

COMMENTARY

Making *in vitro* conditions more reflective of *in vivo* conditions for research on the teleost gastrointestinal tract

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ABSTRACT

To date, the majority of *in vitro* or *ex vivo* fish gastrointestinal research has been conducted under unrealistic conditions. In a living fish, ionic conditions, as well as levels of ammonia, pH, HCO_3^- and P_{CO_2} differ considerably between the different regions of the gastrointestinal tract. These factors also differ from those of the saline often used in gut research. Furthermore, the oxygen gradient from the serosa to the gut lumen is rarely considered: in contrast to the serosa, the lumen is a hypoxic/anoxic environment. In addition, the gut microbiome plays a significant role in gut physiology, increasing the complexity of the *in vivo* gut, but replicating the microbial community for *in vitro* studies is exceptionally difficult. However, there are ways in which we can begin to overcome these challenges. Firstly, the luminal chemistry and P_{O_2} in each gut compartment must be carefully considered. Secondly, although microbiological culture techniques are improving, we must learn how to maintain the microbiome diversity seen *in vivo*. Finally, for *ex vivo* studies, developing mucosal (luminal) solutions that more closely mimic the *in vivo* conditions will better replicate physiological processes. Within the field of mammalian gut physiology, great advances in 'gut-on-chip' devices are providing the tools to better replicate *in vivo* conditions; adopting and adapting this technology may assist in fish gut research initiatives. This Commentary aims to make fish gut physiologists aware of the various issues in replicating the *in vivo* conditions and identifies solutions as well as those areas that require further improvement.

KEY WORDS: Chyme, Stomach, Intestine, Gut microbiome, Mucosal chemistry

Introduction

In vitro gut models have been readily incorporated into mammalian and medical research. Studies using these models, often under symmetrical conditions (i.e. where the same medium bathes both luminal and serosal surfaces), have provided insights into nutrient/drug uptake and factors affecting intestinal epithelial integrity, as well as a better understanding of disease aetiology and prevention (Wang et al., 2023; Kim and Sung, 2024; McCoy et al., 2024). The success of these models has occurred despite knowledge that the gut

is not an isolated organ; its function is regulated by the paracrine and endocrine systems, the diet, infections and commensal microbiota. The adoption of *in vitro* techniques for comparative physiological studies of the gut outside the medical research sphere has been less enthusiastic. There is an obvious reason for this: medical research focuses on one species, *Homo sapiens*; in contrast, comparative physiologists are interested in studying the differences among many species. In addition, within the ~30,000 species of teleost fish, there is much variation: some are herbivores, piscivores, omnivores, planktivores or carnivores; gut function differs between freshwater and seawater environments; some species are stomachless; some have extensive pyloric caecae whereas others do not; the intestine may, or may not, be split into three sections (hind, mid and anterior regions) of varying length. One fish model will not adequately cover these diverse features.

There is a societal and political desire to reduce and replace the use of animals in research. However, for comparative research on gut function, it is difficult to see how it will be possible to mimic whole-organism physiology in a flask for multiple species. Thus, *ex vivo* approaches, such as gut sacs (see Glossary) or the use of Ussing chambers (see Glossary) containing gut epithelia from specific species have been used (Weinrauch et al., 2021). Where *in vitro* models are likely to be particularly useful is in research associated with financially important aquaculture species. In fact, *in vitro* technologies have already been developed for salmonids (Drieschner, et al., 2019a,b). However, most of the published work using *in vitro* systems has made use of culture conditions that do not replicate the situation *in vivo*; thus, our Commentary provides details of the challenges in devising *in vitro* systems that consider variations in luminal chemistry, as well as the presence of commensal microbes and anoxia (Fig. 1A). We focus specifically on the teleost gastrointestinal (GI) tract (see Fig. 1B for an overview of the main cell types present).

Chemistry of the gut lumen

The fluids in the GI tract are part of the external environment from which required substances (e.g. ions and nutrients) are absorbed, and into which waste products may be secreted. Therefore, although it is appropriate in *in vitro* or *ex vivo* work to use physiological saline (a solution designed to mimic the internal environment, i.e. the blood plasma) on the serosal side of the tract, it is not appropriate to use this same solution to mimic the often extreme external environment on the mucosal (i.e. luminal) side. Unfortunately, these are the conditions that have been used in the vast majority of *in vitro* fish studies on gut function.

There is a growing body of literature on the *in vivo* physicochemical conditions of the GI fluids or chyme (see Glossary) in a variety of animals. Here, we consider the freshwater rainbow trout (*Oncorhynchus mykiss*) as an illustration of what is known regarding *in vivo* conditions in the fish gut, because of its importance as a model organism in physiology, aquaculture and

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Glossary

Alkaline tide

This occurs after a meal when the cells of the stomach secrete HCO_3^- into the blood, causing a transient rise in blood pH.

Chyme

Fluid consisting of gastric juices and partially digested food that passes from the stomach and along the various sections of the intestine.

Culturomics

High-throughput cell culture of gut microbes.

Euryhaline

Euryhaline refers to an organism that can tolerate a wide range of salinities.

Gut-associated lymphoid system

A collection of tissues involved in the immune response to specific gut microbes and antigens.

Gut sacs

A technique for measuring properties of the gastrointestinal (GI) tract using segments of the GI removed from the organism. The section of gut sac can either remain in the same orientation or be everted, and then sealed at one end with suture silk and filled at the other end with an indwelling cannula. The sac can be filled with luminal or serosal fluid, and then sealed and placed in a bath of luminal or serosal media, depending on its orientation.

Henderson–Hasselbalch equation

An equation used to calculate the ratio of the concentrations of an acid (A) and its conjugate base at the pH of a chemical solution using the acid dissociation constant, K_a : $\text{pH} = \text{p}K_a + \log_{10}([\text{A}^-]/[\text{HA}])$.

3Rs (Replacement, Reduction and Refinement)

A framework based on the concepts from *The Principles of Humane Experimental Technique* by W. M. S. Russell and R. L. Burch (1959) to improve animal welfare in research.

Solvent drag

A process by which a solvent, such as water, moves across an epithelium by osmosis, taking solutes with it.

Stenohaline

Stenohaline refers to an organism unable to withstand wide variation in salinity.

Two-compartment *in vitro* cell culture model

The culture of cells on a permeable support to generate an *in vitro* epithelium that separates the luminal (apical) from the serosal (basolateral) compartments.

Ussing chamber

An apparatus where either the natural epithelium or a monolayer of cells grown on permeable supports is mounted between two chambers for the measurement of electrical impedance, transepithelial potential and the transport of ions and other molecules across the epithelial membrane.

toxicology (Wood and Bucking, 2011). In research on rainbow trout, Cortland saline (Wolf, 1963) is the solution that has been used most frequently as the basis of the mucosal medium for *in vitro* experiments. Usually, it is pre-equilibrated with air, 0.3% CO_2 (2.3 Torr; where 1 Torr \approx 133 Pa) or 1% CO_2 (7.5 Torr); the latter two CO_2 mixtures may be in air (P_{O_2} = 150–160 Torr) or oxygen (P_{O_2} > 700 Torr). Sometimes, the $[\text{Ca}^{2+}]$ and $[\text{HCO}_3^-]$ are reduced to prevent their precipitation. Table 1 compares the composition of standard Cortland saline with measured physicochemical parameters in the luminal fluids in the stomach, anterior, mid and posterior intestine of fasted trout and those fed recently with commercial trout pellets. It should be noted that fish referred to as ‘fed’ in the literature generally receive commercial pellets that do not reflect a natural food source.

pH

The trout, like many vertebrates (Kong and Singh, 2008) and most but not all fishes (Bakke et al., 2011; Wood, 2019), maintains a very

acidic pH in the luminal fluid of the stomach while fasting. The pH rises considerably after feeding as a result of the buffering action of the ingested food (Table 1, and references therein). This occurs despite a massive activation of HCl secretion by the gastric H^+ , K^+ -ATPase, which creates a reciprocal alkaline tide (see Glossary) in the blood (Bucking and Wood, 2008; Cooper and Wilson, 2008), both of which can persist for more than a day (depending on the species) after feeding. The Cortland saline commonly used *in vitro* has a pH of 7.0–7.8, which is 2–5 pH units above the real *in vivo* pH for the gastric epithelium, which has a 100- to 10,000-fold greater H^+ concentration bathing its mucosal surface. In freshwater fish, the stomach has important roles in the absorption of major ions (Bucking and Wood, 2006a, 2007, 2009), ammonia (Jung et al., 2023b) and nutrient metals such as nickel (Leonard et al., 2009) and copper (Nadella et al., 2011). The availability and ion speciation of these are affected by the low gastric pH. Similarly, low pH is essential for proper activity of gastric proteases (Márquez et al., 2012). *In vitro*, low pH in the luminal fluid can be maintained using buffers such as formic acid (Jung et al., 2003b) or PIPES (Nadella et al., 2011) (the latter of which does not complex metal ions; Kandegedara and Rorabacher, 1999).

