- 1 Precision medicine for pandemics: stratification of COVID-19 molecular
- 2 phenotypes defined by topological analysis of global blood gene expression.
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Abstract:

Precision medicine offers a promising avenue for better therapeutic responses to pandemics such as COVID-19. This study leverages independent patient cohorts in Florence and Liège gathered under the umbrella of the DRAGON consortium for the stratification of molecular phenotypes associated with COVID-19 using topological analysis of global blood gene expression. Whole blood from 173 patients was collected and RNA was sequenced on the Novaseq platform. Molecular phenotypes were defined through topological analysis of gene expression relative to the biological network using the TopMD algorithm. The two cohorts from Florence and Liège allowed for independent validation of the findings in this study. Clustering of the topological maps of differential pathway activation revealed three distinct molecular phenotypes of COVID-19 in the Florence patient cohort, which were also observed in the Liège cohort.

Cluster 1 was characterised by high activation of pathways associated with ESC pluripotency, NRF2, and TGF- $\beta$  receptor signalling. Cluster 2 displayed high activation of pathways including focal adhesion-PI3K-Akt-mTOR signalling and type I interferon induction and signalling, while Cluster 3 exhibited low IRF7-related pathway activation. TopMD was also used with the Drug-Gene Interaction Database (DGldb), revealing pharmaceutical interventions targeting mechanisms across multiple phenotypes and individuals.

The data illustrates the utility of molecular phenotyping from topological analysis of blood gene expression, and holds promise for informing personalised therapeutic strategies not only for COVID-19 but also for Disease X. Its potential transferability across multiple diseases highlights the value in pandemic response efforts, offering insights before large-scale clinical studies are initiated.

### Introduction:

The ongoing challenges of COVID-19, triggered by the emergence of SARS-CoV-2, necessitate a detailed understanding of disease heterogeneity. Despite extensive research characterising the host response to SARS-CoV-2 through pre-clinical (1, 2), and clinical (3-6) functional genomic data, there have been limited approaches that have used data from and encompassed the range of symptom severity, disease heterogeneity and delivered personalised medicine.

Examination of gene expression patterns in blood has been used in previous studies to identify molecular phenotypes associated with different disease profiles in several emerging viral infections including Ebola virus (EBOV) (7) and SARS-CoV-2 (1, 2, 4, 5), as well as more endemic infections such as influenza virus (8). Medical countermeasures focus on either reducing viral load through anti-virals. These target viral biology or modulate the host response to infection to reduce sequalae such as inflammation. For many viruses there is a clear correlation between viral load, disease severity and outcome (survival/death). This is best typified by the Ebola virus where low viral loads correlate with survival and high viral loads correlate with death (9). For SARS-CoV-2 this correlation is less obvious. In animal models of disease, such as the ferret, viral load was correlated with symptomology (10); in humans, there is less data to support an association between viral load and disease. However, studies have shown that severe COVID-19 is associated with dysregulated immune pathology in organs such as the lungs and the respiratory tract (3, 11).

With any emerging viral pathogen, direct acting antivirals take time to develop and trial. Identifying therapeutics that can modulate the host response to reduce symptomology remain a priority. Being able to rapidly characterise aberrations in host pathways that lead to disease and marrying this with therapeutics on the FDA approved list will enhance pandemic preparedness and rapid response. Therefore, a deeper understanding of the host response can be used to guide the selection of host directed medication countermeasures.

The field of digital health and precision medicine is rapidly evolving, with emerging technologies and initiatives aimed at integrating diverse datasets to inform clinical decision-making. In this study we offer a novel way to analyse complex data collected by the DRAGON international consortium which enables rapid identification of targets for treatment by novel and/or re-purposed drugs. Within DRAGON, efforts have been made to harmonise data in digital healthcare, proposing guidelines for the integration of clinical data from various modalities. (12). Additionally, an online platform has been developed to host validated COVID-19 predictive models, facilitating their utilisation by clinicians in real-time decision-making (13). However, challenges persist, as evidenced by the limited success of outcome prediction models for COVID-19 patients based on demographic and comorbidity data, which highlights the need for more sophisticated approaches (14).

While omics data has been instrumental in advancing our understanding of SARS-CoV-2 and COVID-19, its integration into digital health platforms for clinical decisionmaking remains limited (15-17). Traditional molecular phenotyping approaches often provide only shallow insights. In previous work, using topological analysis, we demonstrated how gene expression data derived from whole blood at the time of admission could predict ICU admission (5). However, the current study analysed the blood transcriptomes of patients with COVID-19 as part of the DRAGON-EU consortium and used TopMD, an algorithm that considers all available data across a landscape of pathways, to characterize molecular phenotypes of COVID-19 patients admitted to hospital. Pathways were identified that correlated with clinical disease in the patient cohort. TopMD mapped pathways onto a database containing information on FDA approved drugs and their known gene and pathway interactions to generate a list of potential therapeutics for modulating severe COVID-19. The ability to rapidly identify and therapeutically modulate host pathways responsible for disease with preexisting medical countermeasures will be important in the emergence of novel diseases and future pandemics.

This study describes an analysis of the blood transcriptomes of patients with COVID-19 admitted to hospital in Liège and Florence between February and July 2021, as part of the DRAGON-EU consortium. Alongside collecting blood samples, demographic and clinical observations were recorded; additionally, CT scan data were

obtained for a subset of these patients. We applied an unsupervised approach, in which we characterised the molecular phenotypes of patients within this cohort. We have previously reported the development of a gene signature in patients with COVID-19, predictive of admission to ICU (5). This predictive signature revealed the activation of pathways regulating epidermal growth factor receptor (EGFR) signalling, peroxisome proliferator-activated receptor alpha (PPAR- $\alpha$ ) signalling and transforming growth factor beta (TGF- $\beta$ ) signalling. The observed molecular phenotype aligns with the mechanisms implicated in pulmonary fibrosis, which is also associated with increased severity of disease (18-20).

Methods:

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# Study population and sample collection and ethics

Blood samples were obtained from 132 patients severe enough to require admission because of COVID-19 at Careggi University Hospital, Florence, Italy, and 41 from a pre-defined, separate patient cohort in Liège, between February and July 2021. All patients tested positive nasopharyngeal swab PCR for SARS-CoV-2 infection. Blood samples were collected on Day 0 of hospital admission. The protocol was approved by the ethics committee of the University Hospital of Liège (reference number 2021/89) and the ethics committee of the UNIFI (#18085/OSS). Informed consent was obtained for every participant.

### **Ethical Approval statement**

- The work described has been carried out in accordance with The Code of Ethics of the World Medical Association (Declaration of Helsinki) for experiments involving humans. All procedures were performed in compliance with relevant laws and institutional guidelines and have been approved by the appropriate institutional
- 179 committees. Informed consent was obtained for every participant.
- 180 Clinical data were collected from the patients' electronic medical records by the 181 investigators, and included age, sex, BMI, comorbid conditions etc. The data were 182 then assembled using the Study Data Tabulation Model (SDTM) data format
- developed by the Clinical Data Interchange Standards Consortium (CDISC).

