

1 **Title**

2 **Full title**

3 An evaluation of the effects of localised skin cooling on microvascular, inflammatory,
4 structural, and perceptual responses to sustained mechanical loading of the sacrum: a study
5 protocol.

6 **Short title**

7 Skin cooling and pressure ulcer risks.

8 **Authors**

9 Ralph J. F. H. Gordon^{1*}, Peter R. Worsley², Davide Filingeri¹.

10 **Affiliations**

11 ¹ ThermosenseLab, Skin Sensing Research Group, School of Health Science, University of
12 Southampton, UK.

13 ² PressureLab, Skin Sensing Research Group, School of Health Science, University of
14 Southampton, UK.

15 * Corresponding author

16 E-mail: r.j.f.h.gordon@soton.ac.uk

17 **Abstract**

18 This study protocol aims to investigate how localised cooling influences the skin's
19 microvascular, inflammatory, structural, and perceptual tolerance to sustained mechanical
20 loading at the sacrum, evaluating factors such as morphology, physiology, and perceptual
21 responses. The protocol will be tested on individuals of different age, sex, skin tone and clinical
22 status, using a repeated-measure design with three participants cohorts: i) young healthy
23 (n=35); ii) older healthy (n=35); iii) spinal cord injured (SCI, n=35). Participants will complete
24 three testing sessions during which their sacrum will be mechanically loaded (60 mmHg; 45
25 min) and unloaded (20 min) with a custom-built thermal probe, causing pressure-induced
26 ischemia and post-occlusive reactive hyperaemia. Testing sessions will differ by the probe's
27 temperature, which will be set to either 38°C (no cooling), 24°C (mild cooling), or 16°C (strong
28 cooling). We will measure skin blood flow (via Laser Doppler Flowmetry; 40 Hz); pro- and
29 anti-inflammatory biomarkers in skin sebum (Sebutape); structural skin properties (Optical
30 Coherence Tomography); and ratings of thermal sensation, comfort, and acceptance (Likert
31 Scales); throughout the loading and unloading phases. Changes in post-occlusive reactive
32 hyperaemia will be considered as the primary outcome and data will be analysed for the
33 independent and interactive effects of stimuli's temperature and of participant group on within-
34 and between-subject mean differences (and 95% Confidence Intervals) in peak hyperaemia, by
35 means of a 2-way mixed model ANOVA (or Friedman). Regression models will also be
36 developed to assess the relationship between absolute cooling temperatures and peak
37 hyperaemia. Secondary outcomes will be within- and between-subject mean changes in
38 biomarkers' expression, skin structural and perceptual responses. This analysis will help
39 identifying physiological and perceptual thresholds for the protective effects of cooling from
40 mechanically induced damage underlying the development of pressure ulcers in individuals
41 varying in age and clinical status.

42 **Introduction**

43 Pressure ulcers (PUs) are localised damage to the skin and sub-dermal tissues, resulting from
44 sustained periods of pressure, or pressure in combination with shear forces [1]. In the United
45 Kingdom alone, the annual cost of treating chronic wounds, including PUs, has been estimated
46 at £8.3 billion [2]. Accordingly, an improved understanding of the fundamental mechanisms
47 underlying the physiological tolerance of human skin to mechanical loading could lead to the
48 development of cost-effective, personalised solutions to prevent these wounds and improve
49 patient care and quality of life.

50 Sustained localised mechanical loading on the skin can arise from lying and sitting postures,
51 as well as the prolonged attachment of medical devices, e.g. prosthetics or respiratory masks
52 [3, 4]. Internal tissue deformations will occur as a result of sustained pressure and shear forces
53 that can lead to changes in the physiology of skin and sub-dermal tissue, including ischemia in
54 the blood vasculature, lymphatic impairment, and direct deformation damage [5]. When load
55 is removed, ischemia reperfusion injury may also occur due to the onset of post-occlusive
56 reactive hyperaemia [6]. Reactive hyperaemia can increase the risk of ischemia reperfusion
57 injury by triggering the release of oxygen-derived free radicals with cytotoxic effects, and this
58 can play a role in the pathophysiology of PUs [7]. In addition, microclimate conditions within
59 and around skin tissues strongly influence its tolerance to mechanical loading. For example,
60 elevated temperature and humidity at the skin interface reduces the mechanical stiffness and
61 strength of the skin and can increase its friction coefficient [4]. In contrast, cooling reduces
62 skin tissue's metabolic demands and could increase the skin's physiological tolerance to
63 mechanical damage [4, 8].

64 Evidence that changes in skin temperature could play a role in the tolerance of the skin to
65 mechanical loading and shear came from early animal studies using porcine models, revealing
66 that reduced skin temperature minimises the risk of PU formation through altered

67 microvascular responses [9, 10]. More recently, the protective effective of reducing skin
68 temperature has been demonstrated in rats [11, 12]. While this evidence highlights the potential
69 therapeutic role of skin cooling for protecting tissue health, the mechanisms by which cooling
70 enhances skin tolerance to pressure remain poorly understood in humans [9-11, 13-16].
71 Specifically, it remains unclear whether and to what extent the benefits of lowering skin
72 temperature arise from the individual or combined effects of: 1) preserved microvascular
73 function during mechanical loading and/or attenuated post-occlusive reactive hyperaemia
74 following on pressure release; and 2) downregulation of skin's inflammatory responses to
75 sustained mechanical pressure. Animal studies revealed that local cooling, as well as the
76 stimulation of cold sensitive TRPM8-expressing neurons in dorsal root ganglions, could
77 modulate the skin's inflammatory responses to acute mechanical stress (e.g. pressure loading)
78 [12] and chronic skin damage (e.g. chronic dermatitis) [17], via downregulation of the
79 expression of pro-inflammatory cytokines such as Tumour Necrosis Factor alpha (TNF- α).

80 In addition to its physiological effects, it is well known that localised cooling of the skin can
81 induce cold discomfort, which, if the magnitude of the cooling stimulus is large enough, can
82 limit acceptability and adherence to therapeutic interventions designed to maintain skin health,
83 particularly for vulnerable individuals at risk of PUs such as the elderly [18]. Hence, cold-
84 induced discomfort could provide an obstacle for the adoption of skin cooling as a therapeutic
85 intervention to promote skin integrity in humans. However, there is limited evidence on how
86 the absolute cooling temperature and applied pressure on the skin interact in driving discomfort
87 [19]. Despite these challenges, several support surfaces and therapeutic interfaces have been
88 designed with local and full body cooling applied through microclimate management systems
89 [20-22], although the evidence underlying their efficacy remain limited.