In contrast to the stomach, all sections of the trout intestine are alkaline, with pH increasing progressively from the anterior to posterior sections, and greater alkalisation after feeding (see Table 1, and references therein). The pH of the chyme in the posterior intestine may approach 9.0 in fed trout; again, well outside that of the Cortland saline, but in the opposite direction from that of stomach chyme. In trout and other fish, the cause of the high pH is HCO_3^- secretion by the mucosal epithelium (Wood, 2019). It is well established that the optimal activities of digestive enzymes are very sensitive to luminal fluid pH in salmonids (Krogdahl et al., 2015), as are the intestinal uptake rates of copper (Nadella et al., 2007) and iron ions (Kwong and Niyogi, 2008). It is critical that realistically high pH values are applied on the mucosal surfaces of *in vitro* preparations. Although this can be accomplished using buffers that do not complex metals (Kandegedara and Rorabacher, 1999), it is best done by appropriate manipulations of P_{CO_2} and $[\text{HCO}_3^-]$ (using the Henderson–Hasselbalch equation; see Glossary; Boutilier et al., 1984) to properly mimic the *in vivo* situation.

HCO_3^- concentration

$[\text{HCO}_3^-]$ is negligible in gastric fluids, as a result of the low pH (Table 1). However, the presence of high $[\text{HCO}_3^-]$ in the intestinal fluids as well as the reciprocal variation of $[\text{HCO}_3^-]$ with $[\text{Cl}^-]$ in the intestine have long been known in marine fish (Smith, 1930), including the seawater-acclimated rainbow trout (Wilson et al., 1996). In seawater, $\text{Cl}^-/\text{HCO}_3^-$ exchange serves to absorb Cl^- and secrete HCO_3^- ; this results in the precipitation of divalent cations (Ca^{2+} and Mg^{2+}) and HCO_3^- as carbonates out of solution from the ingested seawater (Walsh et al., 1991), thereby improving the osmotic gradient for water absorption from the lumen into the blood (Grosell, 2006; Wood, 2019). However, only recently has it been shown that luminal $[\text{HCO}_3^-]$ may also be very high in the freshwater-acclimated rainbow trout. The values tend to increase progressively from the anterior to posterior intestine and are especially high after feeding, sometimes exceeding 50 mmol l^{-1} (Table 1). This is true not only in the trout (a euryhaline teleost; see Glossary) but also in several stenohaline freshwater teleosts (see Glossary; Wood and Eom, 2019; Goodrich et al., 2020). There is no obvious role for this high intestinal $[\text{HCO}_3^-]$ in osmoregulation. Rather, the high $[\text{HCO}_3^-]$ balances the high P_{CO_2} in the GI tract,

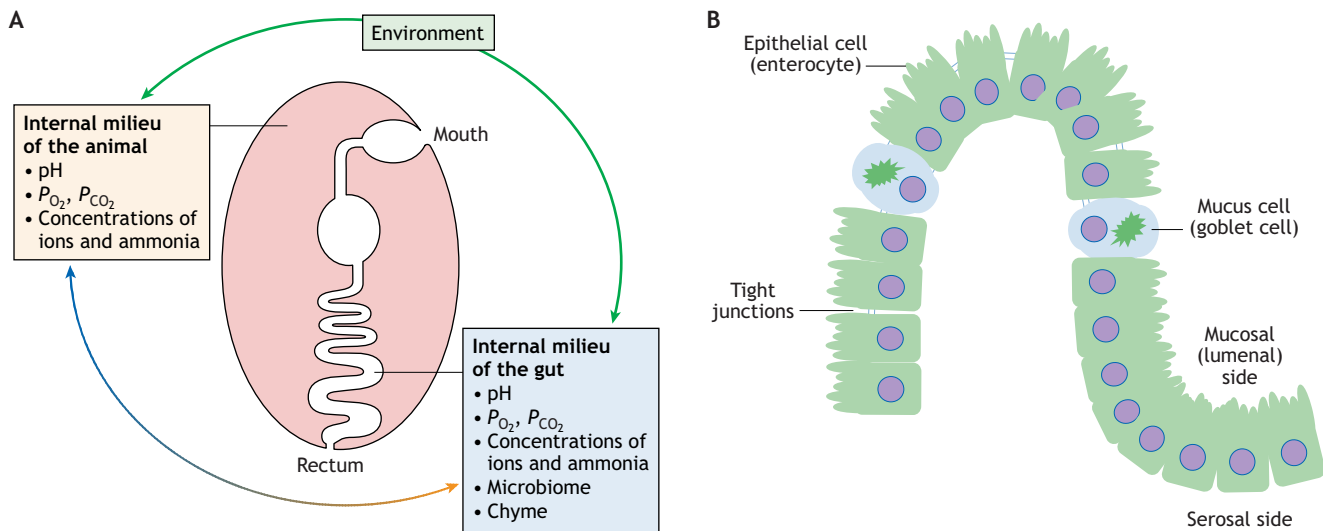


Fig. 1. The fish gastrointestinal tract. (A) Food and water enter the mouth, and waste food and metabolites exit the anus via the rectum. The gut lumen pH, P_{O_2} and P_{CO_2} , as well as the concentration of ions and ammonia, and the microbiome will vary along the gastrointestinal (GI) tract and will differ between species depending on the environment they inhabit and the food resources available. The gradients of P_{O_2} , P_{CO_2} and ammonia concentration between the gut lumen and the body are influenced by the animal's internal milieu. (B) Schematic diagram of the main cells present in the gut lumen.

thereby ensuring an alkaline pH and setting the correct acid–base conditions for enzyme and transporter functions. In conclusion, HCO_3^- levels in standard Cortland saline are far too high for the mucosal solution of the stomach, and too low for the mucosal solution of the intestinal segments, at least after feeding. They should therefore be adjusted accordingly for *in vitro* experiments.

P_{CO_2}

In fish, simple calculations using the Henderson–Hasselbalch equation and measured values of pH and $[HCO_3^-]$ have demonstrated that P_{CO_2} levels in the GI fluids of fish are usually much higher than the 0–7.5 Torr routinely used for Cortland saline (Wood, 2019) (Table 1). This discrepancy increases after feeding and occurs throughout the tract. In fact, P_{CO_2} levels in the GI tract may exceed 50 Torr (10–20 times normal arterial blood levels of 2–4 Torr). This has been confirmed by direct measurements of P_{CO_2} in luminal fluids with micro-optodes both in freshwater rainbow trout (Wood and Eom, 2019) and in marine English sole (Jung et al., 2020). The high P_{CO_2} levels in the chyme result in the diffusion of CO_2 into the enterocytes, cells that are rich in carbonic anhydrase. Subsequently, CO_2 is converted to H^+ and HCO_3^- , thereby ‘refuelling’ the supply of these acid–base equivalents, which are vital for ion, metal and nutrient transport processes, as well as for maintenance of luminal fluid pH (Wood, 2019). Undoubtedly, both the enterocytes and the microbes in the GI tract are adapted to life under high P_{CO_2} . If these conditions are not met at the mucosal surface in *in vitro* preparations, the cells are functioning in a very unnatural environment.

Ammonia

Ammonia is simultaneously a respiratory gas (NH_3), an ion (NH_4^+), a valuable resource for protein production through the glutamine synthesis pathway and a highly toxic nitrogenous waste. A large amount of ammonia is produced by digestive processes, enterocyte metabolism and microbial activity in the GI tract. This accounts for up to 47% of whole-body ammonia production in the post-prandial rainbow trout, and much of this ammonia is absorbed across the tract (Rubino et al., 2014). In trout, total ammonia concentration in gastric and posterior intestinal chyme may exceed 3500 and

1500 $\mu\text{mol l}^{-1}$, respectively (see Table 1, and references therein). By contrast, blood plasma levels of ammonia are generally less than 200 $\mu\text{mol l}^{-1}$, and Cortland saline lacks ammonia entirely. The partitioning of ammonia between NH_4^+ and the more toxic NH_3 is a function of pH (Cameron and Heisler, 1983), with the NH_3 fraction (and therefore P_{NH_3}) increasing as pH increases. Therefore, in trout, P_{NH_3} is essentially zero in gastric chyme and highest in posterior intestinal chyme, where pH is highest (Table 1). As with P_{CO_2} , the enterocytes and microbes are likely to be adapted to these high P_{NH_3} levels and total ammonia concentration *in vivo*; ideally, these conditions should be replicated in the mucosal media during *in vitro* experiments.