# **Chest CT analysis**

- Out of the 173 patients with RNA sequencing data, chest CT data was obtained from
- 186 109 patients using a 128-detector multislice Spiral Computed Tomography (MSCT)
- 187 (Somatom Definition AS, Siemens Healthcare, Erlangen, Germany) applying the
- following parameters: current × exposure time 150 mAs, tube voltage 100 kV, rotation
- 189 time 0.3 s, pitch 1.2 mm, pixel size 0.465 mm, beam collimation 128 × 0.6 mm, both
- slice thickness and reconstruction 1 mm, and reconstruction kernel Bf70 very sharp.
- 191 Axial images were carried out from lung apexes to bases with patient at full inspiration
- 192 mand breath hold. Post-processing, 1-mm-thick sections were reconstructed on
- 193 coronal and sagittal planes oriented on the tracheal plane. Intravenous contrast

medium was not administered. Chest CT images were displayed on a 24-inch medical monitor with a 3-megapixel Barco display (Barco, Kortrijk, Belgium) and 2048 x 1536 resolution. The software programs originally implemented to MSCT were used for image assessment. Images of each patient were evaluated for scan quality considering inspiratory level and motion artifacts. Data pulled out from CT examinations included CO-RADS, chest CT score, dominant pattern, and typical/atypical findings. Specifically:

### CO-RADS

CO-RADS score based on COVID-19 lung involvement and variable from 1 to 5, with higher values reflecting a greater level of suspicion of COVID-19 infection with lung involvement. CO-RADS is a score used to diagnose COVID-19 and does not inevitably reproduce the severity of lung alterations. Low scores corresponded to CT examinations with alterations less likely related to COVID-19 infection. The 5-score CO-RADS scale is as follows: 1: very low level of suspicion; 2: low level of suspicion; 3: equivocal findings; 4: high level of suspicion; 5: very high level of suspicion.

### Chest CT score for lobe involvement

Ranging from 0 to 5, namely 0: 0%; 1: <5%; 2: 5-25%; 3: 26-50%; 4: 51-75%;

5: >75%.

### **Dominant chest CT pattern**

Evaluated in relation to the prevalent alterations among ground-glass opacities, consolidations, ground-glass opacities together with consolidations, crazy-paving, and reverse halo, as defined by the Fleischner Society.

#### **Dominant chest CT distribution**

Lower lobes, upper lobes, peripheral, bronchocentric, dorsal, or diffuse.

# **Additional COVID-19 related findings**

Represented by pleural thickening, vascular enlargement, subpleural sign, halo sign air, bubble sign, perilobular pattern, and subpleural sparing.

### Additional findings not typical for COVID-19

Represented by pleural effusion, pericardial effusion, lymphadenopathy,

cavitation, tree-in-bud, discrete small nodules, isolated lobar/segmental consolidation, atelectasis, and smooth interlobular septal thickening.

### **RNA** extraction

Total RNA was extracted from PAXgene BRT using the PAXgene Blood RNA Kit (PreAnalytix), according to the manufacturer's protocol. Extracted RNA was stored at -80°C until further use. Following the manufacturer's protocols, total RNA was used as input material into the QIAseq FastSelect–rRNA/Globin Kit (Qiagen) protocol to remove cytoplasmic and mitochondrial rRNA and globin mRNA with a fragmentation time of 7 or 15 minutes. Subsequently the NEBNext® Ultra™ II Directional RNA Library Prep Kit for Illumina® (New England Biolabs) was used to generate the RNA libraries, followed by 11 or 13 cycles of amplification and purification using AMPure XP beads. Each library was quantified using Qubit and the size distribution assessed using the Agilent 2100 Bioanalyser and the final libraries were pooled in equimolar ratios. Libraries were sequenced using 150 bp paired-end reads on an Illumina® NovaSeq 6000 (Illumina®, San Diego, USA).

### **Bioinformatics**

Raw fastq files were trimmed using fastp (21). Trimmed paired end sequencing reads were inputted into salmon (v1.5.2) using the -I A –validateMappings –SeqBias –gcBias parameters (22). Quant files generated with salmon were imported into RStudio (4.1.1) using tximport to infer gene expression (23). The edgeR package (3.34.1) was used to normalise and scale sequencing libraries (24). Sequencing reads are available under BioProject ID: PRJNA1085259 on Short Read Archive (SRA).

### Molecular phenotypes mapped by topological analysis

Molecular phenotypes were mapped by topological analysis, using TopMD to measure the shape of global gene expression relative to the biological network (TopMD Patent number GB202306368D0). TopMD works in the following way: The biological network used was an interaction network retrieved from the STRING database (25). The gene nodes of the biological network were assigned vertices according to the measured gene expression. The topological shape, or landscape, of this network is then measured by TopMD's algorithm, clustering differential gene expression hotspots, corresponding to modulated gene pathways. These pathways have 'volume'

comprising the sum of squared differential gene expression of clustered genes, where the most differentially activated pathways have the highest pathway topological volumes. The molecular phenotype is defined as the global profile of volumes of differential pathway activation.

Global biological network
 Annotate gene expression or/and other functional genomic data
 Landscape features are 'hotspots', representing pathway activation.
 Unbiased identification of TopMD Pathways named by GSEA.
 Molecular Phenotype defined as complement of TopMD

Biomarkers identified from within Activated Pathways.

Pathways.

# Drug interactions mapped by topological analysis

Due to the power of TopMD analysis we can group genes depending on their expression values, this means that for each average expression of any cluster of samples, and even on individual samples, we can extrapolate a tailored gene set of activated gene-groups for such expression. These gene-groups can be then compared to other gene sets, as in GSEA, as well against genes activated by specific drugs. To do so, we utilised the Drug-Gene Interaction Database (26) obtained using genes or gene products that are known or predicted to interact with drugs, and compared via a binomial distribution test, the probability that an overlap between such genes and a TopMD gene-group was random. This was measured using a p-value associated with binomial statistic, together with other measures, such as the (Bonferroni) adjusted p-value, a TopMD volume (combining volume of the shape with the statistical significance of the drug-group combination) as well as an activation value, sum of the Log2 fold-change of those genes belonging to both the drug associated gene set and the TopMD group.

### Regression

Regression analysis was carried out using a Logistic regression model with the following optimisation problem:

$$\min_w C \sum_{i=1}^n s_i \left( -y_i \log(\hat{p}(X_i)) - (1-y_i) \log(1-\hat{p}(X_i)) 
ight) + r(w)$$

Where X is the **pathway matrix** and y is the vector of the classification, 0 when the ith sample is in the class considered and 1 otherwise. We considered a regularisation parameter C value of 1. For the penalisation term r(w) for the regression weights w, we considered an ElasticNet penalisation with the **I1 ratio** parameter value of 0.5

$$rac{1-
ho}{2}w^Tw+
ho\|w\|_1$$

The probability the i-th sample with **pathways values** equal to Xi is then:

$$\hat{p}(X_i) = \operatorname{expit}(X_i w + w_0) = rac{1}{1 + \exp(-X_i w - w_0)}$$

- With w0 the intercept. The python module used was **scikit-learn (version 1.4.1)** and the algorithm used LogisticRegression function in the linear model submodule.
- We performed a 70/30 balanced split in the data from both cohorts separately (?), with 10 different splits. For each class we performed the regression based on a different number of pathways, from 1 to 20, ranked in each split separately by their pathway volume. For each regression model so obtained an average score of both training and test splits was carried and the best model was selected using a max-min approach, that is the best model was the one with highest value min(AUC on Train, AUC on Test), to avoid selecting models which were ill-performing on train splits, but instead for random effects very well on test splits.

### **Patient Clustering**

Pathway volumes were plotted on a PCA using PCAtools (v2.14.0), revealing 3 distinct clusters, confirmed by K-means clustering, based on pathway activation against healthy controls. The top ten (10%) of the PCA loadings were then extracted to identify

which pathways were driving cluster separation. To analyse differentially activated pathways between patient clusters, we calculated the average volume, across each cluster, of each pathway relative to the average of all the COVID-19 patients.

# Logistic Regression Receiver Operating Characteristic (LRROC) analysis using patient clusters derived from the patient pathway volume matrix.

The area under the ROC curve (AUC) is a measure of the model's ability to distinguish between classes. A higher AUC indicates better discrimination and, consequently, stronger patient clusters. LRROC for Florence Patients: LRROC analysis was performed exclusively for Florence patients. The patient pathway volume matrix for Florence patients was utilized to train the LRROC model. The output consisted of a Receiver Operating Characteristic (ROC) curve, which depicted the classification performance of patient clusters based on pathway volume. To evaluate the model's generalization capability, the dataset was split into training and testing sets, and separate ROC curves were generated for each.