90 Modelling the relationship between the physiological and perceptual effects of skin cooling
91 could provide an empirical approach to identify a common level of cooling that proves both

92 physiologically beneficial and perceptually acceptable. This could then be translated to inform
93 design parameters for more effective support surfaces and therapeutic interfaces.

94 It should also be recognised that the underlying tolerance to pressure at the skin interface, as
95 well as the physiological and perceptual effects of cooling to pressure-induced damage, may
96 vary as a function of age and comorbidities [3]. These states are associated with changes in
97 skin biophysics and morphology [23], and in thermoregulatory and perceptual sensitivities. For
98 example, ageing is likely to modulate the effects of cooling on tissue tolerance, as aged skin
99 presents a reduced physiological and perceptual sensitivity to cold, due to decreases in both
100 reflex cutaneous vasoconstriction, and density of thermoreceptors [24]. Similarly, the presence
101 of a spinal cord injury (SCI) is associated with autonomic (e.g. impaired control of skin blood
102 flow) and sensory dysfunctions (e.g. perceptual loss below injury level) [25, 26]. Thus, there
103 may be variations between reductions in cold sensitivity and diminished efficacy of therapeutic
104 cooling associated with age and comorbidities.

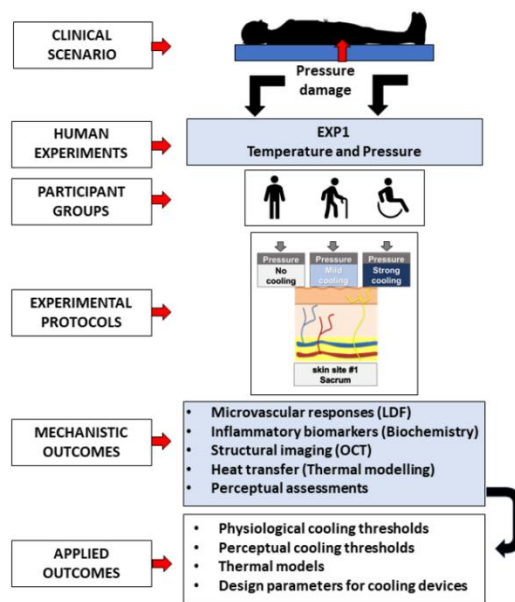
105 This study protocol aims to investigate: 1) how different levels of localised cooling influences
106 the skin's microvascular, inflammatory, structural, and perceptual responses to sustained
107 mechanical loading at the sacrum; and 2) how aging and spinal cord injury may modulate the
108 metabolic, immunological, biophysical, and perceptual pathways underlying the beneficial
109 effects of localised cooling on skin tolerance to mechanical loading. To achieve our aims, we
110 have designed a clinically relevant experiment in healthy young participants and in groups at-
111 risk of PUs (i.e., older and SCI), as detailed in the sections below. The investigation will offer
112 a combination of skin viability, thermal physiology, and non-invasive skin sensing
113 technologies, to develop new basic knowledge on the role of temperature in reducing the risk
114 of skin damage. This will support innovation in the design of healthcare and user-centred
115 technologies, such as mattresses, clothing, and medical devices that can safely interface with

116 the skin and provide some protection from damage. This will unlock the potential of cooling
 117 to the skin that will help maintain skin health across the life course.

118 **Materials and Methods**

119 **Overview**

120 Participants will attend the laboratory within the Clinical Academic Facility located at
 121 Southampton General Hospital (Southampton, UK), to complete three experimental sessions
 122 separated by a minimum of 24 hrs. During the sessions, participants' skin over the sacrum will
 123 be mechanically loaded (60 mmHg; 45 min) and unloaded (minimum tare load: 17.5 mmHg;
 124 20 min) with a custom-built thermal probe, to cause pressure-induced ischemia and post-
 125 occlusive hyperaemia. The study will be a randomised cross-over design, involving three probe
 126 temperatures, which will be set to either 38°C (no cooling), 24°C (mild cooling), or 16°C
 127 (strong cooling). An overview of the study design can be found in Figure. 1.



128

129 **Figure. 1. Experimental design.** We have designed a clinically relevant experiment in healthy young participants
 130 and in groups at-risk of PUs (i.e., elderly and spinal cord injured, SCI), which will determine how different levels
 131 of cooling [i.e. no cooling (38°C), mild cooling (16°C), and strong cooling (16°C)] alter the skin' microvascular,
 132 inflammatory, structural, and perceptual responses to sustained pressure-induced ischemia and reactive
 133 hyperaemia. From an applied standpoint, the research will identify physiological and perceptual cooling
 134 thresholds (i.e. level of cooling, modulations via age and clinical status), which could be used as design parameters
 135 for the development of user-centred medical devices and thermal wearables. LDF: Laser Doppler Flowmetry;
 136 OCT: Optical Coherence Tomography.

137 **Participants**

138 Three participant cohorts will be recruited: i) young healthy (n =35); ii) older healthy (n =35);
139 and iii) spinal cord injured (SCI, n =35). Sample size calculations were performed using
140 Gpower (Gpower 3.1) with an effect size $f = 0.4$ for a repeated-measure ANOVA [parameters:
141 within-between interaction; $\alpha = 0.05$; $\beta = 0.80$; 3 groups; 3 measurements (i.e. peak hyperaemia
142 at 38, 24, 16°C)], based on published data on the mean difference in peak hyperaemia
143 (expressed as percentage of maximal cutaneous vascular conductance, CVCmax) during
144 control conditions ($32.7 \pm 9.4\%$ CVCmax) and during sensory nerve block ($17.3 \pm 6.8\%$
145 CVCmax) [27]. Changes in peak hyperaemia were identified as the primary experimental
146 outcome, as this represents a robust and repeatable microvascular response. The effects of
147 cooling on peak hyperaemia are likely to be similar to those of cutaneous sensory nerve bloc
148 [27], given that cooling also impairs the activity of cutaneous sensory nerves [28]. Hence, it
149 was identified that the 50%-reduction in peak hyperaemia reported by Lorenzo et al. [27] was
150 a large ($f = 0.4$) and physiologically meaningful effect size to evaluate the beneficial effects of
151 cooling on sustained mechanical loading. Based on the data above, we estimated a minimum
152 sample size of 18 participants per group, and we propose testing of 35 individuals per group to
153 allow for sufficient statistical power and to account for up to 50% dropout. Participants will be
154 recruited according to the criteria in Table 1.

