanning EM using a 3View system (Gatan, Abingdon, UK) mounted in a Quanta250 Field Emission Gun Scanning Electron Microscope (FEG-SEM) (ThermoFisher, Eindhoven, The Netherlands). Two-dimensional cross sections of villi were randomly imaged ($n \geq 5$ different villi per biological placenta replicate) across the block face at 15 nm XY pixels.

(c) Image analysis and statistical comparison

Image datasets were imported into Microscopy Image Browser (MIB) [26] as .tiff stacks and segmented by intensity thresholding combined with semi-manual curation. This involved masking regions of interest and thresholding tissue within the bounds of the mask, as detailed in [24]. Labels were further statistically thresholded to clean up erroneous objects by excluding connected components less than 1 k voxels in three dimensions. Labels were quantified for volume, surface area and other morphometric parameters within MIB. Intensity thresholding of villi from microCT datasets resulted in segmented labels where individual villi were sometimes incorrectly connected in two-dimensional slices, however manual separation of every villus across whole datasets was prohibitive and watershed separation ineffective for these datasets. To eliminate the possibility that this connectivity affected accurate surface area quantification, $128 \times 128 \times 128$ voxel subvolumes of villi were cropped from each sample and manually curated (figure 1f,g). The scaling differences between non-curated and curated subvolumes for villus volume (mean \pm s.d., 0.96 ± 0.03) and surface area (1.11 ± 0.07) were extrapolated across the whole tissue volumes to correct for villus connectivity. These scaling differences were not significantly different for humans and equids ($p = 0.35$). Curated subvolumes were also independently compared between species (figure 1h).

Data were evaluated for prior normality and homogeneity assumptions with a Shapiro and Levene's test respectively. Morphometric differences between human and equid placental villi were evaluated using a *t*-test, or a Mann–Whitney *U*-test if the above assumptions were not met. The scipy package [27] run with Python 3.9.12 implemented in Jupyter Notebook 6.4.8 was used for all statistical comparisons. Means and standard deviations were calculated from mean values per biological placenta.

(d) Computational modelling of nutrient transfer

Segmented images were imported into Simpleware ScanIP (P-2019.09; Synopsys, Inc., Mountain View, USA) for mesh generation. Steady state diffusion simulations were performed in COMSOL Multiphysics (v6.0; COMSOL AB, Stockholm, Sweden). Only normalized concentrations were considered, therefore boundary conditions of 1 and 0 mol m⁻³ were applied to the outer villus and inner capillary surface