Major ions and osmolality

In designing mucosal solutions for *in vitro* experiments, it is critical to know the ionic composition of the normal diet to which the animal is adapted, and how this composition changes in the fluid phase of the chyme as it is sequentially processed along the GI tract. For freshwater rainbow trout fed commercial pellets, $[Na^+]$ throughout the tract remains generally similar to or slightly higher than levels in Cortland saline or plasma (see Table 1, and references therein). However, $[Cl^-]$ in gastric chyme is much higher than that of plasma or saline because of HCl secretion in the stomach. $[Cl^-]$ then drops to much lower levels throughout the intestine as a result of Cl^-/HCO_3^- exchange in the intestine (Table 1). Concentrations of K^+ , Ca^{2+} and Mg^{2+} are also elevated in gastric fluid relative to the blood plasma, but unlike $[Cl^-]$, the concentration of these ions remains generally elevated in luminal fluids throughout the intestinal segments; the elevation in concentration is particularly pronounced for Mg^{2+} , for which concentrations may be 10- to 25-fold higher than in plasma or saline. Interestingly, seawater-acclimated rainbow trout fed a diet of commercial pellets exhibit rather similar ranges of major ion concentrations in their GI fluids (Bucking et al., 2011) to those of freshwater trout (reported in Table 1). However, $[Mg^{2+}]$ and $[SO_4^{2-}]$ are much higher in the posterior intestine of fasted trout in seawater compared with those in freshwater (Shehadeh and Gordon, 1969). Of particular note, osmolality of the luminal fluids in fed freshwater trout remains

Table 1. A comparison of physicochemical conditions measured *in vivo* in the gastrointestinal fluids (chyme) of freshwater rainbow trout (*Oncorhynchus mykiss*) with those of the Cortland saline (Wolf, 1963) often used as the luminal fluid for *in vitro* experiments on the physiology of the gastrointestinal tract

Parameter	Cortland saline	Stomach		Anterior intestine		Mid intestine		Posterior intestine		Reference
		Fasted	Fed	Fasted	Fed	Fasted	Fed	Fasted	Fed	
pH	7.0–7.8		4.2–4.7		7.0–7.2		7.7–7.9		7.7–7.8	Clavoni et al. (2023)
			5.3		7.6		7.7		8.0	Jung et al. (2023a)
			3.7		7.9		8.0		8.0	Jung et al. (2023b)
			3.6		7.8		7.9		8.1	Jung et al. (2022)
		2.4	4.2	7.2	7.5	7.3	7.6	7.4	7.7	Wood and Eom (2019)
					7.9		8.3		8.6	Rubino et al. (2014)
		2.8	3.9–5.8	8.2	7.8–8.6	8.3	8.1–8.7	8.3	8.3–9.2	Bucking and Wood (2009)
								8.5–9.0		Shehadeh and Gordon (1969)
[HCO ₃ ⁻] (mmol l ⁻¹)	11.9		1.9		19		21		40	Jung et al. (2023a)
			0.2		33		36		52	Jung et al. (2022)
		0.1*	0.5*	6*	44*	8*	55*	15*	52*	Wood and Eom (2019)
P _{CO₂} (Torr)	0–7.5		126*		12*		16*		15*	Jung et al. (2023a)
			112*		35*		25*		27*	Jung et al. (2022)
		9	20	7	41	8	25	13	28	Wood and Eom (2019)
Total ammonia (μmol l ⁻¹)	0		2200		700		500		1050	Jung et al. (2023a)
			2000		200		250		650	Jung et al. (2023a)
			3500		600		800		1100	Jung et al. (2023b)
			3700		400		450		1050	Jung et al. (2022)
					1068		893		981	Rubino et al. (2014)
					1500–2200		1800–2200		1500–1600	Bucking and Wood (2012)
P _{NH₃} (μTorr)	0		0*		60*		60*		240*	Jung et al. (2023a)
			0*		60*		90*		380*	Jung et al. (2023a)
			0.2*		125*		235*		420*	Jung et al. (2023b)
			0*		100*		200*		520*	Jung et al. (2022)
					390*		839*		1760*	Rubino et al. (2014)
P _{O₂} (Torr)	150–700	0.30	0.50	0.55	0.55	0.60	0.30	0.40	0.40	Jung et al. (2022)
[Na ⁺] (mmol l ⁻¹)	142		14–141		95–155		125–170		135–160	Bucking and Wood (2006a, 2009)
								160–170		Shehadeh and Gordon (1969)
[Cl ⁻] (mmol l ⁻¹)	132		190–220		25–60		20–45		25–50	Bucking and Wood (2006a, 2009)
								15–70		Shehadeh and Gordon (1969)
[K ⁺] (mmol l ⁻¹)	5		9–55		7–12		4–10		6–9	Bucking and Wood (2006a, 2009)
								4–5		Shehadeh and Gordon (1969)
[Ca ²⁺] (mmol l ⁻¹)	3		7–47		3–14		2–8		2–5	Bucking and Wood (2007, 2009)
								2–5		Shehadeh and Gordon (1969)
[Mg ²⁺] (mmol l ⁻¹)	2		13–40		17–32		17–50		12–46	Bucking and Wood (2007, 2009)
								0		Shehadeh and Gordon (1969)
[SO ₄ ²⁻] (mmol l ⁻¹)	2							0		Shehadeh and Gordon (1969)
Total [P _i] (mmol l ⁻¹)	3			1–3 [‡]	4–6 [‡]	1–3 [‡]	4–6 [‡]	1–3 [‡]	4–6 [‡]	Avila et al. (2000)
Osmolality (mOsm)	280		380–770		350–500		350–470		320–410	Bucking and Wood (2006b)

Data for four sections of the tract [stomach, anterior intestine (not including pyloric caecae), mid intestine and posterior intestine] are tabulated for trout fasted for >7 days and for trout recently fed with commercial trout pellets. *Calculated using the Henderson–Hasselbalch equation. †Value for chyme pooled from all intestinal sections.

consistently higher than that in saline or plasma (Table 1). The presence of this *in vivo* mucosal-to-serosal osmotic gradient is important; fluid absorption by the gut *in vitro* is usually assessed under unrealistic conditions where mucosal and serosal media are isosmotic (Genz et al., 2011). If we wish to use *in vitro* protocols to accurately assess the transport of substances (particularly those for which solvent drag – see Glossary – is important), it will be necessary to ensure the appropriate osmolality of the mucosal medium *in vitro*.

Mimicking the *in vivo* environment during *in vitro*/ex vivo experiments – a double-edged sword

Mimicking the variety of *in vivo* ionic, pH, [HCO₃⁻] and ammonia conditions (Table 1) in *in vitro* experiments will involve the use of carefully designed buffered saline; obtaining the correct conditions of oxygen tension and the microbiome poses additional issues. In this section, we discuss potential approaches to address these challenges.

Oxygen challenge

The most egregious deviation from natural conditions in almost all *in vitro* studies to date has been the use of high P_{O₂} (150 to >700 Torr) in the luminal fluid (see Table 1, and references therein). In humans and other vertebrates, the chyme is essentially anoxic, with a steep P_{O₂} gradient across the mucosal epithelium, such that the tip of the villi are nearly anoxic (reviewed by Espey, 2013; Ward et al., 2014; Singhal and Shah, 2020). With an anoxic lumen, oxygen delivery through the blood to the intestine is crucial for its optimal function. Oxygenated blood perfuses from the serosal side of the epithelium and is distributed to the mucosal folds, but this distribution is highly dynamic, depending on physiological state; consequently, the oxygen supply to the mucosa fluctuates widely (Taylor and Grosell, 2009; Seth et al., 2011). During fasting, oxygen supply does not meet the tissue demand, rendering the tip of the villi hypoxic, a state commonly referred to as ‘physiological hypoxia’.

Only recently has luminal fluid P_{O_2} been measured *in vivo* in fish (Jung et al., 2020, 2022); it is less than 1 Torr in all gut sections in both fed and fasted animals (Table 1).

The main approach to oxygenating the tissue during *ex vivo* studies using techniques such as gut sacs and Ussing chambers is to gas the saline with 99.7% O_2 (Wood and Grosell, 2012), 95% O_2 (Bakke-McKellep et al., 2000) or 21% O_2 (i.e. air levels; Sundell et al., 2003). In Ussing chamber studies and two-compartment *in vitro* cell culture models (see Glossary), both the mucosal and serosal side of the epithelium are usually gassed with the same gas mixture. However, N_2 (with or without elevated P_{CO_2}) could easily be used to create an anoxic environment on the mucosal side *in vitro* while oxygen is provided from the serosal side. It is very likely that high values of P_{O_2} on the luminal side of the epithelium (which have been used in virtually all *in vitro* investigations to date) result in severe oxidative stress for both the enterocytes and the resident microbial community, with unknown pathological effects on function.