155

156 **Table 1.** Participant inclusion and exclusion criteria.

Inclusion	Exclusion
18-70 years old (young healthy 18-35 years old; older healthy 55-70 years old).	Young and older health groups only (does not apply to SCI group, see text below): suffering from cardiovascular, metabolic, and neurological disorders and/or comorbidities, e.g., hypertension, diabetes, chronic lung disease.
Male or female (minimum 17 for each sex).	Raynaud's disease.
Skin tone dark, medium, or light (minimum 11 for each tone - assessed via the Fitzpatrick Scale [29]).	Suffering from skin conditions (e.g., eczema).
Healthy groups only: physically active (i.e., performing exercise 1 to 3 times a week).	Under drug therapy affecting thermoregulation (e.g. muscarinic antagonists).
SCI group only: presenting thoracic injury/paraplegia (i.e., injury level within T1-S1).	Smoker or Vaper.
SCI group only: >12 months post-injury and no history of PUs.	

157

158 As identified by the “Guidelines for the conduct of clinical trials for spinal cord injury as
 159 developed by the ICCP Panel” [30], inclusion and exclusion criteria for SCI participants should
 160 consider the confounding effects of various independent variables such as pre-existing or
 161 concomitant medical conditions, other medications, surgical interventions, and rehabilitation
 162 regimens. As it may not be practical or justifiable to limit study enrolment based on factors
 163 such as e.g., rehabilitation regime or sex, all potentially confounding factors will be
 164 comprehensively recorded and considered as potential co-variables in primary and secondary
 165 data analyses.

166 **Experimental procedures**

167 Once screened and recruited, participants will be invited to their pseudo-randomly allocated
 168 experimental session [31]. They will come to the laboratory wearing comfortable, loose-fitting
 169 attire. Upon arrival, participants will be seated whilst they acclimatise to the ambient conditions

170 of the laboratory (22-24 °C; 50% RH) before recording height and body mass (Model 874;
171 Seca GmbH, Hamburg, Germany).

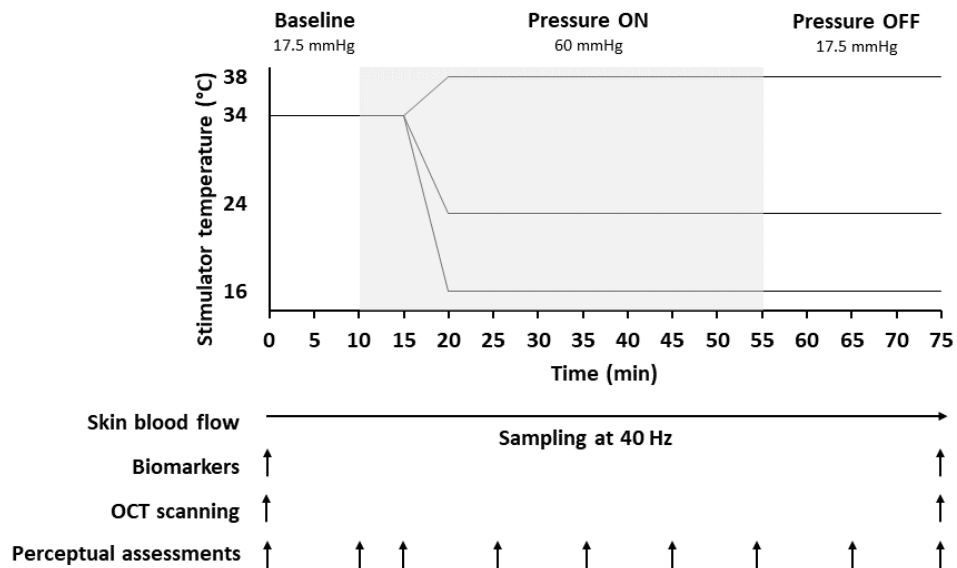
172 Following the pre-experimental checks, participants will lie down in the prone position on a
173 hospital bed. Care will be taken to ensure that participants are as comfortable as possible by
174 providing pillows as necessary to support the pelvic region and upper body, given the length
175 of time (75 minutes) they will need to remain in this position. The addition of the pillows also
176 serves to support the lumbar region by flattening the sacroiliac joint and reducing any
177 pronounced lordosis of the spine while lying in a prone position.

178 First, structural, and functional imaging of the skin of the sacral skin using Optical Coherence
179 Tomography (OCT) will be performed. This will be followed by sampling of skin sebum at the
180 sacrum for subsequent biomarker analysis, using an established methodology involving a 2-
181 minute application time and tweezer extraction to avoid cross-contamination [32]. The
182 investigator will then place a custom-built thermal probe over the sacrum and load it with a
183 weight (see below for details) to achieve pressure of 60 mmHg. The thermal probe consists of
184 a water-perfused set of Peltier elements, which provide local temperature control for a 36-cm²
185 plate. An optic fibre is integrated in the plate, flush to its surface, which allows for the
186 continuous monitoring of skin blood flow via Laser Doppler Flowmetry (LDF). The thermal
187 probe is mounted on a frame with an integrated strain gauge to estimate the force applied to
188 the stimulator. When applied to the skin, the integrated device allows for the manipulation of
189 local skin temperature (range: 0°C to >50°C; variable temperature rates under PID control),
190 applied pressure and the concurrent monitoring of blood flow.

191 To ensure consistent placement between sessions, probe placement will be marked with non-
192 permanent ink. To minimize edge loading, the thermal probe is equipped with a 3D printed
193 sleeve (thermoplastic polymer (Poly lactic acid)) and to ensure uniform pressure distribution

194 across the mechanically loaded sacrum an interface pressure mapping device will be used
 195 (ForeSiteSS, XSensor, Canada). The pressure mapping device comprises a 2*32 sensing array
 196 with a spatial resolution of 12.7mm, operating between 5-256mmHg, with an accuracy of \pm
 197 5%.