Validation of clusters for Liège Patients: The LRROC model trained on Florence patients was validated on Liège patients' data. Using the trained model, an additional ROC curve was generated solely for Liège patients to assess the model's performance in classifying Liège patient clusters based on pathway volume.

### Integration into digital health platform

As a proof of concept, transcriptomics data and TopMD analysis were integrated with a healthcare platform ran by Comunicare (27). This was to highlight the possibilities of integrating omics data into healthcare and digital health platforms. Similar regression analysis of COVID-19 blood transcriptomes, predicting ICU admission, performed within the DRAGON scope (5) generated a linear model which is currently used to generate prediction scores between 0 and 1, using TopMD analysis of each sample submitted. In this way we can present TopMD analysis of individual samples compared to a healthy baseline, which includes pathway activation information, together with a similarity score to the ICU admitted average patient we extracted from previous data.

### Results:

To investigate whether blood transcriptomic analysis coupled with a machine learning approach underpinned by TopMD could be integrated with clinical data, RNA sequencing was performed on peripheral blood obtained from 173 patients from Liège (n=41) and Florence (n=132) gathered under the auspices of the DRAGON consortium. A summary of the patient characteristics is described in Supplementary Table 1. Within this cohort ten patients had fatal disease. As no outcome variables within this cohort had power, an unsupervised approach was undertaken. Out of the 173 patients, 109 patients had matched CT data scored by clinicians. The data is summarised in Supplementary Table 2. The majority of patients had a CORADS score of high and very high, where 26% was equivocal, 4.6% low and 2.8% very low. The CORADS score stands for "COVID-19 Reporting and Data System," which is a classification system used in radiology to assess the likelihood of COVID-19 infection based on chest imaging findings, typically on computed tomography (CT) scans. The score categorizes imaging findings into different levels of suspicion for COVID-19, ranging from very low to very high.

Table 1: Characteristics of 132 patients from Florence included in the study, including lab results at admission.

N	N = 132 <sup>1</sup>
132	
	127 (96%)
	5 (3.8%)
132	60 (50, 68)
132	
	40 (30%)
	92 (70%)
132	
	132 132 132

N		123 (93%)
Υ		9 (6.8%)
Continuous positive airway pressure	132	
N		129 (98%)
Υ		3 (2.3%)
Tracheostomy	132	
N		131 (99%)
Υ		1 (0.8%)
High flow nasal cannula oxygen therapy	132	
N		105 (80%)
Υ		27 (20%)
Hypertension	132	
N		77 (58%)
Υ		55 (42%)
Malnutrition	132	
N		131 (99%)
Υ		1 (0.8%)
Cardiovascular disease	132	
N		119 (90%)
Υ		13 (9.8%)
Respiratory disease	132	
N		118 (89%)
Υ		14 (11%)

Cancer	132	
N		118 (89%)
Υ		14 (11%)
Chronic kidney disease	132	
N		130 (98%)
Υ		2 (1.5%)
Chronic hepatitis	132	
N		130 (98%)
Υ		2 (1.5%)
Cerebrovascular disease	132	
N		125 (95%)
Υ		7 (5.3%)
Chronic hematologic disease	132	
N		129 (98%)
Υ		3 (2.3%)
Diastolic blood pressure (mmHg)	132	79 (70, 85)
Heart rate (BPM)	130	80 (75, 89)
Systolic blood pressure (mmHg)	132	125 (115, 140)
Temperature (°C)	131	36.50 (36.00, 37.20)
Weight (kg)	128	78 (70, 89)
Height (cm)	126	170 (165, 175)
Alanine aminotransferase (U/L)	128	27 (17, 39)
Aspartate aminotransferase (U/L)	64	31 (24, 46)

Bilirubin (mg/dL)	128	0.50 (0.30, 0.60)		
Calcium (mg/dL)	96 4.50 (4.34, 4.63)			
Creatinine (mg/dL)	130	0.83 (0.73, 0.95)		
D-dimer (ng/mL)	91	728 (429, 1,091)		
Direct bilirubin (mg/dL)	44	0.25 (0.17, 0.29)		
Fibrinogen (mg/dL)	117 572 (4			
Fraction of inspired oxygen (%)	127	28 (21, 36)		
Hematocrit (%)	132	42.7 (39.7, 45.8)		
Lactate dehydrogenase (U/L)	118	297 (247, 359)		
Lactic acid (mg/dL)	102	9.0 (7.0, 11.9)		
Leukocytes (10 <sup>9</sup> /L)	132	6.2 (4.6, 7.7)		
Lymphocytes (10 <sup>9</sup> /L)	129	0.90 (0.68, 1.25)		
Neutrophils (10°/L)	129	4.67 (3.08, 6.16)		
Oxygen saturation (%)	109	96.10 (94.20, 97.70)		
Partial pressure oxygen (mmHg)	131	74 (65, 87)		
Partial pressure carbon dioxide (mmHg)	127	36.2 (34.0, 39.0)		
Platelets (10 <sup>9</sup> /L)	132	196 (156, 255)		
Potassium (mmol/L)	128	3.85 (3.50, 4.10)		
Procalcitonin (ug/L)	126	0.09 (0.06, 0.15)		
Prothrombin time (seconds)	127	13.00 (12.30, 13.70)		
Sodium (mmol/L)	129	137 (135, 140)		
Urea nitrogen (g/L)	64 30 (30, 50)			
<sup>1</sup> n (%); Median (IQR)				

# Patients form 3 clusters based on their pathway activation

The RNA sequencing data was used to derive gene expression data (mRNA identification and abundance) which was calculated using Salmon inferred with Tximport in R, where values were converted into log2 counts per million (cpm). TopMD was then employed to calculate the activation of pathways. To identify differences in pathway activation across the cohort, activation data was plotted on a PCA which revealed three distinct clusters of patients (Figure 1). The relationship between clinical observations, demographics and CT scan data in each cluster was explored, and the significant differences are reported in Table 3. Lactic acid was slightly higher in cluster 1 and 2 and lower in cluster 3. A higher proportion of respiratory disease was observed in cluster 2 and the fraction of inspired oxygen was also higher in this cluster. Direct bilirubin was also higher in cluster 2. The majority of those that died from COVID-19 were in cluster 2. CORADS scoring was unable to distinguish between the clusters at a molecular level.

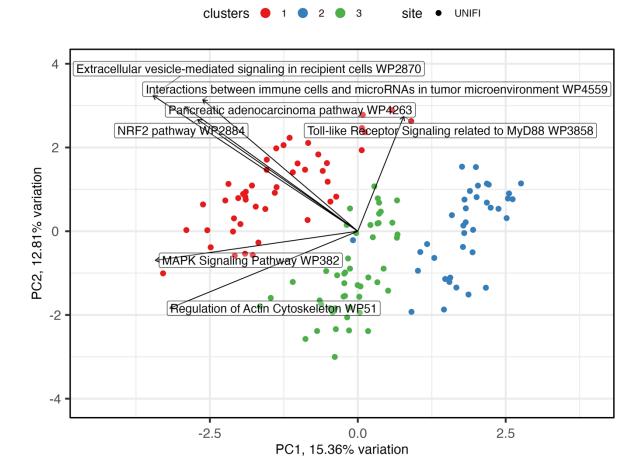


Figure 1:TopMD pathway volumes of each patient in the Florence cohort, calculated from a healthy plotted as a PCA plot. The data reveals three distinct clusters based on pathway activation determined by kmeans.

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Table 2: Patient characteristics that differ between the three clusters in the Florence cohort (p = <0.05).