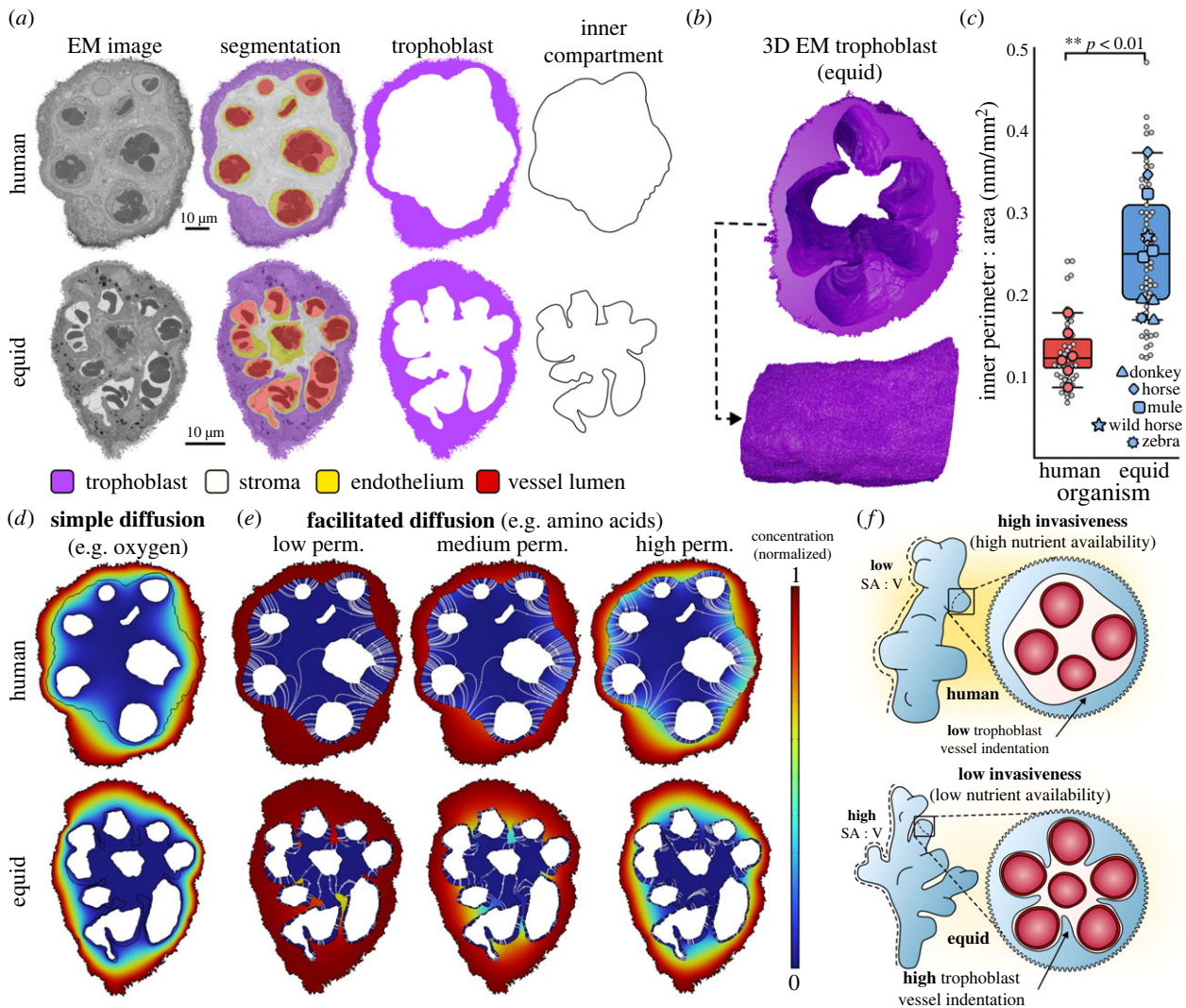


Figure 2. Equid placental villi display deep trophoblastic invaginations and a higher inner perimeter to area ratio ($P:A$) than human villi. (a) Illustrative example of a two-dimensional cross section through human and equid placental villi showing deep trophoblastic basal invaginations in equids but not in humans. (b) Three-dimensional serial block face scanning electron microscopy (SBF-SEM) reconstruction of the trophoblast showing basal invaginations into the stroma in an equid (horse) villus. (c) Electron microscopy of villus cross sections revealed that the inner compartment of equid placental villi has a higher $P:A$ ratio than in human placental villi. Grey points indicate individual villus replicates ($n \geq 5$ different villi per placenta), coloured points indicate means from each whole placenta. (d,e) Illustrative physiological modelling of simple diffusion (d) and facilitated (transporter-mediated) diffusion (e) through human and equid villus cross sections. Concentration gradients shown ranging from red (high) to blue (low). Facilitated diffusion was modelled at low, medium, and high permeabilities of the basal trophoblast membrane (e) with overlaid white lines showing illustrative routes of transfer. (f) Diagrammatic summary of the findings of this study.

respectively. For facilitated diffusion, the normal flux J_{BM} ($\text{mol m}^{-2} \text{s}^{-1}$) at the basal membrane was assumed to be governed by the following equation for a facilitative transporter [28]:

$$J_{BM} = V_{\max} \left(\frac{c_1}{K_m + c_1} - \frac{c_2}{K_m + c_2} \right), \quad (2.1)$$

where V_{\max} ($\text{mol m}^{-2} \text{s}^{-1}$) is the maximum transport rate and K_m (mol m^{-3}) the half maximum rate concentration, set to 0.5, while c_1 is the local intracellular concentration in the trophoblast and c_2 the local extracellular concentration in the stroma on either side of the basal membrane.