Microbiota challenge

As for other animals, the GI microbiome of fish plays a vital part in their health, nutrition and physiology (Xia et al., 2022). However, researchers have generally ignored the contribution of the microbiome because of the complex nature of the GI microbial ecosystem, which creates technical challenges that impede experimentation. Our resulting lack of knowledge regarding the intricate interactions between the microbiome and the host (and their consequences) prevents us from understanding the multifaceted physiology of the GI tract. In fact, the fish host plays a crucial role in shaping its GI microbiome through various physiological processes, including immune responses, gut motility and the secretion of mucus and antimicrobial peptides (e.g. Wiles et al., 2016; Maritan et al., 2024). These host–microbe interactions are essential for maintaining a balanced microbial community and ensuring proper gut function. However, *in vitro* and *ex vivo* models often lack these dynamic interactions (Wiles et al., 2016; Stagaman et al., 2017; Butt and Volkoff, 2019). In the following subsections, we discuss these interactions and how they may be captured in *in vitro* and *ex vivo* work.

Microbial culturing

The elaborate network of bacteria, archaea, fungi and viruses that make up the GI microbiome are notorious for being difficult to isolate and culture using standard laboratory methods (e.g. Lau et al., 2016). Indeed, many of these microorganisms, especially obligate anaerobes and other fastidious species that require specific growth conditions, often fail to grow using standard culturing techniques. For example, the specific nutrient requirements and growth factors necessary for culturing many microbial species are not fully understood, impeding efforts to replicate the native microbiome accurately, although progress is being made (e.g. Lagier et al., 2018; Tao et al., 2023; Średnicka et al., 2023). Furthermore, the interactions among different microbial species in the gut are context dependent, involving symbiotic relationships such as mutualism, commensalism and competition (Pearce et al., 2018; Li et al., 2019). These relationships are difficult to replicate outside the host environment. As a result, the microbial communities cultured and then applied to experimental models may not accurately represent the communities in the fish gut, leading to biased interpretations and conclusions. Indeed, traditional culturing techniques often favour fast-growing, easy-to-culture species, and application of these specific bacteria to intestinal tissues and models results in an incomplete representation of the microbial diversity present in the fish GI tract (reviewed by

Li et al., 2019; additional examples of monocultures and their impacts on non-fish models can be found in Pearce et al., 2018).

Advances in culturing techniques, such as the development of co-culture systems and the use of specialised growth media, have improved the ability to culture a broader range of gut microbes (Yadav et al., 2023). However, these methods are not yet capable of replicating the full diversity of the fish GI microbiome. Continued innovation in culturing techniques, including high-throughput culturing methods, ‘culturomics’ (see Glossary) and the use of gut-mimicking bioreactors, could help overcome these limitations so that the entirety of the community can be selectively applied to tissues for study.

There are further complicating factors in accurately representing the fish gut microbiome *in vitro*. The fish GI microbiome is significantly influenced by the surrounding aquatic environment, including variables such as water temperature and salinity (e.g. Legrand et al., 2020), and diet is another critical influencing factor (e.g. Butt and Volkoff, 2019). Even when complex mixtures of microorganisms are directly applied to *in vitro* models, the resulting communities are not representative of the initial communities; this is probably due to challenges in replicating biological conditions found in living GI systems. Further, microbial communities in the fish GI tract are not static; they continually adapt and evolve in response to environmental pressures and host interactions. This rapid adaptation and evolution is crucial for maintaining a balanced microbiome that can effectively respond to changes in the environment or host health.

Microbe–mucus–epithelial interactions

The microbiota is highly beneficial for host health and produces metabolites that promote and improve health in general and intestinal health in particular. However, if homeostasis is threatened, the same microbiota can turn against the host and cause disease (Ghosh et al., 2021). The intricate balance between health and disease is regulated by the intestinal barriers, including the extrinsic secreted mucus barrier, the intrinsic epithelial barrier and the immune barrier known as the ‘gut-associated lymphoid system’ (see Glossary; Sundh and Sundell, 2015). Within the gut, mucus is continuously secreted, thereby pushing luminal bacteria away from the epithelium towards the lumen, where they can utilise the mucins in the mucus (and undigested biomolecules) as an energy source (Padra et al., 2017). If the mucus layer is compromised, the microbiota can attach to the epithelium, interact with the enterocytes, initiate translocation and stimulate an inflammatory response that weakens epithelial barrier function (Turner, 2009). Thus, mimicking the *in vivo* environment that includes endogenous microbiota will require an intact extrinsic mucus layer to prevent bacteria from interacting with the epithelium. However, preparation of tissues for *ex vivo* experiments (e.g. Ussing chamber) can cause mechanical damage that disrupts the normal status of the mucus layer, and *in vitro* preparations may lack mucus-secreting goblet cells. Thus, when preparing tissues for *ex vivo* studies, care should be taken to keep the protective mucus layer intact. In the future, the development of teleost intestinal organoids containing functional goblet cells could help to ensure that *in vitro* studies better reflect *in vivo* conditions.

Microbial metabolome

The health benefits of the microbiota are mainly provided by the metabolome it produces in the lumen (reviewed by Hou et al., 2022; Ghosh et al., 2021). In humans, there are thousands of microbial metabolites, of which a few have been connected to beneficial intestinal function; these include short-chain fatty acids, indole derivatives, bile acid metabolites, conjugated fatty acids,

polyamines and polyphenolic derivatives (Ghosh et al., 2021). Thus, one approach to capturing the influence of the microbiota in an *in vitro* system may be to mimic the metabolome rather than the microbiota.

In vitro models and development of microfluidic platforms

One of the principles of the 3Rs (Reduce, Refine and Replace; see Glossary) is to develop *in vitro* systems that aim to minimise the use of animals in physiology and toxicology studies. Complex tissues such as the gut are difficult to replicate, but significant progress has been made in recent years, particularly in mammalian gut research, where gut-on-chip devices have been specifically designed to mimic the physical conditions of the gut. The physical properties of the piscine gut are yet to be fully described, but when combined with the correct saline to mimic the chyme, oxygen gradients and microbiomes (or associated metabolites), an *in vitro* replacement that mimics the *in vivo* situation may be possible. Here, we discuss the current range of culture techniques that may, in the future, allow us to recapitulate the *in vivo* conditions in an *in vitro* system.

Fish intestinal primary cell and immortalised cell cultures

Langan et al. (2018a) have described a protocol for gut epithelia cell isolation and primary cell culture for the different regions (pyloric, anterior, mid and posterior) of the rainbow trout gut. Of course, a primary cell culture system still requires donor fish, and within the 3Rs framework it is termed a ‘partial replacement’ for the use of live animals. Furthermore, care has to be taken in the culture conditions. Intestinal cells perform best at 21°C – compared with 15–18°C for rainbow trout gill primary cultures (Stott et al., 2015; Schnell et al., 2016) – and cells from pyloric caecae require MEM media (used in mammalian intestinal cell cultures), whereas the other regions require L-15 media, indicating that the former region requires different ratios of amino acids for sustenance (Bols et al., 1994b). Cells, when cultured on inserts, form epithelia with transepithelial resistance measurements of 60–90 Ω cm², not too dissimilar from

that measured in other salmonids (80–150 Ω cm²; Sundell and Sundh, 2012). Morphologically, the *in vitro* epithelium resembles intact intestinal cells, and it exhibits chemical biotransformation properties (cytochrome P450 activity) in the presence of organic chemicals and pharmaceuticals (Langan et al., 2018b,c). The versatility of this gut primary cell culture system has great potential that has yet to be fully utilised. Preparations from other species would allow for comparative studies, and growth of the epithelia on inserts enables the manipulation of culture conditions to mimic those found *in vivo*.

In addition, a rainbow trout gut cell line, RTgutGC was established by Kawano et al. (2011), and has been predominantly used in the replacement of fish in ecotoxicology experiments. The RTgutGC cell line, similar to the RTgill-W1 cell line (Bols et al., 1994a), has been suggested as a replacement for fish in acute toxicity studies (Schug et al., 2020) and in evaluating the toxicity of novel compounds (e.g. tyre tread particle toxicity; Dufefoi et al., 2024). These cells have also been used as an intestinal barrier model for nanoparticle uptake (Minghetti and Schirmer, 2016; Opršal et al., 2021). They can be cultured on membrane supports and show polarisation, a reasonable transepithelial resistance and apical localisation of the tight-junction protein ZO-1 (Minghetti et al., 2017). Growth of these cells with rainbow trout fibroblasts creates an intestinal–mesenchymal interface that is more representative of the *in vivo* gut epithelium than if the cells are grown on their own. This has been used for electrical impedance measurements (Drieschner et al., 2019a).