Characteristic	N	1, N = $46^1$	<b>2</b> , N = $37^1$	3, $N = 49^1$	p-value <sup>2</sup>
Lactic acid (mg/dL)	102	10.0 (7.7, 13.0)	10.0 (7.2, 12.0)		0.008
Fraction of inspired oxygen (%)	127	28 (21, 36)	32 (27, 40)	28 (21, 29)	0.019
Died	132				0.032
N		46 (100%)	33 (89%)	48 (98%)	
Υ		0 (0%)	4 (11%)	1 (2.0%)	
Respiratory disease	132				0.042
N		44 (96%)	29 (78%)	45 (92%)	
Υ		2 (4.3%)	8 (22%)	4 (8.2%)	
Direct bilirubin (mg/dL)	44	0.20 (0.17, 0.27)	0.28 (0.24, 0.32)	0.20 (0.17, 0.28)	0.047

<sup>&</sup>lt;sup>1</sup> n (%); Median (IQR)

Molecular phenotype, Cluster 1, was characterised by high activation of pathways associated with ESC pluripotency, NRF2, and TGF-β receptor signalling (Figure 2). Molecular phenotype, Cluster 2 displayed high activation of pathways including focal adhesion-PI3K-Akt-mTOR signalling and type I interferon induction and signalling, while Cluster 3 exhibited low IRF7-related pathway activation.

<sup>&</sup>lt;sup>2</sup> Fisher's exact test; Kruskal-Wallis rank sum test; Pearson's Chi-squared test

LRROC analysis was conducted on models trained using 70% of patients from the Florence cohort, with test results evaluated on the remaining 30% of the Florence cohort. The area under the ROC curve (AUCROC) values were found to be 0.84, 0.85, and 0.72 for clusters Cluster 1, Cluster 2, and Cluster 3, respectively. Subsequently, these clusters were validated in the Liège cohort (Supplementary Figure 1), yielding AUCROC values of 0.76, 0.93, and 0.69 for Cluster 1, Cluster 2, and Cluster 3, respectively (Supplementary Figure 2).

# Potential drug candidates are identified for each cluster

To identify potential drug candidates that modulate pathways identified in these patient clusters, TopMD pathway activation was mapped onto the Drug-Gene Interaction Database (Figure 3). This mapping revealed distinct drug targets for each cluster, detailed in the supplementary table 4. This approach has a two-fold benefit: informing potential clinical trials and informing underlying biological mechanisms specific to each cluster. Interestingly, the pattern of pathway activation might also provide insight into the potential benefits or drawbacks of specific therapies, considering a drug's mechanism of action.

While all clusters shared targetable pathways led by genes such as ITGB2, GNAS, and CXCR2, unique targets also emerged. Cluster 1 specifically identified IFNAR1, TGFBR2, and CSF2RB, while cluster 2 added SERPING1 and TLN1. Notably, cluster 3 shared SERPING1 with cluster 2. These findings highlight both commonalities and variations in potential therapeutic targets across the identified patient clusters.

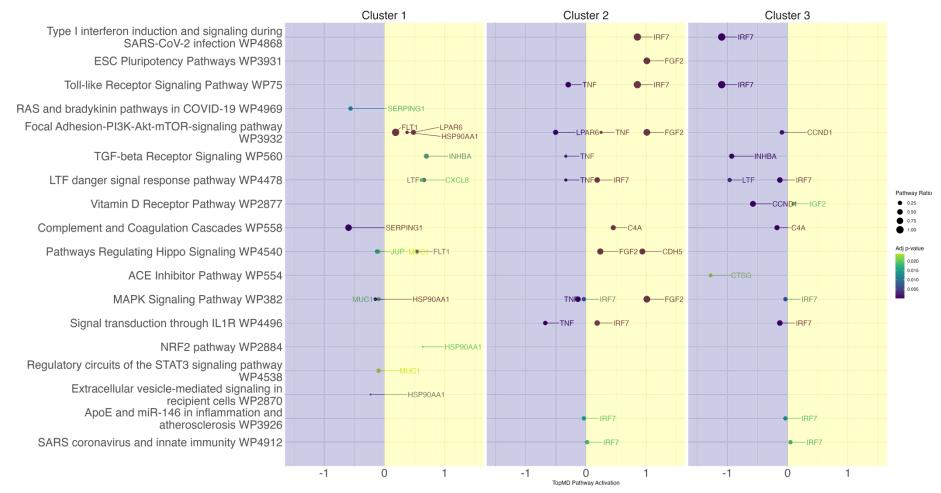


Figure 2: The average pathway volume for each cluster was considered in a TopMD enrichment analysis against the average pathway activation for the whole cohort to identify differentially activated pathways. The enrichment analysis was filtered by adjusted P value, then the top pathways were plotted. The pathways are annotated with the gene that leads the identified pathway. The dots are coloured by adjusted p-value and the size represents the proportion of genes identified within that pathway from TopMD analysis.

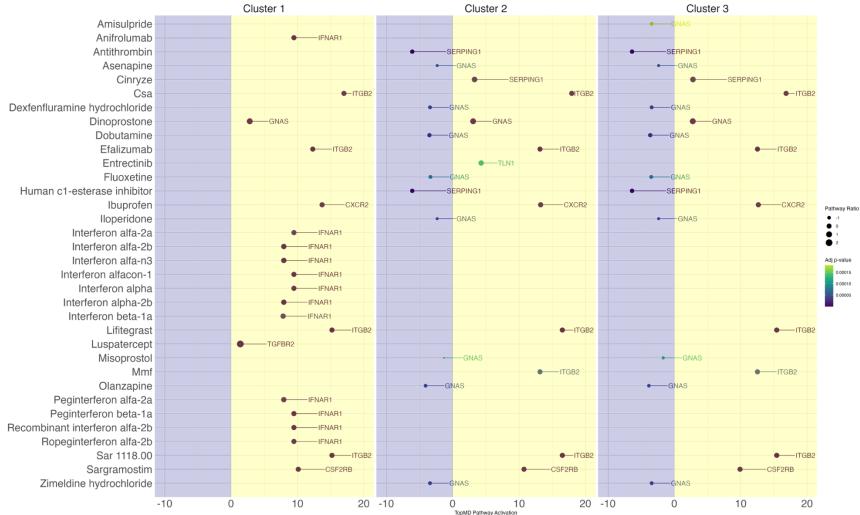


Figure 3: TopMD enrichment analysis was mapped against the Drug-Gene Interaction Database, using a healthy baseline, revealing approved drugs that are known to target genes and their corresponding pathways. The top drug candidates are plotted based on adjusted p-value and pathway volume.

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Identification of pathways in fatal cases where intervention might promote survival Due to limited sample size, we focused on the unsupervised analysis; however, to show utility of investigating pathway activity in individuals, pathway analysis in the 10 deceased patients from the Florence and Liège cohort were observed. Unsurprisingly. these patients exhibited advanced age and high comorbidity rates (cardiovascular: 70%, respiratory: 50%, malnutrition: 40%, hypertension: 90%, cerebrovascular: 30%, chronic hepatitis: 40%). Interestingly, all 10 patients displayed a strong signal for "nonalcoholic fatty liver disease" driven by the NDUFA9 and UQCRC2 genes (Figure 4). Despite this shared pathway, individual analysis revealed heterogeneity among deceased patients. highlighting the complex interplay between COVID-19, comorbidities, and individual demographics on pathway activation. Enrichment analysis identified potential therapeutic targets based on individual pathway activation. All patients displayed potential targets including CXCR2 (Figure 5). Additionally, specific druggable pathways were identified for some patients, including GNAS (multiple patients), ITGB2 (patients 2 & 6), CSF2RB (multiple patients), SERPING1 (5 patients), PIK3CD (patient 5), TGFBR2 (patient 9), and CUL4B (patient 10).