3. Results

MicroCT imaging allowed us to reconstruct and morphometrically quantify the villi of human and equid (donkey, wild horse, horse, zebra and mule) placentas (figure 1d). Quantification of the microCT reconstructions revealed that equid placental villi have a higher surface area to volume ratio ($SA:V$) (73.47 ± 14.91) than human villi (43.34 ± 6.78 , $p < 0.001$) (figure 1e). This was also true when imaged villi were scaled to the same subvolumes and manually curated (figure 1f,g) (82.44 ± 11.54 versus 57.20 ± 3.13) ($p < 0.001$) (figure 1h). Plotting of individual equid placental replicates shows a non-overlapping distribution when compared with human samples, providing justification for the lumped 'equid' category in this study. (Figure 1e,h). At the scale of EM, equid placental villi displayed deep trophoblastic invaginations between blood vessels (figure 2a,b) resulting in deeper vascular indentation than in human villi. This can be quantified by calculating the perimeter to area ratio ($P:A$) of the inner villus

compartment (blood vessels bound by stroma), which was higher in equids (0.24 ± 0.07) than in humans (0.12 ± 0.03) ($p < 0.01$) (figure 2c). This quantitative metric means that the inner compartment is in relatively higher contact with the basal membrane of the trophoblast in equids than in humans. Illustrative modelling of diffusive processes suggests that these protrusions are unlikely to influence simple transfer of gases such as oxygen (figure 2d) but may aid facilitated transporter-mediated transfer of substances such as amino acids (figure 2e). As transporter proteins are embedded in membrane boundaries between tissue layers, this is particularly true for modelled scenarios with low membrane permeabilities (figure 2e). It can also be observed that the deeper vessel indentation in equid than human villi produces shorter diffusion pathways from the trophoblast basal membrane to placental blood vessels, particularly for those in the villus centre (figure 2e).

4. Discussion

Using microCT and EM, we imaged and structurally quantified convergently evolved villi from human and equid placentas. We demonstrate that equid villi have a higher SA:V than human villi, and display basal trophoblastic invaginations which confer deeper blood vessel indentation. We propose that these multiscale structural differences may provide certain advantages to nutrient transfer in equids and may have evolved in response to reduced nutrient availability in less invasive equid placentas (figure 2d–f). If so, our results may challenge the assumption that epitheliochorial placentas are intrinsically ‘less efficient’ at nutrient transfer than more invasive haemochorial placentas and stress the need for resolving placental structures quantitatively across scales. Our study also suggests that qualitative categories for convergently evolved structures in placental biology, such as ‘villi’, may mask hidden biological variation and adaptation. Although in this study numbers of individual equid species were too low to identify intrageneric differences, our methods could also be used in the future to identify differences in the villi between equid species to interrogate these patterns further.

Villous surface area is important in placental nutrient transfer, with decreases in surface area linked to human placental dysfunctions such as pre-eclampsia [29] and anaemia [30]. As a transporting epithelium, the apical and basal membranes of placental villous trophoblasts contain high densities of transporter proteins and are involved in active transport [31]. Therefore, our finding that equid villi have a higher surface area than human villi for a given volume may confer advantages to nutrient transfer. Likewise, the basal invagination and higher blood vessel indentation of the trophoblast seen in equids could provide benefits to basal transfer through moving nutrients into fetal circulation. Indeed, the high blood vessel indentation of epitheliochorial trophoblasts has before been qualitatively noted [32–35]. Modelling of oxygen versus amino acid (simple versus transporter-mediated) diffusion through these structures highlights the potential benefit that basal trophoblast invaginations could confer by ensuring transported solutes are delivered in close proximity to fetal capillaries, reducing diffusion distances and providing additional surface area for membrane transport. While the basal protrusions may facilitate transfer of substrates requiring membrane transport, it is not clear that they will benefit solutes transferred by simple diffusion. That said, the higher SA:V seen in equid villi compared to humans from our microCT data could confer benefits to oxygen transfer through reduced relative diffusion distances to capillaries and the villus centre from the villus exterior.