Gut-on-chip devices

The two-compartment epithelial barrier model allows for independent control of luminal and serosal chamber conditions, including factors such as the microbiome and region-specific ionic composition and P_{O_2} gradients (Table 1). However, it does not recapitulate the dynamic environment of the gut, where, for example, fluid flows over the cells and muscle contractions cause peristalsis. Advancements in mammalian gut-on-chip technologies are starting to engineer devices that mimic this

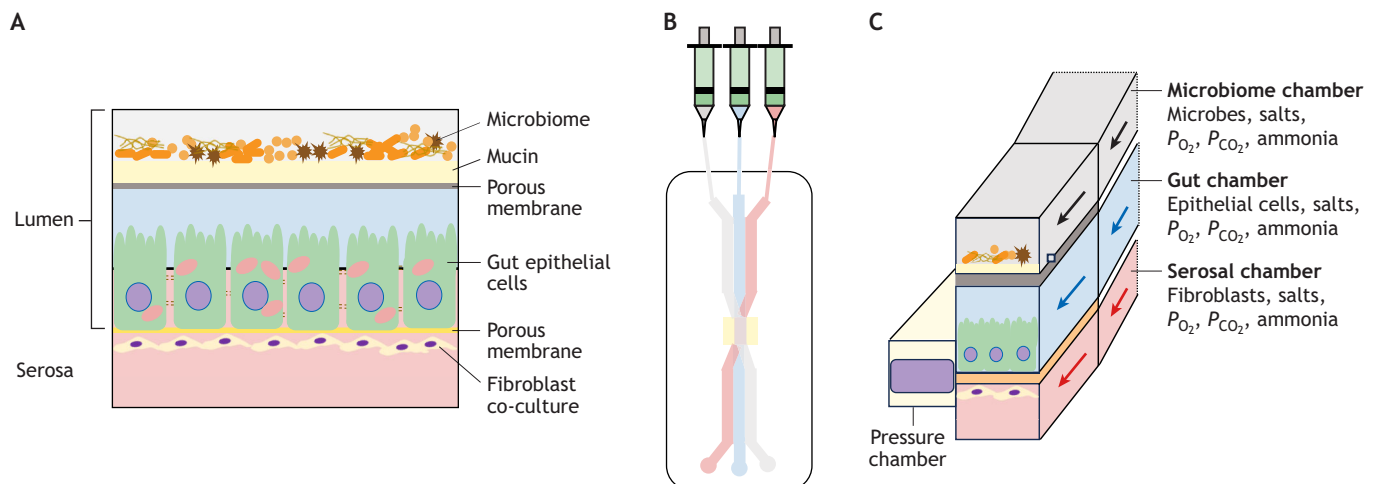


Fig. 2. Conceptual illustrations of a microfluidic teleost gut-on-chip. (A) The top chamber contains the fish GI microbiome grown on a porous membrane covered in mucin, to allow the perfusion of metabolites into the middle chamber, which contains the gut epithelial cells grown on a second membrane. This forms the lumen. In the bottom (serosal) chamber, a layer of companion cells (e.g. fibroblasts) is grown on the other side of the membrane supporting the gut cells. (B) A representation of a microfluidic chip with the three separate chambers: the microbiome (grey channel), gut chamber (blue channel) and serosal chamber (pink channel). Each channel is connected to a syringe pump providing bespoke fluid to each chamber. The gut chamber is connected to a pressure chamber (yellow), which is designed to mimic the pressure changes associated with peristalsis. (C) A cross-section of the three chambers, with the pressure chamber designed to mimic the natural movement of the epithelium and a list of the chamber contents to the right. Based on the design by Marrero et al. (2021).

dynamic environment (Kim and Sung, 2024), and these issues are beginning to be addressed in the fish research community. A RTgutGC gut-on-chip device has been described by Driescher et al. (2019b), in which RTgutGC cells are co-cultured on ultrathin (500 nm) silicon nitride (Si_3N_4) membranes with a gut fibroblast cell line (RTgutF). An open fluidic circuit allows for fluid flow over the cells, and if similar flow conditions are applied to a mammalian intestinal cell line (Caco2 cells), the morphology of the cells alters such that they better represent columnar cells and folds that mimic the intact intestinal epithelium (Kim et al., 2012). Furthermore, stimulation of mechanosensory channels (i.e. Piezo1) in response to movement *in vivo* stimulates the goblet cells to release their contents into the lumen (Xu et al., 2021).

In vitro gut microbiota

One of the major challenges of *in vitro* gut research will be the recapitulation of the gut microbiome. In an attempt to address this in a mammalian cell culture system, Kim et al. (2012) have demonstrated the co-culture of Caco-2 cells with the intestinal bacterium *Lactobacillus rhamnosus*, and this system has been further developed using gut-on-chip devices as a major tool to better understand microbiota interactions with the gut epithelium (e.g. Kim et al., 2015, 2019; Shah et al., 2016; Calatayud et al., 2019). The chip developed by Shah et al. (2016) is a modular microfluidic device that allows for the partitioning of intestinal microbes and Caco-2 cell cultures; specific conditions can be modified within the human cell and microbe chambers, and integrated sensors monitor P_{O_2} in the microchambers and the integrity of the membrane under experimental conditions. Using this system, it has been shown independently that incorporation of dynamic flow into the culture environment induces shear stress that is necessary for morphological changes and polarisation of cell cultures (Fois et al., 2021). Further developments in organoid cell cultures make use of the extracellular matrix as a cell substrate; these systems provide epithelia that better represent the tissues and provide high-throughput platforms (Morelli et al., 2023). Furthermore, Jalili-Firoozinezhad et al. (2019) have developed a method whereby a complex human gut microbiome can be cultured in an anaerobic intestine-on-a-chip, and we are seeing the development of multi-organ-on-chip devices (Ingber, 2022); such devices allow for the analysis of *in vitro* tissue–tissue interactions. This may offer a way forward for *in vitro* studies on fish intestinal function; a potential design for a microfluidic chip consisting of the fish gut epithelium as well as the microbiome is depicted in Fig. 2.

Conclusion

Accurately recapitulating the environments of the different segments of the GI tract from various species of fish will require considerable knowledge of *in vivo* conditions. *In vitro* techniques should enable these conditions to be manipulated; however, almost all *ex vivo/in vitro* fish gut research to date has not carefully considered the unique gut chemistry, oxygen, carbon dioxide and ammonia gradients, physical environment or influence of microbiota and their metabolites. Until this can be achieved, reproducibility and interpretation of the results will require careful reporting of the culture conditions employed. In the future, the applicability of organ-on-chip devices is likely to be restricted to only a few species and is most likely to be used in aquaculture research to help improve productivity. Such investigations may focus on factors that influence the fish GI tract microbiome, nutrient uptake rates and dietary drug delivery. Incorporating realistic GI conditions in *in vitro/ex vivo* research has the potential to greatly

improve our knowledge on basic fish gut physiology; for this to happen, we need greater focus on reporting species-specific luminal chemistries and microbial assemblages and an investment in developing fish-specific microphysiological platforms.