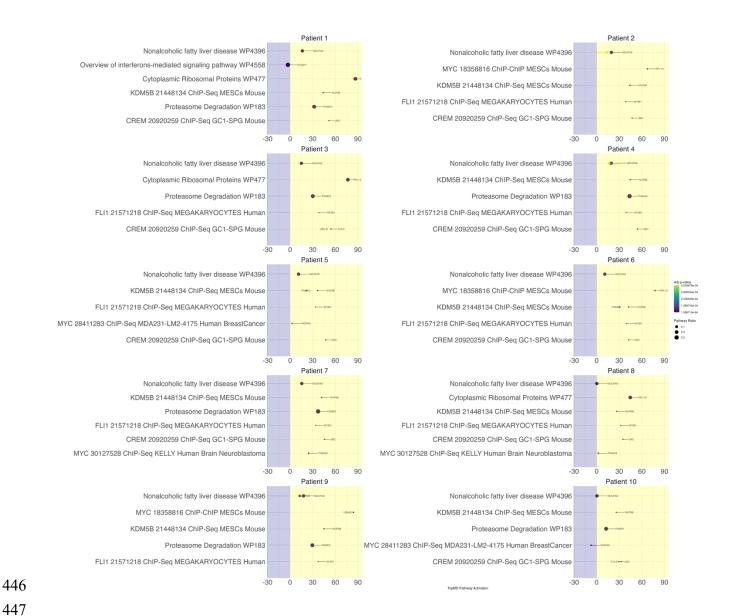


Figure 4: The top 6 pathways enriched in fatal cases within the Florence and Liège cohort using a healthy baseline.

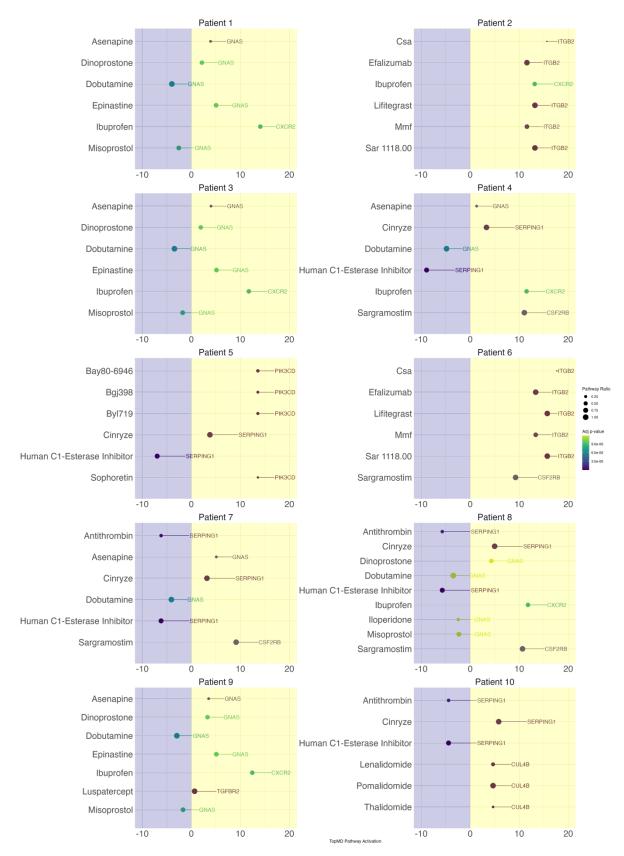


Figure 5: The top significant drug candidates and peak genes that could potentially modulate the phenotype of the 10 fatal cases patients in the Florence and Liège cohort.

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### Discussion:

Traditionally, molecular phenotyping requires data reduction and feature selection, removing biological and technical 'noise', prior to pathway enrichment analysis, but this leads to results which do not accurately represent the molecular phenotypes. Topological analysis of global gene expression finds value in the low abundance transcripts usually discarded as noise, as they represent the 'foothills' of largely activated pathways in a comprehensive molecular landscape. By understanding the molecular phenotype, it is possible to achieve more successful selection of therapeutics, as medicines work at the molecular level as opposed to a clinical level (28).

To redefine predictive models for patient outcomes and health trajectories, there is a growing recognition of the importance of integrating complex datasets. This ranges from biomarkers, clinical parameters to CT scans. For instance, a fully automated AI framework was developed to extract features from chest CT scans for diagnosing COVID-19. The model achieved 85.18% accuracy, enabling rapid and accurate differentiation of COVID-19 from routine clinical conditions, facilitating timely interventions and isolation procedures (29). Similarly, an AI-based analysis named CACOVID-CT was implemented to automatically assess disease severity on chest CT scans. Retrospective analysis of 476 patients revealed that quantitative measurements, such as the percentage of affected lung area (% AA) and CT severity score (CT-SS), correlated strongly with hospital length of stay, ICU admission, mechanical ventilation, and in-hospital mortality. This tool proved effective in identifying patients at higher risk of severe outcomes, facilitating patient management and relieving the workload of radiologists (30).

Our study identified three distinct molecular phenotypes of COVID-19 molecular through topological analysis of global blood gene expression. LRROC analysis demonstrated strong discriminative power of the defined patient clusters tested in the Florence and validated in the Liège cohort. This revealed insights into underlying disease mechanisms, potentially guiding personalised therapeutic approaches.

The analysis using the TopMD algorithm assigned patients to three clusters. Some of the clinical observations aligned with the defined clusters, including lactic acid elevation in cluster 1 and 2 compared to cluster 3. Elevated lactic acid is known to be associated with disease severity and mortality (31). Similarly, cluster 2 showed a higher proportion of respiratory disease and required a higher fraction of inspired oxygen. Additionally, this cluster exhibited elevated direct bilirubin, another potential indicator of disease severity (32). Notably, the majority of those that died from COVID-19 were in cluster 2 (n=4), although the overall number of fatalities in this cohort was small (n=5).

Interestingly, the CORADS scoring system used for chest X-ray/CT severity assessment, couldn't differentiate between the molecular clusters. This suggests different molecular mechanisms might underlie similar clinical presentations, which cannot be identified by CT scan. However, utilising higher resolution CT scan data, such as continuous scoring systems offered by tools like Thirona, might provide more granular insights compared to the categorical data used in this study (30).

Molecular differences were examined between each cluster by considering statistically significant GSEA pathways with highest TopMD pathway volumes (Fig. 2). Cluster 1 displayed a reduction in pathways related to the renin-angiotensin system (RAS) and bradykinin, implicated in COVID-19 pathogenesis (33). Additionally, an increase in focal adhesion pathways, possibly indicating cellular changes related to tissue repair and remodelling. Activation of the complement cascade, led by SERPING1, indicates involvement in the immune response to the virus. Furthermore, an increase in the TGF-β pathway, which regulates inflammation and tissue repair was also identified. Additionally, a high activation of pathways associated with ESC pluripotency, NRF2. and TGF-β receptor signalling. The ESC pluripotency pathway is implicated in tissue repair and regeneration, suggesting a potential compensatory response to tissue damage caused by the virus. NRF2 pathway activation may indicate an antioxidant response to counteract oxidative stress induced by viral infection (34). TGF-β receptor signalling, known for its role in regulating inflammation and fibrosis, may contribute to tissue remodelling and fibrosis observed in severe COVID-19 cases (35). Also, cluster 1 exhibits low activation of pathways related to extracellular vesicle-mediated signalling and complement and coagulation cascades. The decrease in extracellular vesicle-mediated signalling may reflect impaired intercellular communication, while the low activation of complement and coagulation cascades suggests a possible dysregulated immune response and coagulopathy (36).

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In cluster 2, high activation of pathways such as focal adhesion-PI3K-Akt-mTOR signalling and type I interferon induction and signalling was observed, and has been proposed as a potential therapeutic target in SARS-CoV-2 (37, 38) and MERS-CoV (39). Focal adhesion pathway activation may indicate cellular responses to tissue injury or viral invasion, while type I interferon induction and signalling reflect a strong antiviral immune response (40). In contrast, cluster 3 shows opposite activity in IRF7related pathways compared to 2. Additionally, vitamin D receptor activity was observed, which has been implicated in modulating the immune response and may play a role in COVID-19 severity (41-43). Notably, this cluster exhibited low activation of pathways related to TGF-β receptor signalling, IL1R signalling, and LTF danger signal response. The reduced TGF-β receptor signalling suggests decreased fibrotic response and tissue remodelling, while low IL1R signalling may indicate attenuated inflammation (44). The activation of the LTF danger signal response pathway appears to be diminished. Lactoferrin demonstrates antiviral capabilities against various viruses, including coronaviruses (45). It can impede viral replication, disrupt viral attachment and entry, and adjust host immune responses. Lactoferrin's immunomodulatory attributes might aid in tempering excessive inflammation and alleviating cytokine storms observed in severe cases of COVID-19 (46). The decreased activation of the LTF danger signal response pathway could potentially contribute to a weakened interferon response (47).