If these structural adaptations in equid villi are a response to reduced nutrient availability through decreased invasiveness into maternal tissue as we propose, our data could provide evidence supporting the ‘maternal–fetal conflict hypothesis’ in placental evolution. Experimental evidence suggests that placental invasiveness is regulated by the maternal endometrium, such as through the activity of stromal cells, conferring uterine permissiveness [16,36–39]. According to the maternal–fetal conflict hypothesis, epitheliochorial placentation evolved as a way for the mother to constrain nutrients from the fetus in species with precocial energy-intensive young, whereas the genes controlling placental development may be selected for optimal nutrient extraction, potentially at the expense of the mother’s long-term fitness interests. We propose, in accordance with the same hypothesis, that as a result fetal placental villi are pressured to compensate for reduced invasiveness through the evolution of structural adaptations to boost nutrient transfer, demonstrated by our data.

For our hypothesis to be fully supported, further work is needed. Our data highlight the limitations of grouping mammalian placentas into qualitative types and stress the need for multiscale quantitative structural characterization. Structures involved in nutrient transfer in mammalian placentas must also be quantified at other scales. An example of why multiscale analysis is necessary is that while this study finds increased surface area of the basal membrane in horses, use of high-resolution volume EM has identified a different adaptation, basal membrane folds, which increase basal membrane surface area in humans at a scale that is not apparent in the current study [40]. The extent to which similar nanoscale infolding boosts exchange area in equids is unknown, however future investigations should include this metric in a holistic understanding of surface area.

Quantifying exchange areas at the scale of gross morphology would also be particularly informative, as it has been qualitatively observed that there seems to be an inverse correlation between gross morphology and invasiveness, with less invasive placentas occupying a greater relative area of the uterine wall [7,8]. This observation is also true in this study, with haemochorial human placentas being discoid (low relative uterine coverage) and epitheliochorial equid placentas being diffuse (high relative uterine coverage) [41]. Is this inverse correlation also a response to reduced placental invasiveness? While future studies are necessary, we propose that our data provide evidence that epitheliochorial placental villi do exhibit structural adaptations to boost nutrient transfer in response to reduced invasiveness.

5. Conclusion

The mammalian placenta exhibits massive structural diversity, despite having a single evolutionary origin and conserved functions. Mammalian placentas are currently grouped into qualitative ‘types’, which may hide structural variation important for

understanding function, evolution and life-history strategy. Here, we used microCT and EM to quantify structures in placental villi from humans and equids, which have evolved independently and show different levels of invasiveness into maternal tissue. Our study reveals that equid placental villi have a higher SA : V and closer association between the trophoblast basal membrane and the capillary than human villi. We propose that these are structural adaptations to boost nutrient transfer and compensate for reduced invasiveness into maternal tissue in equid placentas. Although further work is needed to fully resolve this hypothesis, these data suggest that mammalian placentas can structurally adapt to boost nutrient transfer efficiency in response to the evolution of reduced maternal nutrient availability and show that not all placental villi are necessarily the same.