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Competing interests

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References

- Avila, E. M., Tu, H., Basantes, S. and Ferraris, R. P. (2000). Dietary phosphorus regulates intestinal transport and plasma concentrations of phosphate in rainbow trout. *J. Comp. Physiol.* **170B**, 201–209. doi:10.1007/s003600050276
- Bakke, A., Glover, C. and Krogdahl, Å. (2011). Feeding, digestion, and absorption of nutrients. In *Fish Physiology: Multifunctional Gut* (ed. M. Grosell, A. P. Farrell and C. J. Brauner), pp. 57–111. Elsevier Inc.
- Bakke-Mckellep, A. M., Nordrum, S., Krogdahl, Å. and Buddington, R. K. (2000). Absorption of glucose, amino acids, and dipeptides by the intestines of Atlantic salmon (*Salmo salar* L.). *Fish Physiol. Biochem.* **22**, 33–44. doi:10.1023/A:1007872929847
- Bols, N. C., Barlian, A., Chirino-Trejo, M., Caldwell, S. J., Goegan, P. and Lee, L. E. J. (1994a). Development of a cell line from primary cultures of rainbow trout, *Oncorhynchus mykiss* (Walbaum), gills. *J. Fish Dis.* **17**, 601–611. doi:10.1093/1994.tb00258.x
- Bols, N. C., Ganassin, R. C., Tom, D. J. and Lee, L. E. (1994b). Growth of fish cell lines in glutamine-free media. *Cytotechnology* **16**, 159–166. doi:10.1007/BF00749903
- Boutiflier, R. G., Heming, T. A. and lwama, G. K. (1984). Appendix: Physicochemical parameters for use in fish respiratory physiology. In *Fish Physiology*, Vol. 10 (ed. W. S. Hoar and D. J. Randall), pp. 403–430. Academic Press.
- Bucking, C. and Wood, C. M. (2006a). Gastrointestinal processing of Na^+ , Cl^- , and K^+ during digestion: implications for homeostatic balance in freshwater rainbow trout. *Am. J. Physiol.* **291R**, 1764–1772. doi:10.1152/ajpregu.00224.2006
- Bucking, C. and Wood, C. M. (2006b). Water dynamics in the digestive tract of freshwater rainbow trout during the processing of a single meal. *J. Exp. Biol.* **209**, 1883–1893. doi:10.1242/jeb.02205
- Bucking, C. and Wood, C. M. (2007). Gastrointestinal transport of Ca^{2+} and Mg^{2+} during the digestion of a single meal in the freshwater rainbow trout. *J. Comp. Physiol.* **177B**, 349–360. doi:10.1007/s00360-006-0134-3
- Bucking, C. and Wood, C. M. (2008). The alkaline tide and ammonia excretion after voluntary feeding in freshwater rainbow trout. *J. Exp. Biol.* **211**, 2533–2541. doi:10.1242/jeb.015610
- Bucking, C. and Wood, C. M. (2009). The effect of postprandial changes in pH along the gastrointestinal tract on the distribution of ions between the solid and fluid phases of chyme in rainbow trout. *Aquacult. Nutr.* **15**, 282–296. doi:10.1111/j.1365-2095.2008.00593.x
- Bucking, C., C. and Wood, C. M. (2012). Digestion of a single meal affects gene expression of ion and ammonia transporters and glutamine synthetase activity in the gastrointestinal tract of freshwater rainbow trout. *J. Comp. Physiol. B.* **182**, 341–350. doi:10.1007/s00360-011-0622-y
- Bucking, C., Fitzpatrick, J. L., Nadella, S. R. and Wood, C. M. (2011). Assimilation of water and dietary ions by the gastrointestinal tract during digestion in seawater-acclimated rainbow trout. *J. Comp. Physiol.* **181B**, 615–630. doi:10.1007/s00360-011-0550-x
- Butt, R. L. and Volkoff, H. (2019). Gut microbiota and energy homeostasis in fish. *Front. Endocrinol.* **10**, 429202. doi:10.3389/fendo.2019.00009
- Calatayud, M., Dezutter, O., Hernandez-Sanabria, E., Hidalgo-Martinez, S., Meysman, F. J. and Van de Wiele, T. (2019). Development of a host-microbiome model of the small intestine. *FASEB J.* **33**, 3985–3996. doi:10.1096/fj.201801414R
- Cameron, J. N. and Heisler, N. (1983). Studies of ammonia in the rainbow trout: Physico-chemical parameters, acid-base behaviour and respiratory clearance. *J. Exp. Biol.* **105**, 107–125. doi:10.1242/jeb.105.1.107