198 Finally, participants will be asked to provide subjective ratings of thermal sensation, comfort,
 199 and acceptability, using Likert scales (detailed below). At this point, the standardised protocol
 200 to cause pressure-induced ischemia and post-occlusive hyperaemia will commence (Figure. 2).



201

202 **Figure. 2. Standardise pressure protocol delivered over the sacrum.** The thermal probe will be used to deliver
 203 a standardised 60mmHg pressure protocol to evoke pronounced tissue ischaemia under 3 thermal conditions, i.e.
 204 a control skin temperature evoking no cooling (i.e. 38°C) and two cooling temperatures of 24°C and 16°C. During
 205 the protocol, a series of non-invasive measurements will be conducted [i.e. skin blood flow via LDF; inflammatory
 206 biomarker sampling from skin sebum; structural and functional imaging via Optical Coherence Tomography
 207 (OCT); perceptual assessment of subjective thermal sensation, comfort, and acceptance] at different time points
 208 (identified in the diagram by \uparrow).

209

210 The protocol consists of three phases; i) a 10-minute baseline stabilisation with minimal
 211 pressure (17.5 mmHg (2.3 kPa)), ii) 45-minute loading phase (60 mmHg/7.9 kPa [15]), and iii)
 212 a 20-minute minimal pressure phase (same as baseline). Skin blood flow will be measured
 213 continuously throughout the three phases of the protocol via Laser Doppler Flowmetry (Moor

214 Instruments, MoorVMS-LDF laser Doppler monitor, UK) with perceptual responses measured
215 at predetermined intervals during the protocol (Figure. 2). During the baseline phase in all
216 conditions, the temperature probe will be set to 34 °C. On starting the loading phase, the
217 thermal probe will be loaded with a 2 kg weight to elicit an equivalent pressure of 60mmHg
218 (7.9 kPa) at the probe interface. The initial 5-minutes of the loading phase will serve as a further
219 baseline measurement for the pressure protocol, to delineate the effects of 60mmHg loading
220 alone on microvascular responses. To this end, once the load is applied to the sacrum, the probe
221 temperature will remain at 34 °C. Following this, the temperature probe will be set to the target
222 temperature for the specific testing session, either 38 °C ($\Delta 13.7$ °C/min), 24 °C ($\Delta 11.3$ °C/min),
223 or 16 °C ($\Delta 9.2$ °C/min) and will be maintained at such temperature until the end of the session.
224 Upon completion of the loading phase, the thermal probe will be unloaded (Figure. 2) whilst
225 maintained in position over the sacrum, to allow skin tissue blood flow reperfusion (which will
226 be continuously monitored by via LDF). The minimal loading phase (comparable to the initial
227 10-minute baseline phase) will last for 20 minutes, after which the thermal probe will be
228 removed to allow for image acquisition via OCT and a final sampling of skin sebum over the
229 sacrum.

230 The sections below provide a detailed description of each measurement undertaken during the
231 protocol.

232 **Measurements**

233 **Skin blood flow**

234 Continuous skin blood flow will be monitored via Laser Doppler Flowmetry at the sacrum.
235 LDF is a non-invasive technique that uses optical probes to measure blood flow velocity in the
236 microvasculature. Typically, tissue thickness is sampled at 1mm implementing the Doppler
237 principle whereby light from a monochromatic laser becomes scattered from moving red blood

238 cells, allowing it to be applied to a wide range of anatomical locations [33]. Given its relative
239 low cost and ease of use, LDF is validated [34] and has been widely used to assess changes in
240 blood flow velocity (as an index of changes in flow) over bony prominences, such as the sacrum
241 [6, 13-15, 35].

242 The optical probe is integrated within the custom-built temperature probe, allowing concurrent
243 manipulation of skin temperature whilst monitoring real-time changes in skin blood flow,
244 during loaded and minimal loaded states. Blood flow during the loading phase will be analysed
245 via spectral analysis of wavelet frequency to investigate temperature-modulated regulatory
246 mechanisms during loading (i.e. changes in 0.1 Hz, 0.04 Hz, and 0.01 Hz frequencies will be
247 associated with myogenic activity of vascular smooth muscles, neurogenic activity of the vessel
248 wall, and vascular endothelium related metabolic activity, respectively) [11]. Blood flow
249 during the minimal loading phase will be used to calculate the baseline skin blood flow (taken
250 as the mean average during the final 3-minutes of the 10-minute baseline period). The average
251 baseline will then be used for normalisation of peak hyperaemia [(peak hyperaemia – average
252 baseline skin blood flow)/average baseline skin blood flow]*100] to investigate its modulation
253 via cooling. Changes in peak hyperaemia represent a robust and repeatable microvascular
254 response [36], which is directly implicated in the pathophysiology of PUs [36]. Secondary
255 perfusion parameters will include the time to peak hyperaemic response and the perfusion area
256 between the skin blood flow response curve and the mean baseline skin blood flow after
257 unloading and during the reperfusion phase. In the measurement of peak hyperaemia skin blood
258 flow will be collected continuously (40 Hz sampling rate) and averaged every 1-minute for the
259 analysis of the temporal dynamics of cooling induced changes in skin blood flow under loading.

260 **Biomarkers**

261 A preselected panel of pro- (IL-1 α , IL-1 β , TNF α , IL-6, IL-8, IFN γ) and anti-inflammatory
262 biomarkers (IL-1RA) will be extracted from skin sebum. Biomarker collection and extraction

263 techniques via application of Sebutape have been optimised in our laboratory [37], to ensure
264 both low abundance and high abundance proteins can be quantified. In brief, Sebutape will be
265 applied to the sacrum for 2-minutes before the samples are extracted using tweezers and a
266 gloved hand to avoid cross contamination. Stored samples will be coded and stored at -80°C
267 prior to analysis using standard ELISA plates for targeted proteins. The extraction of skin
268 inflammatory biomarkers will use chemical and mechanical stimuli to for maximal extraction
269 efficiency. Chemical extraction will involve 0.85 mL of extraction buffer, which consists of
270 PBS + 0.1% Dodecyl maltoside. The tapes will then be shaken with the buffer for 1 hour
271 followed by 5 minutes of sonication. A 0.35 mL aliquot will then be used for total protein
272 analysis. The remaining 0.5 mL will be centrifuged for 10 minutes at a speed of 15 000g at
273 4°C . The supernatants will be discarded and the remaining solution with the pellet briefly
274 vortexed and used for the immunoassay analysis, as prescribed by the manufacturer using MSD
275 U-Plex kits (MesoScale Diagnostics).