Ethics. Human (*Homo sapiens*) placentas ($n = 5$) were collected following delivery from uncomplicated pregnancies with written informed consent and ethical approval from the Southampton and Southwest Hampshire Local Ethics Committee (11/SC/0529). Equid placentas ($n = 12$) were collected non-invasively after foaling at spontaneous placental delivery from healthy full-term pregnancies as part of routine placental checks. Saddlebred horse (*Equus caballus*) placentas ($n = 2$) were collected at the experimental farm of Institut Français du Cheval et de l'Équitation (IFCE), France. Grévy's zebra (*E. grevyi*) placentas ($n = 2$) and a Przewalski's wild horse (*E. ferus*) placenta ($n = 1$) were collected at Marwell Wildlife, UK. Donkey (*E. asinus*) placentas ($n = 3$) and mule (*E. asinus* × *E. caballus*) placentas ($n = 4$) were collected at the Department of Veterinary Sciences at Pisa University, Italy. Sample analysis in Southampton received local approval from the University of Southampton (47770.A1/46381.A2).

Data accessibility. All data (microCT and EM image datasets with segmented labels (.tiff), modelling datasets (.mph), morphometric data (.csv), and metadata (.csv)) associated with this study are freely available to download at Bioimage Archive accession S-BIAD999 [42] under license CC BY 4.0: <https://www.ebi.ac.uk/biostudies/bioimages/studies/S-BIAD999>. Details of image processing and statistical analysis can be found in the Methods section of this paper.

Declaration of AI use. We have not used AI-assisted technologies in creating this article.

Authors' contributions. D.L.: conceptualization, formal analysis, investigation, methodology, visualization, writing—original draft, writing—review and editing; B.G.S.: data curation, formal analysis, investigation, methodology, visualization, writing—review and editing; J.T.: data curation, investigation, methodology, writing—review and editing; S.E.H.: data curation, formal analysis, investigation, methodology, writing—review and editing; O.B.: investigation, methodology, writing—review and editing; P.J.B.: data curation, methodology, writing—review and editing; O.L.K.: data curation, methodology, writing—review and editing; P.G.: data curation, methodology, writing—review and editing; E.D.: investigation, resources, writing—review and editing; D.F.: investigation, resources, writing—review and editing; C.B.: investigation, resources, writing—review and editing; F.C.: investigation, resources, writing—review and editing; J.S.: investigation, resources, writing—review and editing; G.C.-D.: investigation, resources, writing—review and editing; N.J.G.: funding acquisition, supervision, writing—review and editing; P.C.-P.: funding acquisition, supervision, writing—review and editing; R.M.L.: conceptualization, funding acquisition, supervision, writing—original draft, writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. We declare we have no competing interests.