The stratified molecular phenotypes were found to have different expected responses to both medicines used, and medicines not yet used for COVID-19 (Fig. 3). In cluster 1, CSA or cyclosporine has been shown to be safe to use during COVID-19 for the intended use, however, a reduction in hyperinflammation was observed (48). This warrants further investigation as highlighted by others (49). Interferon related therapies that could modulate the pathway activation of cluster 1 were also identified, which have been shown to have positive effects (50-52). Lifitegrast inhibits SARS-CoV-2 *in vitro* (53, 54) By inhibiting TGF- $\beta$  signalling, Luspatercept may help mitigate the excessive inflammatory response and tissue damage seen in severe COVID-19 cases. Similarly,

Sargramostim has shown promise in a small study, but larger trials are needed to confirm these findings (55).

Like, cluster 1, CSA was also identified as a potentially effective treatment for clusters 2 & 3. The mechanisms of actions of other medicines only matched the molecular phenotype of cluster 2. Asenapine, an anti-pyschotic drug identified by others as a potential drug candidate for COVID-19 (56, 57) Cinryze a human c1 esterase inhibitor was also identified, these inhibitors have been shown to improve lung computed tomography scores and increase blood eosinophils, which are indicators of disease recovery, however, time to clinical improvement was not observed (58). Also, for cluster 2, we identified Fluoxetine and other SSRIs such as fluvoxamine which has previously been identified as having potential use for the treatment of COVID-19 and long-COVID (59) Amisulpride was also identified in cluster 3.

To further evaluate the utility of the TopMD algorithm for precision medicine, enrichment analysis was performed on individual data from the 10 fatal cases within the Florence and Liège cohorts. This approach highlights pathway activation specific to each patient, bypassing the need for a whole cohort for deconvolution. All 10 patients showed potential therapeutic targets based on pathway enrichment. CXCR2 and GNAS were commonly activated across patients (Figure 5), suggesting drugs such as Ibuprofen may be able to modulate some pathways associated with their phenotype. For patients 2 and 6, ITGB2 emerged as one of the top druggable pathways. Notably, Lifitegrast has shown to inhibit SARS-CoV-2 *in vitro* (53, 54). Additionally, CSA or cyclosporine, was also identified, which was another compound identified in the cluster analysis.

Multiple patients exhibited CSF2RB enrichment, indicating potential for Sargramostim, a drug shown to reduce mortality and incubation in small COVID-19 study (55). SERPING1 enrichment in 5 patients suggests various approved drugs for pathway modulation, including antithrombin, human c1 esterase inhibitor and cinryze. Patient specific findings were also observed. PIK3CD enrichment in patient 5 suggests Sophoretin as a potentially modulator, with a meta-analysis showing quercetins (including sophoretin), reduce LDH, hospitalisation risk and mortality (60). Patient 9 displayed TGFBR2 enrichment indicating luspatercept as a potential drug (identified

in the cluster analysis) as a potential candidate. Lastly, CUL4B enrichment in patient 10, suggests Thalidomide, Pomalidomide, Lenalidomide for pathway modulation. While Lenalidomide, used to manage multiple myelomas, has been proposed as protective against sever COVID-19 in a case report (61) a clinical trial showed no benefit (62). Thalidomide, although showing no benefit itself (62), remains a subject of discussion for its potential use in COVID-19 (63).

As a proof of concept, TopMD models were integrated into the Comunicare platform (27), a tool developed and configured within the framework of the DRAGON project, aimed at patient empowerment and providing disease management tooling for clinicians and patients. This proof of concept also enables the analysis of clinical data for clinicians in a dedicated dashboard to demonstrate the possibilities of transcriptomics in digital health. As an example, we generated a model that predicts ICU admission based on our previous work (5) as other outcome variables were too low in number. If a clinician has access to transcriptomic data, a csv file can be uploaded to the dashboard and in return activated pathways are returned after running analysis on the TopMD API. While the use of transcriptomics at the bedside is not ready for deployment, we propose that it is a major advance to be able to demonstrate integration of this data into digital health platforms as the growth of precision medicine continues.

This study identified three distinct molecular phenotypes in hospitalised COVID-19 patients, which were not associated with differences in CT scans and clinical observations. However, these molecular phenotypes match the mechanism of action of different medicines, providing the opportunity for biomarker-led stratified medicine. Topological analysis of global gene expression to define a patient's pathway activation map could be useful in future pandemics to aid in treatment decisions before clinical trials can be completed.

Funding:

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# **Contributions:**

- RPR, FS: methodology, RPR, FS: visualisation, RPR, FS, BVE, TK: software, BE:
- project administration, RPR, FS, JG, MH, AS, EG, AFD, LG, AJP, CN: investigation,
- RPR, FS, JG, CH, TP, SW: formal analysis, JG, CN: clinical supervision, RPR, FS,
- 652 AJP, ST, RB, KH, XX, YN, SW: data curation, RPR, FS, EP, PS, JAH, JPS: writing -
- original draft preparation, RPR, FB, BE, BE, JG, MH, AS, EG, AFD, LG, AJP, CN, ST,
- RB, KH, CH, TP, RD, TC, DB, SW, XX, YN, SW, SW, GY, PJS, JAH, JPRS: writing -
- review and editing, JPRS, JAH, PJS, GY, SW: funding acquisition

### **Declarations:**

- RPR is an employee at TopMD Precision Medicine Ltd. JPRS is a founding director,
- 659 CEO, employee, and shareholder in TopMD Precision Medicine Ltd. FS is a founding
- director, CTO, employee, and shareholder in TopMD Precision Medicine Ltd. PS is a

- 661 founding director, employee, and shareholder in TopMD Precision Medicine Ltd. BVE
- is CEO of Comunicare Solutions. TK is CTO of Comunicare Solutions.
- 663 Data availability:

665 Sequencing reads available under SRA bioproject: PRJNA1085259

### References:

- 1. Clark JJ, Penrice-Randal R, Sharma P, Kipar A, Dong X, Pennington SH, et
- al. Sequential infection with influenza A virus followed by severe acute respiratory
- 669 syndrome coronavirus 2 (SARS-CoV-2) leads to more severe disease and
- 670 encephalitis in a mouse model of COVID-19. bioRxiv. 2023:2020.10.13.334532.
- De Neck S, Penrice-Randal R, Clark JJ, Sharma P, Bentley EG, Kirby A, et al.
- The Stereotypic Response of the Pulmonary Vasculature to Respiratory Viral
- 673 Infections: Findings in Mouse Models of SARS-CoV-2, Influenza A and
- 674 Gammaherpesvirus Infections. Viruses [Internet]. 2023; 15(8).
- 675 3. Dorward DA, Russell CD, Um IH, Elshani M, Armstrong SD, Penrice-Randal
- R, et al. Tissue-Specific Immunopathology in Fatal COVID-19. Am J Respir Crit Care Med. 2021;203(2):192-201.
- 4. Legebeke J, Lord J, Penrice-Randal R, Vallejo AF, Poole S, Brendish NJ, et
- al. Evaluating the Immune Response in Treatment-Naive Hospitalised Patients With
- 680 Influenza and COVID-19. Front Immunol. 2022;13:853265.
- 5. Penrice-Randal R, Dong X, Shapanis AG, Gardner A, Harding N, Legebeke J,
- et al. Blood gene expression predicts intensive care unit admission in hospitalised
- 683 patients with COVID-19. Front Immunol. 2022;13:988685.
- 684 6. Russell CD, Valanciute A, Gachanja NN, Stephen J, Penrice-Randal R,
- 685 Armstrong SD, et al. Tissue Proteomic Analysis Identifies Mechanisms and Stages
- of Immunopathology in Fatal COVID-19. Am J Respir Cell Mol Biol. 2021.
- 687 7. Liu X, Speranza E, Muñoz-Fontela C, Haldenby S, Rickett NY, Garcia-Dorival
- I, et al. Transcriptomic signatures differentiate survival from fatal outcomes in
- humans infected with Ebola virus. Genome Biology. 2017;18(1):4.
- 690 8. McClain MT, Constantine FJ, Nicholson BP, Nichols M, Burke TW, Henao R,
- et al. A blood-based host gene expression assay for early detection of respiratory
- of viral infection: an index-cluster prospective cohort study. Lancet Infect Dis.
- 693 2021;21(3):396-404.
- 694 9. Carroll MW, Matthews DA, Hiscox JA, Elmore MJ, Pollakis G, Rambaut A, et
- al. Temporal and spatial analysis of the 2014-2015 Ebola virus outbreak in West
- 696 Africa. Nature. 2015;524(7563):97-101.
- 697 10. Watson RJ, Tree J, Fotheringham SA, Hall Y, Dong X, Steeds K, et al. Dose-
- Dependent Response to Infection with Ebola Virus in the Ferret Model and Evidence
- of Viral Evolution in the Eye. Journal of virology. 2021;95(24):e0083321.
- 700 11. Russell CD, Valanciute A, Gachania NN, Stephen J, Penrice-Randal R,
- 701 Armstrong SD, et al. Tissue proteomic analysis identifies mechanisms and stages of
- immunopathology in fatal COVID-19. American journal of respiratory cell and
- 703 molecular biology. 2022;66(2):196-205.
- 704 12. Nan Y, Ser JD, Walsh S, Schönlieb C, Roberts M, Selby I, et al. Data
- harmonisation for information fusion in digital healthcare: A state-of-the-art
- systematic review, meta-analysis and future research directions. Information Fusion.
- 707 2022;82:99-122.
- 13. Halilaj I, Chatterjee A, van Wijk Y, Wu G, van Eeckhout B, Oberije C, et al.
- 709 Covid19Risk.ai: An Open Source Repository and Online Calculator of Prediction
- 710 Models for Early Diagnosis and Prognosis of Covid-19. BioMed. 2021;1(1):41-9.
- 711 14. Chatterjee A, Wu G, Primakov S, Oberije C, Woodruff H, Kubben P, et al. Can
- 712 predicting COVID-19 mortality in a European cohort using only demographic and
- comorbidity data surpass age-based prediction: An externally validated study. PLOS
- 714 ONE. 2021;16(4):e0249920.

- 715 15. Cen X, Wang F, Huang X, Jovic D, Dubee F, Yang H, et al. Towards precision
- medicine: Omics approach for COVID-19. Biosaf Health. 2023;5(2):78-88.
- 717 16. Teodori L, Osimani B, Isidoro C, Ramakrishna S. Mass versus personalized
- medicine against COVID-19 in the "system sciences" era. Cytometry Part A.
- 719 2022;101(12):995-9.
- 720 17. Wang Z, He Y. Precision omics data integration and analysis with
- interoperable ontologies and their application for COVID-19 research. Briefings in
- 722 Functional Genomics. 2021;20(4):235-48.
- 723 18. Venkataraman T, Coleman CM, Frieman MB. Overactive Epidermal Growth
- 724 Factor Receptor Signaling Leads to Increased Fibrosis after Severe Acute
- Respiratory Syndrome Coronavirus Infection. Journal of virology. 2017;91(12).
- 726 19. Venkataraman T, Frieman MB. The role of epidermal growth factor receptor
- 727 (EGFR) signaling in SARS coronavirus-induced pulmonary fibrosis. Antiviral Res.
- 728 2017;143:142-50.
- 729 20. Vagapova ER, Lebedev TD, Prassolov VS. Viral fibrotic scoring and drug
- 730 screen based on MAPK activity uncovers EGFR as a key regulator of COVID-19
- 731 fibrosis. Scientific Reports. 2021;11(1):11234.
- 732 21. Chen S, Zhou Y, Chen Y, Gu J. fastp: an ultra-fast all-in-one FASTQ
- 733 preprocessor. Bioinformatics. 2018;34(17):i884-i90.
- 734 22. Patro R, Duggal G, Love MI, Irizarry RA, Kingsford C. Salmon provides fast
- and bias-aware quantification of transcript expression. Nature Methods.
- 736 2017;14(4):417-9.
- 737 23. Soneson C, Love MI, Robinson MD. Differential analyses for RNA-seq:
- transcript-level estimates improve gene-level inferences. F1000Res. 2015;4:1521.
- 739 24. Robinson MD, McCarthy DJ, Smyth GK. edgeR: a Bioconductor package for
- 740 differential expression analysis of digital gene expression data. Bioinformatics.
- 741 2010;26(1):139-40.
- 742 25. Szklarczyk D, Kirsch R, Koutrouli M, Nastou K, Mehryary F, Hachilif R, et al.
- 743 The STRING database in 2023: protein-protein association networks and functional
- enrichment analyses for any sequenced genome of interest. Nucleic Acids Res.
- 745 2023;51(D1):D638-d46.
- 746 26. Freshour SL, Kiwala S, Cotto KC, Coffman AC, McMichael JF, Song JJ, et al.
- 747 Integration of the Drug-Gene Interaction Database (DGldb 4.0) with open
- crowdsource efforts. Nucleic Acids Res. 2021;49(D1):D1144-d51.
- 749 27. Duquenne JB, Corhay JL, Louis R, Van Cauwenberge H. [Feasibility and
- 750 effectiveness study of a simplified mobile self-education and self-monitoring
- application for patients with severe chronic obstructive pulmonary disease]. Rev Med
- 752 Liege. 2022;77(2):110-7.
- 753 28. Farahani M, Niknam Z, Mohammadi Amirabad L, Amiri-Dashatan N, Koushki
- M, Nemati M, et al. Molecular pathways involved in COVID-19 and potential
- pathway-based therapeutic targets. Biomedicine & Pharmacotherapy.
- 756 2022;145:112420.
- 757 29. Guiot J, Vaidyanathan A, Deprez L, Zerka F, Danthine D, Frix A-N, et al.
- 758 Development and Validation of an Automated Radiomic CT Signature for Detecting
- 759 COVID-19. Diagnostics. 2021;11(1):41.
- Guiot J, Maes N, Winandy M, Henket M, Ernst B, Thys M, et al. Automatized
- lung disease quantification in patients with COVID-19 as a predictive tool to assess
- hospitalization severity. Frontiers in Medicine. 2022;9.