276 **Skin imaging**

277 Skin imaging via Optical Coherence Tomography using VivoSight® device with dynamic
278 OCT processing software (Michelson Diagnostics Ltd., Maidstone, Kent, UK) with a Class 1M
279 (EN 60825-1) laser source of near-infrared wavelength (1305 nm). A total of 120 images with
280 $50\ \mu\text{m}$ spacing will be acquired as a $6 \times 6 \times 2\ \text{mm}^3$ (width \times length \times depth) stack. This
281 technique is non-inferior to punch biopsy for skin characterization [38] and will allow to non-
282 invasively characterise the skin's epidermal (i.e. thickness, stratum corneum hydration,
283 collagen density) and blood perfusion properties (i.e. vascular plexus density and diameter)
284 prior to and following the thermomechanical manipulations. This evaluation will also be
285 relevant to model group differences in baseline skin anatomy and biophysics that may underlie
286 the differential effects of cooling with ageing and SCI. The OCT probe will be placed gently

287 on the skin, maintaining a static position during acquisition. Spacers at the probe interface will
288 be used to optimise the focal point of the epidermis during scanning.

289 **Perceptual assessments**

290 Perceptual assessments of participants' local thermal and comfort sensations will be assessed
291 via Likert scales, to establish time-dependent changes in subjective perceptions of cooling [39].
292 The Likert scales for thermal sensation, thermal comfort, and thermal acceptance were created
293 based on the recommendations of Schweiker et al., [40], i.e. using a ruler to draw a 100 mm
294 horizontal line the anchors were then spaced evenly along the line. Thermal sensation consisted
295 of a 7-point scale from 1 (cold) to 7 (hot) with 4 as neutral. Thermal comfort used a 5-point
296 scale ranging from 1 (comfortable) to 5 (extremely uncomfortable) and thermal acceptance
297 used a 4-point scale ranging from 1 (clearly acceptable) to 4 (clearly unacceptable). Perceptual
298 sampling will occur at pre-determine time points throughout the entire pressure protocol
299 (Figure. 2). This evaluation will establish ageing- and SCI-induced changes in peripheral
300 neurosensory function, as well as the relationship between the physiological and perceptual
301 effects of cooling during mechanical loading.

302 **Statistical analyses**

303 Data will be assessed for normality of distribution (Kolmogorov–Smirnov test), and then
304 analysed for the independent and interactive effects of pressure stimuli's temperature (i.e. 3
305 levels: 38, 24, 16 °C), and of participant group (i.e. 3 levels: young, older, SCI) on within- and
306 between-subject mean differences in peak hyperaemia (N=35 with 95 % Confidence Intervals).
307 This will be conducted using a 2-way mixed model ANOVA or Friedman, depending on the
308 data distribution (parametric or non-parametric, respectively). Post-hoc analyses will be
309 performed between pressure stimuli's temperatures and participant groups based on the
310 presence of main effects and using Tukey's test. The use of regression models will evaluate the
311 relationship between absolute cooling temperatures and peak hyperaemia during pressure

312 stimuli. This approach is designed to identify “physiological thresholds” for the protective
313 effects of cooling under mechanical loading. Mean thresholds will subsequently be determined
314 and their inter-individual variability across the cohort. For the perceptual data, thermal
315 discomfort will identify “Uncomfortable” on the Likert Scale as the threshold onset of
316 discomfort. Regression analyses will be used to interrogate the relationship between pressure
317 stimuli’s cooling temperature and local discomfort, for each participant. This approach will
318 also enable the assessment of “perceptual cooling thresholds” for cold-induced discomfort at
319 two absolute cold temperatures (i.e. 24 and 16 °C), and their individual variability.
320 Physiological and perceptual thresholds will be combined to produce common cooling
321 parameters that will accommodate different cooling characteristics. Regarding the secondary
322 experimental outcomes, these will be within- and between-subject changes in biomarkers’
323 expression, skin structural and biophysical properties (i.e. imaging parameters and skin
324 friction), and subjective thermal perceptions, as a function of pressure stimuli temperature (i.e.
325 3 levels: 38, 24, 16°C), time (i.e. varying levels depending on variables’ sampling rate), and of
326 participant group (i.e. 3 levels: young, older, SCI). Data will be first assessed for normality of
327 distribution (Kolmogorov–Smirnov test), and then analysed for the independent and interactive
328 effects of pressure and stimuli temperature, time, and participant group by means of a 3- way
329 mixed model ANOVA (or Friedman). Post-hoc analyses will be performed between pressure
330 stimuli temperatures, time, and participant groups based the presence of main effects and using
331 Tukey’s test. Group-related co-variables associated with sex, skin tone, and clinical status
332 (applicable to SCI participants only, e.g. rehabilitation status) will be considered in all analyses
333 to interpret the proportion of variance unexplained by the main effects (i.e. temperature, time,
334 and group) and their interactions.

335 **Data management**

336 Data management will be in line with the University of Southampton' policy on data quality,
337 which forms part of the University's Information Governance Framework and demonstrates
338 compliance with its obligations under the Data Protection Legislation. Therefore, the study will
339 comply with the requirements of the Data Protection Act 2018 and the University of
340 Southampton's Ethics Committee (ERGO) policies. This project involves human participants
341 and will be conducted in line with the University's Policy on the Ethical Conduct of Research
342 and Studies involving Human Participants, and the Medical Research Council's policies on
343 ethics and data sharing. Data will be fully anonymised at the earliest opportunity and before
344 being made available open access in the University's data repository. All data that supports
345 publications will be deposited and will be citable using a persistent identifier (DOI). Original
346 hardcopies of study documents (e.g. consent forms) will be stored securely for ten years from
347 completion of the project within a locked office at the University or scanned, encrypted and
348 securely stored on the University's IT system.