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References

- Burton GJ, Fowden AL, Thornburg KL. 2016 Placental origins of chronic disease. *Physiol. Rev.* **96**, 1509–1565. (doi:10.1152/physrev.00029.2015)
- Cooke LDF, Tumbarello DA, Harvey NC, Sethi JK, Lewis RM, Cleal JK. 2021 Endocytosis in the placenta: an undervalued mediator of placental transfer. *Placenta* **113**, 67–73. (doi:10.1016/j.placenta.2021.04.014)
- Wildman DE, Chen CY, Erez O, Grossman LI, Goodman M, Romero R. 2006 Evolution of the mammalian placenta revealed by phylogenetic analysis. *Proc. Natl Acad. Sci. USA* **103**, 3203–3208. (doi:10.1073/pnas.0511344103)
- Furukawa S, Kuroda Y, Sugiyama A. 2014 A comparison of the histological structure of the placenta in experimental animals. *J. Toxicol. Pathol.* **27**, 11–18. (doi:10.1293/tox.2013-0060)
- Carter AM. 2018 Comparative placentation. In *Encyclopedia of reproduction*, pp. 129–136. London, UK: Academic Press.
- Chavatte-Palmer P, Tarrade A. 2016 Placentation in different mammalian species. *Ann. Endocrinol.-Paris* **77**, 67–74.
- Burton GJ. 2022 Placental types. In *Benirschke's pathology of the human placenta*, pp. 23–38, 7th edn. Cham, Switzerland: Springer Nature.
- Laundon D, Gostling NJ, Sengers BG, Chavatte-Palmer P, Lewis RM. 2023 Placental evolution from a three-dimensional and multiscale structural perspective. *Evolution* **78**, 13–25. (doi:10.1093/evolut/qpaa209)
- Capellini I, Venditti C, Barton RA. 2011 Placentation and maternal investment in mammals. *Am. Nat.* **177**, 86–98. (doi:10.1086/657435)
- Capellini I, Nunn CL, Barton RA. 2015 Microparasites and placental invasiveness in eutherian mammals. *PLoS ONE* **10**, e0132563. (doi:10.1371/journal.pone.0132563)
- Kazemian A, Hooshmandabbasi R, Schraner EM, Boos A, Klisch K. 2019 Evolutionary implications of fetal and maternal microvillous surfaces in epitheliochorial placentae. *J. Morphol.* **280**, 615–622.
- Upham NS, Esselstyn JA, Jetz W. 2019 Inferring the mammal tree: species-level sets of phylogenies for questions in ecology, evolution, and conservation. *PLoS Biol.* **17**, e3000494.
- Vogel P. 2005 The current molecular phylogeny of eutherian mammals challenges previous interpretations of placental evolution. *Placenta* **26**, 591–596. (doi:10.1016/j.placenta.2004.11.005)
- Carter AM, Mess A. 2007 Evolution of the placenta in eutherian mammals. *Placenta* **28**, 259–262. (doi:10.1016/j.placenta.2006.04.010)
- Elliot MG, Crespi BJ. 2009 Phylogenetic evidence for early hemochorial placentation in eutheria. *Placenta* **30**, 949–967. (doi:10.1016/j.placenta.2009.08.004)
- Mika K, Whittington CM, McAllan BM, Lynch VJ. 2022 Gene expression phylogenies and ancestral transcriptome reconstruction resolves major transitions in the origins of pregnancy. *eLife* **11**, e74297.
- Elliot MG. 2016 Oxidative stress and the evolutionary origins of preeclampsia. *J. Reprod. Immunol.* **114**, 75–80.