- Ciavoni, E., Nederlof, M., Rooijackers, J., Schrama, J. W. and Philip, A. J. P. (2023). Effect of dietary macronutrient composition and buffering capacity on chyme characteristics and digestion kinetics in the gastrointestinal tract of freshwater rainbow trout (*Oncorhynchus mykiss*). *Aquaculture* **574**, 739674. doi:10.1016/j.aquaculture.2023.739674
- Cooper, C. A. and Wilson, R. W. (2008). Post-prandial alkaline tide in freshwater rainbow trout: effects of meal anticipation on recovery from acid-base and ion regulatory disturbances. *J. Exp. Biol.* **211**, 2542-2550. doi:10.1242/jeb.015586
- Drieschner, C., Vo, N. T. K., Schug, H., Burkard, M., Bols, N. C., Renaud, P. and Schirmer, K. (2019a). Improving a fish intestinal barrier model by combining two rainbow trout cell lines: epithelial RTgutGC and fibroblastic RTgutF. *Cytotechnology* **71**, 835-848. doi:10.1007/s10616-019-00327-0
- Drieschner, C., Könemann, S., Renaud, P. and Schirmer, K. (2019b). Fish-gut-on-chip: development of a microfluidic bioreactor to study the role of the fish intestine in vitro. *Lab. Chip* **19**, 3268-3276. doi:10.1039/C9LC00415G
- Dudefoi, W., Ferrari, B. J. D., Breider, F., Masset, T., Leger, G., Vermeirssen, E., Bergmann, A. J. and Schirmer, K. (2024). Evaluation of tire tread particle toxicity to fish using rainbow trout cell lines. *Sci. Total Environ.* **912**, 168933. doi:10.1016/j.scitotenv.2023.168933
- Espey, M. G. (2013). Role of oxygen gradients in shaping redox relationships between the human intestine and its microbiota. *Free Radic. Biol. Med.* **55**, 130-140. doi:10.1016/j.freeradbiomed.2012.10.554
- Fois, C. A. M., Schindeler, A., Valtchev, P. and Dehghani, F. (2021). Dynamic flow and shear stress as key parameters for intestinal cells morphology and polarization in an organ-on-a-chip model. *Biomed. Microdevices* **23**, 55. doi:10.1007/s10544-021-00591-y
- Genz, J., McDonald, M. D. and Grosell, M. (2011). Concentration of MgSO₄ in the intestinal lumen of *Opsanus beta* limits osmoregulation in response to acute hypersalinity stress. *Am. J. Physiol.* **300**, R895-R909. doi:10.1152/ajpregu.00299.2010
- Goodrich, H. R., Bayley, M., Birgersson, L., Davison, W. G., Johannsson, O. E., Kim, A. B., Le My, P., Tinh, T. H., Thanh, P. N., Thanh, H. D. T. et al. (2020). Understanding the gastrointestinal physiology and responses to feeding in air-breathing Anabantiform fishes. *J. Fish Biol.* **96**, 986-1003. doi:10.1111/jfb.14288
- Ghosh, S., Samuel Whitley, C., Haribabu, B. and Rao Jala, V. (2021). Regulation of intestinal barrier function by microbial metabolites. *Cell Mol. Gastroenterol. Hepatol* **11**, 1463-1482. doi:10.1016/j.jcmgh.2021.02.007
- Grosell, M. (2006). Intestinal anion exchange in marine fish osmoregulation. *J. Exp. Biol.* **209**, 2813-2827. doi:10.1242/jeb.02345
- Hou, K., Wu, Z. X., Chen, X. Y., Wang, J. Q., Zhang, D., Xiao, C., Zhu, D., Koya, J. B., Wei, L., Li, J. et al. (2022). Microbiota in health and diseases. *Signal Transduct. Target. Ther.* **7**, 135. doi:10.1038/s41392-022-00974-4
- Ingber, D. E. (2022). Human organs-on-chips for disease modelling, drug development and personalized medicine. *Nat. Rev. Genet.* **23**, 467-491. doi:10.1038/s41576-022-00466-9
- Jalili-Firoozinezhad, S., Gazzaniga, F. S., Calamari, E. L., Camacho, D. M., Fadel, C. W., Bein, A., Swenor, B., Nestor, B., Cronce, M. J., Tovaglieri, A. et al. (2019). A complex human gut microbiome cultured in an anaerobic intestine-on-a-chip. *Nat. Biomed. Engineer.* **3**, 520-531. doi:10.1038/s41551-019-0397-0
- Jung, E. H., Eom, J., Brauner, C. J., Martinez-Ferreras, F. and Wood, C. M. (2020). The gaseous gastrointestinal tract of a seawater teleost, the English sole (*Parophrys vetulus*). *Comp. Biochem. Physiol.* **247A**, 110743. doi:10.1016/j.cbpa.2020.110743
- Jung, E. H., Brauner, C. J. and Wood, C. M. (2022). Post-prandial respiratory gas and acid-base profiles in the gastrointestinal tract and its venous drainage in freshwater rainbow trout (*Oncorhynchus mykiss*) and seawater English sole (*Parophrys vetulus*). *Comp. Biochem. Physiol.* **265A**, 111123. doi:10.1016/j.cbpa.2021.111123
- Jung, E. H., Brauner, C. E. and Wood, C. M. (2023a). Do extreme postprandial levels of oxygen, carbon dioxide, and ammonia in the digestive tract equilibrate with the bloodstream in the freshwater rainbow trout (*Oncorhynchus mykiss*)? *J. Comp. Physiol.* **193B**, 193-205. doi:10.1007/s00360-023-01475-8
- Jung, E. H., Nguyen, J., Nelson, C., Brauner, C. J. and Wood, C. M. (2023b). Ammonia transport is independent of PNH₃ gradients across the gastrointestinal epithelia of the rainbow trout: a role for the stomach. *J. Exp. Zool.* **339A**, 180-192. doi:10.1002/jez.2670
- Kandedgedara, A. and Rorabacher, D. B. (1999). Noncomplexing tertiary amines as "better" buffers covering the range of pH 3–11. Temperature dependence of their acid dissociation constants. *Anal. Chem.* **71**, 3140-3144. doi:10.1021/ac9902594
- Kawano, A., Haiduk, C., Schirmer, K., Hanner, R., Lee, L. E. J., Dixon, B. and Bols, N. C. (2011). Development of a rainbow trout intestinal epithelial cell line and its response to lipopolysaccharide. *Aquacul. Nutr.* **17**, e241-e252. doi:10.1111/j.1365-2095.2010.00757.x
- Kim, R. and Sung, J. H. (2024). Recent advances in gut- and gut-organ-axis-on-a-chip models. *Adv. Healthc. Mater.* **20**, e2302777. doi:10.1002/adhm.202302777
- Kim, H. J., Li, H., Collins, J. J. and Ingber, D. E. (2015). Contributions of microbiome and mechanical deformation to intestinal bacterial overgrowth and inflammation in a human gut-on-a-chip. *Proc. Natl. Acad. Sci. USA* **113**, E7-E15. doi:10.1073/pnas.1522193112
- Kim, R., Attayek, P. J., Wang, Y., Furtado, K. L., Tamayo, R., Sims, C. E. and Allbritton, N. L. (2019). An in vitro intestinal platform with a self-sustaining oxygen gradient to study the human gut/microbiome interface. *Biofabrication* **12**, 015006. doi:10.1088/1758-5090/ab446e
- Kim, H. J., Huh, D., Hamilton, G. and Ingber, D. E. (2012). Human gut-on-a-chip inhabited by microbial flora that experiences intestinal peristalsis-like motions and flow. *Lab. Chip* **12**, 2165-2174. doi:10.1039/c2lc40074j
- Kong, F. and Singh, R. P. (2008). Disintegration of solid foods in human stomach. *J. Food Sci.* **73**, R67-R80. doi:10.1111/j.1750-3841.2008.00766.x
- Krogdahl, Å., Sundby, A. and Holm, H. (2015). Characteristics of digestive processes in Atlantic salmon (*Salmo salar*). Enzyme pH optima, chyme pH, and enzyme activities. *Aquaculture* **449**, 27-36. doi:10.1016/j.aquaculture.2015.02.032
- Kwong, R. W. and Niyogi, S. (2008). An *in vitro* examination of intestinal iron absorption in a freshwater teleost, rainbow trout (*Oncorhynchus mykiss*). *J. Comp. Physiol.* **178B**, 963-975. doi:10.1007/s00360-008-0279-3
- Lagier, J. C., Dubourg, G., Million, M., Cadoret, F., Bilen, M., Fenollar, F., Levasseur, A., Rolain, J. M., Fournier, P. E. and Raoult, D. (2018). Culturing the human microbiota and culturomics. *Nat. Rev. Microbiol.* **16**, 540-550. doi:10.1038/s41579-018-0041-0
- Langan, L. M., Owen, S. F. and Jha, A. N. (2018a). Establishment and long-term maintenance of primary intestinal epithelial cells cultured from the rainbow trout, *Oncorhynchus mykiss*. *Biol. Open* **7**, bio032870. doi:10.1242/bio.032870
- Langan, L. M., Arossa, S., Owen, S. F. and Jha, A. N. (2018b). Assessing the impact of benzo[a]pyrene with the *in vitro* fish gut model: An integrated approach for eco-genotoxicological studies. *Mutat. Res. Genet. Toxicol. Environ. Mutagen* **826**, 53-64. doi:10.1016/j.mrgentox.2017.12.009
- Langan, L. M., Owen, S. F., Trznadel, M., Dodd, N. J. F., Jackson, S. K., Purcell, W. M. and Jha, A. N. (2018c). Spheroid size does not impact metabolism of the β -blocker propranolol in 3D intestinal fish model. *Front. Pharmacol.* **9**, 947. doi:10.3389/fphar.2018.00947
- Lau, J. T., Whelan, F. J., Herath, I., Lee, C. H., Collins, S. M., Bercik, P. and Surette, M. G. (2016). Capturing the diversity of the human gut microbiota through culture-enriched molecular profiling. *Genome Med.* **8**, 1-10. doi:10.1186/s13073-015-0257-9
- Legrand, T. P., Wynne, J. W., Weyrich, L. S. and Oxley, A. P. (2020). A microbial sea of possibilities: current knowledge and prospects for an improved understanding of the fish microbiome. *Rev. Aquacult.* **12**, 1101-1134. doi:10.1111/raq.12375
- Leonard, E. M., Nadella, S. R., Bucking, C. and Wood, C. M. (2009). Characterization of dietary Ni uptake in the rainbow trout, *Oncorhynchus mykiss*. *Aquat. Toxicol.* **95**, 205-216. doi:10.1016/j.aquatox.2009.05.002
- Li, X., Ringø, E., Hosenifair, S. H., Lauzon, H. L., Birkbeck, H. and Yang, D. (2019). The adherence and colonization of microorganisms in fish gastrointestinal tract. *Rev. Aquacult.* **11**, 603-618. doi:10.1111/raq.12248
- Maritan, E., Quagliarello, A., Frago, E., Patarnello, T. and Martino, M. E. (2024). The role of animal hosts in shaping gut microbiome variation. *Phil. Trans. R. Soc. B* **379**, 20230071. doi:10.1098/rstb.2023.0071
- Márquez, L., Robles, R., Morales, G. A. and Moyano, F. J. (2012). Gut pH as a limiting factor for digestive proteolysis in cultured juveniles of the gilthead sea bream (*Sparus aurata*). *Fish Physiol. Biochem.* **38**, 859-869. doi:10.1007/s10695-011-9573-1
- Marrero, D., Pujol-Vila, F., Vera, D., Gabriel, G., Illa, X., Elizalde-Torrent, A., Alvarez, M. and Villa, R. (2021). Gut-on-a-chip: Mimicking and monitoring the human intestine. *Biosen. Bioelect.* **181**, 113156. doi:10.1016/j.bios.2021.113156
- McCoy, R., Oldroyd, S., Yang, W., Wang, K., Hoven, D., Bulmer, D., Zilbauer, M. and Owens, R. M. (2024). In Vitro models for investigating intestinal host-pathogen interactions. *Adv. Sci.* **11**, e2306727. doi:10.1002/advs.202306727
- Minghetti, M. and Schirmer, K. (2016). Effect of media composition on bioavailability and toxicity of silver and silver nanoparticles in fish intestinal cells (RTgutGC). *Nanotoxicology* **10**, 1526-1534. doi:10.1080/17435390.2016.1241908
- Minghetti, M., Drieschner, C., Bramaz, N., Schug, H. and Schirmer, K. (2017). A fish intestinal epithelial barrier model established from the rainbow trout (*Oncorhynchus mykiss*) cell line, RTgutGC. *Cell. Biol. Toxicol.* **33**, 539-555. doi:10.1007/s10565-017-9385-x
- Morelli, M., Kurek, D., Ng, C. P. and Queiroz, K. (2023). Gut-on-a-Chip Models: current and future perspectives for host-microbial interactions research. *Biomedicines* **11**, 619. doi:10.3390/biomedicines11020619
- Nadella, S. R., Grosell, M. and Wood, C. M. (2007). Mechanisms of dietary Cu uptake in freshwater rainbow trout: evidence for Na⁺-assisted Cu transport and a specific metal carrier in the intestine. *J. Comp. Physiol.* **177B**, 433-446. doi:10.1007/s00360-006-0142-3
- Nadella, S. R., Hung, C. Y. and Wood, C. M. (2011). Mechanistic characterization of gastric copper transport in rainbow trout. *J. Comp. Physiol.* **181B**, 27-41. doi:10.1007/s00360-010-0510-x
- Opršal, J., Knotek, P., Zickler, G. A., Sigg, L., Schirmer, K., Pouzar, M. and Geppert, M. (2021). Cytotoxicity, accumulation and translocation of silver and silver sulfide nanoparticles in contact with Rainbow Trout intestinal cells. *Aquat. Toxicol.* **237**, 105869. doi:10.1016/j.aquatox.2021.105869
- Padra, J. T., Sundh, H., Sundell, K., Venkatakrishnan, V., Jin, C., Samuelsson, T., Karlsson, N. G. and Lindén, S. K. (2017). *Aeromonas salmonicida* growth in response to Atlantic salmon mucins differs between epithelial sites, is governed by sialylated and N-acetylhexosamine-containing O-glycans and is affected by Ca²⁺. *Infect. Immun.* **85**, e00189-17. doi:10.1128/IAI.00189-17