349 **Ethical considerations and declarations**

350 The project will involve testing healthy individuals aged 18 to 70 years, and those with a SCI,
351 and will be conducted in line with Southampton University Code of Practice for Research and
352 will comply with the *Declaration of Helsinki*. Participants will provide written informed
353 consent, and relevant personal information (e.g. skin's perceptual sensitivity to cooling). All
354 experiments will pose low risks to participants, and not greater than what they face in their
355 daily living (e.g. undergoing a GP examination of skin sensitivity); yet a set of mitigation
356 measures to manage these risks have been developed. For example, there is a risk of discomfort
357 from skin cooling. This will be akin to having a cold pack applied on the skin following an
358 injury. Mitigation measures such as on-going skin temperature monitoring, checks of
359 subjective wellbeing, and active skin re-warming, will be in place. The skin will also be

360 checked for blanching erythema. All research methods for evaluating physiological
361 thermoregulatory and vasomotor responses (e.g. recording of skin temperature via
362 thermocouple microsensors, skin blood flow via Laser Doppler Flowmetry) will be non-
363 invasive (e.g. sensors applied to participants' skin with hypoallergenic medical tape), and they
364 pose low risk.

365 Ethical approval for the stated measurements and procedures has been granted by the
366 University of Southampton's Ethics Committee (ERGO 88984).

367 **Status and timeline of the study**

368 At the time of publication pilot testing and technical development of the protocol have been
369 completed. Formal recruitment commenced on 16th January 2024. The project is being
370 supported by a Medical Council Research grant (MR/X019144/1) and has a lifespan of 42
371 months from March 2023.

372 **Discussion**

373 Skin damage, leading to PUs, can affect any individual who experience prolonged periods of
374 immobility, ranging from newborn babies to older adults. In addition, with the recent Covid-
375 19 pandemic, there has been an enormous increase in the number of hospitalized patients with
376 novel respiratory diseases and the associated healthcare management team, who have
377 developed skin damage from prolonged use of personal protective medical devices [41]. Thus,
378 the proposed research which addresses the physiological tolerance of human skin to prolonged
379 mechanical loading is both important in improving scientific knowledge and timely to societal
380 demands.

381 **Strengths of the planned study**

382 A purposefully sampled group of human volunteers has the unique advantage of targeting
383 patient-relevant, physiological, and perceptual mechanisms, which would otherwise be

384 inaccessible via in vitro skin constructs or in vivo animal models. Animal studies involving
385 mice and porcine models of mechanically induced PU formation have been developed to
386 investigate how the temperature of loaded skin modulates skin damage [9, 35]. Similarly, in
387 vitro skin models have been used to investigate drug delivery for wound healing [42]. Yet, the
388 translational value of these animal and in vitro models to human participants is limited. This is
389 because differences exist in skin morpho-physiology, immunology, genetics, and
390 thermoregulatory control amongst these models [43]. Importantly, these previous approaches
391 do not allow for the evaluation of the perceptual effects of thermal interventions on loaded
392 skin, critical when considering the adoption in different care settings. Localised skin cooling
393 induces cold discomfort [44], which greatly limits acceptability and adherence to therapeutic
394 interventions designed to maintain skin health, particularly for vulnerable individuals such as
395 the elderly [18]. Hence, a human-centric approach can help identifying optimal levels of
396 cooling that can provide a physiological effect within the acceptable range of perceived
397 comfort. This is critical to develop “user-centred” therapeutic approaches that are both effective
398 and comfortable.

399 Some animal data indicates that in association with reducing local metabolic demands,
400 reducing skin temperature during applied mechanical loading could preserve metabolic and
401 myogenic components of skin blood flow [11]. This would protect the skin against pressure
402 induced ischemia and reduce the potential of tissue necrosis. Local cooling could also reduce
403 the magnitude of reactive hyperaemia following a period of pressure-induced ischaemia [6].
404 Reactive hyperaemia can increase the risk of ischemia reperfusion injury by triggering the
405 release of oxygen-derived free radicals with cytotoxic effects, and this can play an equivalent
406 role in the pathophysiology of PUs as sustained pressure [7]. This has been demonstrated with
407 cooling of cardiac tissues to protect from ischemia-reperfusion injury in animal models [45].

408 This project will generate novel insights on temperature-modulated skin tolerance in vivo,
409 which will be relevant to skin physiologists, bioengineers, and clinicians such as dermatologists
410 and intensive care nurses, to better understand the physiological processes and the potential
411 benefits of cooling strategies to minimise the individual PU risk in a clinical setting.

412 Establishing the physiological and perceptual relationships between cooling and skin tolerance
413 to pressure will help inform the design of public health interventions to protect vulnerable
414 groups at risk of PUs such as the elderly. The research will improve quality of life in individuals
415 who are at risk of PUs. Improved therapeutic interventions will reduce discomfort and lower
416 the incidence of injuries, thus reducing the financial burden on healthcare providers (cost for
417 NHS wound care ~£8Bn/yr). In addition, effective technologies which provide cooling can be
418 maintained in-situ for prolonged periods, decreasing the demand for repeat interventions.
419 Applications of the physiological and perceptual thresholds will inform the design of user-
420 centred medical devices and wearables, including support surfaces and garments delivering
421 cooling to the skin at a level and rate that is both beneficial and comfortable. This knowledge
422 will be relevant to material and textile engineers engaged in the design of healthcare and
423 medical device products.

424 **Limitations**

425 The study has been robustly designed but is not without limitation, which relates to the
426 measurement of skin blood flow perfusion and reactive hyperaemia. The measurement of blood
427 perfusion using LDF provides high temporal resolution, however; limited spatial resolution is
428 offered [46], and the optical probe must be placed directly over the area of interest to detect
429 blood flow changes. Thus, the surface area of the optical probe in contact with the skin is
430 limited by the probe size (which is smaller than the thermally stimulated area. Using a single
431 probe could also present limitations due to potential high variability due to a lack of
432 homogeneity in skin morphology and inter-variability of participant anatomy, although single

433 site LDF measurements in the forearm has been shown to be reliable [47]. Future studies should
434 therefore consider methodological advancement to facilitate LDF assessments at multiple sites
435 under concurrent thermal and mechanical stimulation.