18. Gluckman PD, Morel PCH, Ambler GR, Brier BH, Blair HT, Mccutcheon SN. 1992 Elevating maternal insulin-like growth factor-I in mice and rats alters the pattern of fetal growth by removing maternal constraint. *J. Endocrinol.* **134**, R1–R3. (doi:10.1677/joe.0.134R001)
19. Allen WR, Wilsher S, Turnbull C, Stewart F, Ousey J, Rosedale PD, Fowden AL. 2002 Influence of maternal size on placental, fetal and postnatal growth in the horse. I. Development *in utero*. *Reproduction* **123**, 445–453. (doi:10.1530/rep.0.1230445)
20. Haig D. 1993 Genetic conflicts in human pregnancy. *Q Rev. Biol.* **68**, 495–532. (doi:10.1086/418300)
21. Klich K, Mess A. 2007 Evolutionary differentiation of cetartiodactyl placentae in the light of the viviparity-driven conflict hypothesis. *Placenta* **28**, 353–360. (doi:10.1016/j.placenta.2006.03.014)
22. Wang X, Miller DC, Harman R, Antczak DF, Clark AG. 2013 Paternally expressed genes predominate in the placenta. *Proc. Natl Acad. Sci. USA* **110**, 10 705–10 710. (doi:10.1073/pnas.1308998110)
23. Lewis RM, Cleal JK, Hanson MA. 2012 Review: Placenta, evolution and lifelong health. *Placenta* **33**, S28–S32. (doi:10.1016/j.placenta.2011.12.003)
24. Laundon D, Katsamenis OL, Thompson J, Goggin P, Chatelet DS, Chavatte-Palmer P, Gostling NJ, Lewis RM. 2023 Correlative multiscale microCT-SBF-SEM imaging of resin-embedded tissue. *Methods Cell Biol.* **177**, 241–267.
25. Katsamenis OL *et al.* 2023 A high-throughput 3D X-ray histology facility for biomedical research and preclinical applications. *Wellcome Open Res.* **8**, 366.
26. Belevich I, Joensuu M, Kumar D, Vihinen H, Jokitalo E. 2016 Microscopy Image Browser: a platform for segmentation and analysis of multidimensional datasets. *PLoS Biol.* **14**, e1002340. (doi:10.1371/journal.pbio.1002340)
27. Virtanen P *et al.* 2020 SciPy 1.0: fundamental algorithms for scientific computing in Python. *Nat. Methods* **17**, 261–272. (doi:10.1038/s41592-019-0686-2)
28. Panitchob N, Widdows KL, Crocker IP, Johnstone ED, Please CP, Sibley CP, Glazier JD, Lewis RM, Sengers BG. 2016 Computational modelling of placental amino acid transfer as an integrated system. *Bba-Biomembranes* **1858**, 1451–1461. (doi:10.1016/j.bbamem.2016.03.028)
29. Egbor M, Ansari T, Morris N, Green CJ, Sibbons PD. 2006 Morphometric placental villous and vascular abnormalities in early- and late-onset pre-eclampsia with and without fetal growth restriction. *Int. J. Obstet. Gy.* **113**, 580–589. (doi:10.1111/j.1471-0528.2006.00882.x)
30. Reshetnikova OS, Burton GJ, Teleshova OV. 1995 Placental histomorphometry and morphometric diffusing capacity of the villous membrane in pregnancies complicated by maternal iron-deficiency anemia. *Am. J. Obstet. Gynecol.* **173**, 724–727. (doi:10.1016/0002-9378(95)90330-5)
31. Burton GJ. 2022 Basic structure of a placental villus. In *Benirschke's pathology of the human placenta*, pp. 23–38, 7th edn. Cham, Switzerland: Springer Nature.
32. Samuel CA, Allen WR, Steven DH. 1976 Studies on the equine placenta II. Ultrastructure of the placental barrier. *J. Reprod. Fert.* **48**, 257–264.
33. Enders AC, Blankenship TN. 1999 Comparative placental structure. *Adv. Drug Deliver. Rev.* **38**, 3–15. (doi:10.1016/S0169-409x(99)00003-4)
34. Carter AM, Enders A. 2004 Comparative aspects of trophoblast development and placentation. *Reprod. Biol. Endocrinol.* **2**, 1–15.
35. Wooding FBP, Burton GJ. 2008 *Comparative placentation: structures, functions and evolution*. Berlin, Germany: Springer.
36. Samuel CA, Perry JS. 1972 The ultrastructure of pig trophoblast transplanted to an ectopic site in the uterine wall. *J. Anat.* **113**, 139–149.
37. Graham CH, Lala PK. 1991 Mechanism of control of trophoblast invasion *in situ*. *J. Cell. Physiol.* **148**, 228–234. (doi:10.1002/jcp.1041480207)
38. Kshitiz *et al.* 2019 Evolution of placental invasion and cancer metastasis are causally linked. *Nat. Ecol. Evol.* **3**, 1743. (doi:10.1038/s41559-019-1046-4).
39. Wagner GP, Kshitiz, Dighe A, Levchenko A. 2022 The coevolution of placentation and cancer. *Annu. Rev. Anim. Biosci.* **10**, 259–279. (doi:10.1146/annurev-animal-020420-031544)
40. Tashev SA *et al.* 2022 Folding of the syncytiotrophoblast basal plasma membrane increases the surface area available for exchange in human placenta. *Placenta* **117**, 57–63. (doi:10.1016/j.placenta.2021.11.002)
41. Baur R. 1977 Morphometry of the placental exchange area. *Adv. Anat. Embryol. Cell Biol.* **53**, 3–65. (doi:10.1007/978-3-642-66603-2)
42. Laundon D *et al.* 2024 Data from: Convergently evolved placental villi show multiscale structural adaptations to differential placental invasiveness. (Bioimage Archive, S-BIAD999). Bioimage Archive. (<https://www.ebi.ac.uk/biostudies/bioimages/studies/S-BIAD999>)