- Pearce, S. C., Coia, H. G., Karl, J. P., Pantoja-Feliciano, I. G., Zachos, N. C. and Raciocot, K. (2018). Intestinal *in vitro* and *ex vivo* models to study host-microbiome interactions and acute stressors. *Front. Physiol.* **9**, 419368. doi:10.3389/fphys.2018.01584
- Rubino, J. G., Zimmer, A. M. and Wood, C. M. (2014). An *in vitro* analysis of intestinal ammonia handling in fasted and fed freshwater rainbow trout (*Oncorhynchus mykiss*). *J. Comp. Physiol.* **184B**, 91-105. doi:10.1007/s00360-013-0781-0
- Russell, W. M. S. and Burch, R. L. (1959). *The Principles of Humane Experimental Technique*. London, UK: Methuen & Co Ltd.
- Schnell, S., Stott, L. C., Hogstrand, C., Wood, C. M., Kelly, S. P., Pärt, P., Owen, S. F. and Bury, N. R. (2016). Procedures for the reconstruction, primary culture and experimental use of rainbow trout gill epithelia. *Nat. Protoc.* **11**, 490-498. doi:10.1038/nprot.2016.029
- Schug, H., Maner, J., Hülskamp, M., Begnaud, F., Debonneville, C., Berthaud, F., Gimeno, S. and Schirmer, K. (2020). Extending the concept of predicting fish acute toxicity *in vitro* to the intestinal cell line RTgutGC. *ALTEX* **37**, 37-46.
- Sethi, H., Axelsson, M. and Farrell, A. P. (2011). The circulation and metabolism of the gastrointestinal tract. *Fish Physiol.* **30**, 351-393. doi:10.1016/S1546-5098(10)03009-8
- Shah, P., Fritz, J. V., Glaab, E., Desai, M. S., Greenhalgh, K., Frachet, A., Niegowska, M., Estes, M., Jäger, C., Seguin-Devaux, C. et al. (2016). A microfluidics-based *in vitro* model of the gastrointestinal human-microbe interface. *Nat. Commun.* **7**, 11535. doi:10.1038/ncomms11535
- Shehadeh, Z. H. and Gordon, M. S. (1969). The role of the intestine in salinity adaptation of the rainbow trout, *Salmo gairdneri*. *Comp. Biochem. Physiol.* **30**, 397-418. doi:10.1016/0010-406X(69)92011-8
- Singhal, R. and Shah, Y. M. (2020). Oxygen battle in the gut: Hypoxia and hypoxia-inducible factors in metabolic and inflammatory responses in the intestine. *J. Biol. Chem.* **295**, 10493-10505. doi:10.1074/jbc.REV120.011188
- Smith, H. W. (1930). The absorption and excretion of water and salts by marine teleosts. *Am. J. Physiol.* **93**, 480-505. doi:10.1152/ajplegacy.1930.93.2.480
- Średnicka, P., Roszko, M. Ł., Popowski, D., Kowalczyk, M., Wójcicki, M., Emanowicz, P., Szczepańska, M., Kotyrba, D. and Juszczuk-Kubiak, E. (2023). Effect of *in vitro* cultivation on human gut microbiota composition using 16S rDNA amplicon sequencing and metabolomics approach. *Sci. Rep.* **13**, 3026. doi:10.1038/s41598-023-29637-2
- Stagaman, K., Burns, A. R., Guillemin, K. and Bohannon, B. J. (2017). The role of adaptive immunity as an ecological filter on the gut microbiota in zebrafish. *ISME J.* **11**, 1630-1639. doi:10.1038/ismej.2017.28
- Stott, L. C., Schnell, S., Hogstrand, C., Owen, S. F. and Bury, N. R. (2015). A primary fish gill cell culture model to assess pharmaceutical uptake and efflux: evidence for passive and facilitated transport. *Aquat. Toxicol.* **159**, 127-137. doi:10.1016/j.aquatox.2014.12.007
- Sundell, K., Jutfelt, F., Agustsson, T., Olsen, R. E., Sandblom, E., Hansen, T. and Björnsson, B. T. (2003). Intestinal transport mechanisms and plasma cortisol levels during normal and out-of-season parr-smolt transformation of Atlantic salmon, *Salmo salar*. *Aquaculture* **222**, 265-285. doi:10.1016/S0044-8486(03)00127-3
- Sundell, K. S. and Sundh, H. (2012). Intestinal fluid absorption in anadromous salmonids: importance of tight junctions and aquaporins. *Front. Physiol.* **3**, 388. doi:10.3389/fphys.2012.00388
- Sundh, H. and Sundell, K. S. (2015). Environmental impacts on fish mucosa. In *Mucosal Health in Aquaculture* (ed. E. Peatman), pp. 171-197. Elsevier.
- Tao, X., Huang, W., Pan, L., Sheng, L., Qin, Y., Chen, L., Yu, L., Wu, G., Wan, J. and Li, H. (2023). Optimizing *ex vivo* culture conditions to study human gut microbiome. *ISME Commun* **3**, 1-10. doi:10.1038/s43705-022-00209-1
- Taylor, J. R. and Grosell, M. (2009). The intestinal response to feeding in seawater gulf toadfish, *Opsanus beta*, includes elevated base secretion and increased epithelial oxygen consumption. *J. Exp. Biol.* **212**, 3873-3881. doi:10.1242/jeb.034579
- Turner, J. R. (2009). Intestinal mucosal barrier function in health and disease. *Nat. Rev. Immunol.* **9**, 799-809. doi:10.1038/nri2653
- Walsh, P. J., Blackwelder, P., Gill, K. A., Danulat, E. and Mommsen, T. P. (1991). Carbonate deposits in marine fish intestines: a new source of biomineralization. *Limnol. Oceanogr.* **36**, 1227-1232. doi:10.4319/lo.1991.36.6.1227
- Wang, H., Xu, C., Tan, M. and Su, W. (2023). Advanced gut-on-chips for assessing carotenoid absorption, metabolism, and transport. *Crit. Rev. Food Sci. Nutr.* **14**, 1-19. doi:1080/10408398.2023.2293250
- Ward, J. B. J., Keely, S. J. and Keely, S. J. (2014). Oxygen in the regulation of intestinal epithelial transport. *J. Physiol.* **592**, 2473-2489. doi:10.1113/jphysiol.2013.270249
- Weinrauch, A. M., Hoogenboom, J. L. and Anderson, W. G. (2021). A review of reductionist methods in fish gastrointestinal tract physiology. *Comp. Biochem. Physiol.* **254B**, 110571. doi:10.1016/j.cbpb.2021.110571
- Wiles, T. J., Jemielita, M., Baker, R. P., Schlomann, B. H., Logan, S. L., Ganz, J., Melancon, E., Eisen, J. S., Guillemin, K. and Parthasarathy, R. (2016). Host gut motility promotes competitive exclusion within a model intestinal microbiota. *PLoS Biol.* **14**, e1002517. doi:10.1371/journal.pbio.1002517
- Wilson, R. W., Gilmour, K. M., Henry, R. P. and Wood, C. M. (1996). Intestinal base excretion in the seawater-adapted rainbow trout: a role in acid-base balance? *J. Exp. Biol.* **199**, 2331-2343. doi:10.1242/jeb.199.10.2331
- Wolf, K. (1963). Physiological salines for freshwater teleosts. *Progress. Fish Cult.* **25**, 135-140. doi:10.1577/1548-8659(1963)25[135:PSFFT]2.0.CO;2
- Wood, C. M. (2019). Internal spatial and temporal CO₂ dynamics: Fasting, feeding, drinking, and the alkaline tide. In *Fish Physiology: Carbon Dioxide*, Vol. 37 (ed. M. Grosell, P. L. Munday, A. P. Farrell and C. J. Brauner), pp. 245-286. Elsevier.
- Wood, C. M. and Bucking, C. (2011). The role of feeding in salt and water balance. In *Fish Physiology: The Multifunctional Gut of Fish*, Vol. 30 (ed. M. Grosell, A. P. Farrell and C. J. Brauner), pp. 165. San Diego: Academic Press.
- Wood, C. M. and Eom, J. (2019). The internal CO₂ threat to fish: high PCO₂ in the digestive tract. *Proc. R. Soc. B Biol. Sci.* **286**, 20190832. doi:10.1098/rspb.2019.0832
- Wood, C. M. and Grosell, M. (2012). Independence of net water flux from paracellular permeability in the intestine of *Fundulus heteroclitus*, a euryhaline teleost. *J. Exp. Biol.* **215**, 508-517. doi:10.1242/jeb.060004
- Xia, H., Chen, H., Cheng, X., Yin, M., Yao, X., Ma, J., Huang, M., Chen, G. and Liu, H. (2022). Zebrafish: an efficient vertebrate model for understanding role of gut microbiota. *Mol. Med.* **28**, 161. doi:10.1186/s10020-022-00579-1
- Xu, Y., Bai, T., Xiong, Y., Liu, C., Liu, Y., Hou, X. and Song, J. (2021). Mechanical stimulation activates Piezo1 to promote mucin2 expression in goblet cells. *J. Gastroent. Hepatol* **36**, 3127-3139. doi:10.1111/jgh.15596
- Yadav, A., Ahlawat, S. and Sharma, K. K. (2023). Culturing the unculturables: strategies, challenges, and opportunities for gut microbiome study. *J. Appl. Microbiol.* **134**, lxad280. doi:10.1093/jambio/lxad280