436 **Conclusions**

437 We have designed a clinically relevant set of experiments in healthy young participants and in
438 groups at-risk of PUs, to determine how different levels of cooling alter the skin' microvascular,
439 inflammatory, structural, and perceptual responses to a) sustained pressure induced ischemia;
440 b) post-occlusive reactive hyperaemia. The outcomes of this project will help identifying the
441 metabolic, immunological, biophysical, and perceptual pathways underlying the potential
442 beneficial effects of cooling on skin tolerance to loading in distinct cohorts to fundamentally
443 change our understanding of normal and pathological skin function. This knowledge will be
444 translated to support innovation of assistive thermal technologies that maintain skin health
445 across the life course.

446 **References**

447

- 448 1. Kottner J, Cuddigan J, Carville K, Balzer K, Berlowitz D, Law S, et al. Prevention and
449 treatment of pressure ulcers/injuries: The protocol for the second update of the international
450 Clinical Practice Guideline 2019. *J Tissue Viability*. 2019;28(2):51-8.
- 451 2. Guest JF, Fuller GW, Vowden P. Cohort study evaluating the burden of wounds to the
452 UK's National Health Service in 2017/2018: update from 2012/2013. *BMJ Open*.
453 2020;10(12):e045253.
- 454 3. Coleman S, Nixon J, Keen J, Wilson L, McGinnis E, Dealey C, et al. A new pressure
455 ulcer conceptual framework. *J Adv Nurs*. 2014;70(10):2222-34.

- 456 4. Kottner J, Black J, Call E, Gefen A, Santamaria N. Microclimate: A critical review in
457 the context of pressure ulcer prevention. *Clin Biomech (Bristol, Avon)*. 2018;59:62-70.
- 458 5. Worsley PR, Crielaard H, Oomens CWJ, Bader DL. An evaluation of dermal
459 microcirculatory occlusion under repeated mechanical loads: Implication of lymphatic
460 impairment in pressure ulcers. *Microcirculation*. 2020;27(7):e12645.
- 461 6. Tzen YT, Brienza DM, Karg PE, Loughlin PJ. Effectiveness of local cooling for
462 enhancing tissue ischemia tolerance in people with spinal cord injury. *J Spinal Cord Med*.
463 2013;36(4):357-64.
- 464 7. Tsuji S, Ichioka S, Sekiya N, Nakatsuka T. Analysis of ischemia-reperfusion injury in
465 a microcirculatory model of pressure ulcers. *Wound Repair Regen*. 2005;13(2):209-15.
- 466 8. Clark M. Microclimate: Rediscovering an Old Concept in the Aetiology of Pressure
467 Ulcers. In: Romanelli M, Clark M, Gefen A, Ciprandi G, editors. *Science and Practice of*
468 *Pressure Ulcer Management*. London: Springer London; 2018. p. 103-10.
- 469 9. Kokate JY, Leland KJ, Held AM, Hansen GL, Kveen GL, Johnson BA, et al.
470 Temperature-modulated pressure ulcers: a porcine model. *Arch Phys Med Rehabil*.
471 1995;76(7):666-73.
- 472 10. Iazzo PaKGLaKJYaLKJaHGaSE. Prevention of pressure ulcers by focal cooling:
473 Histological assessment in a porcine model. *Wounds*. 1995;7:161-9.
- 474 11. Jan YK, Lee B, Liao F, Foreman RD. Local cooling reduces skin ischemia under
475 surface pressure in rats: an assessment by wavelet analysis of laser Doppler blood flow
476 oscillations. *Physiol Meas*. 2012;33(10):1733-45.
- 477 12. Bernard Lee and Siribhinya Benyajati and Jeffrey AWaY-KJ. Effect of local cooling
478 on pro-inflammatory cytokines and blood flow of the skin under surface pressure in rats:
479 Feasibility study. *Journal of Tissue Viability*. 2014;23(2):69-77.

- 480 13. Lachenbruch C. Skin cooling surfaces: estimating the importance of limiting skin
481 temperature. *Ostomy Wound Manage.* 2005;51(2):70-9.
- 482 14. Bergstrand S, Lindberg LG, Ek A-C, Lindén M, Lindgren M. Blood flow measurements
483 at different depths using photoplethysmography and laser Doppler techniques. *Skin Research
484 and Technology.* 2009;15(2):139-47.
- 485 15. Tzen YT, Brienza DM, Karg P, Loughlin P. Effects of local cooling on sacral skin
486 perfusion response to pressure: implications for pressure ulcer prevention. *J Tissue Viability.*
487 2010;19(3):86-97.
- 488 16. Lee B, Benyajati S, Woods JA, Jan YK. Effect of local cooling on pro-inflammatory
489 cytokines and blood flow of the skin under surface pressure in rats: feasibility study. *J Tissue
490 Viability.* 2014;23(2):69-77.
- 491 17. Wang W, Wang H, Zhao Z, Huang X, Xiong H, Mei Z. Thymol activates TRPM8-
492 mediated Ca(2+) influx for its antipruritic effects and alleviates inflammatory response in
493 Imiquimod-induced mice. *Toxicol Appl Pharmacol.* 2020;407:115247.
- 494 18. Ledger L, Worsley P, Hope J, Schoonhoven L. Patient involvement in pressure ulcer
495 prevention and adherence to prevention strategies: An integrative review. *Int J Nurs Stud.*
496 2020;101:103449.
- 497 19. Filingeri D, Redortier B, Hodder S, Havenith G. Thermal and tactile interactions in the
498 perception of local skin wetness at rest and during exercise in thermo-neutral and warm
499 environments. *Neuroscience.* 2014;258:121-30.
- 500 20. Worsley PR, Bader DL. A modified evaluation of spacer fabric and airflow
501 technologies for controlling the microclimate at the loaded support interface. *Textile Research
502 Journal.* 2019;89(11):2154-62.