- 763 31. Carpenè G, Onorato D, Nocini R, Fortunato G, Rizk JG, Henry BM, et al.
- Blood lactate concentration in COVID-19: a systematic literature review. Clin Chem
- 765 Lab Med. 2022;60(3):332-7.
- 766 32. Chen W, Liu H, Yang G, Wang W, Liu Q, Huang C, et al. Effect of Direct
- 767 Bilirubin Level on Clinical Outcome and Prognoses in Severely/Critically III Patients
- 768 With COVID-19. Front Med (Lausanne). 2022;9:843505.
- 769 33. Garvin MR, Alvarez C, Miller JI, Prates ET, Walker AM, Amos BK, et al. A
- 770 mechanistic model and therapeutic interventions for COVID-19 involving a RAS-
- mediated bradykinin storm. Elife. 2020;9.
- 772 34. Lee C. Therapeutic Modulation of Virus-Induced Oxidative Stress via the Nrf2-
- 773 Dependent Antioxidative Pathway. Oxid Med Cell Longev. 2018;2018:6208067.
- 774 35. Chen J, Wu W, Wang W, Tang Y, Lan H-Y. Role of TGF-β Signaling in
- 775 Coronavirus Disease 2019. Integrative Medicine in Nephrology and Andrology.
- 776 2022;9(1):9.
- 777 36. Conway EM, Mackman N, Warren RQ, Wolberg AS, Mosnier LO, Campbell
- RA, et al. Understanding COVID-19-associated coagulopathy. Nat Rev Immunol.
- 779 2022;22(10):639-49.
- 780 37. Fattahi S, Khalifehzadeh-Esfahani Z, Mohammad-Rezaei M, Mafi S, Jafarinia
- 781 M. PI3K/Akt/mTOR pathway: a potential target for anti-SARS-CoV-2 therapy.
- 782 Immunol Res. 2022;70(3):269-75.
- 783 38. Khezri MR, Varzandeh R, Ghasemnejad-Berenji M. The probable role and
- therapeutic potential of the PI3K/AKT signaling pathway in SARS-CoV-2 induced
- coagulopathy. Cell Mol Biol Lett. 2022;27(1):6.
- 786 39. Kindrachuk J, Ork B, Hart BJ, Mazur S, Holbrook MR, Frieman MB, et al.
- 787 Antiviral potential of ERK/MAPK and PI3K/AKT/mTOR signaling modulation for
- 788 Middle East respiratory syndrome coronavirus infection as identified by temporal
- 789 kinome analysis. Antimicrob Agents Chemother. 2015;59(2):1088-99.
- 790 40. Channappanavar R, Perlman S. Pathogenic human coronavirus infections:
- 791 causes and consequences of cytokine storm and immunopathology. Semin
- 792 Immunopathol. 2017;39(5):529-39.
- 793 41. Azmi A, Rismani M, Pourmontaseri H, Mirzaii E, Niknia S, Miladpour B. The
- role of vitamin D receptor and IL-6 in COVID-19. Mol Genet Genomic Med.
- 795 2023;11(7):e2172.
- 796 42. Hurst EA, Mellanby RJ, Handel I, Griffith DM, Rossi AG, Walsh TS, et al.
- 797 Vitamin D insufficiency in COVID-19 and influenza A, and critical illness survivors: a
- 798 cross-sectional study. BMJ Open. 2021;11(10):e055435.
- 799 43. Evans RM, Lippman SM. Shining Light on the COVID-19 Pandemic: A
- 800 Vitamin D Receptor Checkpoint in Defense of Unregulated Wound Healing. Cell
- 801 Metabolism. 2020;32(5):704-9.
- 802 44. Biernacka A, Dobaczewski M, Frangogiannis NG. TGF-β signaling in fibrosis.
- 803 Growth Factors. 2011;29(5):196-202.
- 804 45. Puddu P, Valenti P, Gessani S. Immunomodulatory effects of lactoferrin on
- antigen presenting cells. Biochimie. 2009;91(1):11-8.
- 806 46. Zimecki M, Actor JK, Kruzel ML. The potential for Lactoferrin to reduce SARS-
- 807 CoV-2 induced cytokine storm. Int Immunopharmacol. 2021;95:107571.
- 808 47. Actor JK, Hwang SA, Kruzel ML. Lactoferrin as a natural immune modulator.
- 809 Curr Pharm Des. 2009;15(17):1956-73.
- 810 48. Blumberg EA, Noll JH, Tebas P, Fraietta JA, Frank I, Marshall A, et al. A
- phase I trial of cyclosporine for hospitalized patients with COVID-19. JCI Insight.
- 812 2022;7(11).

- 813 49. Devaux CA, Melenotte C, Piercecchi-Marti MD, Delteil C, Raoult D.
- 814 Cyclosporin A: A Repurposable Drug in the Treatment of COVID-19? Front Med
- 815 (Lausanne). 2021;8:663708.
- 816 50. Kamyshnyi A, Koval H, Kobevko O, Buchynskyi M, Oksenych V, Kainov D, et
- al. Therapeutic Effectiveness of Interferon-α2b against COVID-19 with Community-
- 818 Acquired Pneumonia: The Ukrainian Experience. Int J Mol Sci. 2023;24(8).
- 819 51. Reis G, Moreira Silva EAS, Medeiros Silva DC, Thabane L, Campos VHS,
- Ferreira TS, et al. Early Treatment with Pegylated Interferon Lambda for Covid-19.
- 821 New England Journal of Medicine. 2023;388(6):518-28.
- 822 52. Monk PD, Marsden RJ, Tear VJ, Brookes J, Batten TN, Mankowski M, et al.
- 823 Safety and efficacy of inhaled nebulised interferon beta-1a (SNG001) for treatment
- of SARS-CoV-2 infection: a randomised, double-blind, placebo-controlled, phase 2
- trial. The Lancet Respiratory Medicine. 2021;9(2):196-206.
- 826 53. Shen X-R, Geng R, Li Q, Chen Y, Li S-F, Wang Q, et al. ACE2-independent
- 827 infection of T lymphocytes by SARS-CoV-2. Signal Transduction and Targeted
- 828 Therapy. 2022;7(1):83.
- 54. Day CJ, Bailly B, Guillon P, Dirr L, Jen FE, Spillings BL, et al. Multidisciplinary
- 830 Approaches Identify Compounds that Bind to Human ACE2 or SARS-CoV-2 Spike
- Protein as Candidates to Block SARS-CoV-2-ACE2 Receptor Interactions. mBio.
- 832 2021;12(2).
- 833 55. Paine R, Chasse R, Halstead ES, Nfonoyim J, Park DJ, Byun T, et al. Inhaled
- 834 Sargramostim (Recombinant Human Granulocyte-Macrophage Colony-Stimulating
- Factor) for COVID-19-Associated Acute Hypoxemia: Results of the Phase 2,
- Randomized, Open-Label Trial (iLeukPulm). Mil Med. 2022;188(7-8):e2629-38.
- 837 56. Ku KB, Shin HJ, Kim HS, Kim BT, Kim SJ, Kim C. Repurposing Screens of
- 838 FDA-Approved Drugs Identify 29 Inhibitors of SARS-CoV-2. J Microbiol Biotechnol.
- 839 2020;30(12):1843-53.
- 840 57. Rajput A, Thakur A, Rastogi A, Choudhury S, Kumar M. Computational
- identification of repurposed drugs against viruses causing epidemics and pandemics
- via drug-target network analysis. Comput Biol Med. 2021;136:104677.
- 843 58. Mansour E, Palma AC, Ulaf RG, Ribeiro LC, Bernardes AF, Nunes TA, et al.
- 844 Safety and Outcomes Associated with the Pharmacological Inhibition of the Kinin-
- 845 Kallikrein System in Severe COVID-19. Viruses. 2021;13(2).
- 846 59. Hashimoto K. Overview of the potential use of fluvoxamine for COVID-19 and
- long COVID. Discov Ment Health. 2023;3(1):9.
- 848 60. Ziaei S, Alimohammadi-Kamalabadi M, Hasani M, Malekahmadi M, Persad E,
- Heshmati J. The effect of guercetin supplementation on clinical outcomes in COVID-
- 19 patients: A systematic review and meta-analysis. Food Science & Nutrition.
- 851 2023;11(12):7504-14.

- 852 61. Al Sbihi A, Manasrah N, Sano D. Can Lenalidomide Protect against Severe
- 853 COVID-19 Symptoms in Multiple Myeloma Patients? A Case Series and Review of
- the Literature. Eur J Case Rep Intern Med. 2022;9(3):003216.
- 855 62. Amra B, Ashrafi F, Torki M, Hashemi M, Shirzadi M, Soltaninejad F, et al.
- 856 Thalidomide for the Treatment of COVID-19 Pneumonia: A Randomized Controlled
- 857 Clinical Trial. Adv Biomed Res. 2023;12:14.
- 858 63. Sundaresan L, Giri S, Singh H, Chatterjee S. Repurposing of thalidomide and
- its derivatives for the treatment of SARS-coV-2 infections: Hints on molecular action.
- 860 Br J Clin Pharmacol. 2021;87(10):3835-50.