- 503 21. van Leen M, Halfens R, Schols J. Preventive Effect of a Microclimate-Regulating
504 System on Pressure Ulcer Development: A Prospective, Randomized Controlled Trial in Dutch
505 Nursing Homes. *Adv Skin Wound Care*. 2018;31(1):1-5.
- 506 22. Denzinger M, Rothenberger J, Held M, Joss L, Ehnert S, Kolbensschlag J, et al. A
507 quantitative study of transepidermal water loss (TEWL) on conventional and microclimate
508 management capable mattresses and hospital beds. *J Tissue Viability*. 2019;28(4):194-9.
- 509 23. Holbrook KA, Odland GF. Regional differences in the thickness (cell layers) of the
510 human stratum corneum: an ultrastructural analysis. *J Invest Dermatol*. 1974;62(4):415-22.
- 511 24. Johnson JM, Minson CT, Kellogg DL, Jr. Cutaneous vasodilator and vasoconstrictor
512 mechanisms in temperature regulation. *Compr Physiol*. 2014;4(1):33-89.
- 513 25. Forsyth P, Miller J, Pumpa K, Thompson KG, Jay O. Independent Influence of Spinal
514 Cord Injury Level on Thermoregulation during Exercise. *Med Sci Sports Exerc*.
515 2019;51(8):1710-9.
- 516 26. Zeilig G, Enosh S, Rubin-Asher D, Lehr B, Defrin R. The nature and course of sensory
517 changes following spinal cord injury: predictive properties and implications on the mechanism
518 of central pain. *Brain*. 2012;135(Pt 2):418-30.
- 519 27. Lorenzo S, Minson CT. Human cutaneous reactive hyperaemia: role of BKCa channels
520 and sensory nerves. *J Physiol*. 2007;585(Pt 1):295-303.
- 521 28. De Jong RH, Hershey WN, Wagman IH. Nerve conduction velocity during
522 hypothermia in man. *Anesthesiology*. 1966;27(6):805-10.
- 523 29. Fitzpatrick, Thomas. Soleil et peau [Sun and skin]. *J Méd Esthétique*. 1975;2:33-4.
- 524 30. Fawcett JW, Curt A, Steeves JD, Coleman WP, Tuszynski MH, Lammertse D, et al.
525 Guidelines for the conduct of clinical trials for spinal cord injury as developed by the ICCP
526 panel: spontaneous recovery after spinal cord injury and statistical power needed for
527 therapeutic clinical trials. *Spinal Cord*. 2007;45(3):190-205.

- 528 31. Urbaniak GC, Plous S. Resaerch Randomizer (Version 4.0) [Computer software] 2013
529 [Available from: <http://www.randomizer.org>.
- 530 32. Jayabal H, Bader DL, Worsley P. Development of an Efficient Extraction Methodology
531 to Analyse Potential Inflammatory Biomarkers from Sebum. *Skin Pharmacol Physiol.*
532 2023;36(1):38-50.
- 533 33. Humeau A, Steenbergen W, Nilsson H, Strömberg T. Laser Doppler perfusion
534 monitoring and imaging: novel approaches. *Med Biol Eng Comput.* 2007;45(5):421-35.
- 535 34. Wright CI, Kroner CI, Draijer R. Non-invasive methods and stimuli for evaluating the
536 skin's microcirculation. *J Pharmacol Toxicol Methods.* 2006;54(1):1-25.
- 537 35. Lachenbruch C, Tzen YT, Brienza DM, Karg PE, Lachenbruch PA. The relative
538 contributions of interface pressure, shear stress, and temperature on tissue ischemia: a cross-
539 sectional pilot study. *Ostomy Wound Manage.* 2013;59(3):25-34.
- 540 36. Hoogendoorn I, Reenalda J, Koopman B, Rietman JS. The effect of pressure and shear
541 on tissue viability of human skin in relation to the development of pressure ulcers: a systematic
542 review. *J Tissue Viability.* 2017;26(3):157-71.
- 543 37. Soetens JFJ, Worsley PR, Bader DL, Oomens CWJ. Investigating the influence of
544 intermittent and continuous mechanical loading on skin through non-invasive sampling of IL-
545 1 α . *J Tissue Viability.* 2019;28(1):1-6.
- 546 38. Adan F, Nelemans PJ, Essers BAB, Brinkhuizen T, Dodemont SRP, Kessels J, et al.
547 Optical coherence tomography versus punch biopsy for diagnosis of basal cell carcinoma: a
548 multicentre, randomised, non-inferiority trial. *Lancet Oncol.* 2022;23(8):1087-96.
- 549 39. Typolt O, Filingeri D. Evidence for the involvement of peripheral cold-sensitive
550 TRPM8 channels in human cutaneous hygro-sensation. *Am J Physiol Regul Integr Comp*
551 *Physiol.* 2020;318(3):R579-r89.

- 552 40. Schweiker M, André M, Al-Atrash F, Al-Khatiri H, Alprianti RR, Alsaad H, et al.
553 Evaluating assumptions of scales for subjective assessment of thermal environments – Do
554 laypersons perceive them the way, we researchers believe? *Energy and Buildings*.
555 2020;211:109761.
- 556 41. Évora SA, Abiakam N, Jayabal H, Worsley PR, Zhang Z, S AJ, et al. Characterisation
557 of superficial corneocytes in skin areas of the face exposed to prolonged usage of respirators
558 by healthcare professionals during COVID-19 pandemic. *J Tissue Viability*. 2023;32(2):305-
559 13.
- 560 42. Ud-Din S, Bayat A. Non-animal models of wound healing in cutaneous repair: In silico,
561 in vitro, ex vivo, and in vivo models of wounds and scars in human skin. *Wound Repair Regen*.
562 2017;25(2):164-76.
- 563 43. Zomer HD, Trentin AG. Skin wound healing in humans and mice: Challenges in
564 translational research. *J Dermatol Sci*. 2018;90(1):3-12.
- 565 44. Cotter JD, Taylor NA. The distribution of cutaneous sudomotor and alliesthesial
566 thermosensitivity in mildly heat-stressed humans: an open-loop approach. *The Journal of*
567 *physiology*. 2005;565:335-45.
- 568 45. Olivecrona GK, Götberg M, Harnek J, Van der Pals J, Erlinge D. Mild hypothermia
569 reduces cardiac post-ischemic reactive hyperemia. *BMC Cardiovasc Disord*. 2007;7:5.
- 570 46. Strömberg T, Sjöberg F, Bergstrand S. Temporal and spatiotemporal variability in
571 comprehensive forearm skin microcirculation assessment during occlusion protocols.
572 *Microvasc Res*. 2017;113:50-5.
- 573 47. Roustit M, Blaise S, Millet C, Cracowski JL. Reproducibility and methodological
574 issues of skin post-occlusive and thermal hyperemia assessed by single-point laser Doppler
575 flowmetry. *Microvasc Res*. 2010;79(2):102-8.

576